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## INTRODUCTION

- Chronic implants cause an inflammatory response that can negatively impact the function of the implant. However, current methods to assess inflammation are terminal involving histology.
- Implant stimulation could exacerbate inflammation that in turn, can impact the efficacy of DBS therapy requiring frequent tuning by an expert.
- Glial fibrillary acidic protein (GFAP), expressed in reactive astrocytes, is a widely used biomarker to assess brain tissue response to inflammatory stressors.
- Peripheral blood monocytes (PBMCs) are recruited to the brain to clear inflammation debris and can re-enter the blood stream.
- We propose here a non-invasive method for assessing the GFAP response in brain tissue using blood-draw that examines the peripheral blood monocytes

## METHODS

- We used the following 4 experimental cohorts (n=3 rats in each cohort implanted with 4 teflon-insulated tungsten electrode arrays) to assess GFAP levels in PBMC extracts:

Cohort 1: surgery, craniotomy, no implant

Cohort 2: surgery, implant, no electrical stimulation

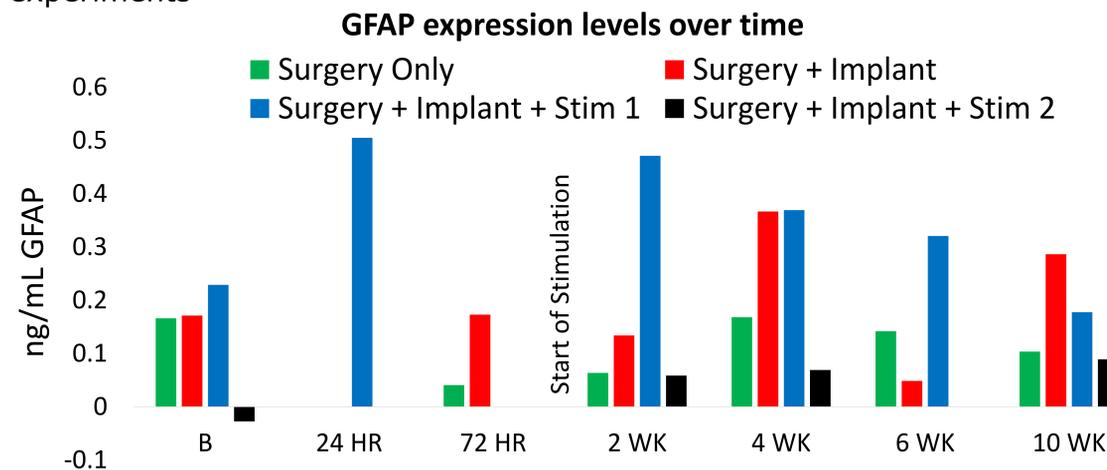
Cohort 3: surgery, implant, electrical stimulation, **Stim1** - 9 nC/phase and 25  $\mu\text{C}/\text{cm}^2$  with a GSA of 35,000  $\mu\text{m}^2$  (below Shannon limit)

Cohort 4: surgery, implant, electrical stimulation, **Stim2** - 36 nC/phase and 1834  $\mu\text{C}/\text{cm}^2$  with a GSA of 1900  $\mu\text{m}^2$  (above Shannon limit)

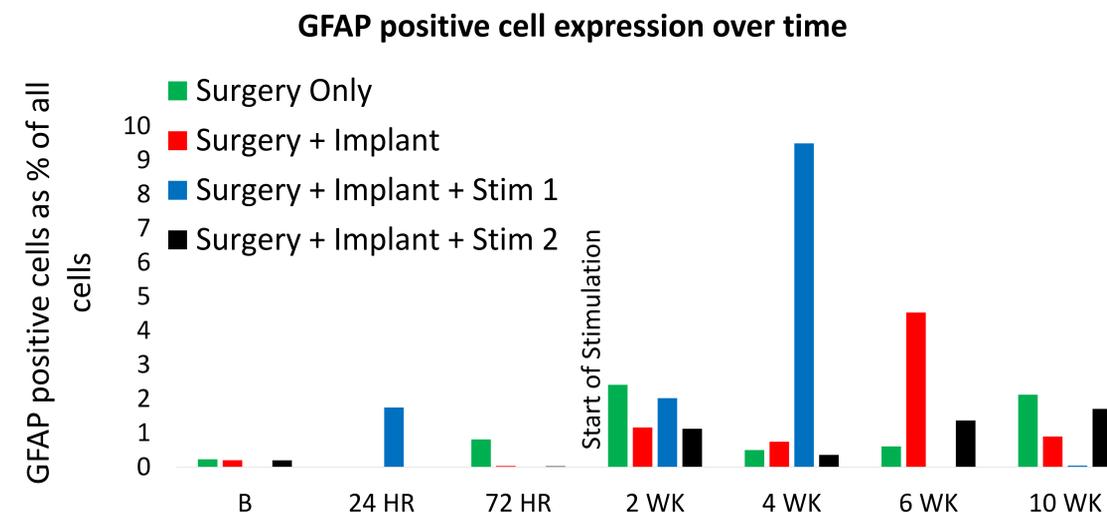
- For each rat ~1 ml of blood was drawn 2 weeks before and at various times after surgery, followed by a terminal bleed 10 weeks after surgery
- Cohorts in experimental conditions 3 & 4 were stimulated every day for 1 hr for the first 4 weeks
- Stimulation conditions were biphasic, charge-balanced at 100 Hz with amplitude of 180  $\mu\text{A}$  and 50  $\mu\text{s}/\text{phase}$  (condition 3) or 360  $\mu\text{A}$  and 100  $\mu\text{s}/\text{phase}$  (condition 4)
- Blood samples were analyzed by a) fluorescence-activated cell sorting (FACS) to measure the number of GFAP containing cells and b) single-sided ELISA (ELISA) to determine GFAP levels in extracts of peripheral blood monocytes at ZelosDX, Tucson, AZ
- Electrical impedance spectroscopy (EIS) and cyclic voltammetry (CV) were performed periodically

