# BDNF-based synaptic repair as a disease-modifying strategy for neurodegenerative diseases

#### Bai Lu<sup>1</sup>, Guhan Nagappan<sup>1</sup>, Xiaoming Guan<sup>1</sup>, Pradeep J. Nathan<sup>2,3</sup> and Paul Wren<sup>1</sup>

Abstract | Increasing evidence suggests that synaptic dysfunction is a key pathophysiological hallmark in neurodegenerative disorders, including Alzheimer's disease. Understanding the role of brain-derived neurotrophic factor (BDNF) in synaptic plasticity and synaptogenesis, the impact of the *BDNF* Val66Met polymorphism in Alzheimer's disease-relevant endophenotypes — including episodic memory and hippocampal volume — and the technological progress in measuring synaptic changes in humans all pave the way for a 'synaptic repair' therapy for neurodegenerative diseases that targets pathophysiology rather than pathogenesis. This article reviews the key issues in translating BDNF biology into synaptic repair therapies.

#### Disease-modifying therapies

Medical therapies that address the cause of the disease either directly or indirectly and thereby modify the course of the disease: that is, slow down, halt or reverse disease progression.

Symptomatic treatment

A medical therapy that only relieves or controls the disease symptoms but not its cause per se.

<sup>1</sup>GlaxoSmithKline, R&D China, Shanghai, 201203, China.
<sup>2</sup>Medicines Discovery and Development, GlaxoSmithKline, Clinical Unit Cambridge, Cambridge, CB2 2GG, UK.
<sup>3</sup>Brain Mapping Unit, Department of Psychiatry, University of Cambridge, Cambridge, CB2 OSZ, UK.
Correspondence to B.L.
e-mail: <u>bai.b.lu@gsk.com</u>
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Published online 15 May 2013 Over the past 15 years, there has been significant advancement in neuroscience research, as evidenced by the dramatic increase in the number of papers published. Despite great progress in understanding the molecular mechanisms underlying nervous system disorders, such findings have not been effectively translated into developing disease-modifying therapies for neurological and psychiatric diseases. As the average human life expectancy continues to increase<sup>1</sup>, the prevalence of age-related neurodegenerative diseases, particularly Alzheimer's disease (AD)<sup>2</sup> and Parkinson's disease (PD)<sup>3</sup>, increases concomitantly, affecting millions worldwide.

In the development of disease-modifying drugs (as opposed to symptomatic treatment) for neurodegenerative diseases, efforts have so far primarily focused on a 'toxin-reducing' approach. This approach is based on the idea that removing the causes — such as amyloid- $\beta$  (A $\beta$ ) aggregates, amyloid plaques, tau aggregates and neurofibrillary tangles in AD<sup>4</sup> — would halt disease progression. Emerging data from several Phase III clinical studies targeting the amyloid cascade suggest that this approach is ineffective, at least in patients at an advanced stage of the disease<sup>5-7</sup>.

In this Review, we highlight evidence that the progression of neurodegenerative disorders is more tightly associated with synapse degeneration rather than with 'toxin build-up'. This suggests that synapse loss is a major pathophysiological hallmark shared by all neurodegenerative diseases and leads to the proposal that effective therapies should target this pathophysiological feature of neurodegenerative diseases rather than their pathogenesis. We review data showing that brainderived neurotrophic factor (BDNF), in addition to its pro-survival effects, has powerful synaptic effects — promoting synaptic transmission, synaptic plasticity and synaptic growth (also called synaptogenesis) — and we propose a paradigm-shifting, BDNF-based 'synaptic repair' strategy for neurodegenerative diseases. Thus, a combination of BDNF–neurotrophic receptor tyrosine kinase TRKB (also known as NTRK2) pathway modulators and more reliable and sensitive methods to measure synaptic changes in humans *in vivo* could pave the way for developing effective disease-modifying therapies.

#### Challenges in neurodegeneration therapy

Many factors have contributed to the lack of success in the development of disease-modifying medicines for neurodegenerative disorders. First, the underlying disease mechanisms are complex and poorly understood. Although several mechanisms and pathways have been implicated in neurodegenerative diseases — including accumulation of neurotoxic substances, inflammation, lipid metabolism, oxidative stress, autophagy, protein degradation and mitochondrial dysfunction<sup>8</sup> — it remains unclear whether they are the cause of the disease or the consequence of the primary and/or secondary damage. Consequently, therapies based on some of these individual mechanisms have not been clinically successful. Second, given that accumulation of misfolded toxic proteins in the brain is considered

#### Phase III

A trial conducted to gather new information about the safety and effectiveness of a particular therapy in a larger group of patients than that used in Phase II clinical trials.

#### Synaptic plasticity

Activity-dependent modulation of synaptic structure and/or function.

#### Synaptogenesis

The formation of new synapses between neurons, and the maturation and stabilization of existing synapses. It is also termed synaptic growth. to be a key pathogenic factor for neurodegenerative diseases, efforts to develop disease-modifying therapies for AD and other neurodegenerative diseases have thus far followed a toxin-reducing approach9. However, clinical studies to date have shown that lowering toxic proteins, such as  $A\beta$  in AD, reduced amyloid plaque pathology but failed to improve the clinical outcome<sup>5,6</sup>. Third, although many pharmacological agents showed beneficial effects in various animal models<sup>10-13</sup> of AD, few have translated into clinical efficacy<sup>14,15</sup>. It therefore remains a challenge to develop animal models with predictive value. Last, the lack of qualified biomarkers and robust clinical measurements hampers accurate and early diagnosis, patient stratification and early evaluation of therapeutic efficacy of new medications. Thus, clinical trials designed to evaluate disease modification are usually lengthy and involve a large number of patients.

An emerging idea in the toxin-reducing approach is to lower the levels of pathological toxins much earlier in the disease or to prevent the formation of protein aggregates that are thought to cause the disease in order to halt neuronal loss (BOX 1). This idea is based on the premise that genetic and/or environmental factors may trigger pathological mechanisms very early in the disease process that lead to sequential and/or parallel primary and secondary damage. At the time of diagnosis, the accumulation of molecular and cellular disturbances in the brain may have already led to profound pathophysiological changes - which may or may not be dependent on the continued presence of pathological factors - that give rise to distinct components of the clinical syndrome<sup>16</sup> (FIG. 1a). A significant hurdle in this 'early toxin-reducing' approach is that changes in brain function in the asymptomatic stage of the disease often take years to occur and are small and highly variable<sup>17</sup>. The lack of sensitive measures and qualified biomarkers of such changes makes it extremely challenging to identify patient cohorts and to demonstrate clinical efficacy (FIG. 1b). Therefore, a clinical trial that can truly measure the efficacy of a treatment that prevents

#### Box 1 | Prevention trials in Alzheimer's disease

In an attempt to ultimately test the therapeutic value of treatment strategies based on the amyloid hypothesis, three new trials are underway to investigate the effectiveness of anti-amyloid therapies in patients before they exhibit clinical symptoms<sup>199</sup>. The Alzheimer's Prevention Initiative (API) and the Dominantly Inherited Alzheimer Network (DIAN) trials focus on cohorts carrying mutations associated with early-onset Alzheimer's disease, whereas the Anti-Amyloid Treatment of Asymptomatic Alzheimer's (A4) trial will enrol healthy subjects who display positive amyloid brain scans. These studies will offer, for the first time, the opportunity to test amyloid-based therapies in asymptomatic subjects. Substantial hurdles and limitations continue to exist, although there is optimism for such early prevention or intervention trials. For example, the lack of qualified biomarkers for early diagnosis makes it challenging to select cohorts of asymptomatic individuals for trials. Also, it is unclear how early a 'prevention' therapy has to begin to halt neuron loss. For instance, in subjects who carry an autosomal dominant mutation that increases the risk of Alzheimer's disease, deposition of amyloid- $\beta$  in the brain or changes in the levels of amyloid- $\beta$  in cerebrospinal fluid could happen as early as 15–25 years before any clinical symptoms become evident<sup>17,200,201</sup>. Nevertheless, results from these preventive trials will help to determine the direction of future Alzheimer's disease drug research and development.

toxin generation and accumulation will undoubtedly be long and require large patient cohorts, which are both expensive and difficult to manage.

#### Synaptic deficits: a pathophysiological hallmark

Given the current lack of success in the development of drugs that target the pathogenesis of a neurodegenerative disease, one could consider a different approach: to identify drugs that target the pathophysiology (FIG. 1a). This idea is based on the proposal by Lewis and Sweet regarding therapeutic approaches for schizophrenia<sup>16</sup>. These authors suggested that although targeting pathogenesis may be a suitable approach for prophylactic treatment, targeting the pathophysiology may be a better approach for therapeutic intervention in a disease that is already present. In the case of neurodegenerative diseases, targeting the pathophysiology would involve identifying and targeting the pathophysiological mechanisms that underlie the clinical syndromes.

Synapse degeneration is a major pathophysiological hallmark in neurodegenerative diseases. For example, substantial evidence indicates that in AD, there is a decrease in the number of synapses, which occurs later than AB accumulation and correlates with disease progression<sup>18–20</sup>. Several mechanistic mouse models<sup>21</sup> of AD show age-dependent deficits in hippocampal long-term potentiation (LTP)<sup>22,23</sup>, which correlate with the impairment in hippocampus-dependent memory<sup>22</sup>. In vivo two-photon imaging in the triple transgenic 3xTg-AD mice<sup>21</sup> expressing yellow fluorescent protein revealed that the progressive loss of dendritic spines in the somatosensory cortex coincided with the accumulation of soluble A $\beta$  and hyperphosphorylated tau (at 13 months of age), whereas a reduction in dendritic spine density in the hippocampus became evident only when amyloid plaques and hyperphosphorylated tau were abundant (at 15 months of age)<sup>24</sup>. Light and electron microscopic assessment of synaptic density in another transgenic model, Tg2576 (based on the human amyloid precursor protein (APP) 695 isoform with K670N and M671L mutations<sup>25</sup>), revealed region- and age-dependent synapse loss<sup>26</sup>. Furthermore, application of synthetically prepared Aß oligomers to hippocampal slices induces LTP deficits<sup>27</sup> and spine loss<sup>28</sup>, and synaptic deficits often occur in the proximity of A $\beta$  plaques<sup>26</sup>. Together, these data provide a link between the pathogenic trigger (Aß accumulation) and pathophysiological manifestation (synapse loss) of the disease.

Electron microscopic analysis of autopsied brain tissue from patients with mild to moderate AD<sup>29-31</sup> within 2–4 years after clinical onset demonstrated progressive synapse loss in the hippocampus, the frontal and inferior parietal cortex and the entorhinal cortex<sup>32,33</sup>. In the hippocampal CA1 region, progressive synapse loss has been reported (18% in subjects with mild cognitive impairment (MCI) to 55% in subjects with mild AD)<sup>33</sup>. Consistent with synapse loss, multiple studies using 2-deoxyglucose or fluorodeoxyglucose (FDG) positron emission tomography (PET) have reported an antecedent decline in cerebral glucose use decades before the diagnosis of AD<sup>34</sup>. Unbiased stereological counting of

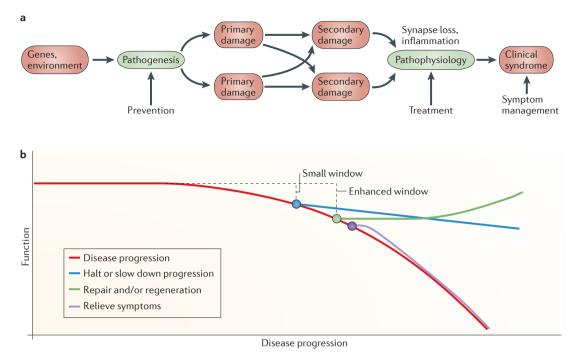


Figure 1 | Disease progression and its underlying pathogenic and pathophysiological processes. Advantages of focusing on pathophysiology rather than pathogenesis for the treatment of neurodegenerative diseases. a Pathogenesis is the primary event (that is, it is the cause of or a contributor to the disease initiation process) triggered by genetic and/or environmental factors. Preventive therapies are expected to be efficacious if they are aimed at controlling pathogenesis (for example, active or passive amyloid-targeting therapies for Alzheimer's disease). The pathogenic process leads to a cascade of primary and secondary damage over a long period of time, ultimately resulting in pathophysiological changes that are manifested as the clinical syndrome. Targeting these pathophysiological changes may be an effective strategy for the treatment of neurodegenerative diseases; here, the changes to be targeted include synapse loss and inflammation. **b** | Graphical representation of disease progression (specifically, the functional deficits associated with disease progression). Different modes of intervention may alter disease progression: disease-modifying treatments can prevent, slow down or halt disease progression if they target either the causal pathogenic mechanisms early (blue line, with small window to demonstrate efficacy) or the driver pathophysiological mechanisms (green line, with larger window to show efficacy). Treatments that only relieve clinical symptoms do not halt or slow disease progression (purple line). It is important to note that early intervention is critical for targeting pathogenesis, whereas repair or regeneration therapies could start relatively later. Part a is modified, with permission, from REF. 16 © (2009) American Society for Clinical Investigation.

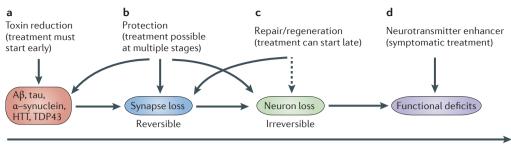
synapses also revealed that synapse degeneration was evident at the MCI stage and was strongly correlated with deficits in episodic memory (that is, delayed recall)<sup>32</sup>. In Huntington's disease, synapse degeneration occurs shortly after the build-up of the aggregated mutant huntingtin protein in the striatum but before neuronal loss, correlating with the progression of cognitive deficits<sup>35</sup>. Similarly, an impairment of synaptic plasticity (such as LTP and long-term depression (LTD)) in the striatum could account for the onset and the progression of motor and cognitive symptoms of PD<sup>36</sup>.

#### Synaptic repair therapy

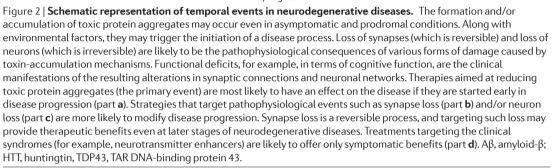
We suggest that this pathophysiological hallmark of neurodegenerative diseases — synapse degeneration should be considered as a target for disease-modifying treatments for these diseases. Specifically, such treatments could aim for neuronal repair, especially synaptic repair and regeneration. This proposal is based on several facts. First, synaptic dysfunction and synapse loss, unlike neuronal loss, are reversible (FIG. 2). Indeed,

synapses are highly dynamic and plastic<sup>37-39</sup>, such that dysfunctional synapses can be repaired and new synapses can be formed. This is important, as synaptic integrity and function are the foundation of neuronal circuits and are essential for maintaining physiological and behavioural functions<sup>40</sup>. Second, as synaptic dysfunction and loss are a point of convergence in most complex neurological diseases35,41-44, therapeutic intervention at the level of synaptic structure and function may be beneficial in multiple neurological diseases regardless of the type or origin of the toxic insult. Last, and most importantly, the window for therapeutic intervention based on synaptic repair and regeneration is longer than that for toxin-clearance approaches, and such interventions can thus be applied at a relatively late stage of the disease to slow or halt progression (FIG. 2) when early diagnosis remains a challenge.

