

AMPK activation, a preventive therapeutic target in the transition from cardiac injury to heart failure

Christophe Beauloye¹, Luc Bertrand¹, Sandrine Horman¹, and Louis Hue^{1,2*}

¹Institut de Recherche Expérimentale et Clinique, Pôle de Recherche Cardio-Vasculaire, Université catholique de Louvain, Brussels, Belgium; and ²HORM Unit, de Duve Institute, Université catholique de Louvain, Brussels, Belgium

Received 9 November 2010; revised 6 January 2011; accepted 27 January 2011; online publish-ahead-of-print 1 February 2011

Abstract

Heart failure is a progressive muscular disorder leading to a deterioration of the heart characterized by a contractile dysfunction and a chronic energy deficit. As a consequence, the failing heart is unable to meet the normal metabolic and energy needs of the body. The transition between compensated left ventricular hypertrophy and the de-compensated heart is multifactorial, although metabolic disturbances are considered to play a significant role. In this respect, the AMP-activated protein kinase (AMPK) could be a potential target in heart failure development. AMPK senses the energy state of the cell and orchestrates a global metabolic response to energy deprivation. We briefly review here the current knowledge about the chronic energy deficit of the failing heart, as well as the role of AMPK in energy homeostasis and in the control of non-metabolic targets in relation to cardiac hypertrophy and heart failure. The relative importance of energetic and non-metabolic effects in the potential cardioprotective action of AMPK is discussed.

Keywords

AMPK • Heart failure • Hypertrophy

This article is part of the Spotlight Issue on: Metabolic Remodelling in Heart Failure

1. Introduction

Heart failure (HF) is a multifactorial, progressive, and disabling syndrome, characterized by symptoms resulting from ventricular dysfunction, either diastolic (impaired relaxation) or systolic (impaired contraction). The most common cause of HF is coronary artery disease and myocardial infarction, often linked with co-morbidities (e.g. hypertension, diabetes).

HF is in fact a muscular disorder leading to a progressive deterioration of the heart, characterized by contractile dysfunction linked to chronic energy deficit.^{1–5} It is also a systemic disease in which a neurohormonal response of the body activates the renin–angiotensin and adrenergic systems, and leads to left ventricular remodelling and dilatation. Consequently, the increased wall stress enhances local oxygen consumption and worsens the energy deficiency and contractile dysfunction. The heart enters a vicious circle, which precipitates the clinical evolution and aggravates the prognosis. In addition, repeated ischaemic attacks decrease the number of functional cardiomyocytes and lead to maladapted cardiac remodelling.

Restoration of muscle contraction by inotropic support acting on beta-adrenergic signalling fails to improve the short- and long-term prognosis.⁶ In contrast, the current HF therapies that inhibit the renin–angiotensin system and the catecholamine response,

dramatically improve the prognosis of HF patients.⁷ Although they do provide benefits, HF remains a life-threatening condition. Alternative therapies that could improve the energetic state and disrupt the vicious circle of the failing heart are of particular interest.

In this context, the AMP-activated protein kinase (AMPK) appears as a potential therapeutic target. AMPK is a highly conserved eukaryotic protein serine/threonine kinase that senses the energy status of the cell and coordinates a global metabolic response to energy deprivation.^{8,9} The question then arises of its potential beneficial metabolic and/or therapeutic effects in the energy-deprived failing heart. In this chapter, we summarize the current knowledge on the energetic state of the failing heart, the biochemical characteristics of AMPK, and its role in energy homeostasis. We also briefly describe non-metabolic, anti-proliferative, anti-fibrotic, and angiogenic effects of AMPK that are independent of energy but related to cardiac hypertrophy and HF. The relative importance of these effects in the potential cardioprotective action of AMPK is discussed.

2. Energetic state of the failing heart

There is a continuum between compensated left ventricular hypertrophy and de-compensated HF. The pathological transition is probably

* Corresponding author. Tel: +322 764 7576, Fax: +322 764 7507, Email: louis.hue@uclouvain.be

multifactorial, although metabolic disturbances are considered to play a significant role.^{1,2}

Energy depletion characterizes the failing heart, although the extent of the deficit may vary depending on the stage of HF.^{1,2,4,10} In several models of left ventricular hypertrophy as well as in patients suffering from HF, measurements of the changes in cardiac energy charge by NMR techniques revealed significant decreases in phosphorylation potential (decreased phosphocreatine and ATP and increased ADP concentrations) that were proportional to the degree of hypertrophy and could be used as predictors of mortality.^{2,11–14} The low energetic potential of the failing heart affects contraction and relaxation, both of which depend on ATP. Clearly, energy deprivation results from the decreased ability of the failing heart to produce ATP from the available substrates—‘the failing heart: an engine out of fuel’.³ Actually the defect affects all steps of energy production and includes disturbances in substrate utilization, mitochondrial oxidative capacity, and ATP transfer.^{1,2} Moreover, this does not exclude abnormal ATP utilization by non-contractile biochemical systems.

With regard to substrate utilization, fatty acids are preferred substrates. Their oxidation inhibits glucose uptake, whereas glucose together with insulin inhibits fatty acid oxidation. This reciprocal metabolic control, known as the Randle cycle,^{2,15,16} is perturbed in HF. In animal models of HF, the failing heart favours glucose utilization at the expense of fatty acids. The changes are however progressive and depend on the stage of HF.⁴ At early stages, fatty acid oxidation is either unchanged or even slightly increased, whereas at more advanced stages, fatty acid oxidation is clearly limited. The metabolic shift corresponds to fundamental changes in the expression of genes controlling fatty acid oxidation and mitochondrial biogenesis, thus leading to metabolic inflexibility and lack of substrate adaptability to the energy needs.^{2,4,17,18} It is also interesting to note that overexpression of GLUT1 protects against contractile dysfunction and prevents pressure overload-induced HF.¹⁹ It also rescues the contractile dysfunction in peroxisomal proliferator-activated receptor (PPAR) alpha null hearts submitted to high workload.²⁰ However, increased glucose uptake and oxidation in transgenic mice overexpressing GLUT1 decreases fatty acid oxidation and remodels metabolism towards glucose utilization, but at the same time increases oxidative stress and results in cardiac dysfunction when these mice are fed a high-fat diet.²¹

Mitochondrial dysfunction has emerged as a characteristic feature of the failing heart and contributes to energy deprivation. The failing heart contains more mitochondria, which are however reduced in size and display ultrastructural abnormalities.^{18,22–24} Their electron transfer chain complexes and oxidative phosphorylation capacity are decreased.^{1,2,18} Fatty acid oxidation is especially affected in severe HF with a decreased content of enzymes, such as carnitine palmitoyl transferase 1 (CPT1) and acyl-CoA dehydrogenases that control fatty acid oxidation. This deficient fatty acid oxidation is explained by a decreased expression of peroxisome proliferator-activated receptor gamma co-activator (PGC1 alpha),²⁵ which is probably the most important transcriptional factor involved in heart mitochondrial biogenesis.

On top of a deficient oxidative mitochondrial capacity, the failing heart is also unable to couple energy production to utilization through the compartmentalized creatine kinase system. In the failing heart, decreased content and isoform alteration of this energy transfer system concur to an inefficient adaptation of energy production to utilization.¹

3. AMPK, a metabolic master switch and more

AMPK senses the energy status of the cell—the fuel gauge of the cell²⁶—and orchestrates an integrated metabolic response to energy deprivation in order to conserve ATP via short- and long-term metabolic control. AMPK is therefore regarded as a metabolic master switch in normal and pathological conditions.⁸

3.1 AMPK structure

AMPK is a heterotrimeric complex containing a catalytic (alpha) and two regulatory (beta and gamma) subunits. Each subunit has multiple isoforms (alpha 1 and 2, beta 1 and 2, and gamma 1, 2, and 3) giving 12 possible combinations of holoenzyme, which are present in murine and human hearts. The catalytic alpha subunit contains the protein kinase domain and a threonine residue (Thr172) whose phosphorylation by upstream kinases is responsible for AMPK activation.⁸ In mouse hearts, AMPK alpha-2 accounts for 60–80% of total AMPK activity, whereas in human hearts both alpha-1 and alpha-2 catalytic subunits equally contribute to the total AMPK activity.²⁷ The beta subunit acts as a scaffold for the other two subunits. It also contains a glycogen-binding domain, whose physiological role might be to control glycogen metabolism.²⁸ The beta-2 isoform is the main isoform expressed in the heart.²⁹ The gamma subunit contains three AMP-binding domains, one of which binds a non-exchangeable nucleotide, whereas the others can bind AMP or ATP, with however a lower affinity for the latter.³⁰ Mutations in the gamma-2 subunit cause glycogen accumulation and lead to cardiac arrhythmias, also called Wolff–Parkinson–White syndrome.³¹