## RESULTS

- In cohort 3, there was a 3- to 4-fold increase in GFAP expression at 24 hours and 2 weeks post-implantation compared to baseline.
- The levels of GFAP were found to decrease progressively at 4-, 6-, and 10-weeks post-implantation in cohorts 2 and 4.
- Results of GFAP in PBMC extracts in all the cohorts are consistent with terminal GFAP assessments in brain tissue using immunohistochemistry techniques.
- FACS analysis of the percentage of monocytes in the blood showed a similar GFAP expression profile over time of implantation, but did not show any significant differences between cohorts 3 & 4
- EIS and CV measurements showed consistent electrode function during experiments

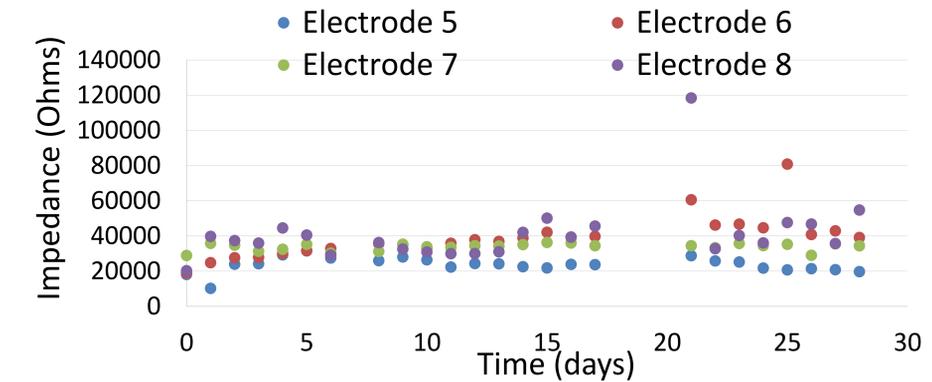


**Figure 1:** Mean GFAP expression levels in PBMC extracts over time in different cohorts.

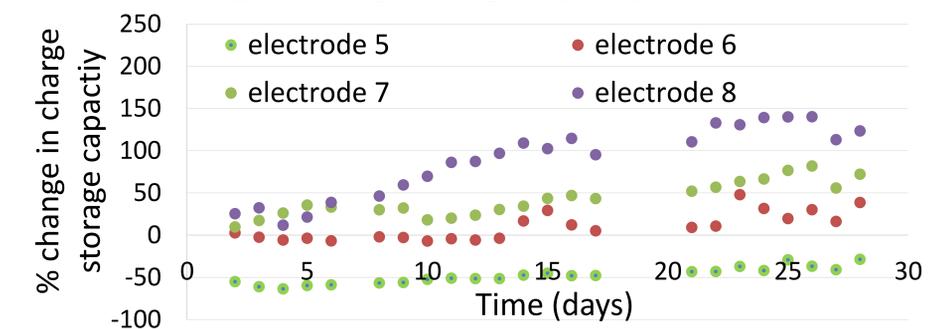


**Figure 2:** Percentage of GFAP positive cells (determined using FACS) over time in different cohorts.

## Electrode impedances as a function of time



## Change in Charge Storage Capacity Over Time



**Figure 3:** Typical impedances at 1 kHz (top) and charge storage capacity (bottom) in an array over the duration of a chronic experiment show no significant changes.

- Electrical impedances at 1 kHz and charge storage capacities did not change significantly during the experiment indicating the integrity of the neural interface.
- The proposed novel assay involving PBMCs allows for multiple GFAP assessments at 2-week intervals in longitudinal rodent experiments for the first time.
- This will allow us, for the first time, to track changes in *expression of GFAP in rodent brain tissue* due to any interventions introduced during the experiment.

## CONCLUSION

In conclusion, we have shown that the novel blood-based assay is capable of detecting trace amounts of GFAP in response to microelectrode implantation and DBS activation in rodent peripheral blood monocytes in chronic experiments

## ACKNOWLEDGEMENTS

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