Synapses are regulated at the level of synaptic transmission, synaptic plasticity and synaptic growth. Accordingly, therapeutics targeting synaptic dysfunction could involve three different approaches (BOX 2).



Disease progression



Conceptually, it is important to distinguish short-term processes that last from seconds to minutes, such as synaptic transmission and short-term plasticity, from long-term processes that last for hours, days or even longer, such as long-term plasticity and synaptogenesis<sup>45</sup>. Although therapies aimed at enhancing synaptic transmission may be useful for certain disease indications, for neurodegenerative diseases, where synapse loss and synaptic dysfunction are progressive, simple enhancement of synaptic transmission would probably not be able to halt the deterioration of neuronal circuits. Such an approach could at best provide symptomatic management. A disease-modifying agent should induce long-lasting functional changes (for example, LTP) and/or structural changes (for example, synaptogenesis, which involves increasing the number of synapses or enlargement of existing synapses) at synapses to slow, halt or reverse disease progression. Of note, substantial evidence suggests that long-term plasticity, such as LTP, leads to synaptic growth<sup>46-48</sup>.

#### BDNF: a potent synaptic repair molecule

Of all the molecules involved in synapse biology, BDNF (a member of the neurotrophin family), is by far the best studied and arguably the only one that has been associated with synaptic regulation in humans<sup>49–51</sup>. Substantial *in vitro* and *in vivo* evidence supports the pro-survival functions of BDNF on neurons under various pathological conditions (BOX 3).

BDNF is widely expressed in the CNS, and its expression is reduced in various pathological conditions<sup>52–56</sup>. The neurotrophic function of BDNF is primarily mediated by the TRKB receptor. In the adult brain, the main function of BDNF is to enhance synaptic transmission, facilitate synaptic plasticity and promote synaptic growth (FIG. 3). Given that these are exactly the three processes that could be targeted in synaptic repair therapy (BOX 2), the BDNF– TRKB pathway is a particularly suitable candidate to test the feasibility of the proposed 'synaptic repair' strategy. BDNF effects on synaptic transmission and plasticity. An early study showed that fast perfusion of BDNF onto adult rat hippocampal slices rapidly potentiated basal synaptic transmission at CA1 synapses<sup>57</sup>. Similarly, intrahippocampal infusion of a high dose of BDNF induced a lasting potentiation of basal synaptic transmission in the hippocampal dentate gyrus in vivo<sup>58</sup>. However, replication of these results remained a significant challenge59-63. Some studies showed that slow perfusion of BDNF on hippocampal slices from neonatal rats facilitated classic early-phase LTP (E-LTP) at hippocampal synapses without affecting basal synaptic transmission<sup>59,60,64,65</sup>. In parallel, in adult rat hippocampal slices, slow perfusion of BDNF converted short-term synaptic potentiation induced by a weak tetanic stimulation into LTP. This LTP-promoting effect of BDNF was further validated in studies using BDNF knockout mice66,67 and TRKB knockout mice68,69. A recent study not only resolved the longstanding debate on whether BDNF facilitates basal synaptic transmission but also provided new insights into the mechanism of BDNF signalling<sup>70</sup>. Specifically, acute BDNF application results in a rapid increase in BDNF levels and a transient activation of the TRKB receptor and its downstream signalling pathways, leading to a rapid enhancement in synaptic transmission. By contrast, slow perfusion of BDNF elicits a gradual increase in BDNF levels and a sustained activation of TRKB signalling, resulting in an increase in the magnitude of LTP70.

BDNF also plays a crucial part in late-phase LTP (L-LTP)<sup>71,72</sup>. First, L-LTP-inducing, strong theta-burst stimulation triggers not only the secretion<sup>73–75</sup> but also the synthesis of BDNF in hippocampal and cortical neurons<sup>76</sup>. Second, BDNF is required for L-LTP induction, as genetically<sup>66</sup> or pharmacologically<sup>77,78</sup> reducing the levels of BDNF blocks L-LTP. Third, an increase in BDNF levels is sufficient for the maintenance of L-LTP:

#### Early-phase LTP

(E-LTP). Early-phase long-term potentiation (LTP) is a sustained increase in synaptic efficacy that is induced by brief, high-frequency tetanic stimulation and lasts for 1–2 hours. It does not require gene transcription or protein synthesis.

#### Late-phase LTP

(L-LTP). Late-phase long-term potentiation (LTP) is a long-lasting increase in synaptic efficacy that is induced by strong, multiple tetanic stimuli and lasts for several hours or even days. It is both transcription- and translation-dependent and is often accompanied by morphological changes at the stimulated synapses.

#### Box 2 | Three aspects of synaptic modulation

There are three aspects to the modulation of synapse function. The first is enhancement of synaptic transmission. One way to do this is to increase neurotransmitter concentrations at the synaptic cleft by enhancing transmitter release or blocking transmitter degradation and/or reuptake. For example, donepezil, an acetylcholinesterase inhibitor that blocks the metabolism of acetylcholine, enhances transmission at the cholinergic synapses. It is a front-line drug for managing cognitive symptoms in patients with mild to moderate Alzheimer's disease<sup>15</sup>. Levodopa, a dopamine precursor that can be taken up by neurons and converted to dopamine and that is the first-line treatment for Parkinson's disease, is another example. An alternative approach is to activate and/or enhance postsynaptic function by activating postsynaptic receptors directly with an agonist or by modulating receptor signalling or function. Indeed, several dopamine receptor agonists are used to treat parkinsonian symptoms<sup>202</sup>. In both cases, the enhancement of synaptic function is transient, and synaptic deficits ensue after cessation of the treatment. Furthermore, these drugs cannot stop or slow down disease progression and are therefore categorized as 'symptomatic' rather than 'disease-modifying' treatments.

The second aspect to the modulation of synapse function involves facilitation of synaptic plasticity. Synaptic plasticity, by strict definition, is a change in synaptic strength in response to a brief increase in neuronal activity. It is different from simple enhancement of synaptic transmission induced by a chemical agent or drug as highlighted above. Numerous studies over the past two decades have demonstrated that synaptic plasticity mediates diverse brain functions, ranging from memory and emotion to fine motor control and executive function. Long-term potentiation (LTP), the most extensively studied form of synaptic plasticity, is thought to be the cellular mechanism underlying memory. LTP deficits are consistently reported in transgenic animal models of Alzheimer's disease<sup>23</sup>. Facilitation of LTP in animal models is often considered to be an important criterion in the selection of candidate drugs for neurodegenerative diseases. The key to synaptic plasticity is its activity-dependence. Thus, an attractive feature of plasticity-modulating agents is that they may preferentially alter synapses that are actively engaged in brain functions (for example, episodic memory) relevant to the disease.

The third aspect to the modulation of synapse function involves stimulation of synaptic growth (also termed synaptogenesis). Synaptic growth is a highly dynamic process that persists throughout adulthood. Unlike neuronal loss, which is irreversible, disease-associated loss of synaptic connectivity could be rescued through the growth of new terminals and/or dendritic spines. In addition, degenerating synapses could be 'stabilized' through strengthening of the existing pre- and postsynaptic structures and increasing the levels of synaptic proteins. A unique feature of the synaptogenesis-promoting agents, such as brain-derived neurotrophic factor, is that they may elicit long-lasting effects even after their withdrawal.

application of BDNF after E-LTP-inducing, weak theta-burst stimulation resulted in sustained L-LTP in hippocampal slices<sup>79</sup>. Similarly, in VP16-cyclic AMPresponsive element-binding protein (CREB) mice (in which BDNF levels are raised) the E-LTP-inducing, weak tetanus is able to induce L-LTP, which could be reversed by the BDNF scavenger TRKB-specific immunoglobulin G<sup>80</sup>. Furthermore, when all protein synthesis is blocked, application of BDNF after theta-burst stimulation is sufficient to maintain L-LTP79. Fourth, a critical step in L-LTP is the extracellular conversion of proBDNF to mature BDNF through a tissue plasminogen activator (tPA)-plasmin-dependent mechanism. The L-LTP impairment observed in mice lacking tPA or plasminogen could be completely rescued by mature BDNF perfusion but not by proBDNF perfusion<sup>79</sup>. Fifth, recent evidence suggests that TRKB at a particular synapse may act as a tag to capture BDNF<sup>81</sup>. This synaptic tagging mechanism is thought to ensure synapse-specific expression of L-LTP.

BDNF effects on synaptic growth. BDNF also promotes synapse formation<sup>82</sup> by regulating axonal branching<sup>83</sup>, dendritic growth<sup>84</sup> and activity-dependent synapse refinement<sup>85</sup>. Here, we focus on BDNF regulation of synaptic growth, which is defined as an increase in the number and/or the size of synapses. For example, treatment of postnatal hippocampal slice cultures with BDNF for 2-3 days increased the spine density in CA1 pyramidal neurons<sup>86</sup> and enhanced the expression of synaptic proteins<sup>87</sup>. In general, chronic exposure to BDNF increases spine motility<sup>88</sup>, which in turn increases the potential to form new synapses. Interestingly, fast delivery of BDNF enlarged the size of mushroom spines, whereas slow perfusion of BDNF induced more thin spines. This is suggestive of the consolidation of existing synapses and the formation of new synapses, respectively<sup>70</sup>.

Heterozygous BDNF knockout mice showed reduced hippocampal expression of synaptic proteins such as synaptobrevin and a reduction in the number of synaptic vesicles docked at the active zone<sup>89</sup>. Analysis of TRKB knockout mice revealed that, in addition to the changes in vesicle docking and synaptic protein expression, there was a substantial reduction in synaptic density (17–39% reduction) in mossy fibre terminals in the dentate gyrus<sup>90</sup>. Transgenic mice overexpressing BDNF had an increased number of synapses (63%) and increased synaptic vesicle docking in area CA1 (REF. 91). A study in which BDNF was re-expressed in neurons derived from BDNF knockout mice showed that BDNF increased the number of synapses within 16 hours<sup>92</sup>.

Although functional changes at synapses generally precede structural alterations, the two might be intricately linked through activity-dependent BDNF secretion<sup>46</sup>. Repetitive pairing of synaptic stimulation (through glutamate photo-uncaging) and postsynaptic spiking induced both LTP and a gradual enlargement of spine heads. Blockade of BDNF–TRKB signalling prevented the spine head enlargement, whereas synaptic stimulation plus the addition of exogenous BDNF induced spine enlargement in the absence of postsynaptic spikes<sup>46</sup>. These results suggest that activity-dependent BDNF secretion mediates both LTP and synaptic growth, but at different timescales.

BDNF effects on learning and memory. BDNF regulation of synaptic plasticity and synaptic growth suggests that it has a crucial role in cognitive functions. Indeed, a reduction of hippocampal BDNF levels through either genetic or pharmacological means not only impaired LTP and reduced the number of synapses but also caused deficits in the formation and consolidation of hippocampus-dependent memory<sup>93-95</sup>. Similar effects were observed when the level of TRKB or its activity was manipulated<sup>68,96,97</sup>. Furthermore, infusion of a BDNF antisense oligonucleotide into the hippocampus in rats several hours after learning impaired memory retention<sup>98</sup>. Conversely, overexpression of TRKB improved memory and occluded LTP99. Behavioural experiments have also shown a role for BDNF in episodic memory, fear memory extinction<sup>100</sup>, motor learning<sup>51</sup> and mood

#### Box 3 | Pro-survival effects of BDNF

In vitro, brain-derived neurotrophic factor (BDNF) prevents neuronal death induced by several different types of insults, including ischaemia due to oxygen-, glucose- or serum-deprivation<sup>203,204</sup>, oxidative stress (50 µM H<sub>2</sub>O<sub>2</sub>)<sup>205</sup>, glutamate toxicity<sup>206</sup> and toxic proteins such as amyloid- $\beta^{207}$ . The neuroprotective effects of BDNF have also been demonstrated in vivo in animal models of ischaemia and stroke<sup>208,209</sup>, for oxidative stress associated with Parkinson's disease<sup>210,211</sup>, for glutamate toxicity associated with seizures<sup>212</sup> and in an amyloid- $\beta$  overexpression animal model of Alzheimer's disease<sup>106,207</sup>. The readers are referred to an excellent review for detailed accounts of the neuroprotective role of BDNF in neurological and psychiatric diseases<sup>104</sup>. Perhaps surprisingly, conditional deletion of either Bdnf<sup>67</sup> or the neurotrophic receptor tyrosine kinase Trkb<sup>213</sup> gene in the adult mouse brain did not lead to obvious changes in neuronal number or brain morphology, suggesting that the pro-survival functions of BDNF are manifested primarily when neurons are under stress. Thus, BDNF may serve as a homeostatic regulator, eliciting neuroprotective functions only when neurons are damaged in disease conditions. The pro-survival effects of BDNF, together with its synapse-enhancing properties, make the BDNF-TRKB pathway an attractive therapeutic target for neurodegenerative diseases. Nerve growth factor (NGF), which is another neurotrophin that is expressed at low levels in the brain, has also been shown to promote the survival of cholinergic neurons in the brain. The therapeutic potential of NGF has been reviewed elsewhere<sup>214</sup>

> control<sup>101–103</sup>. Moreover, BDNF can protect synapses against various toxic insults in animal models of neurodegenerative diseases, such as AD, Huntington's disease, amyotrophic lateral sclerosis (ALS) and PD<sup>104</sup>. In a transgenic mouse model of AD (APP/PS1), inhibition of TRKB signalling exacerbated the spatial memory deficit, whereas overexpression of TRKB rescued spatial memory<sup>105</sup>. Remarkably, BDNF has been shown to protect and/or repair hippocampal neurons and synapses despite A $\beta$  build-up and neuronal toxicity in a mouse model of AD<sup>106</sup> and to rescue plasticity deficits induced by synthetic A $\beta$  oligomers in rat hippocampal slices *ex vivo*<sup>107</sup>, suggesting its potential as a therapeutic even in the presence of pathogenic factors.

#### The BDNF Val66Met polymorphism

The study of BDNF function has greatly benefited from the identification of the single-nucleotide polymorphism (SNP) in the gene encoding BDNF in humans that converts a valine to methionine at codon 66 (Val66Met)<sup>108</sup>. The *BDNF* Val66Met polymorphism does not alter the expression or processing of proBDNF or the structure of mature BDNF<sup>108</sup>. Rather, the BDNF<sup>Met</sup> protein results in impairment in the dendritic trafficking and synaptic localization of the protein and, most importantly, an 18–30% reduction in activity-dependent BDNF secretion<sup>108,109</sup>. The *BDNF* Val66Met polymorphism is associated with alterations in brain structure, network and function in healthy humans and has been implicated in several neurological and psychiatric disorders.