3.2 Control of AMPK activity

3.2.1 Biochemical mechanisms of activation

AMPK is activated when AMP concentration increases as a result of insufficient ATP production or unmatched energy demand. AMPK can also be activated independently of adenine nucleotides, by changes in calcium concentrations as well as by increased production of reactive oxygen species (ROS). Whatever the stimulus, AMPK activation requires phosphorylation by upstream kinases of a threonine residue (Thr172) located in the activation loop of the alpha catalytic subunit. At least two pathways lead to AMPK activation by phosphorylation of Thr172.^{8,32,33} The first one senses energy depletion and is mediated by AMP and LKB1 (Peutz–Jeghers protein), which seems to be specific for AMPK alpha-2, because in heart from LKB1 KO mice, AMPK alpha-2, but not AMPK alpha-1 activation is abrogated.^{34,35} The AMPK alpha-1 kinase acting under these conditions is not known. The second activation pathway is triggered by increased calcium concentration and is mediated by calcium/calmodulin-dependent protein kinase kinase-beta, which phosphorylates Thr172.^{36–38} Although this protein kinase is present in heart, the demonstration of its participation in AMPK activation has not been reported.

3.2.2 AMPK activation following ATP depletion

Conditions leading to changes in AMP concentrations and AMPK activation are directly related to changes in ATP concentrations, which adenylate kinase translates into relatively larger changes in AMP. Accordingly, ischaemia and mitochondrial inhibitors activate AMPK within a few minutes^{39,40} (Figure 1). Increased ATP demand also leads to AMPK activation, especially when combined with decreased

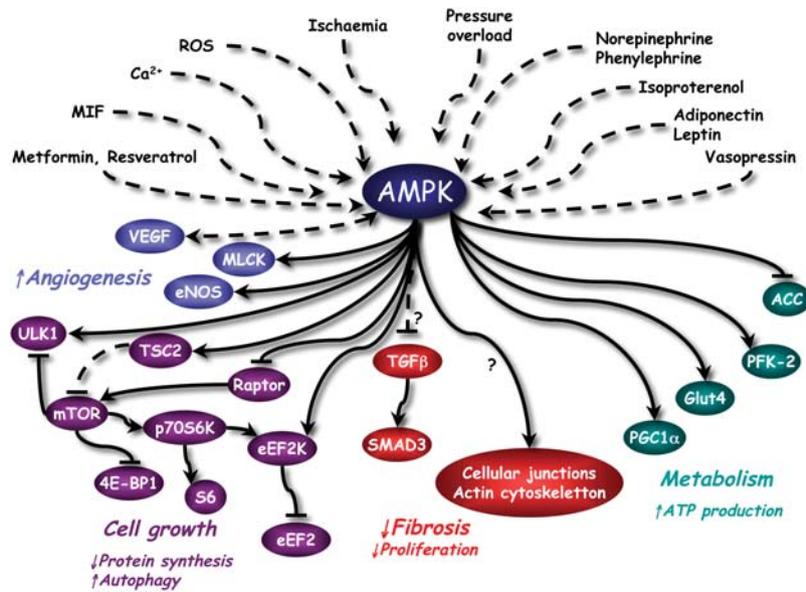


Figure 1 Upstream stimulatory factors and downstream targets of AMPK in the heart. Dashed lines correspond to indirect mechanisms. Question mark signifies that this pathway has not been studied in the heart. Abbreviations: 4E-BP1, 4E binding protein-1;¹⁴⁰ ACC, acetyl-CoA carboxylase;¹⁴¹ eEF2K, eukaryotic elongation factor 2 kinase;^{87,142,143} eNOS, endothelial nitric oxide synthase;^{92–94} Glut4, glucose transporter 4;¹⁴⁴ MIF, migration inhibitory factor;⁵⁰ MLCK, myosin light chain kinase;⁴⁴ mTOR, mammalian target of rapamycin;¹⁴⁵ p70S6K, p70 ribosomal S6 protein kinase;¹⁴⁶ PGC1 α , peroxisome proliferator-activated receptor gamma co-activator alpha;⁷⁷ PFK-2, 6-phosphofructo-2-kinase;³⁹ ROS, reactive oxygen species;^{70–72} S6, ribosomal S6 protein;¹⁴⁷ TGF β , transforming growth factor beta;¹²² TSC2, tuberous sclerosis factor 2;⁸⁵ ULK1, uncoordinated51-like kinase 1;^{88–90} VEGF, Vascular endothelial growth factor.⁹⁵

ATP supply, as is the case in contracting skeletal muscle during intense exercise. Remarkably, this is not the case in perfused hearts subjected to high workload, presumably because this organ can adapt its ATP supply to increased energy demand.⁴¹

3.2.3 Control of AMPK by hormones and agonists

Norepinephrine, phenylephrine, isoproterenol, or vasopressin activate heart AMPK^{42–45} (Figure 1 and Table 1). Interestingly, AMPK activation in response to certain cytokines protects the heart against pressure overload or ischaemic injury. The cardioprotective effects of adiponectin^{46–48} and leptin,⁴⁹ and of the macrophage migration inhibitory factor (MIF), a proinflammatory component released by the ischaemic heart,⁵⁰ are mediated, at least in part,⁵¹ by AMPK activation. The precise mechanism of AMPK activation by these agents is however not clear. In addition, AMPK activation also counteracts the angiotensin II-induced hypertrophy.⁵² On the other hand, certain hormones inhibit AMPK activation. Insulin antagonizes AMPK activation independently of adenosine, via a hierarchical mechanism whereby phosphorylation by protein kinase B (PKB) of a serine residue (Ser485) in the AMPK α subunit prevents subsequent phosphorylation of Thr172 by LKB1.⁵⁵ Angiotensin II has also been reported to inactivate AMPK by a still unknown mechanism.⁵² But interestingly, if AMPK is activated by a pharmacological agent, it then inhibits cardiac hypertrophy⁵² and vascular smooth muscle proliferation induced by angiotensin II.⁵⁶

3.2.4 AMPK activation by pharmacological agents

Among the substances and drugs known to activate AMPK (Figure 1), AICA (5-amino-4-imidazole-carboxamide) riboside has

been widely used. It is an analogue of adenosine that, in certain cells, is phosphorylated to the corresponding nucleotide, ZMP, which mimics several effects of AMP, including AMPK activation.^{57,58} Its use to activate AMPK in cardiomyocytes is not recommended, because of its poor metabolism in these cells,^{39,59} and of several unwanted side effects.^{60,61} Other more specific tools, such as the Abbott compound A762669, should be preferred.^{62,63} Metformin, the most prescribed anti-Type 2 diabetic drug, and its more potent but toxic analogue, phenformin, as well as thiazolidinediones, another class of anti-diabetic drugs, are known to activate AMPK.^{64,65} Their initial metabolic effect is to inhibit mitochondrial respiration, thus decreasing ATP production.^{66,67} It should however be noted that several effects of metformin are thought to be mediated by p38-MAPK or by an inhibition of mTORC1, independently of AMPK.^{68,69} Finally, AMPK could be redox-sensitive: hydrogen peroxide and increased production of ROS activate AMPK,^{70,71} possibly by oxidation of two cysteine residues in the α subunit of AMPK.⁷²

A mechanism connecting caloric restriction and AMPK activation has been described recently. SIRT1, a member of the sirtuin family of NAD-dependent protein deacetylases, is activated by nutrient deprivation and by resveratrol, a cardioprotective polyphenol of red wine. It was initially thought that the anti-ageing effects of resveratrol were mediated by SIRT1.⁷³ However, resveratrol also activates AMPK, probably by inhibiting mitochondrial respiration,⁷⁴ and recent evidence indicates that the protective effect of resveratrol on mitochondrial function is mediated by AMPK, whereas SIRT1 would act downstream of AMPK by de-acetylating PGC1 α .⁷⁵

Table 1 Mechanisms of modulation of AMPK activity by hormones and pharmacological agents

Cardiac stimuli	AMPK activity	Mechanisms for modulation of AMPK activity	References
Norepinephrine, Phenylephrine	↑	?	42
Isoproterenol	↑	Phosphorylation of LKB1	43
Vasopressin	↑	?	44,45
Adiponectin	↑	Binding of APPL1 with AMPK α 2	46–48
Leptin	↑	?	49
MIF	↑	?	50
H ₂ O ₂ ROS	↑	Inhibition of respiratory chain, AMP↑ ATP↓	70,71
Insulin	↓	Phosphorylation of Ser 485/491	55
Angiotensin	↓	?	52
AICAr	↑	ZMP accumulation	57,58
A762669	↑	Allosteric stimulation	62
Metformin/Phenformin	↑	Inhibition of respiratory chain, AMP↑ ATP↓	64,65
Resveratrol	↑	Inhibition of ATP synthase, AMP↑ ATP↓	74

A762669, Abbott compound A-762669; AICAr, 5-aminoimidazole-4-carboxamide ribonucleoside; APPL1, Adaptor protein, phosphotyrosine interaction, PH domain and leucine zipper containing 1; LKB1, also called STK11, serine/threonine kinase 11; MIF, macrophage inhibitory factor; ROS, reactive oxygen species; ZMP, 5-aminoimidazole-4-carboxamide ribonucleotide.