*Effects on hippocampal volume.* A reduction in BDNF secretion may affect dendritic and axonal growth, leading to changes in volume of certain brain areas. Structural MRI revealed a small but significant bilateral reduction (~10%) in the grey matter volume of the hippocampus, amygdala and neocortex in BDNF<sup>Met/Met</sup> carriers (that is, BDNF<sup>Val/Met</sup> individuals and BDNF<sup>Met/Met</sup> individuals) compared with BDNF<sup>Val/Val</sup> individuals<sup>110–112</sup>.

A longitudinal study revealed a twofold higher incidence of age-related reductions in hippocampal volume in healthy subjects carrying the *BDNF*<sup>Met</sup> allele compared with healthy *BDNF*<sup>Val/Val</sup> individuals<sup>113</sup>. However, a metaanalysis suggested that the effect of the *BDNF*<sup>Met</sup> allele on hippocampal volume may be overestimated owing to underpowered studies<sup>114</sup>.

A reduction in brain volume has been associated with (susceptibility to) brain illnesses. Indeed, hippocampal volume is consistently reduced in *BDNF*<sup>Met</sup> carriers compared with *BDNF*<sup>Val/Val</sup> patients with major depressive disorder, independently of age<sup>115,116</sup>. Perhaps surprisingly, in patients with multiple sclerosis, the *BDNF*<sup>Met</sup> genotype was associated with the preservation of grey matter volume and was inversely correlated with autoimmune-induced lesions<sup>117</sup>, suggesting that the *BDNF*<sup>Met</sup> genotype may be protective in certain diseases. Adult healthy *BDNF*<sup>Met</sup> carriers exhibit structural phenotypes similar to those seen in AD — a reduction in the thickness of temporal lobe structures, including the entorhinal cortex, and in white matter tracts that connect temporoparietal and temporofrontal areas<sup>118</sup>.

Effects on cognitive performance and neuronal network activity. Since the initial report<sup>108</sup>, numerous studies have reported changes in brain activation and function associated with the BDNF<sup>Val/Met</sup> polymorphism. However, the findings have not been consistent, probably owing to small sample sizes or population differences in factors such as ethnicity, age and gender. For example, functional MRI experiments have revealed reduced hippocampal activation during the encoding or retrieval of episodic memory in *BDNF*<sup>Met</sup> carriers compared with BDNF<sup>Val/Val</sup> subjects<sup>108,110</sup> even when performance levels were matched<sup>106</sup>. By contrast, when effects on successful memory-related activation were examined, BDNFMet carriers showed a greater engagement of the hippocampus and other medial temporal lobe areas during encoding and retrieval, potentially suggesting that there is neural inefficiency in memory-specific networks<sup>119</sup>. Excess hippocampal activation may contribute to memory impairment and may be associated with widespread degenerative processes in prodromal AD120,121. BDNFMet carriers exhibit impaired performance in various memory tasks (including episodic, visuospatial and working memory)<sup>108,122-125</sup>, which is consistent with the role of BDNF in LTP and hippocampus-dependent memories71. However, many studies have failed to observe robust and consistent effects of the BDNF<sup>Val/Met</sup> genotype on various cognitive functions<sup>126</sup>, probably because the tasks are relatively insensitive to a small reduction in BDNF secretion.

Brain stimulation studies using transcranial magnetic stimulation, transcranial direct current stimulation or paired associative stimulation have revealed impairments in cortical excitability or plasticity in *BDNF*<sup>Met</sup> carriers<sup>50,127</sup>, although other studies could not replicate these findings<sup>128–131</sup>. These inconsistencies may be due to methodological or inter-subject differences in response to brain stimulation. Moreover, *in vivo* electrophysiology, which is perhaps the best measure of synaptic and circuit function in the brain, was influenced by the *BDNF*<sup>Val/Met</sup> genotype.

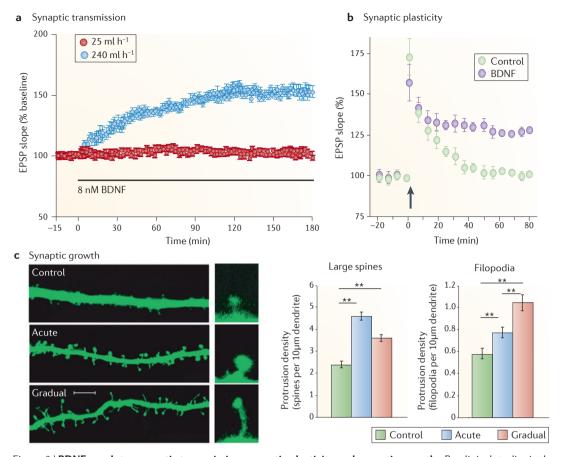


Figure 3 | BDNF regulates synaptic transmission, synaptic plasticity and synaptic growth. Preclinical studies in the past two decades have demonstrated the role of brain-derived neurotrophic factor (BDNF) in enhancing synaptic transmission<sup>57,70</sup>, modulating synaptic plasticity<sup>59</sup> and also in promoting synaptic growth<sup>220</sup>. a | Effects of BDNF on basal synaptic transmission. Acute application of recombinant BDNF (8 nM) to hippocampal slices from 8-week-old mice at a fast (blue, 240 ml h<sup>-1</sup>) but not slow (red, 25 ml h<sup>-1</sup>) perfusion rate rapidly enhances synaptic transmission at CA1–Schaffer collateral synapses, **b** | Effects of BDNF on hippocampal long-term potentiation (LTP). Hippocampal slices from 2-week-old rats were incubated in BDNF (2 nM) for 2.5-4 hours before electrophysiological recordings were made. Theta-burst stimulation (indicated by the arrow) applied to CA1–Schaffer-collateral synapses induced LTP in BDNF-treated slices but not in control slices. c | Effects of BDNF on synaptic growth. Rat hippocampal neurons transfected with green fluorescent protein in cultures examined 20 days after a 1-day treatment with BDNF (1 nM) showed increases in spine density (mushroom and filopodia) in general (left panel). Acute application of BDNF preferentially increases the number of large spines (middle panel), whereas a gradual increase of BDNF stimulates spine motility and preferentially increases the number of filopodia (right panel). The asterisks indicate statistically significant differences. EPSP, excitatory postsynaptic potential. Parts a and c are modified, with permission, from REF. 70 © (2010) Macmillan Publishers Ltd. All rights reserved. Part b is modified, with permission, from REF. 59 © (1996) Macmillan Publishers Ltd. All rights reserved.

Specifically, studies using resting electroencephalography (EEG) revealed a general increase in slow-wave activity (theta and delta power) but a decrease in fast-wave activity (alpha power) in  $BDNF^{Met}$  subjects compared with  $BDNF^{Val}$  subjects, suggesting that there is an increase in inhibitory and/or a decrease in excitatory synaptic activity in the cortex<sup>132</sup>. Studies using event-related potentials have also shown that  $BDNF^{Met}$  carriers exhibit impairments in synchronization processes that underlie error processing during an error-related negativity task<sup>133</sup> and in attention (that is, P300 latency increase and amplitude reduction)<sup>125</sup>.

Overall, the *BDNF* Val66Met polymorphism may serve as a useful tool to elucidate genotype-phenotype relationships in humans. Given the variability and inconsistency in past studies, future investigations need to focus on a subgroup of carefully phenotyped subjects (BOX 4).

The BDNF Val66Met polymorphism in neurodegenerative diseases. Attempts to establish a direct association of the *BDNF* Val66Met polymorphism with the risk of AD, age of disease onset or disease progression have been inconclusive. For instance, *BDNF*<sup>Met</sup> carriers may have an increased risk of early-onset AD, whereas *BDNF*<sup>Val</sup> carriers may have an increased risk of late-onset AD<sup>118</sup>. *BDNF*<sup>Val</sup> seems to delay the age of onset of AD and reduce the risk of AD in female apolipoprotein E  $\epsilon 4$  (*APOE*  $\epsilon 4$ ) carriers of Han Chinese origin<sup>134</sup> but not

#### Box 4 | Outstanding questions about the BDNF Val66Met polymorphism

The extensive characterization of the brain-derived neurotrophic factor (BDNF) Val66Met polymorphism offers a good foundation to address a number of key questions in future studies. First, the vast majority of studies published so far compared BDNF<sup>Val/Va</sup> carriers with BDNF<sup>Met</sup> carriers, owing to the rarity of BDNF<sup>Met/Met</sup> homozygotes in Caucasian individuals<sup>108,215</sup>. Gene dosage studies, by comparing all three genotypes (BDNF<sup>Val/Val</sup>, BDNF<sup>Val/Met</sup> and BDNF<sup>Met/Met</sup>), would reveal whether the phenotypic effects are truly associated with the BDNF polymorphism. The high occurrence of BDNF<sup>Met/Met</sup> in Asian populations<sup>134,216</sup> may offer an opportunity for such studies. Second, it is imperative to quantitatively measure the effect of the BDNF genotype on several endophenotypes: for example, hippocampal volume or cognition in the same subjects. Assuming that an alteration in BDNF trafficking or secretion correlates with changes in synaptic function, such a study will provide the most sensitive and reliable measure for synaptic changes to be used in clinical trials. Third, studying possible epistasis between the BDNF<sup>Val/Met</sup> genotype and other common polymorphisms may offer insights into disease risk and progression that may not be revealed by assessing the effect of one single-nucleotide polymorphism alone. This could be powerful when the effects of interactions with a disease-risk gene (for example, the gene encoding apolipoprotein E4) on endophenotypes relevant to a disease (for example, episodic memory or hippocampal volume in Alzheimer's disease) are examined. Fourth, most studies published so far have revealed negative effects of the BDNF<sup>Met</sup> allele on brain functions. Given that the BDNF Val66Met polymorphism has been selected during evolution<sup>215</sup>, it may have some beneficial effects. For example, the Met allele appears to be protective against grey matter damage in multiple sclerosis<sup>217</sup>, childhood-onset bipolar disorder<sup>152</sup> and obsessive compulsive disorder<sup>218,219</sup>. Last, differences in cognitive functions associated with the BDNF Val66Met polymorphism in adults may result from cumulative changes during decades of brain development or result from functional changes in synapses and neuronal circuitry. It is widely believed that developmental alterations of neuronal networks may be difficult to restore by pharmacological intervention; however, when the neuronal circuits formed during development remain unaltered, functional modulation of synapses can be used to restore network function. Pharmacological interventions that increase BDNF expression or secretion will not only help to distinguish these possibilities but also offer hope for therapies for diseases that result from synaptic dysfunction.

#### Gene dosage

A linear relationship between the number of genes (or alleles), the gene (or allele) product and the resulting effect (the phenotype).

#### Epistatic

The effect of one gene or gene product influencing the effect of other genes or gene products.

#### Endophenotypes

Intrinsic phenotypes that are relevant to a disease but not evident without a test. A good endophenotype must be tightly associated with the disease and display familial association even in non-diseased relatives with a higher odds ratio than in the general population.

#### Phase I

Phase I trials are typically conducted in healthy volunteers or in patients in a closely monitored clinic to evaluate safety, tolerability and pharmacokinetics of a new investigational drug. in other ethnic populations. In addition, *BDNF*<sup>Met/Met</sup> and *BDNF*<sup>Val/Met</sup> subjects with AD showed a higher risk for depression (with odds ratios threefold and twofold, respectively) compared with *BDNF*<sup>Val/Val</sup> subjects<sup>135</sup>.

Any effects of the polymorphism on cognitive performance and disease should become evident in longitudinal studies. Indeed, a recent study demonstrated a faster and more robust decline in episodic memory and hippocampal volume over a 36-month period in healthy elderly individuals with both the BDNFMet genotype and high amyloid load (but not in those with the BDNF<sup>Met</sup> genotype or a high A $\beta$  load alone)<sup>136</sup>. More importantly, patients with MCI (who have a high  $A\beta$  load) with the BDNF<sup>Met</sup> genotype also exhibit a faster and more robust decline in these two measures compared with BDNF<sup>Val/Val</sup> patients with MCI<sup>136</sup>. Thus, the BDNF<sup>Met</sup> genotype may accelerate the progression of AD. Consistent with this finding, inhibition of TRKB signalling exacerbated spatial memory impairment in APP/PS1 mice (a model of AD) but had no effect in wild-type mice<sup>105</sup>. If these findings in humans are validated in independent cohorts, they should help to develop a strategy for patient stratification so that clinical studies require fewer patients and can be of shorter duration. Future studies should also examine whether the faster decline in episodic memory and hippocampal volume in the BDNF<sup>Met</sup> MCI subpopulation can be reversed by pharmacological interventions.

Emerging findings also provide evidence for an association of the *BDNF* Val66Met polymorphism with PD: the *Gene*PD study<sup>137</sup> revealed an association of the polymorphism with the risk and age of onset of familial PD<sup>137-139</sup>, which was especially strong in patients with PD who had cognitive impairments<sup>140</sup>. Epistatic interactions between the *BDNF*<sup>Met</sup> allele and the most common variant of the gene encoding leucine-rich repeat kinase 2 (the G2385R variant) increases the risk of PD in Asian populations. The overall odds ratio increased from 3.2 to 4.0 and to over 6, if the age of PD onset was >60 years<sup>141</sup>. These emerging results continue to substantiate the role of BDNF in neurodegenerative diseases.

BDNF<sup>Val/Met</sup> knock-in mice as a tool. To study the BDNF Val66Met polymorphism in well-controlled conditions and to determine whether the endophenotypes seen in BDNF<sup>Met</sup> carriers could be rescued genetically or pharmacologically, a knock-in mouse line was generated. The mice have a point mutation in the endogenous mouse Bdnf, resulting in a Bdnf<sup>Met</sup> genotype<sup>109</sup> instead of the Bdnf<sup>Val</sup> genotype. Neurons derived from Bdnf<sup>Met/Met</sup> mice showed reduced activity-dependent BDNF secretion<sup>109</sup>. Bdnf<sup>Met/Met</sup> mice had normal total brain BDNF levels, a smaller hippocampus (associated with reduced dendritic complexity) and impairments in hippocampus-dependent contextual memory<sup>100</sup>. In addition, hippocampal slices derived from *Bdnf*<sup>Met/Met</sup> mice exhibited a deficit in NMDA receptor (NMDAR)-dependent LTP but not in basal glutamatergic transmission<sup>142</sup>. NMDAR-dependent LTD was also reduced, whereas metabotropic glutamate receptor-dependent LTD remained intact. These results suggest that activity-dependent BDNF secretion may be selectively involved in NMDA-dependent forms of synaptic plasticity. Moreover, Bdnf<sup>Met/Met</sup> mice showed anxiety-like behaviour that was resistant to treatment with antidepressants such as fluoxetine<sup>109</sup> and ketamine<sup>143</sup>. A different mouse knock-in line in which the mouse Bdnf coding region was replaced by the human BDNF<sup>Val</sup> or BDNF<sup>Met</sup> sequence also exhibited synaptic deficits<sup>144</sup>. Although BDNF<sup>Met</sup> knock-in mice capture some phenotypes associated with the human BDNF polymorphism, it is important to assess whether the deficits in BDNFMet knock-in mice can be rescued by pharmacological intervention in vivo. If so, BDNFMet knock-in mice could serve as a translational preclinical model not only to study synaptic dysfunction but also to use in translational drug discovery for CNS diseases.