3.3 AMPK targets

3.3.1 Metabolic targets

When activated, AMPK aims at restoring the cellular energy charge by switching off anabolic ATP-consuming pathways, while switching on catabolic ATP-producing pathways. It does so by phosphorylating key metabolic enzymes and transcription factors^{8,9} (Figure 1). Transcription activation could be mediated through histone H2B phosphorylation.⁷⁶ Biosynthetic processes, such as gluconeogenesis, glycogen synthesis, lipogenesis, cholesterol synthesis, and protein synthesis are inhibited, whereas glucose utilization, fatty acid oxidation, and mitochondrial biogenesis are stimulated.^{8,9,40} AMPK does not affect the mitochondrial oxidative capacity in the short term. It does however stimulate mitochondrial biogenesis via activation of PGC1 α ,⁷⁷ which is particularly relevant to the energy-deficient failing heart.

3.3.2 Anti-stress effects of AMPK

Recent evidence suggests that AMPK could inhibit (i) endoplasmic stress, although the mechanism remains to be elucidated;⁷⁸ (ii) cJUN kinase activation;⁷⁹ and (iii) oxidative stress in several cellular models, including cardiomyocytes. The latter could result from the phosphorylation and activation of the forkhead transcription factor 3, which reduces ROS levels by inducing anti-oxidant systems including thioredoxin.^{80,81} Finally, AMPK has been reported to inhibit glucose-induced oxidative stress and NADPH oxidase activation in endothelial cells.⁸²

3.3.3 Control of protein synthesis, cell growth, and autophagy

AMPK inhibits the mammalian target of rapamycin (mTOR) pathway, which controls protein synthesis and cell growth^{9,40,83} (Figure 1). This effect is relevant to cardiac hypertrophy. It is mediated by the phosphorylation of upstream controlling elements, such as tuberous sclerosis complex 2 (TSC2) and/or Raptor.^{84,85} Downstream of mTOR, p70 ribosomal S6 protein kinase (p70S6K), and 4E-binding protein-1 (4EBP1) are involved in protein translation and cell growth.^{83,86} In addition, AMPK directly phosphorylates eukaryotic elongation factor

2 kinase (eEF2K), thereby inhibiting protein elongation through eEF2 phosphorylation.⁸⁷ Moreover, AMPK has recently been shown to promote autophagy. It directly phosphorylates and activates ULK1, an initiator of autophagy that is inactivated by mTOR.^{88–91} Taken together these data indicate that AMPK inhibits protein synthesis and cell growth and stimulates autophagy.

3.3.4 AMPK and the vascular system

In endothelial cells, AMPK is activated by VEGF and controls eNOS activation.^{92,93} eNOS is also known to be a direct target of AMPK in cardiomyocytes,⁹⁴ although the role of this phosphorylation is not known. In vascular smooth muscle, direct phosphorylation of myosin light chain kinase by AMPK allows this protein kinase to participate in the control of vascular tone.⁴⁴ Interestingly, in skeletal muscle and cardiomyocytes, AMPK activation induces VEGF expression and secretion. The latter plays an important role in muscular adaptation to exercise and coordinates angiogenesis to hypertrophy in response to pressure overload.⁹⁵

4. From hypertrophy to failure

4.1 Cardiac hypertrophy

AMPK is activated in models of chronic pressure overload.⁹⁶ This activation is responsible for an increase in glucose metabolism and probably acts as a negative feed-back on hypertrophy by inhibiting the mTOR pathway. Indeed, inhibition of mTOR by its specific inhibitor rapamycin or partial ablation of mTOR blocked p70S6K activation and counteracted the development of cardiac hypertrophy (Tables 2 and 3).^{97,98} Moreover, pharmacological activation of AMPK inhibits the mTOR pathway and attenuates the development of hypertrophy.^{99–101} In addition, in AMPK α -2 null mice, cardiac hypertrophy induced by isoproterenol or aortic constriction is significantly larger than in controls and is correlated with p70S6K activation.^{102,103} Furthermore, the cardio-specific deletion of LKB1 led to hypertrophy, correlated with a stimulation of mTOR signalling and reduced

Table 2 Changes in cardiac function in AMPK transgenic/KO mice and effects of pharmacological agents

Animal model	Surgery	AMPK activity	Pharmacological treatment	Downstream effects	References
Wild-type (mouse)	TAC	↑	Compound C (inhibits AMPK)	↑ Systolic dysfunction ↑ Hypertrophy, ↓ capillary formation	95
Wild-type (mouse)	CAL	↑	Metformin (activates AMPK)	↓ Systolic dysfunction ↓ Hypertrophy ↑ eNOS phosphorylation ↑ PGC-1 α expression, ↑ Mitochondrial respiration	135
AMPK α 2-DN(D157A)	CAL	↓	Metformin (no AMPK activation)	No cardioprotective effect	135
AMPK α 2-KO	TAC	↓	—	↑ Systolic dysfunction ↑ Hypertrophy, fibrosis	103
AMPK α 2-KO	—	↓	Isoproterenol	↑ Hypertrophy ↑ p70S6K activity	102
LKB1-KO (cardiac-specific)	—	↓	—	↑ Systolic dysfunction ↑ Hypertrophy, fibrosis	105
ObR-KO	CAL	↓	AICAr (activates AMPK)	↓ Systolic dysfunction, ↓ hypertrophy ↓ Inflammation, fibrosis, apoptosis	49
APN/KO	TAC	↓	—	↑ Systolic dysfunction ↑ Hypertrophy, ↓ capillary formation	95
Wild-type (dog)	Rapid ventricular pacing	↑	Metformin AICAr (activates AMPK)	↓ Systolic dysfunction ↑ eNOS phosphorylation ↑ Plasma NO levels ↓ Apoptosis	136

AICAr, 5-aminoimidazole-4-carboxamide ribonucleoside; APN, adiponectin; CAL, coronary artery ligation; DN, dominant negative; eNOS, endothelial nitric oxide synthase; KO, knock out; ObR, leptin receptor; p70S6K, p70-ribosomal S6 kinase; PGC-1 α , peroxisome proliferator-activated receptor gamma coactivator 1-alpha; TAC, trans-aortic constriction.

Table 3 AMPK activity in cardiac pathologies

Cardiac pathologies	AMPK activity	Putative role of AMPK	References
Hypertrophy(Left ventricular pressure overload)	↑	Chronically activated, protective	96,101–103,105,132
Ischaemia (<i>ex vivo</i> , <i>in vivo</i>)	↑	Acutely activated, protective	34,35,39,48,53,55,79,127–129
Hypertrophic cardiomyopathy (HCM) associated with Wolf–Parkinson–White syndrome	↑ or ↓	Chronically (in)activated, deleterious	See reference ¹²⁵ for a review

AMPK phosphorylation and could be prevented by overexpressing a constitutively active form of AMPK or by inhibiting mTOR with rapamycin.^{104,105} Interestingly, the hypertrophic response to pressure overload is amplified in adiponectin-deficient mice, which exhibit diminished AMPK activity.¹⁰⁶

4.2 Transition from cardiac hypertrophy to HF

AMPK and its upstream kinase LKB1 not only antagonizes the hypertrophic response, it also delays the transition to HF, as demonstrated by studies resorting to genetic manipulations of these protein kinases. Under normal conditions, the crucial role played by LKB1 in maintaining cardiac function stems from the phenotype following LKB1 deletion. In these LKB1 deficient hearts, the lack of AMPK alpha-2 subunit activation increases mTOR signalling, decreases energy efficiency and VEGF expression, and impairs cardiac function.^{104,105,107}

Under pathological conditions, as in a model of chronic pressure overload, the lack of AMPK alpha-2 exacerbates hypertrophy and favours the transition to HF.¹⁰³ In addition, disruption of the coordination between angiogenesis and hypertrophy is another crucial factor in the pathological transition to HF.¹⁰⁸ And in adiponectin-deficient animals, the lack of AMPK activation by this hormone exacerbates the transition to HF in pressure overloaded hypertrophied hearts due to an angiogenesis deficiency (*Tables 2 and 3*).¹⁰⁹

As regards energy depletion, AMPK activation, which is expected in energy deficient hearts, is not sufficient to maintain ATP. Several reports indicate that expression of PPAR alpha and PGC1 alpha is decreased and may explain the low-energetic state of the failing heart.²⁵ However, although PGC1 alpha down-regulation contributes to energy deficiency, it does not suffice by itself to induce HF. In two genetic models of PGC1 alpha deficiency, the overall metabolic disturbances did not lead to HF, except when the hearts were submitted to chronic haemodynamic overloads.^{110–112} Thus energy deprivation

alone is not sufficient to cause HF but may contribute to the mal-adaptative response of the heart. It follows that the pathological transition from compensated cardiac hypertrophy to HF implies more than energy depletion.