#### **BDNF-based therapeutic strategy**

The preclinical and human studies summarized above point to BDNF as a pro-survival molecule (BOX 3) as well as a synaptic repair molecule for neurodegenerative diseases. However, results from clinical studies using BDNF as a therapeutic agent have not been encouraging. To date, five clinical trials using BDNF have been conducted (four in ALS and one in diabetic neuropathy<sup>145</sup>), and the results have been inconclusive. In a Phase I/II<sup>146</sup> openlabel trial for ALS, subcutaneously administered BDNF showed a delay in the percentage of forced vital capacity decline and an improvement in walking time, whereas

#### Forced vital capacity

The amount of air that can be forcibly exhaled from the lungs after a deep breath, which can be measured with a spirometer

#### Phase II

A trial conducted primarily to evaluate the effectiveness of a drug in people who have a certain disease or condition. Safety continues to be evaluated in the clinical setting. Initial Phase II efficacy studies are also referred to as proof-of-concept studies.

#### ALSFRS

Amyotrophic Lateral Sclerosis Functional Rating Scale (ALSFRS-R is the revised version) is a validated measure (scores 0–48) that aids the assessment of disability of the patients with motor neuron diseases based on a questionnaire that asks about daily activities and how much help the patients need along with disease-specific symptoms.

#### Nanoparticle

A microscopic particle with a diameter of less than 100 nm. Here, it refers to the liposomes or exosomes that carry drug substances into the brain.

#### Trojan horse

A strategy to deliver drugs to target sites that are normally inaccessible. The drug is fused to a molecule or encapsulated in a cell or nanoparticle that can readily cross the blood– brain barrier.

#### Nose-to-brain

Delivery or transport of drugs, cells or cargoes into the brain intranasally through the olfactory or trigeminal neuronal pathway. This delivery route limits systemic exposure and bypasses the blood-brain barrier. a Phase II/III trial did not replicate these benefits<sup>143</sup>. In another Phase I/II placebo-controlled, double-blind trial<sup>144</sup> and in an as yet unpublished Phase III trial<sup>145</sup> using intrathecal administration, BDNF showed no clinical benefits on survival or on the ALS functional rating scale (ALSFRS) score<sup>145,147</sup>. Given the beneficial effects of BDNF on neuronal functions observed in preclinical studies, how can we explain these clinical failures? One explanation could be that BDNF is cleared rapidly in vivo and does not easily penetrate into the spinal cord parenchyma. Although a dose-dependent increase in BDNF levels in cerebrospinal fluid (CSF) was reported after intrathecal delivery<sup>148</sup>, there was no clinical evidence that BDNF had reached the target site (that is, the ventral horn of the spinal cord). In fact, none of the trial design included measurements of 'target engagement' (that is, TRKB activation or downstream signalling) or of an immediate downstream pharmacodynamic response following BDNF administration. In short, these four trials in ALS did not unequivocally test the BDNF-TRKB mechanism. Thus, it may be premature to conclude that BDNF is ineffective as a therapy for ALS 145,149,150.

BDNF as a therapeutic molecule. The problem described above - namely, the lack of evidence that BDNF has been delivered to the right tissues or activated TRKB is not unique to BDNF-based trials. A recent analysis by Pfizer indicates that 43% of Pfizer programmes that were terminated because of a negative outcome in Phase II (testing proof-of-concept) had not adequately tested the drug's mechanism of action<sup>151</sup>. In other words, in these trials, it was unclear whether the pharmacological agents under investigation had reached the target tissue in the disease at therapeutic concentrations, or whether they had engaged the proposed target. A 'three pillars of survival'151 theory that is based on this analysis proposes that three specific conditions (the 'pillars') should be met to increase the likelihood that a candidate drug 'survives' a Phase II trial. The three pillars are: first, that the drug is delivered to the target site over a desired period of time; second, that the drug binds to the target under investigation at the required level; and third, that drug-induced modulation of the target results in a pharmacodynamic effect.

Two additional problems have hampered the translation of the beneficial effects of stimulating BDNF–TRKB signalling from bench to bedside: the first is the inability to deliver BDNF across the blood–brain barrier (BBB); and the second is the poor bioavailability of BDNF owing to its physiochemical properties.

Regarding the first problem, several approaches are actively being pursued to deliver BDNF into the CNS. BDNF can be delivered to the required site of action through invasive procedures (by using a catheter or implantable pumps<sup>152</sup> or through implantation of biodegradable polyethylene glycol-based hydrogel device containing poly(lactic-co-glycolic acid) (PLGA) microparticles encapsulated with BDNF<sup>153,154</sup>). Although these approaches may be suitable for the short-term treatment of localized acute CNS injuries, they are of limited benefit in chronic neurological diseases because patients would have to undergo these invasive procedures repeatedly, which can lead to secondary complications.

Non-invasive approaches such as nanoparticle-, Trojan horse- and nose-to-brain-mediated delivery of BDNF into the brain are also being explored. Trojan horse technology involves conjugating BDNF to molecules that can readily cross the BBB. Emerging evidence suggests that preferential uptake of BDNF into the CNS can be achieved by conjugating BDNF to ligands that bind to certain receptors in endothelial cells that facilitate transcytosis or to antibodies directed against these receptors. For instance, BDNF conjugated to a monoclonal antibody against the human insulin receptor exhibited a ~tenfold increase in brain concentrations of BDNF and a 100-fold increase in the mean residence time of BDNF in the circulation without altering the blood glucose level<sup>155</sup>. Similar results have been obtained by conjugating BDNF to an antibody targeting the transferrin receptor<sup>156</sup>. A recent study revealed the significance of the antibody binding kinetics in Trojan horse-mediated CNS delivery<sup>157</sup>. Other Trojan horse carriers or ligands include low-density lipoprotein receptor-related protein 1 (LRP1)<sup>158</sup>, diphtheria toxin receptor<sup>159</sup> or single-chain domain antibodies isolated from llama<sup>160</sup>. Current Trojan horse approaches lack tissue specificity and suffer from potential interference with the endogenous function of the transcytosis receptors. (For comprehensive reviews on the delivery of proteins, including BDNF, into the CNS, see REFS 161,162).

The nose-to-brain (or intranasal) route is an alternative way to deliver macromolecules into the brain parenchyma. The advantages of the nose-to-brain route include simple and non-invasive administration, rapid delivery to the brain, minimal systemic exposure to the drug and the option for repeated dosing. Nose-to-brain delivery of BDNF not only resulted in an increase in brain parenchymal concentrations within ~30 minutes but also induced activation of the TRKB receptor and its downstream phosphoinositide 3-kinase-AKT pathway163. In a rat middle cerebral artery occlusion model, nose-to-brainadministered BDNF 2 hours after the ischaemic insult was neuroprotective<sup>164</sup>. Further characterizations will be required to overcome the challenges associated with regulating the delivery of the intended dose to achieve region-selective delivery.

The second hurdle is the poor bioavailability and stability of BDNF (it has a half-life of few minutes in rat plasma<sup>165</sup> and a few hours in sheep CSF<sup>152</sup>). BDNF in the circulation is primarily cleared by the liver owing to its basic isoelectric pH. PEGylation of BDNF has been shown to enhance its stability (>60%) without affecting its bioactivity or mean time in the circulation<sup>166</sup>. Recombinant engineering methods should be considered to generate a mutant form of BDNF with higher stability, neutral isoelectric pH and enhanced brain penetration.

Development of drugs targeting the BDNF-TRKB pathway. In addition to BDNF itself as potential medicine, several strategies could be used to manipulate BDNF-TRKB signalling, including small-molecule

lable 1   Summary of approaches for targeting the BDNF-TRKB pathway			
Approach	Advantages	Challenges	Examples
Small-molecule neurotrophic receptor tyrosine kinase TRKB agonist, modulator and/or transactivator	<ul> <li>Systemic administration and delivery across the blood–brain barrier for CNS actions</li> <li>TRK selectivity (not targeting the p75NTR pathway)</li> <li>Transactivation through other tyrosine kinase receptors, G protein-coupled receptors and metal ions</li> </ul>	<ul> <li>Identifying small molecules that can induce TRKB dimerization</li> <li>Finding a true TRKB agonist and/or modulator that directly binds and activates TRKB</li> <li>Off-target effects</li> </ul>	<ul> <li>7,8 -dihydroxyflavone<sup>167</sup></li> <li>LM-22 series<sup>168</sup></li> <li>De-oxygedunin<sup>174</sup></li> <li>Antidepressants (fluoxetine)<sup>184</sup></li> <li>N-acetylserotonin<sup>221</sup></li> <li>Adenosine 2A receptor agonist<sup>222,223</sup></li> <li>Apomorphine<sup>224</sup></li> <li>Zinc<sup>225</sup></li> </ul>
Biopharmaceutical TRKB agonist and/or modulator	<ul> <li>Specificity, fewer off-target effects</li> <li>Better pharmacokinetics and bioavailability (compared with brain-derived neurotrophic factor (BDNF))</li> <li>Receptor selectivity (not targeting TRKA, TRKC or the pan-neurotrophin receptor p75NTR)</li> <li>Low-affinity may limit hyperactivation and desensitization of TRKB</li> </ul>	Delivery into CNS	<ul> <li>Peptidomimetics<sup>171-173,226</sup></li> <li>Agonist antibodies<sup>169,176,177,227</sup></li> <li>RNA aptamer<sup>228</sup></li> </ul>
Small molecules that enhance the transcription, translation or secretion of endogenous BDNF	<ul> <li>Limits pharmacology to physiologically relevant BDNF-expressing cells</li> <li>Regulation at subcellular location (dendritic and/or synaptic synthesis and release)</li> <li>Modulates endogenous mechanisms, limiting clinical adverse events</li> </ul>	<ul> <li>Achieving promoter-specific activation of transcription</li> <li>Increasing translation or inhibiting degradation of BDNF without affecting other molecules</li> <li>Affecting secretion of growth factors, chemokines or cytokines</li> <li>Generating a sufficient increase of extracellular BDNF to induce prolonged TRKB activation with a full spectrum of downstream signalling necessary for synaptic growth</li> </ul>	<ul> <li>Antidepressants<sup>181</sup></li> <li>Memantine<sup>188</sup></li> <li>Ampakines<sup>229-232</sup></li> <li>Rolipram<sup>233</sup></li> <li>Fingolimod<sup>234</sup></li> <li>CEP-1347 (REF. 235)</li> <li>Exercise<sup>180,236</sup></li> <li>Acetylcholinesterase inhibitors (donepezil, galantamine and huperzine A)<sup>237,238</sup></li> <li>Cystamine or cysteamine<sup>239</sup></li> </ul>

#### Table 1 | Summary of approaches for targeting the BDNF-TRKB pathway

TRKB agonists or modulators<sup>167,168</sup>, TRKB agonistic antibodies<sup>169,170</sup> or peptidomimetics<sup>171-173</sup>, or small molecules that stimulate endogenous BDNF expression or BDNF-TRKB signalling (see TABLE 1 for a summary). The discovery of a specific and selective small-molecule TRKB agonist or activator has remained a significant challenge, primarily because of a lack of distinct binding pockets for small molecules in the extracellular domain of TRKB, and the requirement of a dimeric ligand to form extensive protein-protein interactions to activate TRKB. Several recent studies<sup>167,168,174</sup> reported the identification of small-molecule agonists or modulators of TRKB based on the crystal structure of BDNF and even demonstrated binding of such molecules to the extracellular domain of TRKB using biochemical or biophysical methods. However, in the absence of the co-crystal structure of the proclaimed TRKB-bound agonists or activators and the difficulties in reproducing the results175 one must be cautious about designating them as 'true' agonists or activators. Nevertheless, the in vivo effects of these molecules could be true due to 'indirect' activation of TRKB (for example, through transactivation) or due to 'off-target' effects.

Agonist antibodies and peptidomimetics have been shown to activate the TRKB receptor in a dose-dependent manner both *in vitro* and *in vivo*, although their binding affinities, potency and magnitude of TRKB activation are not comparable with those of BDNF<sup>169,176,177</sup>. The agonistic antibodies are both selective and specific to TRKB: that is, they do not bind or activate TRKA (also known as NTRK1) or TRKC (also known as NTRK3) receptors, nor the pan-neurotrophin receptor p75NTR (also known as NGFR). Obtaining the ligand–TRKB co-crystallographic structures would provide further evidence for such non-BDNF-mediated pharmacological activation of the TRKB receptor and may even uncover novel ways to activate the receptor. However, the difficulty in CNS delivery limits the use of TRKB agonistic antibodies or peptidomimetics as a therapy for neurodegenerative diseases.

Physical exercise has been shown to enhance endogenous levels of BDNF<sup>178</sup>. However, a systematic analysis of the effect of exercise on circulating BDNF levels in healthy subjects revealed that the BDNF increase (~10% above the baseline plasma BDNF levels) was mostly transient, with levels returning to baseline within 10–60 minutes<sup>179</sup>. Consistent with this, BDNF protein and mRNA levels increase in the hippocampus of young and middle-aged rats in a transient manner after voluntary exercise<sup>180</sup>.

An even more promising strategy is to pharmacologically enhance the expression of endogenous BDNF in the brain using a small molecule approach. Indeed, chronic but not acute administration of antidepressant drugs has been shown to enhance hippocampal *BDNF* expression<sup>181</sup>. However, it seems that the magnitude and duration of the increase in BDNF levels are not sufficient to ensure TRKB activation<sup>182</sup>. Acute and chronic antidepressant treatment can also increase TRKB signalling. This effect is transient, independent of BDNF and incomplete (that is, tyrosine phosphorylation occurs at the autophosphorylation site and phospholipase Cy site but not the SHC binding site of TRKB), possibly through a transactivation mechanism<sup>183,184</sup>. Thus, it is unclear whether the classic antidepressants could have a meaningful impact on the BDNF-TRKB pathway to promote synaptic growth. The NMDAR channel blocker ketamine has been shown to produce rapid (within hours) antidepressant actions in treatment-resistant patients185. Intriguingly, ketamine administration in animals induced a rapid but transient increase in cortical BDNF expression<sup>186</sup> and an increase in synaptic protein expression by activating the mammalian target of rapamycin pathway, and these effects were associated with an increase in dendritic spines and synaptic transmission<sup>187</sup>. Future studies should determine whether the synaptogenic effect of ketamine is truly mediated by ketamineinduced BDNF expression.

Memantine, a drug commonly used to treat patients with moderate to severe AD, has also been shown to increase BDNF expression in the limbic cortex in preclinical models<sup>188</sup>. However, the increase in BDNF expression was only observed at a dose of 50 mg per kg, which is predicted to be toxic in humans, and the effect was marginal (~25%) at a non-toxic dose<sup>188</sup>. Donepezil and galantamine, the acetylcholinesterase inhibitors commonly used to treat cognitive deficits in MCI and early AD, have also been shown to increase serum BDNF levels<sup>189</sup>, although it remains unclear whether they increase BDNF in the human brain. These molecules exhibit properties similar to that of memantine, activating the AKT pathway but not the mitogen-activated protein kinase (MAPK) pathway<sup>190</sup>. Given that activation of the MAPK pathway is essential for BDNF-mediated regulation of spine growth<sup>191</sup>, these drugs may not promote synaptic growth.