4.3 Remodelling and fibrosis

Cardiac remodelling occurs following any form of cardiac injury¹¹³ and remodelling of the extracellular matrix (ECM) contributes to contractile dysfunction.¹¹⁴ It develops in response to increased ventricular walls tension and to different hormones (including angiotensin, catecholamines, and endothelins) and inflammatory cytokines (IL1-b, TGF-b, TNF-a, IL-6...^{115,116}). Myocardial fibrosis is a pathological entity of ECM remodelling, which contributes to HF by increasing myocardial stiffness and reducing pumping capacity.¹¹⁷ The synthesis and turnover regulation of ECM components constitute the primary role of cardiac fibroblasts (CFs), which represent 26–63% of cells within the myocardium of mouse and rat, respectively.^{118,119}

Angiotensin II is a critical mediator of cardiomyocyte hypertrophy and cardiac fibrosis.¹²⁰ AMPK could interfere with this phenomenon by inhibiting the angiotensin II-induced stimulation of proliferation via a cross-talk with extracellular signal-regulated kinase (ERK), as shown in CFs.¹²¹ Interaction with myodifferentiation has also been studied in mesangial cells in which AMPK inhibits TGF-b-induced smad3-dependent transcription.¹²² Finally, AMPK could alter cell–cell or ECM–cell communication in the heart by modulating assembly of cellular junctions, as it does in epithelial kidney cells.^{123,124} Together, these *in vitro* results suggest that AMPK activators might have therapeutic potential for HF, in terms of cardiac fibrosis.

5. Cardio-protective effects of AMPK

5.1 Protection against ischaemia/reperfusion injury

The cardioprotective effect of AMPK in ischaemic hearts is well documented and involves a stimulation of glucose uptake and glycolysis.^{39,125} However, during reperfusion with fatty acids, AMPK

favours fatty acid oxidation, which inhibits glucose oxidation and may decrease cardiac efficiency.^{126,127} In mice lacking AMPK alpha-2 or expressing a cardio-specific dominant negative mutant of the same subunit, the ischaemia-induced stimulation of glucose uptake and glycolysis was inhibited leading to ATP depletion and ischaemic contracture, which were obviously not prevented by the residual activity of the AMPK alpha-1.^{35,128–130} Similarly, the infarct size following coronary ligation was larger in mice expressing a dominant negative AMPK than in controls (Tables 3 and 4).¹³¹

5.2 Potential protection by hormones and pharmacological activators against transition to HF

Several indirect arguments indicate that AMPK could prevent the deleterious effects of hypertrophy on cardiac metabolism and function. Leptin has been reported to protect against cardiac injury in the failing heart by increasing STAT-3 and AMPK activation. It diminished cardiac hypertrophy, inflammation, and cardiac dysfunction.⁴⁹ Similarly, adiponectin could prevent the transition between cardiac hypertrophy to HF by promoting an AMPK-dependent angiogenic regulatory axis⁹⁵ and/or by inhibiting NF-kappaB activation.¹³² It has also been reported that the metabolic changes and hypertrophy induced by angiotensin II in cultured H9C2 cardiomyocytes are prevented by AMPK activation.⁵²

A large number of papers report the beneficial effects of metformin (Table 2). For example, clinical studies have shown that metformin is cardioprotective and improves outcomes in patients with HF.^{133,134} In addition, metformin exerts beneficial effects on cardiac function and survival in murine models of HF.¹³⁵ The effects of MF are mediated, at least in part, by activation of AMPK^{40,51,65} and, interestingly, the cardioprotective effects of MF on murine models of HF are mediated by AMPK activation.¹³⁵ In dogs, metformin attenuated oxidative stress-induced cardiomyocyte apoptosis and prevented the progression of HF along with AMPK activation.¹³⁶ In addition, metformin could also affect the fibrotic response induced by pressure overload.¹³⁷ The protection results from an inhibition of the

Table 4 Changes in metabolism and function in hearts of AMPK transgenic/KO mice submitted to ischaemia

Animal model	Surgery/perfusion	AMPK activation	Pharmacological treatment	Downstream effects	References
AMPK α 2-DN (D157A)	No flow ischaemia (<i>ex vivo</i>)	↓	—	↓ Glucose uptake More rapid ischaemic contracture	130
AMPK α 2-DN (D157A)	CAL	↓	Metformin	↑ Myocardial infarct size	131
AMPK α 2-DN (K45R)	No flow ischaemia	↓	—	↓ Glucose uptake, glycolysis ↓ Myocardial function recovery	128
AMPK α 2-KO	No flow ischaemia	↓	—	↓ Glycogen content, ↓ glycolytic flux More rapid ischaemic contracture No effect on myocardial function recovery	35
AMPK α 2-KO	Low-flow ischaemia	↓	—	↓ Glucose uptake More rapid ischaemic contracture Delayed post-ischaemic contractile recovery	129

CAL, coronary artery ligation.

TGF-beta-smad-signalling pathway but is however independent of AMPK activation.¹³⁷

6. Conclusions and perspectives

Despite its known involvement in energy homeostasis, AMPK activation fails to restore energy balance in the failing heart. Once de-compensated, the heart takes little advantage from AMPK, probably because other deficiencies have brought the failing heart to a point of no return. Clearly, the cardio-protective effects of AMPK activators are obtained on the long term by preventing or delaying the pathological transition from hypertrophy to HF. Whether these beneficial effects only result from energetic recovery of the heart remains to be demonstrated. We speculate that they could instead result from non-metabolic effects, which include anti-proliferative, anti-fibrotic, and angiogenic effects of AMPK.

The evidence for cardioprotective effects of AMPK is only circumstantial and indirect. It relies on the use of pharmacological drugs, with off-target effects, and on the phenotype analysis of mice with whole-body deletion of AMPK alpha subunits. To validate AMPK as a potential target in HF progression, new AMPK-specific, and ideally heart-specific, AMPK activators are needed but remain to be discovered. Similarly, our understanding of the importance of AMPK in the transition between hypertrophy and HF would benefit from the study of mice with a heart-specific deletion of AMPK specific isoforms. In addition, a comprehensive and comparative analysis of the various AMPK isoforms expressed in mice and humans could help to improve an AMPK-mediated therapeutic approach.¹³⁸ Hopefully, the tools to achieve these goals do not seem out of reach.

Finally and as stated in the introduction, HF is a many-sided muscular disorder in which chronic energy deficit is but one aspect. Dysfunctions of calcium handling and of the contractile machinery are integral parts of HF and go together with energy deficit. And the lack of coordination between contraction, calcium, and energy in HF has been appropriately called 'failing complexity'.¹³⁹ As far as we know, the pathological transition to HF is multifactorial and cannot be reduced to a single preponderant disturbed event. To understand this transition and hence to develop a coherent therapeutic approach, the temporal changes of these disturbances should be analysed by systems level integration.^{4,139}

Conflict of interest: none declared.

Funding

S.H. and L.B. are Research Associates of the Fonds National de la Recherche Scientifique, Belgium.