Thus, several factors must be considered when using pharmacological agents to stimulate endogenous BDNF expression. First, the molecule must induce BDNF expression at non-toxic doses and induce sufficiently high concentrations of extracellular BDNF to activate TRKB. Second, the molecule should induce sustained levels of TRKB activation with the full spectrum of downstream signalling that is necessary for synaptic growth. Third, the molecule should have synaptogenic effects: namely, facilitating L-LTP, promoting dendritic spine growth and/or enhancing synaptic protein expression. Any existing drugs with these properties could be considered for synaptic repair therapy.

General challenges for BDNF-based synaptic repair therapies. Patient heterogeneity makes it difficult to perform a well-controlled clinical study with a small number of patients, especially in the more common neurodegenerative diseases such as AD and PD. A proper investigation of a targeted mechanism — and hence the drug efficacy — is best performed in a clinically homogeneous patient cohort. For instance, patients with AD who have the BDNF<sup>Met</sup> genotype have more severe endophenotypes (in terms of, for example, episodic memory and hippocampal volume) or faster disease progression<sup>136</sup>. Such patients could therefore be selected for clinical trials to enhance the sensitivity of detecting drug efficacy or to shorten the length of the trial. Recent studies have shown an epistatic interaction between the BDNF<sup>Met</sup> and APOE4 alleles on disease progression in preclinical AD<sup>192,193</sup> as well as between the BDNF<sup>Met</sup> allele and high Aβ amyloid levels in prodromal AD<sup>136</sup>. Similar genetic interaction studies may identify new traits that could predict disease risk, age at onset and/or progression of neurodegenerative diseases, and therefore be used for patient stratification.

In addition to the three pillars of survival highlighted above, we have proposed a fourth pillar: to demonstrate the efficacy of drugs early in clinical studies using sensitive and reliable biomarkers<sup>194</sup>. In the case of synaptic repair therapy, this would involve measuring synaptic dysfunction and repair or regeneration in vivo, both preclinically and clinically. Unfortunately, the commonly used preclinical measures of synaptic plasticity, such as hippocampal LTP and performance in the Morris water maze test, are not translatable in humans. Conversely, methods that indirectly measure in vivo synaptic function, such as functional MRI and FDG-PET, are technically challenging in animals, and their spatial and temporal resolution is too low in humans. Although cognitive measures such as episodic memory are relevant to synaptic function, they are subjective, highly variable and too insensitive to monitor drug efficacy. Finally, for a drug to achieve disease modification, it would have to demonstrate long-term effects on synaptic plasticity and synaptogenesis that persist after drug withdrawal rather than a transient effect such as an increase in synaptic transmission.

Non-invasive in vivo electrophysiological methods may be able to measure synaptic function in both animals and humans. Recent studies have shown that genetic perturbations of the BDNF-TRKB pathway that are known to alter hippocampal synaptic networks decrease gamma oscillatory activity in hippocampal slices, reflecting desynchronization of neuronal activity within the hippocampal synaptic circuits<sup>195,196</sup>. Thus, BDNF-induced changes in synaptic connectivity in the cortex could be recorded using surface EEG, which probes the spatial and temporal summation of synchronous current flow through postsynaptic dendrites of cortical pyramidal neurons. Indeed, human BDNFMet carriers exhibit a slower EEG profile132 and abnormal event-related potential activity and/or synchrony in cognitive tasks125. Interestingly, some of these EEG and event-related potential changes are associated with hippocampal and frontal activation<sup>125</sup>, and are correlated with MCI-to-AD conversion and AD progression<sup>197,198</sup>. To ascertain the translational value of using EEG to measure synaptic changes, future studies should assess the EEG phenotypes of *BDNF*<sup>Met</sup> knock-in mice.

#### **Conclusions and future directions**

It is generally thought that a toxin-reducing approach may have beneficial outcomes in neurodegenerative diseases if it is started early, but the lack of sensitive biomarkers for disease progression and drug efficacy makes such early intervention studies extremely challenging. Increasing evidence suggests that in the case of progressive neurodegenerative disorders such as AD and PD, it might be more effective to treat the pathophysiology that directly underlies the clinical syndromes than to target the pathogenesis. Synaptic dysfunction seems to be a key pathophysiological feature for all neurodegenerative disorders. Given that synapse loss is reversible and predictive of disease progression, targeting mechanisms that stabilize and protect, or repair and regenerate synapses would enable clinical intervention at both early and late stages of the disease. Although the success of such synaptic repair approaches can ultimately only be measured by assessing their clinical efficacy, confidence in the potential of synaptic repair therapy will be strengthened if synaptic dysfunction and repair and/or regeneration can be measured reliably in the clinic.

Our understanding of BDNF–TRKB biology and the role of BDNF in synaptic plasticity and synaptic growth should now be translated into disease-modifying therapies for neurodegenerative disorders. BDNF is by far the best known synaptogenic molecule and perhaps the only one that has been associated with synaptic regulation in humans. Unlike most of the existing drugs that target synaptic transmission or plasticity<sup>59</sup>, BDNF also promotes synaptic growth, and this can form a basis for a disease-modifying therapy. Animal experiments have established the role of BDNF in cognitive functions. More importantly, BDNF is neuroprotective and can repair synaptic deficits, despite the build-up of toxic proteins, in animal models of neurodegeneration<sup>106</sup>. The discovery of the BDNF Val66Met polymorphism, which influences synaptic localization and activity-dependent secretion of BDNF, provides an unprecedented opportunity to assess how changes in synaptic function in humans influence endophenotypes that are relevant to neurodegeneration. A systematic comparison of imaging, electrophysiological and behavioural findings in the three BDNF<sup>Val/Met</sup> genotypes should be able to identify suitable (that is, reliable and sensitive) measures of BDNF-induced synaptogenic effects in carefully phenotyped or at-risk human subjects. A better understanding of epistatic interactions between the BDNFVal66Met polymorphism and 'disease genes' in the endophenotypes of PD or AD may aid the development of strategies for patient stratification. Taken together, a combination of activating the BDNF pathway and a more reliable and sensitive method to measure the resulting synaptic changes could pave the way for the development of disease-modifying therapies.

- Salomon, J. A. *et al.* Healthy life expectancy for 187 countries, 1990–2010: a systematic analysis for the Global Burden Disease Study 2010. *Lancet* 380, 2144–2162 (2013).
- Brookmeyer, R., Johnson, E., Ziegler-Graham, K. & Arrighi, H. M. Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement.* 3, 186–191 (2007).
- Dorsey, E. R. *et al.* Projected number of people with Parkinson disease in the most populous nations, 2005 through 2030. *Neurology* 68, 384–386 (2007).
- Alzheimer, A., Stelzmann, R. A., Schnitzlein, H. N. & Murtagh, F. R. An English translation of Alzheimer's 1907 paper, "Über eine eigenartige Erkankung der Hirnrinde". *Clin. Anat.* 8, 429–431 (1995).
- Gilman, S. *et al.* Clinical effects of Aβ immunization (AN1792) in patients with AD in an interrupted trial. *Neurology* 64, 1553–1562 (2005).
- Mullard, A. Sting of Alzheimer's failures offset by upcoming prevention trials. *Nature Rev. Drug Discov.* 11, 657–660 (2012).
- Herrmann, N., Chau, S. A., Kircanski, I. & Lanctot, K. L. Current and emerging drug treatment options for Alzheimer's disease: a systematic review. *Drugs* **71**, 2031–2065 (2011).
- Drugs **71**, 2031–2065 (2011).
   Querfurth, H. W. & LaFerla, F. M. Alzheimer's disease. N. Engl. J. Med. **362**, 329–344 (2010).
- Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P. & Kivipelto, M. Alzheimer's disease: clinical trials and drug development. *Lancet Neurol.* 9, 702–716 (2010).
- Perez, S. E. *et al.* Dimebon alters hippocampal amyloid pathology in 3xTg-AD mice. *Int. J. Physiol. Pathophysiol. Pharmacol.* 4, 115–127 (2012).
- Zago, W. *et al.* Neutralization of soluble, synaptotoxic amyloid β species by antibodies is epitope specific. *J. Neurosci.* **32**, 2696–2702 (2012).
   Romberg, C., Mattson, M. P., Mughal, M. R.,
- Romberg, C., Mattson, M. P., Mughal, M. R., Bussey, T. J. & Saksida, L. M. Impaired attention in the 3xTgAD mouse model of Alzheimer's disease: rescue by donepezil (Aricept). *J. Neurosci.* **31**, 3500–3507 (2011).
- Lemere, C. A. & Masliah, E. Can Alzheimer disease be prevented by amyloid-β immunotherapy? *Nature Rev. Neurol.* 6, 108–119 (2010).
- 14. Hellweg, R., Wirth, Y., Janetzky, W. & Hartmann, S. Efficacy of memantine in delaying clinical worsening in

Alzheimer's disease (AD): responder analyses of nine clinical trials with patients with moderate to severe AD. *Int. J. Geriatr. Psychiatry* **27**, 651–656 (2011).

- Rodda, J., Morgan, S. & Walker, Z. Are cholinesterase inhibitors effective in the management of the behavioral and psychological symptoms of dementia in Alzheimer's disease? A systematic review of randomized, placebo-controlled trials of donepezil, rivastigmine and galantamine. *Int. Psychogeriatr.* 21, 813–824 (2009).
- Lewis, D. A. & Sweet, R. A. Schizophrenia from a neural circuitry perspective: advancing toward rational pharmacological therapies. *J. Clin. Invest.* **119**, 706–716 (2009).

In this review, the authors emphasize the importance of separating pathogenesis from pathophysiology and suggest that therapeutic interventions for complex diseases such as schizophrenia might be more effective by focusing on pathophysiology, which is closer to clinical features.

- Villemagne, V. L. *et al.* Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *Lancet Neurol.* **12**, 357–367 (2013).
   A study that provides an estimate of changes in rates of Aβ deposition, cerebral atrophy and cognitive decline in healthy subjects, patients with MCI and patients with AD. Projections based on this longitudinal measurement suggest a protracted (~ 17 years) preclinical phase of AD.
- Terry, R. D. *et al.* Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. *Ann. Neurol.* **30**, 572–580 (1991).
- Shankar, G. M. & Walsh, D. M. Alzheimer's disease: synaptic dysfunction and Aβ. *Mol. Neurodegener.* 4, 48 (2009).
- 20. Selkoe, D. J. Alzheimer's disease is a synaptic failure. *Science* **298**, 789–791 (2002).
- Oddo, S. *et al.* Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Aβ and synaptic dysfunction. *Neuron* 39, 409–421 (2003).
   A transgenic mouse overexpressing APP<sub>Swe</sub>, PS1<sub>M146V</sub> and tau<sub>P501L</sub> shows age-dependent relationship between Aβ, synaptic dysfunction and tangle formation. This model recapitulates the

'amyloid cascade' hypothesis in which A $\beta$ accumulation precedes tau pathology. Impairments in synaptic transmission and LTP occur as a consequence of A $\beta$  but prior to tau pathology.

- Chapman, P. F. *et al.* Impaired synaptic plasticity and learning in aged amyloid precursor protein transgenic mice. *Nature Neurosci.* 2, 271–276 (1999).
- Marchetti, C. & Marie, H. Hippocampal synaptic plasticity in Alzheimer's disease: what have we learned so far from transgenic models? *Rev. Neurosci.* 22, 373–402 (2011).
- Bittner, T. *et al.* Multiple events lead to dendritic spine loss in triple transgenic Alzheimer's disease mice. *PLoS ONE* 5, e15477 (2010).
- Hsiao, K. *et al.* Correlative memory deficits, Aβ elevation, and amyloid plaques in transgenic mice. *Science* 274, 99–102 (1996).
- Dong, H., Martin, M. V., Chambers, S. & Csernansky, J. G. Spatial relationship between synapse loss and β-amyloid deposition in Tg2576 mice. J. Comp. Neurol. 500, 311–321 (2007).
- Shankar, G. M. *et al.* Amyloid-β protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nature Med.* 14, 837–842 (2008).
- Shankar, G. M. *et al.* Natural oligomers of the Alzheimer amyloid-β protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. *J. Neurosci.* 27, 2866–2875 (2007).
- Davies, C. A., Mann, D. M., Sumpter, P. Q. & Yates, P. O. A quantitative morphometric analysis of the neuronal and synaptic content of the frontal and temporal cortex in patients with Alzheimer's disease. *J. Neurol. Sci.* 78, 151–164 (1987).
- 30. Scheff, S. W., DeKosky, S. T. & Price, D. A. Quantitative assessment of cortical synaptic density in Alzheimer's disease. *Neurobiol. Aging* 11, 29–37 (1990). An electron microscopic investigation of the post-mortem AD brain that showed widespread reduction in synapse number and a compensatory increase in synapse size in different layers of the cortical regions.
- Masliah, E., Mallory, M., Hansen, L., DeTeresa, R. & Terry, R. D. Quantitative synaptic alterations in the human neocortex during normal aging. *Neurology* 43, 192–197 (1993).

- Scheff, S. W., Price, D. A., Schmitt, F. A. & Mufson, E. J. Hippocampal synaptic loss in early Alzheimer's disease and mild cognitive impairment. *Neurobiol. Aging* 27, 1372–1384 (2006).
- Scheff, S. W., Price, D. A., Schmitt, F. A., DeKosky, S. T. & Mufson, E. J. Synaptic alterations in CA1 in mild Alzheimer disease and mild cognitive impairment. *Neurology* 68, 1501–1508 (2007).
- Chen, K. *et al.* Characterizing Alzheimer's disease using a hypometabolic convergence index. *Neuroimage* 56, 52–60 (2011).
- Milnerwood, A. J. & Raymond, L. A. Early synaptic pathophysiology in neurodegeneration: insights from Huntington's disease. *Trends Neurosci.* 33, 513–523 (2010).
- Picconi, B., Piccoli, G. & Calabresi, P. Synaptic dysfunction in Parkinson's disease. *Adv. Exp. Med. Biol.* 970, 553–572 (2012).
- Trachtenberg, J. T. *et al.* Long-term *in vivo* imaging of experience-dependent synaptic plasticity in adult cortex. *Nature* **420**, 788–794 (2002).
- Matsuzaki, M., Honkura, N., Ellis-Davies, G. C. & Kasai, H. Structural basis of long-term potentiation in single dendritic spines. *Nature* 429, 761–766 (2004).
- Bourne, J. N. & Harris, K. M. Nanoscale analysis of structural synaptic plasticity. *Curr. Opin. Neurobiol.* 22, 372–382 (2011).
- Xu, T. *et al.* Rapid formation and selective stabilization of synapses for enduring motor memories. *Nature* 462, 915–919 (2009).
- Bagetta, V., Chiglieri, V., Sgobio, C., Calabresi, P. & Picconi, B. Synaptic dysfunction in Parkinson's disease. *Biochem. Soc. Trans.* 38, 493–497 (2010).
- Schulz-Schaeffer, W. J. The synaptic pathology of α-synuclein aggregation in dementia with Lewy bodies, Parkinson's disease and Parkinson's disease dementia. Acta Neuropathol. 120, 131–143 (2010).
- Galvin, J. E., Uryu, K., Lee, V. M. & Trojanowski, J. Q. Axon pathology in Parkinson's disease and Lewy body dementia hippocampus contains α-, β-, and γ-synuclein. Proc. Natl Acad. Sci. USA 96, 13450–13455 (1999).
- DiProspero, N. A. *et al*. Early changes in Huntington's disease patient brains involve alterations in cytoskeletal and synaptic elements. *J. Neurocytol.* 33, 517–533 (2004).
   Mayford, M., Siegelbaum, S. A. & Kandel, E. R.
- Mayford, M., Siegelbaum, S. A. & Kandel, E. R. Synapses and memory storage. *Cold Spring Harb. Perspect. Biol.* 4, a005751 (2012).
- Tanaka, J. *et al.* Protein synthesis and neurotrophindependent structural plasticity of single dendritic spines. *Science* **319**, 1683–1687 (2008).
- Maletic-Savatic, M., Malinow, R. & Svoboda, K. Rapid dendritic morphogenesis in CA1 hippocampal dendrites induced by synaptic activity. *Science* 283, 1923–1927 (1999).
- Engert, F. & Bonhoeffer, T. Dendritic spine changes associated with hippocampal long-term synaptic plasticity. *Nature* 399, 66–70 (1999).
- Kleim, J. A. *et al. BDNF* val66met polymorphism is associated with modified experience-dependent plasticity in human motor cortex. *Nature Neurosci.* 9, 735–737 (2006).
- Cheeran, B. et al. A common polymorphism in the brain-derived neurotrophic factor gene (*BDNF*) modulates human cortical plasticity and the response to rTMS. J. Physiol. 586, 5717–5725 (2008).
- Fritsch, B. et al. Direct current stimulation promotes BDNF-dependent synaptic plasticity: potential implications for motor learning. *Neuron* 66, 198–204 (2010).

# This paper demonstrates the use of a non-invasive approach — transcranial direct current stimulation — to alter BDNF secretion in human brain *in vivo*, suggesting that this approach improves motor learning by inducing LTP that is dependent on BDNF secretion in the motor cortex.

- Ferrer, I., Goutan, E., Marin, C., Rey, M. J. & Ribalta, T. Brain-derived neurotrophic factor in Huntington disease. *Brain Res.* 866, 257–261 (2000).
   Zuccato, C. *et al.* Loss of huntingtin-mediated *BDNF*
- Zuccato, C. *et al.* Loss of huntingtin-mediated *BDNF* gene transcription in Huntington's disease. *Science* 293, 493–498 (2001).
- Durany, N. *et al.* Brain-derived neurotrophic factor and neurotrophin-3 levels in Alzheimer's disease brains. *Int. J. Dev. Neurosci.* 18, 807–813 (2000).
- Hock, C., Heese, K., Hulette, C., Rosenberg, C. & Otten, U. Region-specific neurotrophin imbalances in Alzheimer disease: decreased levels of brain-derived neurotrophic factor and increased levels of nerve

growth factor in hippocampus and cortical areas. *Arch. Neurol.* **57**, 846–851 (2000).

- Phillips, H. S. *et al.* BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron* 7, 695–702 (1991).
- Kang, H. & Schuman, E. M. Long-lasting neurotrophininduced enhancement of synaptic transmission in the adult hippocampus. *Science* 267, 1658–1662 (1995). This is the first report to show, using rat hippocampal slices, a pharmacological effect of BDNF on synaptic transmission.
- Messaoudi, E., Bårdsen, K., Srebro, B. & Bramham, C. R. Acute intrahippocampal infusion of BDNF induces lasting potentiation of synaptic transmission in the rat dentate gyrus. *J. Neurophysiol.* 79, 496–499 (1998).
- Figurov, A., Pozzo-Miller, L. D., Olafsson, P., Wang, T. & Lu, B. Regulation of synaptic responses to highfrequency stimulation and LTP by neurotrophins in the hippocampus. *Nature* 381, 706–709 (1996).
   This is the first report to show a pharmacological effect of BDNF on LTP.
- Patterson, S. L. *et al.* Recombinant BDNF rescues deficits in basal synaptic transmission and hippocampal LTP in BDNF knockout mice. *Neuron* 16, 1137–1145 (1996).
- Tanaka, T., Saito, H. & Matsuki, N. Inhibition of GABA<sub>A</sub> synaptic responses by brain-derived neurotrophic factor (BDNF) in rat hippocampus. *J. Neurosci.* **17**, 2959–2966 (1997).
- Frerking, M., Malenka, R. C. & Nicoll, R. A. Brainderived neurotrophic factor (BDNF) modulates inhibitory, but not excitatory, transmission in the CA1 region of the hippocampus. *J. Neurophysiol.* 80, 3383–3386 (1998).
- Gottschalk, W., Pozzo-Miller, L. D., Figurov, A. & Lu, B. Presynaptic modulation of synaptic transmission and plasticity by brain-derived neurotrophic factor in the developing hippocampus. *J. Neurosci.* 18, 6830–6839 (1998).
- Akaneya, Y., Tsumoto, T., Kinoshita, S. & Hatanaka, H. Brain-derived neurotrophic factor enhances long-term potentiation in rat visual cortex. *J. Neurosci.* 17, 6707–6716 (1997).
- Huber, K. M., Sawtell, N. B. & Bear, M. F. Brainderived neurotrophic factor alters the synaptic modification threshold in visual cortex. *Neuropharmacology* 37, 571–579 (1998).
- Korte, M. *et al.* Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc. Natl Acad. Sci. USA* **92**, 8856–8860 (1995).
- Monteggia, L. M. *et al.* Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proc. Natl Acad. Sci. USA* 101, 10827–10832 (2004).
- Minichiello, L. *et al.* Essential role for TrkB receptors in hippocampus-mediated learning. *Neuron* 24, 401–414 (1999).
- Xu, B. *et al.* The role of brain-derived neurotrophic factor receptors in the mature hippocampus: modulation of long-term potentiation through a presynaptic mechanism involving TrkB. *J. Neurosci.* 20, 6888–6897 (2000).
- 70. Ji, Y. et al. Acute and gradual increases in BDNF concentration elicit distinct signaling and functions in neurons. Nature Neurosci. 13, 302–309 (2010). This study resolves a long-term dispute: whether BDNF enhances basal synaptic transmission. It demonstrates that a gradual increase in BDNF concentration facilitates LTP, whereas a rapid rise of BDNF concentration increases basal synaptic transmission. Thus, a single extracellular factor could induce distinct signalling and functions based on how it is delivered.
- Lu, Y., Christian, K. & Lu, B. BDNF: a key regulator for protein synthesis-dependent LTP and long-term memory? *Neurobiol. Learn. Mem.* 89, 312–323 (2008).
- Korte, M., Kang, H., Bonhoeffer, T. & Schuman, E. A role for BDNF in the late-phase of hippocampal long-term potentiation. *Neuropharmacology* 37, 553–559 (1998).
- Balkowiec, A. & Katz, D. M. Activity-dependent release of endogenous brain-derived neurotrophic factor from primary sensory neurons detected by ELISA in situ. J. Neurosci. 20, 7417–7423 (2000)
- Balkowiec, A. & Katz, D. M. Cellular mechanisms regulating activity-dependent release of native brainderived neurotrophic factor from hippocampal neurons. J. Neurosci. 22, 10399–10407 (2002).

- Nagappan, G. *et al.* Control of extracellular cleavage of ProBDNF by high frequency neuronal activity. *Proc. Natl Acad. Sci. USA* **106**, 1267–1272 (2009)
- Natl Acad. Sci. USA 106, 1267–1272 (2009).
   Patterson, S. L., Grover, L. M., Schwartzkroin, P. A. & Bothwell, M. Neurotrophin expression in rat hippocampal slices: a stimulus paradigm inducing LTP in CA1 evokes increases in BDNF and NT-3 mRNAs. Neuron 9, 1081–1088 (1992).
- Neuron 9, 1081–1088 (1992).
  77. Kang, H., Welcher, A. A., Shelton, D. & Schuman, E. M. Neurotrophins and time: different roles for TrkB signaling in hippocampal long-term potentiation. *Neuron* 19, 653–664 (1997).
- Chen, G., Kolbeck, R., Barde, Y. A., Bonhoeffer, T. & Kossel, A. Relative contribution of endogenous neurotrophins in hippocampal long-term potentiation. *J. Neurosci.* **19**, 7983–7990 (1999).
- Pang, P. T. *et al.* Cleavage of proBDNF by tPA/plasmin is essential for long-term hippocampal plasticity. *Science* **306**, 487–491 (2004).
- Barco, A. et al. Gene expression profiling of facilitated L-LTP in VP16-CREB mice reveals that BDNF is critical for the maintenance of LTP and its synaptic capture. *Neuron* 48, 123–137 (2005).
- Lu, Y. et al. TkB as a potential synaptic and behavioral tag. J. Neurosci. **31**, 11762–11771 (2011)
- tag. J. Neurosci. 31, 11762–11771 (2011).
  Park, H. & Poo, M. M. Neurotrophin regulation of neural circuit development and function. *Nature Rev. Neurosci.* 14, 7–23 (2013).
  A recent review summarizing progress in the

#### understanding of the cellular and molecular mechanisms underlying BDNF regulation of neural circuits in the brain.

- Cohen-Cory, S. & Fraser, S. E. Effects of brain-derived neurotrophic factor on optic axon branching and remodelling *in vivo*. *Nature* **378**, 192–196 (1995).
- McAllister, A. K., Lo, D. C. & Katz, L. C. Neurotrophins regulate dendritic growth in developing visual cortex. *Neuron* 15, 791–803 (1995).
- Cabelli, R. J., Hohn, A. & Shatz, C. J. Inhibition of ocular dominance column formation by infusion of NT-4/5 or BDNF. *Science* 267, 1662–1666 (1995).
- Tyler, W. J. & Pozzo-Miller, L. Miniature synaptic transmission and BDNF modulate dendritic spine growth and form in rat CA1 neurones. *J. Physiol.* 553, 497–500 (2003).

An important study that clearly demonstrates the pharmacological effect of BDNF on synaptic growth; that is, it found an increase in the number of dendritic spines.

- Tartaglia, N. *et al.* Protein synthesis-dependent and independent regulation of hippocampal synapses by brain-derived neurotrophic factor. *J. Biol. Chem.* 276, 37585–37593 (2001).
- Horch, H. W., Kruttgen, A., Portbury, S. D. & Katz, L. C. Destabilization of cortical dendrites and spines by BDNF. *Neuron* 23, 353–364 (1999).
- Pozzo-Miller, L. D. *et al.* Impairments in high-frequency transmission, synaptic vesicle docking, and synaptic protein distribution in the hippocampus of BDNF knockout mice. *J. Neurosci.* **19**, 4972–4983 (1999).
- Otal, R., Martinez, A. & Soriano, E. Lack of TrkB and TrkC signaling alters the synaptogenesis and maturation of mossy fiber terminals in the hippocampus. *Cell Tissue Res.* **319**, 349–358 (2005).
- Aguado, F. *et al.* BDNF regulates spontaneous correlated activity at early developmental stages by increasing synaptogenesis and expression of the K<sup>+</sup>/ Cl<sup>-</sup> co-transporter KCC2. *Development* **130**, 1267–1280 (2003).
- Singh, B. et al. Altered balance of glutamatergic/ GABAergic synaptic input and associated changes in dendrite morphology after BDNF expression in BDNFdeficient hippocampal neurons. J. Neurosci. 26, 7189–7200 (2006).
- Ma, Y. L., Wang, H. L., Wu, H. C., Wei, C. L. & Lee, E. H. Brain-derived neurotrophic factor antisense oligonucleotide impairs memory retention and inhibits long-term potentiation in rats. *Neuroscience* 82, 957–967 (1998).
- Mu, J. S., Li, W. P., Yao, Z. B. & Zhou, X. F. Deprivation of endogenous brain-derived neurotrophic factor results in impairment of spatial learning and memory in adult rats. *Brain Res.* 835, 259–265 (1999).
- Linnarsson, S., Bjorklund, A. & Ernfors, P. Learning deficit in BDNF mutant mice. *Eur. J. Neurosci.* 9, 2581–2587 (1997).
- 2581–2587 (1997).
   Chen, X. *et al.* A chemical-genetic approach to studying neurotrophin signaling. *Neuron* 46, 13–21 (2005).
- 97. Johnson, A. W. *et al.* The brain-derived neurotrophic factor receptor TrkB is critical for the acquisition but

not expression of conditioned incentive value. *Eur. J. Neurosci.* **28**, 997–1002 (2008).

- Alonso, M., Vianna, M. R., Izquierdo, I. & Medina, J. H. Signaling mechanisms mediating BDNF modulation of memory formation *in vivo* in the hippocampus. *Cell. Mol. Neurobiol.* 22, 663–674 (2002).
- Koponen, E. *et al.* Transgenic mice overexpressing the full-length neurotrophin receptor trkB exhibit increased activation of the trkB–PLCγ pathway, reduced anxiety, and facilitated learning. *Mol. Cell Neurosci.* 26, 166–181 (2004).
- Soliman, F. *et al.* A genetic variant BDNF polymorphism alters extinction learning in both mouse and human. *Science* **327**, 863–866 (2010).
   Martinowich, K. & Lu, B. Interaction between BDNF
- and serotonin: role in mood disorders. Neuropsychopharmacology 33, 73–83 (2008). 102. Martinowich, K., Manji, H. & Lu, B. New insights into
- 102. Martinowich, K., Manji, H. & Lu, B. New insights into BDNF function in depression and anxiety. *Nature Neurosci.* **10**, 1089–1093 (2007).
- Rakofsky, J. J., Ressler, K. J. & Dunlop, B. W. BDNF function as a potential mediator of bipolar disorder and post-traumatic stress disorder comorbidity. *Mol. Psychiatry* 17, 22–35 (2012).
- Psychiatry 17, 22–35 (2012).
  Nagahara, A. H. & Tuszynski, M. H. Potential therapeutic uses of BDNF in neurological and psychiatric disorders. *Nature Rev. Drug Discov.* 10, 209–219 (2011).

## A comprehensive review that discusses the role of BDNF in multiple neurological and psychiatric disorders and its therapeutic use thereof.

- Kemppainen, S. *et al.* Impaired TrkB receptor signaling contributes to memory impairment in APP/PS1 mice. *Neurobiol. Aging* 33,1122.e23–1122.e39 (2012).
- 106. Nagahara, A. H. *et al.* Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nature Med.* 15, 331–337 (2009).

A systematic investigation of the neuroprotective effects of BDNF in APP transgenic mice, aged rats and brain lesioned non-human primates, using a therapeutic modality that ameliorated synapse loss, neuronal atrophy and restored cognitive deficits without directly altering the level of toxins (specifically, amyloid).

- 107. Żeng, Y., Zhao, D. & Xie, C. W. Neurotrophins enhance CaMKII activity and rescue amyloid-β-induced deficits in hippocampal synaptic plasticity. *J. Alzheimers Dis.* 21, 823–831 (2010).
- Egan, M. F. *et al.* The BDNF val66met polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. *Cell* **112**, 257–269 (2003).