References

- Ventura-Clapier R, Garnier A, Veksler V. Energy metabolism in heart failure. *J Physiol* 2004;**555**:1–13.
- Stanley WC, Recchia FA, Lopaschuk GD. Myocardial substrate metabolism in the normal and failing heart. *Physiol Rev* 2005;**85**:1093–1129.
- Neubauer S. The failing heart—an engine out of fuel. *N Engl J Med* 2007;**356**:1140–1151.
- Turer AT, Malloy CR, Newgard CB, Podgoreanu MV. Energetics and metabolism in the failing heart: important but poorly understood. *Curr Opin Clin Nutr Metab Care* 2010;**13**:458–465.
- Mudd JO, Kass DA. Tackling heart failure in the twenty-first century. *Nature* 2008;**451**:919–928.
- DiBianco R, Shabetai R, Kostuk W, Moran J, Schlant RC, Wright R. A comparison of oral milrinone, digoxin, and their combination in the treatment of patients with chronic heart failure. *N Engl J Med* 1989;**320**:677–683.
- Effect of enalapril on survival in patients with reduced left ventricular ejection fractions and congestive heart failure. The SOLVD Investigators. *N Engl J Med* 1991;**325**:293–302.
- Hardie DG. AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol* 2007;**8**:774–785.
- Hue L, Rider MH. The AMP-activated protein kinase: more than an energy sensor. *Essays Biochem* 2007;**43**:121–137.
- Ashrafian H, Frenneaux MP, Opie LH. Metabolic mechanisms in heart failure. *Circulation* 2007;**116**:434–448.
- Jameel MN, Zhang J. Myocardial energetics in left ventricular hypertrophy. *Curr Cardiol Rev* 2009;**5**:243–250.
- Nascimben L, Ingwall JS, Pauletto P, Friedrich J, Gwathmey JK, Saks V et al. Creatine kinase system in failing and nonfailing human myocardium. *Circulation* 1996;**94**:1894–1901.
- Neubauer S, Horn M, Cramer M, Harre K, Newell JB, Peters W et al. Myocardial phosphocreatine-to-ATP ratio is a predictor of mortality in patients with dilated cardiomyopathy. *Circulation* 1997;**96**:2190–2196.
- Neubauer S, Newell JB, Ingwall JS. Metabolic consequences and predictability of ventricular fibrillation in hypoxia. A 31P- and 23Na-nuclear magnetic resonance study of the isolated rat heart. *Circulation* 1992;**86**:302–310.
- Hue L, Taegtmeyer H. The Randle cycle revisited: a new head for an old hat. *Am J Physiol Endocrinol Metab* 2009;**297**:E578–591.
- Lopaschuk GD, Ussher JR, Folmes CD, Jaswal JS, Stanley WC. Myocardial fatty acid metabolism in health and disease. *Physiol Rev* 2010;**90**:207–258.
- Nikolaidis LA, Sturzu A, Stolarski C, Elahi D, Shen YT, Shannon RP. The development of myocardial insulin resistance in conscious dogs with advanced dilated cardiomyopathy. *Cardiovasc Res* 2004;**61**:297–306.
- Rosca MG, Hoppel CL. Mitochondria in heart failure. *Cardiovasc Res* 2010;**88**:40–50.
- Liao R, Jain M, Cui L, D'Agostino J, Aiello F, Luptak I et al. Cardiac-specific overexpression of GLUT1 prevents the development of heart failure attributable to pressure overload in mice. *Circulation* 2002;**106**:2125–2131.
- Luptak I, Balschi JA, Xing Y, Leone TC, Kelly DP, Tian R. Decreased contractile and metabolic reserve in peroxisome proliferator-activated receptor-alpha-null hearts can be rescued by increasing glucose transport and utilization. *Circulation* 2005;**112**:2339–2346.
- Yan J, Young ME, Cui L, Lopaschuk GD, Liao R, Tian R. Increased glucose uptake and oxidation in mouse hearts prevent high fatty acid oxidation but cause cardiac dysfunction in diet-induced obesity. *Circulation* 2009;**119**:2818–2828.
- Huss JM, Kelly DP. Mitochondrial energy metabolism in heart failure: a question of balance. *J Clin Invest* 2005;**115**:547–555.
- Marin-Garcia J, Goldenthal MJ. Mitochondrial centrality in heart failure. *Heart Fail Rev* 2008;**13**:137–150.
- Murray AJ, Edwards LM, Clarke K. Mitochondria and heart failure. *Curr Opin Clin Nutr Metab Care* 2007;**10**:704–711.
- Ventura-Clapier R, Garnier A, Veksler V. Transcriptional control of mitochondrial biogenesis: the central role of PGC-1alpha. *Cardiovasc Res* 2008;**79**:208–217.
- Hardie DG, Carling D. The AMP-activated protein kinase—fuel gauge of the mammalian cell? *Eur J Biochem* 1997;**246**:259–273.
- Cheung PC, Salt IP, Davies SP, Hardie DG, Carling D. Characterization of AMP-activated protein kinase gamma-subunit isoforms and their role in AMP binding. *Biochem J* 2000;**346**(Pt 3):659–669.
- McBride A, Ghilagaber S, Nikolaev A, Hardie DG. The glycogen-binding domain on the AMPK beta subunit allows the kinase to act as a glycogen sensor. *Cell Metab* 2009;**9**:23–34.
- Thornton C, Snowden MA, Carling D. Identification of a novel AMP-activated protein kinase beta subunit isoform that is highly expressed in skeletal muscle. *J Biol Chem* 1998;**273**:12443–12450.
- Xiao B, Heath R, Saiu P, Leiper FC, Leone P, Jing C et al. Structural basis for AMP binding to mammalian AMP-activated protein kinase. *Nature* 2007;**449**:496–500.
- Blair E, Redwood C, Ashrafian H, Oliveira M, Broxholme J, Kerr B et al. Mutations in the gamma(2) subunit of AMP-activated protein kinase cause familial hypertrophic cardiomyopathy: evidence for the central role of energy compromise in disease pathogenesis. *Hum Mol Genet* 2001;**10**:1215–1220.
- Kahn BB, Alquier T, Carling D, Hardie DG. AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab* 2005;**1**:15–25.
- Witters LA, Kemp BE, Means AR. Chutes and Ladders: the search for protein kinases that act on AMPK. *Trends Biochem Sci* 2006;**31**:13–16.
- Sakamoto K, Zarrinpashneh E, Budas GR, Pouleur AC, Dutta A, Prescott AR et al. Deficiency of LKB1 in heart prevents ischemia-mediated activation of AMPKalpha2 but not AMPKalpha1. *Am J Physiol Endocrinol Metab* 2006;**290**:E780–E788.
- Zarrinpashneh E, Carjaval K, Beauloye C, Ginion A, Mateo P, Pouleur AC et al. Role of the alpha2-isoform of AMP-activated protein kinase in the metabolic response of the heart to no-flow ischemia. *Am J Physiol Heart Circ Physiol* 2006;**291**:H2875–H2883.

36. Hurley RL, Anderson KA, Franzone JM, Kemp BE, Means AR, Witters LA. The Ca^{2+} /calmodulin-dependent protein kinase kinases are AMP-activated protein kinase kinases. *J Biol Chem* 2005;**280**:29060–29066.
37. Hawley SA, Pan DA, Mustard KJ, Ross L, Bain J, Edelman AM *et al*. Calmodulin-dependent protein kinase kinase-beta is an alternative upstream kinase for AMP-activated protein kinase. *Cell Metab* 2005;**2**:9–19.
38. Woods A, Dickerson K, Heath R, Hong SP, Momcilovic M, Johnstone SR *et al*. Ca^{2+} /calmodulin-dependent protein kinase kinase-beta acts upstream of AMP-activated protein kinase in mammalian cells. *Cell Metab* 2005;**2**:21–33.
39. Marsin AS, Bertrand L, Rider MH, Deprez J, Beauloye C, Vincent MF *et al*. Phosphorylation and activation of heart PFK-2 by AMPK has a role in the stimulation of glycolysis during ischaemia. *Curr Biol* 2000;**10**:1247–1255.
40. Hardie DG. AMP-activated protein kinase as a drug target. *Annu Rev Pharmacol Toxicol* 2007;**47**:185–210.
41. Beauloye C, Marsin AS, Bertrand L, Vanoverschelde JL, Rider MH, Hue L. The stimulation of heart glycolysis by increased workload does not require AMP-activated protein kinase but a wortmannin-sensitive mechanism. *FEBS Lett* 2002;**531**:324–328.
42. Xu M, Zhao YT, Song Y, Hao TP, Lu ZZ, Han QD *et al*. Alpha1-adrenergic receptors activate AMP-activated protein kinase in rat hearts. *Sheng Li Xue Bao* 2007;**59**:175–182.
43. Jaswal JS, Lund CR, Keung W, Beker DL, Rebeyka IM, Lopaschuk GD. Isoproterenol stimulates 5'-AMP-activated protein kinase and fatty acid oxidation in neonatal hearts. *Am J Physiol Heart Circ Physiol* 2010;**299**:H1135–H1145.
44. Horman S, Morel N, Vertommen D, Hussain N, Neumann D, Beauloye C *et al*. AMP-activated protein kinase phosphorylates and desensitizes smooth muscle myosin light chain kinase. *J Biol Chem* 2008;**283**:18505–18512.
45. Saeedi R, Saran VV, Wu SS, Kume ES, Paulson K, Chan AP *et al*. AMP-activated protein kinase influences metabolic remodeling in H9c2 cells hypertrophied by arginine vasopressin. *Am J Physiol Heart Circ Physiol* 2009;**296**:H1822–1832.
46. Fang X, Palanivel R, Cresser J, Schram K, Ganguly R, Thong FS *et al*. An APPL1-AMPK signaling axis mediates beneficial metabolic effects of adiponectin in the heart. *Am J Physiol Endocrinol Metab* 2010;**299**:E721–E729.
47. Hopkins TA, Ouchi N, Shibata R, Walsh K. Adiponectin actions in the cardiovascular system. *Cardiovasc Res* 2007;**74**:11–18.
48. Shibata R, Sato K, Pimentel DR, Takemura Y, Kihara S, Ohashi K *et al*. Adiponectin protects against myocardial ischemia-reperfusion injury through AMPK- and COX-2-dependent mechanisms. *Nat Med* 2005;**11**:1096–1103.
49. McGaffin KR, Witham WG, Yester KA, Romano LC, O'Doherty RM, McTiernan CF *et al*. Cardiac-specific leptin receptor deletion exacerbates ischaemic heart failure in mice. *Cardiovasc Res* 2011;**89**:60–71.
50. Miller EJ, Li J, Leng L, McDonald C, Atsumi T, Bucala R *et al*. Macrophage migration inhibitory factor stimulates AMP-activated protein kinase in the ischaemic heart. *Nature* 2008;**451**:578–582.
51. Wang Y, Tao L, Yuan Y, Lau WB, Li R, Lopez BL *et al*. Cardioprotective effect of adiponectin is partially mediated by its AMPK-independent antinflammatory action. *Am J Physiol Endocrinol Metab* 2009;**297**:E384–E391.
52. Stuck BJ, Lenski M, Bohm M, Laufs U. Metabolic switch and hypertrophy of cardiomyocytes following treatment with angiotensin II are prevented by AMP-activated protein kinase. *J Biol Chem* 2008;**283**:32562–32569.
53. Beauloye C, Marsin AS, Bertrand L, Krause U, Hardie DG, Vanoverschelde JL *et al*. Insulin antagonizes AMP-activated protein kinase activation by ischemia or anoxia in rat hearts, without affecting total adenine nucleotides. *FEBS Lett* 2001;**505**:348–352.
54. Kovacic S, Soltys CL, Barr AJ, Shiojima I, Walsh K, Dyck JR. Akt activity negatively regulates phosphorylation of AMP-activated protein kinase in the heart. *J Biol Chem* 2003;**278**:39422–39427.
55. Horman S, Vertommen D, Heath R, Neumann D, Mouton V, Woods A *et al*. Insulin antagonizes ischemia-induced Thr172 phosphorylation of AMP-activated protein kinase alpha-subunits in heart via hierarchical phosphorylation of Ser485/491. *J Biol Chem* 2006;**281**:5335–5340.
56. Nagata D, Takeda R, Sata M, Satonaka H, Suzuki E, Nagano T *et al*. AMP-activated protein kinase inhibits angiotensin II-stimulated vascular smooth muscle cell proliferation. *Circulation* 2004;**110**:444–451.
57. Corton JM, Gillespie JG, Hawley SA, Hardie DG. 5-aminoimidazole-4-carboxamide ribonucleoside. A specific method for activating AMP-activated protein kinase in intact cells? *Eur J Biochem* 1995;**229**:558–565.
58. Henin N, Vincent MF, Gruber HE, Van den Berghe G. Inhibition of fatty acid and cholesterol synthesis by stimulation of AMP-activated protein kinase. *FASEB J* 1995;**9**:541–546.
59. Javaux F, Vincent MF, Wagner DR, van den Berghe G. Cell-type specificity of inhibition of glycolysis by 5-amino-4-imidazolecarboxamide riboside. Lack of effect in rabbit cardiomyocytes and human erythrocytes, and inhibition in FTO-2B rat hepatoma cells. *Biochem J* 1995;**305**(Pt 3):913–919.
60. Guigas B, Taleux N, Foretz M, Detaille D, Andreelli F, Viollet B *et al*. AMP-activated protein kinase-independent inhibition of hepatic mitochondrial oxidative phosphorylation by AICA riboside. *Biochem J* 2007;**404**:499–507.
61. Guigas B, Bertrand L, Taleux N, Foretz M, Wiernsperger N, Vertommen D *et al*. 5-Aminoimidazole-4-carboxamide-1-beta-D-ribofuranoside and metformin inhibit hepatic glucose phosphorylation by an AMP-activated protein kinase-independent effect on glucokinase translocation. *Diabetes* 2006;**55**:865–874.
62. Guigas B, Sakamoto K, Taleux N, Reyna SM, Musi N, Viollet B *et al*. Beyond AICA riboside: in search of new specific AMP-activated protein kinase activators. *IUBMB Life* 2009;**61**:18–26.
63. Cool B, Zinker B, Chiou W, Kifle L, Cao N, Perham M *et al*. Identification and characterization of a small molecule AMPK activator that treats key components of type 2 diabetes and the metabolic syndrome. *Cell Metab* 2006;**3**:403–416.
64. Fryer LG, Parbu-Patel A, Carling D. The Anti-diabetic drugs rosiglitazone and metformin stimulate AMP-activated protein kinase through distinct signaling pathways. *J Biol Chem* 2002;**277**:25226–25232.
65. Zhou G, Myers R, Li Y, Chen Y, Shen X, Shen X, Fenyk-Melody J *et al*. Role of AMP-activated protein kinase in mechanism of metformin action. *J Clin Invest* 2001;**108**:1167–1174.
66. Owen MR, Doran E, Halestrap AP. Evidence that metformin exerts its anti-diabetic effects through inhibition of complex 1 of the mitochondrial respiratory chain. *Biochem J* 2000;**348**(Pt 3):607–614.
67. El-Mir MY, Nogueira V, Fontaine E, Averet N, Rigoulet M, Leverve X. Dimethylbiguanide inhibits cell respiration via an indirect effect targeted on the respiratory chain complex I. *J Biol Chem* 2000;**275**:223–228.
68. Kalender A, Selvaraj A, Kim SY, Gulati P, Brule S, Viollet B *et al*. Metformin, independent of AMPK, inhibits mTORC1 in a rag GTPase-dependent manner. *Cell Metab* 2010;**11**:390–401.
69. Saeedi R, Parsons HL, Wambolt RB, Paulson K, Sharma V, Dyck JR *et al*. Metabolic actions of metformin in the heart can occur by AMPK-independent mechanisms. *Am J Physiol Heart Circ Physiol* 2008;**294**:H2497–H2506.
70. Emerling BM, Weinberg F, Snyder C, Burgess Z, Mutlu GM, Viollet B *et al*. Hypoxic activation of AMPK is dependent on mitochondrial ROS but independent of an increase in AMP/ATP ratio. *Free Radic Biol Med* 2009;**46**:1386–1391.
71. Choi SL, Kim SJ, Lee KT, Kim J, Mu J, Birnbaum MJ *et al*. The regulation of AMP-activated protein kinase by H(2)O(2). *Biochem Biophys Res Commun* 2001;**287**:92–97.
72. Zmijewski JW, Banerjee S, Bae H, Friggeri A, Lazarowski ER, Abraham E. Exposure to hydrogen peroxide induces oxidation and activation of AMP-activated protein kinase. *J Biol Chem* 2010;**285**:33154–33164.
73. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG *et al*. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 2003;**425**:191–196.
74. Gledhill JR, Montgomery MG, Leslie AG, Walker JE. Mechanism of inhibition of bovine F1-ATPase by resveratrol and related polyphenols. *Proc Natl Acad Sci U S A* 2007;**104**:13632–13637.
75. Um JH, Park SJ, Kang H, Yang S, Foretz M, McBurney MW *et al*. AMP-activated protein kinase-deficient mice are resistant to the metabolic effects of resveratrol. *Diabetes* 2010;**59**:554–563.
76. Bungard D, Fuerth BJ, Zeng PY, Faubert B, Maas NL, Viollet B *et al*. Signaling kinase AMPK activates stress-promoted transcription via histone H2B phosphorylation. *Science* 2010;**329**:1201–1205.
77. Jager S, Handschin C, St-Pierre J, Spiegelman BM. AMP-activated protein kinase (AMPK) action in skeletal muscle via direct phosphorylation of PGC-1alpha. *Proc Natl Acad Sci U S A* 2007;**104**:12017–12022.
78. Dong Y, Zhang M, Liang B, Xie Z, Zhao Z, Asfa S *et al*. Reduction of AMP-activated protein kinase alpha2 increases endoplasmic reticulum stress and atherosclerosis *in vivo*. *Circulation* 2010;**121**:792–803.
79. Qi D, Hu X, Wu X, Merk M, Leng L, Bucala R *et al*. Cardiac macrophage migration inhibitory factor inhibits JNK pathway activation and injury during ischemia/reperfusion. *J Clin Invest* 2009;**119**:3807–3816.
80. Li XN, Song J, Zhang L, LeMaire SA, Hou X, Zhang C *et al*. Activation of the AMPK-FOXO3 pathway reduces fatty acid-induced increase in intracellular reactive oxygen species by upregulating thioredoxin. *Diabetes* 2009;**58**:2246–2257.
81. Hou X, Song J, Li XN, Zhang L, Wang X, Chen L *et al*. Metformin reduces intracellular reactive oxygen species levels by upregulating expression of the antioxidant thioredoxin via the AMPK-FOXO3 pathway. *Biochem Biophys Res Commun* 2010;**396**:199–205.
82. Ceolotto G, Gallo A, Papparella I, Franco L, Murphy E, Iori E *et al*. Rosiglitazone reduces glucose-induced oxidative stress mediated by NAD(P)H oxidase via AMPK-dependent mechanism. *Arterioscler Thromb Vasc Biol* 2007;**27**:2627–2633.
83. Bertrand L, Horman S, Beauloye C, Vanoverschelde JL. Insulin signalling in the heart. *Cardiovasc Res* 2008;**79**:238–248.
84. Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS *et al*. AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell* 2008;**30**:214–226.
85. Inoki K, Li Y, Xu T, Guan KL. Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. *Genes Dev* 2003;**17**:1829–1834.
86. Proud CG. Signalling to translation: how signal transduction pathways control the protein synthetic machinery. *Biochem J* 2007;**403**:217–234.
87. Horman S, Browne G, Krause U, Patel J, Vertommen D, Bertrand L *et al*. Activation of AMP-activated protein kinase leads to the phosphorylation of elongation factor 2 and an inhibition of protein synthesis. *Curr Biol* 2002;**12**:1419–1423.