A pioneering study linking a human genetic polymorphism, *BDNF* Val66Met, to a reduction in activity-dependent BDNF secretion. This study explained the changes in hippocampal function and episodic memory associated with the polymorphism and triggered many subsequent studies on the impact of the *BDNF* Val66Met polymorphism on various cognitive functions and brain disorders in humans.

- Chen, Z. Y. *et al.* Genetic variant BDNF (Val66Met) polymorphism alters anxiety-related behavior. *Science* **314**, 140–143 (2006).
- Pezawas, L. *et al.* The brain-derived neurotrophic factor val66met polymorphism and variation in human cortical morphology. *J. Neurosci.* 24, 10099–10102 (2004).
- Szeszko, P. R. *et al.* Brain-derived neurotrophic factor val66met polymorphism and volume of the hippocampal formation. *Mol. Psychiatry* **10**, 631–636 (2005).
- 112. Yang, X. et al. Impact of brain-derived neurotrophic factor Val66Met polymorphism on cortical thickness and voxel-based morphometry in healthy Chinese young adults. PLoS ONE 7, e37777 (2012).
- 113. Sanchez, M. M. *et al.* BDNF polymorphism predicts the rate of decline in skilled task performance and hippocampal volume in healthy individuals. *Transl. Psychiatry* 1, e51 (2011).
- Molendijk, M. L. *et al.* A systematic review and metaanalysis on the association between BDNF val<sup>66</sup>met and hippocampal volume — a genuine effect or a winners curse? *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 731–740 (2012).
   Campbell, S., Marriott, M., Nahmias, C. &
- 115. Campbell, S., Marriott, M., Nahmias, C. & MacQueen, G. M. Lower hippocampal volume in patients suffering from depression: a meta-analysis. *Am. J. Psychiatry* **161**, 598–607 (2004).

- 116. Videbech, P. & Ravnkilde, B. Hippocampal volume and depression: a meta-analysis of MRI studies. *Am. J. Psychiatry* **161**, 1957–1966 (2004)
- *J. Psychiatry* **161**, 1957–1966 (2004).
   Zivadinov, R. *et al.* Preservation of gray matter volume in multiple sclerosis patients with the Met allele of the *rs6265* (Val66Met) SNP of brain-derived neurotrophic factor. *Hum. Mol. Genet.* **16**, 2659– 2668 (2007).
   Voineskos, A. N. *et al.* The brain-derived neurotrophic.
- 118. Voineskos, A. N. et al. The brain-derived neurotrophifactor Val66Met polymorphism and prediction of neural risk for Alzheimer disease. Arch. Gen. Psychiatry 68, 198–206 (2011).
- 119. Dennis, N. A. et al. Brain-derived neurotrophic factor val66met polymorphism and hippocampal activation during episodic encoding and retrieval tasks. *Hippocampus* 21, 980–989 (2011).
- Putcha, D. et al. Hippocampal hyperactivation associated with cortical thinning in Alzheimer's disease signature regions in non-demented elderly adults. J. Neurosci. **31**, 17680–17688 (2011).
   Yassa, M. A. et al. High-resolution structural and
- 121. Yassa, M. A. *et al.* High-resolution structural and functional MRI of hippocampal CA3 and dentate gyrus in patients with amnestic Mild Cognitive Impairment. *Neuroimage* 51, 1242–1252 (2010).
- 122. Dempster, E. et al. Association between BDNF val66 met genotype and episodic memory. Am. J. Med. Genet. B Neuropsychiatr. Genet. 134B, 73–75 (2005).
- 123. Ho, B. C. *et al.* Cognitive and magnetic resonance imaging brain morphometric correlates of brainderived neurotrophic factor Val66Met gene polymorphism in patients with schizophrenia and healthy volunteers. *Arch. Gen. Psychiatry* 63, 731–740 (2006).
- Miyajima, F. et al. Brain-derived neurotrophic factor polymorphism Val66Met influences cognitive abilities in the elderly. *Genes Brain Behav.* 7, 411–417 (2008).
   Schofield, P. R. et al. Disturbances in selective
- 125. Schofield, P. R. *et al.* Disturbances in selective information processing associated with the BDNF Val66Met polymorphism: evidence from cognition, the P300 and fronto-hippocampal systems. *Biol. Psychol.* **80**, 176–188 (2009).
- 126. Mandelman, S. D. & Grigorenko, E. L. BDNF Val66Met and cognition: all, none, or some? A metaanalysis of the genetic association. *Genes Brain Behav.* 11, 127–136 (2012).
- 127. Cirillo, J., Hughes, J., Ridding, M., Thomas, P. Q. & Semmler, J. G. Differential modulation of motor cortex excitability in *BDNF* Met allele carriers following experimentally induced and use-dependent plasticity. *Eur. J. Neurosci.* **36**, 2640–2649 (2012).
- 128. Antal, A. *et al.* Brain-derived neurotrophic factor (*BDNF*) gene polymorphisms shape cortical plasticity in humans. *Brain Stimul.* **3**, 230–237 (2010).
- 129. Witte, A. V. *et al.* Interaction of BDNF and COMT polymorphisms on paired-associative stimulationinduced cortical plasticity. *J. Neurosci.* **32**, 4553–4561 (2012).
- 130. Li Voti, P. *et al.* Correlation between cortical plasticity, motor learning and BDNF genotype in healthy subjects. *Exp. Brain Res.* **212**, 91–99 (2011).
- Di Lazzaro, V. *et al.* The effects of prolonged cathodal direct current stimulation on the excitatory and inhibitory circuits of the ipsilateral and contralateral motor cortex. *J. Neural Transm.* **119**, 1499–1506 (2012).
- 132. Gatt, J. M. *et al.* Association between BDNF Val66Met polymorphism and trait depression is mediated via resting EEG alpha band activity. *Biol. Psychol.* **79**, 275–284 (2008).
- Beste, C. et al. The role of the BDNF Val66Met polymorphism for the synchronization of error-specific neural networks. J. Neurosci. **30**, 10727–10733 (2010).
- 134. Bian, J. T., Zhang, J. W., Zhang, Z. X. & Zhao, H. L. Association analysis of brain-derived neurotrophic factor (*BDNF*) gene 196 A/G polymorphism with Alzheimer's disease (AD) in mainland Chinese. *Neurosci. Lett.* 387, 11–16 (2005).
- Borroni, B. et al. Role of BDNF Val66Met functional polymorphism in Alzheimer's disease-related depression. *Neurobiol. Aging* **30**, 1406–1412 (2009).
- 136. Lim, Y. Y. et al. Modulation of Aβ amyloid-related cognitive decline by brain-derived neurotrophic factor Val66Met polymorphism in preclinical Alzheimer's disease. Neurobiol. Aging (in the press). A longitudinal study demonstrating an epistatic relationship between the BDNF Val66Met polymorphism and amyloid deposition in both preclinical and clinical phases of AD. Subjects with high levels of Aβ combined with the BDNF<sup>Met</sup>

polymorphism showed a faster decline in hippocampal volume and episodic memory.

- 137. Karamohamed, S. *et al.* BDNF genetic variants are associated with onset age of familial Parkinson disease: *GenePD* Study. *Neurology* 65, 1823–1825 (2005).
- Momose, Y. *et al.* Association studies of multiple candidate genes for Parkinson's disease using single nucleotide polymorphisms. *Ann. Neurol.* **51**, 133–136 (2002).
- 139. Parsian, A., Sinha, R., Racette, B., Zhao, J. H. & Perlmutter, J. S. Association of a variation in the promoter region of the brain-derived neurotrophic factor gene with familial Parkinson's disease. *Parkinsonism Relat. Disord.* **10**, 213–219 (2004).
- 140. Guerini, F. R. *et al.* BDNF Val66Met polymorphism is associated with cognitive impairment in Italian patients with Parkinson's disease. *Eur. J. Neurol.* 16, 1240–1245 (2009).
- 141. Liu, J. et al. Brain-derived neurotrophic factor (BDNF) genetic polymorphism greatly increases risk of leucinerich repeat kinase 2 (LRRK2) for Parkinson's disease. Parkinsonism Relat. Disord. 18, 140–143 (2012).
- 142. Ninan, I. et al. The BDNF Val66Met polymorphism impairs NMDA receptor-dependent synaptic plasticity in the hippocampus. J. Neurosci. 30, 8866–8870 (2010).
- 143. Liu, R. J. *et al.* Brain-derived neurotrophic factor Val66Met allele impairs basal and ketaminestimulated synaptogenesis in prefrontal cortex. *Biol. Psychiatry* **71**, 996–1005 (2012).
  144. Cao, L. *et al.* Genetic modulation of BDNF signaling
- 144. Cao, L. *et al.* Genetic modulation of BDNF signaling affects the outcome of axonal competition *in vivo*. *Curr. Biol.* **17**, 911–921 (2007).
- 145. Thoenen, H. & Sendtner, M. Neurotrophins: from enthusiastic expectations through sobering experiences to rational therapeutic approaches. *Nature Neurosci.* 5, 1046–1050 (2002).
- 146. Bradley, W. G. Miami, F. L. & the BDNF Trial Group. A Phasel/II study of recombinant human brain-derived neurotrophic factor in patients with amyotrophic lateral sclerosis. *Ann. Neurol.* **38**, 971 (1995).
- 147. [No authors listed.] A controlled trial of recombinant methionyl human BDNF in ALS: The BDNF Study Group (Phase III). *Neurology* 52, 1427–1433 (1999).
- 148. Ochs, G. et al. A phase I/II trial of recombinant methionyl human brain derived neurotrophic factor administered by intrathecal infusion to patients with amyotrophic lateral sclerosis. Amyotroph. Lateral Scler. Other Motor. Neuron Disord. 1, 201–206 (2000).
- 149. Apfel, S. C. Is the therapeutic application of neurotrophic factors dead? *Ann. Neurol.* **51**, 8–11 (2002).
- Henriques, A., Pitzer, C. & Schneider, A. Neurotrophic growth factors for the treatment of amyotrophic lateral sclerosis: where do we stand? *Front. Neurosci.* 4, 32 (2010).
- 151. Morgan, P. et al. Can the flow of medicines be improved? Fundamental pharmacokinetic and pharmacological principles toward improving Phase II survival. Drug Discov. Today 17, 419–424 (2012).
- Dittrich, F. *et al.* Pharmacokinetics of intrathecally applied BDNF and effects on spinal motoneurons. *Exp. Neurol.* 141, 225–239 (1996).
- 153. Lampe, K. J., Kern, D. S., Mahoney, M. J. & Bjugstad, K. B. The administration of BDNF and GDNF to the brain via PLGA microparticles patterned within a degradable PEG-based hydrogel: protein distribution and the glial response. *J. Biomed. Mater. Res. A* 96, 595–607 (2011).
- 154. Bertram, J. P., Rauch, M. F., Chang, K. & Lavik, E. B. Using polymer chemistry to modulate the delivery of neurotrophic factors from degradable microspheres: delivery of BDNF. *Pharm. Res.* 27, 82–91 (2010).
- 155. Boado, R. J., Zhang, Y. & Pardridge, W. M. Genetic engineering, expression, and activity of a fusion protein of a human neurotrophin and a molecular Trojan horse for delivery across the human blood– brain barrier. *Biotechnol. Bioeng.* **97**, 1376–1386 (2007).
- 156. Żhang, Y. & Pardridge, W. M. Conjugation of brainderived neurotrophic factor to a blood-brain barrier drug targeting system enables neuroprotection in regional brain ischemia following intravenous injection of the neurotrophin. *Brain Res.* 889, 49–56 (2001).
- 157. Yu, Y. J. *et al.* Boosting brain uptake of a therapeutic antibody by reducing its affinity for a transcytosis target. *Sci. Transl. Med.* **3**, 84ra44 (2011).
- Demeule, M. *et al.* Identification and design of peptides as a new drug delivery system for the brain. *J. Pharmacol. Exp. Ther.* **324**, 1064–1072 (2008).

- 159. Gaillard, P. J., Visser, C. C. & de Boer, A. G. Targeted delivery across the blood–brain barrier. *Expert Opin. Drug Deliv.* 2, 299–309 (2005).
- 160. Muruganandam, A., Tanha, J., Narang, S. & Stanimirovic, D. Selection of phage-displayed llama single-domain antibodies that transmigrate across human blood-brain barrier endothelium. *FASEB J.* 16, 240–242 (2002).
- 161. Geral, C., Angelova, Á. & Lesieur, S. From molecular to nanotechnology strategies for delivery of neurotrophins: emphasis on brain-derived neurotrophic factor (BDNF). *Pharmaceutics* 5, 127–167 (2013).

#### A comprehensive review describing the approaches to deliver BDNF and other proteins into the CNS using nanotechnology.

- 162. Gabathuler, R. Approaches to transport therapeutic drugs across the blood–brain barrier to treat brain diseases. *Neurobiol. Dis.* **37**, 48–57 (2010).
- 163. Alcala-Barraza, S. R. *et al.* Intranasal delivery of neurotrophic factors BDNF, CNTF, EPO, and NF4 to the CNS. *J. Drug Target* **18**, 179–190 (2010).
- 164. Jiang, Y. et al. Intranasal brain-derived neurotrophic factor protects brain from ischemic insult via modulating local inflammation in rats. *Neuroscience* **172**, 398–405 (2011).
- 165. Poduslo, J. F. & Curran, G. L. Permeability at the blood-brain and blood-nerve barriers of the neurotrophic factors: NGF, CNTF, NT-3, BDNF. Brain Res. Mol. Brain Res. 36, 280–286 (1996).
- Res. Mol. Brain Res. 36, 280–286 (1996).
  166. Sakane, T. & Pardridge, W. M. Carboxyl-directed pegylation of brain-derived neurotrophic factor markedly reduces systemic clearance with minimal loss of biologic activity. *Pharm. Res.* 14, 1085–1091 (1997).
- Jang, S. W. et al. A selective TrkB agonist with potent neurotrophic activities by 7,8-dihydroxyflavone. Proc. Natl Acad. Sci. USA 107, 2687–2692 (2010).
- 168. Massa, S. M. *et al.* Small molecule BDNF mimetics activate TrkB signaling and prevent neuronal degeneration in rodents. *J. Clin. Invest.* **120**, 1774–1785 (2010).
- 169. Qian, M. D. *et al.* Novel agonist monoclonal antibodies activate TrkB receptors and demonstrate potent neurotrophic activities. *J. Neurosci.* 26, 9394–9403 (2006).
- 170. Lin, J. C. *et al.* Appetite enhancement and weight gain by peripheral administration of TrkB agonists in nonhuman primates. *PLoS ONE* **3**, e1900 (2008).
- O'Leary, P. D. & Hughes, R. A. Design of potent peptide mimetics of brain-derived neurotrophic factor. *J. Biol. Chem.* 278, 25738–25744 (2003).
- 172. Fletcher, J. M. & Hughes, R. A. Novel monocyclic and bicyclic loop mimetics of brain-derived neurotrophic factor. J. Pept. Sci. 12, 515–524 (2006).
- 173. Cardenas-Aguayo Mdel, C., Kazim, S. F., Grundke-Iqbal, I. & Iqbal, K. Neurogenic and neurotrophic effects of BDNF peptides in mouse hippocampal primary neuronal cell cultures. *PLoS ONE* 8, e53596 (2013).
- 174. Jang, S. W. *et al.* Deoxygedunin, a natural product with potent neurotrophic activity in mice. *PLoS ONE* **5**, e11528 (2010).
- 175. Lanz, T. Å. *et al*. Development of an assay to identify activators of TrkB signaling using human induced pluripotent stem cell derived neurons. *Soc. Neurosci. Abstr.* 435.08 (14 Nov 2011, Washington D.C.).
- Tsao, D. *et al.* TrkB agonists ameliorate obesity and associated metabolic conditions in mice. *Endocrinology* **149**, 1038–1048 (2008).
   Xu, L., Zhang, Y., Cohen, S. B. & DiPetrillo, K. TrkB
- Xu, L., Zhang, Y., Cohen, S. B. & DiPetrillo, K. TrkB agonist antibody dose-dependently raises blood pressure in mice with diet-induced obesity. *Am. J. Hypertens.* 23, 732–736 (2010).
   Zoladz, J. A. & Pilc, A. The effect of physical activity on
- Zoladz, J. A. & Pilc, A. The effect of physical activity on the brain derived neurotrophic factor: from animal to human studies. *J. Physiol. Pharmacol.* **61**, 533–541 (2010).
- 179. Knaepen, K., Goekint, M., Heyman, E. M. & Meeusen, R. Neuroplasticity — exercise-induced response of peripheral brain-derived neurotrophic factor: a systematic review of experimental studies in human subjects. *Sports Med.* **40**, 765–801 (2010).
- Adlard, P. A., Perreau, V. M. & Cotman, C. W. The exercise-induced expression of BDNF within the hippocampus varies across life-span. *Neurobiol. Aging* 26, 511–520 (2005).
- Nibuya, M., Morinobu, S. & Duman, R. S. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. J. Neurosci. 15, 7539–7547 (1995).