88. Lee JW, Park S, Takahashi Y, Wang HG. The association of AMPK with ULK1 regulates autophagy. *PLoS One* 2010;**5**:e15394.
89. Jung CH, Jun CB, Ro SH, Kim YM, Otto NM, Cao J et al. ULK-Atg13-FIP200 complexes mediate mTOR signaling to the autophagy machinery. *Mol Biol Cell* 2009;**20**:1992–2003.
90. Hosokawa N, Sasaki T, Iemura S, Natsume T, Hara T, Mizushima N. Atg101, a novel mammalian autophagy protein interacting with Atg13. *Autophagy* 2009;**5**:973–979.
91. Ganley IG, Lam du H, Wang J, Ding X, Chen S, Jiang X. ULK1.ATG13.FIP200 complex mediates mTOR signaling and is essential for autophagy. *J Biol Chem* 2009;**284**:12297–12305.
92. Reihill JA, Ewart MA, Hardie DG, Salt IP. AMP-activated protein kinase mediates VEGF-stimulated endothelial NO production. *Biochem Biophys Res Commun* 2007;**354**:1084–1088.
93. Chen ZP, Mitchellhill KI, Michell BJ, Stapleton D, Rodriguez-Crespo I, Witters LA et al. AMP-activated protein kinase phosphorylation of endothelial NO synthase. *FEBS Lett* 1999;**443**:285–289.
94. Li J, Hu X, Selvakumar P, Russell RR III, Cushman SW, Holman GD et al. Role of the nitric oxide pathway in AMPK-mediated glucose uptake and GLUT4 translocation in heart muscle. *Am J Physiol Endocrinol Metab* 2004;**287**:E834–E841.
95. Shimano M, Ouchi N, Shibata R, Ohashi K, Pimentel DR, Murohara T et al. Adiponectin deficiency exacerbates cardiac dysfunction following pressure overload through disruption of an AMPK-dependent angiogenic response. *J Mol Cell Cardiol* 2010;**49**:210–220.
96. Tian R, Musi N, D'Agostino J, Hirshman MF, Goodyear LJ. Increased adenosine monophosphate-activated protein kinase activity in rat hearts with pressure-overload hypertrophy. *Circulation* 2001;**104**:1664–1669.
97. Shioi T, McMullen JR, Tarnavski O, Converso K, Sherwood MC, Manning WJ et al. Rapamycin attenuates load-induced cardiac hypertrophy in mice. *Circulation* 2003;**107**:1664–1670.
98. McMullen JR, Sherwood MC, Tarnavski O, Zhang L, Dorfman AL, Shioi T et al. Inhibition of mTOR signaling with rapamycin regresses established cardiac hypertrophy induced by pressure overload. *Circulation* 2004;**109**:3050–3055.
99. Chan AY, Dolinsky VW, Soltys CL, Viollet B, Baksh S, Light PE et al. Resveratrol inhibits cardiac hypertrophy via AMP-activated protein kinase and Akt. *J Biol Chem* 2008;**283**:24194–24201.
100. Chan AY, Soltys CL, Young ME, Proud CG, Dyck JR. Activation of AMP-activated protein kinase inhibits protein synthesis associated with hypertrophy in the cardiac myocyte. *J Biol Chem* 2004;**279**:32771–32779.
101. Li HL, Yin R, Chen D, Liu D, Wang D, Yang Q et al. Long-term activation of adenosine monophosphate-activated protein kinase attenuates pressure-overload-induced cardiac hypertrophy. *J Cell Biochem* 2007;**100**:1086–1099.
102. Zarrinpashneh E, Beauloye C, Ginion A, Pouleur AC, Havaux X, Hue L et al. AMP-Kalpa2 counteracts the development of cardiac hypertrophy induced by isoproterenol. *Biochem Biophys Res Commun* 2008;**376**:677–681.
103. Zhang P, Hu X, Xu X, Fasset J, Zhu G, Viollet B et al. AMP activated protein kinase-alpha2 deficiency exacerbates pressure-overload-induced left ventricular hypertrophy and dysfunction in mice. *Hypertension* 2008;**52**:918–924.
104. Jessen N, Koh HJ, Folmes CD, Wagg C, Fujii N, Lofgren B et al. Ablation of LKB1 in the heart leads to energy deprivation and impaired cardiac function. *Biochim Biophys Acta* 2010;**1802**:593–600.
105. Ikeda Y, Sato K, Pimentel DR, Sam F, Shaw RJ, Dyck JR et al. Cardiac-specific deletion of LKB1 leads to hypertrophy and dysfunction. *J Biol Chem* 2009;**284**:35839–35849.
106. Shibata R, Ouchi N, Ito M, Kihara S, Shiojima I, Pimentel DR et al. Adiponectin-mediated modulation of hypertrophic signals in the heart. *Nat Med* 2004;**10**:1384–1389.
107. Thomson DM, Hancock CR, Evanson BG, Kenney SG, Malan BB, Mongillo AD et al. Skeletal muscle dysfunction in muscle-specific LKB1 knockout mice. *J Appl Physiol* 2010;**108**:1775–1785.
108. Shiojima I, Sato K, Izumiya Y, Schiekofer S, Ito M, Liao R et al. Disruption of coordinated cardiac hypertrophy and angiogenesis contributes to the transition to heart failure. *J Clin Invest* 2005;**115**:2108–2118.
109. Liao Y, Takashima S, Maeda N, Ouchi N, Komamura K, Shimomura I et al. Exacerbation of heart failure in adiponectin-deficient mice due to impaired regulation of AMPK and glucose metabolism. *Cardiovasc Res* 2005;**67**:705–713.
110. Arany Z, He H, Lin J, Hoyer K, Handschin C, Toka O et al. Transcriptional coactivator PGC-1 alpha controls the energy state and contractile function of cardiac muscle. *Cell Metab* 2005;**1**:259–271.
111. Arany Z, Novikov M, Chin S, Ma Y, Rosenzweig A, Spiegelman BM. Transverse aortic constriction leads to accelerated heart failure in mice lacking PPAR-gamma coactivator 1alpha. *Proc Natl Acad Sci U S A* 2006;**103**:10086–10091.
112. Leone TC, Lehman JJ, Finck BN, Schaeffer PJ, Wende AR, Boudina S et al. PGC-1alpha deficiency causes multi-system energy metabolic derangements: muscle dysfunction, abnormal weight control and hepatic steatosis. *PLoS Biol* 2005;**3**:e101.
113. Swynghedauw B. Molecular mechanisms of myocardial remodeling. *Physiol Rev* 1999;**79**:215–262.
114. Graham HK, Horn M, Trafford AW. Extracellular matrix profiles in the progression to heart failure. European Young Physiologists Symposium Keynote Lecture-Bratislava 2007. *Acta Physiol (Oxf)* 2008;**194**:3–21.
115. Rosenkranz S. TGF-beta1 and angiotensin networking in cardiac remodeling. *Cardiovasc Res* 2004;**63**:423–432.
116. Ferrario CM, Strawn WB. Role of the renin-angiotensin-aldosterone system and proinflammatory mediators in cardiovascular disease. *Am J Cardiol* 2006;**98**:121–128.
117. Sugihara N, Genda A, Shimizu M, Suematsu T, Kita Y, Minamoto M et al. Diastolic dysfunction and its relation to myocardial fibrosis in essential hypertension. *J Cardiol* 1988;**18**:353–361.
118. Banerjee I, Fuseler JW, Price RL, Borg TK, Baudino TA. Determination of cell types and numbers during cardiac development in the neonatal and adult rat and mouse. *Am J Physiol Heart Circ Physiol* 2007;**293**:H1883–H1891.
119. Pelouch V, Dixon IM, Golfman L, Beamish RE, Dhalla NS. Role of extracellular matrix proteins in heart function. *Mol Cell Biochem* 1993;**129**:101–120.
120. Diez J. Mechanisms of cardiac fibrosis in hypertension. *J Clin Hypertens (Greenwich)* 2007;**9**:546–550.
121. Du J, Guan T, Zhang H, Xia Y, Liu F, Zhang Y. Inhibitory crosstalk between ERK and AMPK in the growth and proliferation of cardiac fibroblasts. *Biochem Biophys Res Commun* 2008;**368**:402–407.
122. Mishra R, Cool BL, Laderoute KR, Foretz M, Viollet B, Simonson MS. AMP-activated protein kinase inhibits transforming growth factor-beta-induced Smad3-dependent transcription and myofibroblast transdifferentiation. *J Biol Chem* 2008;**283**:10461–10469.
123. Zheng B, Cantley LC. Regulation of epithelial tight junction assembly and disassembly by AMP-activated protein kinase. *Proc Natl Acad Sci U S A* 2007;**104**:819–822.
124. Zhang L, Li J, Young LH, Caplan MJ. AMP-activated protein kinase regulates the assembly of epithelial tight junctions. *Proc Natl Acad Sci U S A* 2006;**103**:17272–17277.
125. Kim AS, Miller EJ, Young LH. AMP-activated protein kinase: a core signalling pathway in the heart. *Acta Physiol (Oxf)* 2009;**196**:37–53.
126. Dyck JR, Lopaschuk GD. AMPK alterations in cardiac physiology and pathology: enemy or ally? *J Physiol* 2006;**574**:95–112.
127. Folmes CD, Wagg CS, Shen M, Clanachan AS, Tian R, Lopaschuk GD. Suppression of 5'-AMP-activated protein kinase activity does not impair recovery of contractile function during reperfusion of ischemic hearts. *Am J Physiol Heart Circ Physiol* 2009;**297**:H313–H321.
128. Russell RR III, Li J, Coven DL, Pypaert M, Zechner C, Palmeri M et al. AMP-activated protein kinase mediates ischemic glucose uptake and prevents posts ischemic cardiac dysfunction, apoptosis, and injury. *J Clin Invest* 2004;**114**:495–503.
129. Carvajal K, Zarrinpashneh E, Szarsoi O, Joubert F, Athea Y, Mateo P et al. Dual cardiac contractile effects of the alpha2-AMPK deletion in low-flow ischemia and reperfusion. *Am J Physiol Heart Circ Physiol* 2007;**292**:H3136–H3147.
130. Xing Y, Musi N, Fujii N, Zou L, Luptak I, Hirshman MF et al. Glucose metabolism and energy homeostasis in mouse hearts overexpressing dominant negative alpha2 subunit of AMP-activated protein kinase. *J Biol Chem* 2003;**278**:28372–28377.
131. Calvert JW, Gundewar S, Jha S, Greer JJ, Bestermann WH, Tian R et al. Acute metformin therapy confers cardioprotection against myocardial infarction via AMPK-eNOS-mediated signaling. *Diabetes* 2008;**57**:696–705.
132. Wang C, Li L, Zhang ZG, Fan D, Zhu Y, Wu LL. Globular adiponectin inhibits angiotensin II-induced nuclear factor kappaB activation through AMP-activated protein kinase in cardiac hypertrophy. *J Cell Physiol* 2010;**222**:149–155.
133. Home PD, Pocock SJ, Beck-Nielsen H, Curtis PS, Gomis R, Hanefeld M et al. Rosiglitazone evaluated for cardiovascular outcomes in oral agent combination therapy for type 2 diabetes (RECORD): a multicentre, randomised, open-label trial. *Lancet* 2009;**373**:2125–2135.
134. Eurich DT, Majumdar SR, McAlister FA, Tsuyuki RT, Johnson JA. Improved clinical outcomes associated with metformin in patients with diabetes and heart failure. *Diabetes Care* 2005;**28**:2345–2351.
135. Gundewar S, Calvert JW, Jha S, Toedt-Pingel I, Ji SY, Nunez D et al. Activation of AMP-activated protein kinase by metformin improves left ventricular function and survival in heart failure. *Circ Res* 2009;**104**:403–411.
136. Sasaki H, Asanuma H, Fujita M, Takahama H, Wakeno M, Ito S et al. Metformin prevents progression of heart failure in dogs: role of AMP-activated protein kinase. *Circulation* 2009;**119**:2568–2577.
137. Xiao H, Ma X, Feng W, Fu Y, Lu Z, Xu M et al. Metformin attenuates cardiac fibrosis by inhibiting the TGFbeta1-Smad3 signalling pathway. *Cardiovasc Res* 2010;**87**:504–513.
138. Kim M, Tian R. Targeting AMPK for cardiac protection: opportunities and challenges. *J Mol Cell Cardiol* 2010; doi:10.1016/j.jmcc.2010.12.004. Published online ahead of print 13 December 2010.
139. Brutsaert DL. Cardiac failure: quo vadis? *Eur J Heart Fail* 2010;**12**:785–788.
140. Dreyer HC, Fujita S, Cadenas JG, Chinkes DL, Volpi E, Rasmussen BB. Resistance exercise increases AMPK activity and reduces 4E-BP1 phosphorylation and protein synthesis in human skeletal muscle. *J Physiol* 2006;**576**:613–624.

141. Kudo N, Gillespie JG, Kung L, Witters LA, Schulz R, Clanachan AS et al. Characterization of 5'AMP-activated protein kinase activity in the heart and its role in inhibiting acetyl-CoA carboxylase during reperfusion following ischemia. *Biochim Biophys Acta* 1996;**1301**:67–75.
142. Horman S, Beauloye C, Vertommen D, Vanoverschelde JL, Hue L, Rider MH. Myocardial ischemia and increased heart work modulate the phosphorylation state of eukaryotic elongation factor-2. *J Biol Chem* 2003;**278**:41970–41976.
143. Browne GJ, Finn SG, Proud CG. Stimulation of the AMP-activated protein kinase leads to activation of eukaryotic elongation factor 2 kinase and to its phosphorylation at a novel site, serine 398. *J Biol Chem* 2004;**279**:12220–12231.
144. Russell RR III, Bergeron R, Shulman GI, Young LH. Translocation of myocardial GLUT-4 and increased glucose uptake through activation of AMPK by AICAR. *Am J Physiol* 1999;**277**:H643–H649.
145. Bolster DR, Crozier SJ, Kimball SR, Jefferson LS. AMP-activated protein kinase suppresses protein synthesis in rat skeletal muscle through down-regulated mammalian target of rapamycin (mTOR) signaling. *J Biol Chem* 2002;**277**:23977–23980.
146. Krause U, Bertrand L, Hue L. Control of p70 ribosomal protein S6 kinase and acetyl-CoA carboxylase by AMP-activated protein kinase and protein phosphatases in isolated hepatocytes. *Eur J Biochem* 2002;**269**:3751–3759.
147. Dubbelhuis PF, Meijer AJ. Hepatic amino acid-dependent signaling is under the control of AMP-dependent protein kinase. *FEBS Lett* 2002;**521**:39–42.