- 182. Altar, C. A., Whitehead, R. E., Chen, R., Wortwein, G. & Madsen, T. M. Effects of electroconvulsive seizures and antidepressant drugs on brain-derived neurotrophic factor protein in rat brain. *Biol. Psychiatry* 54, 703–709 (2003).
- 183. Saarelainen, T. *et al.* Activation of the TrkB neurotrophin receptor is induced by antidepressant drugs and is required for antidepressant-induced behavioral effects. *J. Neurosci.* 23, 349–357 (2003)
- 184. Rantamaki, T. *et al.* Antidepressant drugs transactivate TrkB neurotrophin receptors in the adult rodent brain independently of BDNF and monoamine transporter blockade. *PLoS ONE* 6, e20567 (2011).
- 185. Zarate, C. A. Jr *et al.* A randomized trial of an *N*-methyl-D-aspartate antagonist in treatmentresistant major depression. *Arch. Gen. Psychiatry* **63**, 856–864 (2006).
- 186. Autry, A. E. et al. NMDA receptor blockade at rest triggers rapid behavioural antidepressant responses. *Nature* 475, 91–95 (2011). This study shows that low doses of NMDA receptor antagonists such as ketamine rapidly increase the synthesis of BDNF in the mouse brain through post-transcriptional mechanisms.
- Li, N. *et al.* mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science* **329**, 959–964 (2010).
- 188. Marvanova, M. *et al.* The neuroprotective agent memantine induces brain-derived neurotrophic factor and trkB receptor expression in rat brain. *Mol. Cell Neurosci.* 18, 247–258 (2001).
- 189. Leyhe, T., Stransky, E., Eschweiler, G. W., Buchkremer, G. & Laske, C. Increase of BDNF serum concentration during donepezil treatment of patients with early Alzheimer's disease. *Eur. Arch. Psychiatry Clin. Neurosci.* 259, 124–128 (2008).
- 190. Autio, H. *et al.* Acetylcholinesterase inhibitors rapidly activate Trk neurotrophin receptors in the mouse hippocampus. *Neuropharmacology* **61**, 1291–1296.
- 191. Alonso, M., Medina, J. H. & Pozzo-Miller, L ERK1/2 activation is necessary for BDNF to increase dendritic spine density in hippocampal CA1 pyramidal neurons. *Learn. Mem.* **11**, 172–178 (2004).
- 192. Hashimoto, R. *et al.* Effect of the brain-derived neurotrophic factor and the apolipoprotein E polymorphisms on disease progression in preclinical Alzheimer's disease. *Genes Brain Behav.* 8, 43–52 (2009).
- 193. Adamczuk, K. et al. Polymorphism of brain derived neurotrophic factor influences β amyloid load in cognitively intact apolipoprotein Εε4 carriers. *Neuroimage Clin.* (in the press).
- 194. Blin, O., Davies, C. H. & Lu, B. Clinical innovation for neurodegenerative diseases. *Clin. Invest.* 2, 663–665 (2012).
- 195. Huang, Y. & Morozov, A. Hippocampal deletion of BDNF gene attenuates gamma oscillations in area CA1 by up-regulating 5-HT3 receptor. *PLoS ONE* 6, e16480 (2011).
- 196. Zheng, K. et al. TrkB signaling in parvalbumin-positive interneurons is critical for gamma-band network synchronization in hippocampus. Proc. Natl Acad. Sci. USA 108, 17201–17206 (2011).
- 197. Rossini, P. M. *et al.* Conversion from mild cognitive impairment to Alzheimer's disease is predicted by sources and coherence of brain electroencephalography rhythms. *Neuroscience* 143, 793–803 (2006).
- 198. Lai, C. L., Lin, R. T., Liou, L. M. & Liu, C. K. The role of event-related potentials in cognitive decline in Alzheimer's disease. *Clin. Neurophysiol.* **121**, 194–199 (2010).
- 199. Miller, G. Alzheimer's research. Stopping Alzheimer's before it starts. *Science* **337**, 790–792 (2012).
- Bateman, R. J. *et al.* Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N. Engl. J. Med.* 367, 795–804 (2012).
- Reiman, E. M. *et al.* Brain imaging and fluid biomarker analysis in young adults at genetic risk for autosomal dominant Alzheimer's disease in the presenilin 1 E280A kindred: a case-control study. *Lancet Neurol.* 11, 1048–1056 (2012).
- 202. Alonso Canovas, A. *et al.* Dopaminergic agonists in Parkinson's disease. *Neurologia* 2 Jul 2011(doi:10.1016/j.nrl.2011.04.012).
- 203. Tong, L. & Perez-Polo, R. Brain-derived neurotrophic factor (BDNF) protects cultured rat cerebellar granule neurons against glucose deprivation-induced apoptosis. J. Neural Transm. 105, 905–914 (1998).
- 204. Ferenz, K. B. *et al.* Nerve growth factor and brainderived neurotrophic factor but not granulocyte

colony-stimulating factor, nimodipine and dizocilpine, require ATP for neuroprotective activity after oxygenglucose deprivation of primary neurons. *Brain Res.* **1448**, 20–26 (2012).

- Lee, B. *et al.* The CREB/CRE transcriptional pathway: protection against oxidative stress-mediated neuronal cell death. *J. Neurochem.* **108**, 1251–1265 (2009).
- 206. Lindholm, D., Dechant, G., Heisenberg, C. P. & Thoenen, H. Brain-derived neurotrophic factor is a survival factor for cultured rat cerebellar granule neurons and protects them against glutamate-induced neurotoxicity. *Eur. J. Neurosci.* 5, 1455–1464 (1993).
- Arancibia, S. *et al.* Protective effect of BDNF against β-amyloid induced neurotoxicity *in vitro* and *in vivo* in rats. *Neuropiol. Dis.* **31**, 316–326 (2008).
- Ferrer, I. *et al.* Brain-derived neurotrophic factor reduces cortical cell death by ischemia after middle cerebral artery occlusion in the rat. *Acta Neuropathol.* **101**, 229–238 (2001).
- 209. Müller, H. D. *et al.* Brain-derived neurotrophic factor but not forced arm use improves long-term outcome after photothrombotic stroke and transiently upregulates binding densities of excitatory glutamate receptors in the rat brain. *Stroke* **39**, 1012–1021 (2008).
- 210. Frim, D. M. *et al.* Implanted fibroblasts genetically engineered to produce brain-derived neurotrophic factor prevent 1-methyl-4-phenylpyridinium toxicity to dopaminergic neurons in the rat. *Proc. Natl Acad. Sci. USA* **91**, 5104–5108 (1994).
- Shults, C. W., Kimber, T. & Altar, C. A. BDNF attenuates the effects of intrastriatal injection of 6-hydroxydopamine. *Neuroreport* 6, 1109–1112 (1995).
- Tandon, P., Yang, Y., Das, K., Holmes, G. L. & Stafstrom, C. E. Neuroprotective effects of brainderived neurotrophic factor in seizures during development. *Neuroscience* **91**, 293–303 (1999).
- Luikart, B. W. *et al.* TrkB has a cell-autonomous role in the establishment of hippocampal Schaffer collateral synapses. *J. Neurosci.* 25, 3774–3786 (2005).
- 214. Čattaneo, A., Capsoni, S. & Paoletti, F. Towards non invasive nerve growth factor therapies for Alzheimer's disease. J. Alzheimers Dis. 15, 255–283 (2008).
- 215. Petryshen, T. L. *et al.* Population genetic study of the brain-derived neurotrophic factor (*BDNF*) gene. *Mol. Psychiatry* **15**, 810–815 (2010).
- 216. Shimizu, E., Hashimoto, K. & Iyo, M. Ethnic difference of the BDNF 196G/A (val66met) polymorphism frequencies: the possibility to explain ethnic mental traits. Am. J. Med. Genet. B Neuropsychiatr. Genet. 126B, 122–123 (2004).
- 217. Ramasamy, D. P. et al. Effect of Met66 allele of the BDNF rs6265 SNP on regional gray matter volumes in patients with multiple sclerosis: a voxel-based morphometry study. Pathophysiology 18, 53–60 (2011)
- Hall, D., Dhilla, A., Charalambous, A., Gogos, J. A. & Karayiorgou, M. Sequence variants of the brainderived neurotrophic factor (*BDNF*) gene are strongly associated with obsessive-compulsive disorder. *Am. J. Hum. Genet.* **73**, 370–376 (2003).
- Geller, B. *et al.* Linkage disequilibrium of the brainderived neurotrophic factor Val66Met polymorphism in children with a prepubertal and early adolescent bipolar disorder phenotype. *Am. J. Psychiatry* 161, 1698–1700 (2004).
- Martinez, A. *et al.* TrkB and TrkC signaling are required for maturation and synaptogenesis of hippocampal connections. *J. Neurosci.* 18, 7336–7350 (1998).
- Jang, S. W. et al. N-acetylserotonin activates TrkB receptor in a circadian rhythm. Proc. Natl Acad. Sci. USA 107, 3876–3881 (2010).
- Lee, F. S. & Chao, M. V. Activation of Trk neurotrophin receptors in the absence of neurotrophins. *Proc. Natl Acad. Sci. USA* 98, 3555–3560 (2001).
- 223. Lee, F. S., Rajagopal, R., Kim, A. H., Chang, P. C. & Chao, M. V. Activation of Trk neurotrophin receptor signaling by pituitary adenylate cyclase-activating polypeptides. *J. Biol. Chem.* **277**, 9096–9102 (2002).
- 224. Swift, J. L. et al. Quantification of receptor tyrosine kinase transactivation through direct dimerization and surface density measurements in single cells. Proc. Natl Acad. Sci. USA 108, 7016–7021 (2011).
- 225. Huang, Y. Z., Pan, E., Xiong, Z. Q. & McNamara, J. O. Zinc-mediated transactivation of TrkB potentiates the hippocampal mossy fiber–CA3 pyramid synapse. *Neuron* 57, 546–558 (2008).

- Fletcher, J. M. *et al.* Design of a conformationally defined and proteolytically stable circular mimetic of brain-derived neurotrophic factor. *J. Biol. Chem.* 283, 33375–33383 (2008).
- 227. Peter, J.-C. et al. Anti-trkb antibodies as pharmacological tools to study the function of the TrkB receptor and its role in the regulation of food intake. *Pharmacological* 1–14 (2013)
- Pharmacologia 4, 1–14 (2013).
  228. Huang, Y. Z. *et al.* RNA aptamer-based functional ligands of the neurotrophin receptor, TrkB. *Mol. Pharmacol.* 82, 623–635 (2012).
- 229. Lauterborn, J. C., Lynch, G., Vanderklish, P., Arai, A. & Gall, C. M. Positive modulation of AMPA receptors increases neurotrophin expression by hippocampal and cortical neurons. *J. Neurosci.* 20, 8–21 (2000).
- Lauterborn, J. C. *et al.* Chronic elevation of brainderived neurotrophic factor by ampakines. *J. Pharmacol. Exp. Ther.* **307**, 297–305 (2003).
- 231. Lauterborn, J. C. *et al.* Ampakines cause sustained increases in brain-derived neurotrophic factor signaling at excitatory synapses without changes in

AMPA receptor subunit expression. *Neuroscience* **159**, 283–295 (2009).

- Legutko, B., Li, X. & Skolnick, P. Regulation of BDNF expression in primary neuron culture by LY392098, a novel AMPA receptor potentiator. *Neuropharmacology* 40, 1019–1027 (2001).
- Nibuya, M., Nestler, E. J. & Duman, R. S. Chronic antidepressant administration increases the expression of cAMP response element binding protein (CREB) in rat hippocampus. J. Neurosci, 16, 2365–2372 (1996).
- 234. Deogracias, R. et al. Fingolimod, a sphingosine-1 phosphate receptor modulator, increases BDNF levels and improves symptoms of a mouse model of Rett syndrome. Proc. Natl Acad. Sci. USA 109, 14230–14235 (2012).
- Apostol, B. L. *et al.* CEP-1347 reduces mutant huntingtin-associated neurotoxicity and restores BDNF levels in R6/2 mice. *Mol. Cell Neurosci.* 39, 8–20 (2008).
- 236. Rasmussen, P. et al. Evidence for a release of brainderived neurotrophic factor from the brain during exercise. *Exp. Physiol.* **94**, 1062–1069 (2009).

- Kotani, S., Yamauchi, T., Teramoto, T. & Ogura, H. Pharmacological evidence of cholinergic involvement in adult hippocampal neurogenesis in rats. *Neuroscience* 142, 505–514 (2006).
- Wang, Z. F., Tang, L. L., Yan, H., Wang, Y. J. & Tang, X. C. Effects of huperzine A on memory deficits and neurotrophic factors production after transient cerebral ischemia and reperfusion in mice. *Pharmacol. Biochem. Behav.* 83, 603–611 (2006).
- Borrell-Pages, M. *et al.* Cystamine and cysteamine increase brain levels of BDNF in Huntington disease via HSJ1b and transglutaminase. *J. Clin. Invest.* **116**, 1410–1424 (2006).

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