

Preliminary Report: Determine the effectiveness of Xtreme Bio[®] in Inactivating African swine fever virus

Objective:

To determine the effectiveness of XTREME BIO[®] (XB) in inactivating the infectivity of African swine fever virus (ASFV).

In vitro phase:

Note: All ASFV work were performed in BSL3 laboratory facility.

Materials needed:

- PB-derived macrophages at day 3 to 5 in culture in 96-well plate
- Cell culture media, phosphate-buffered saline (PBS) and water for dilution
- 160mm Petri dishes, Multichannel tips, 96-well and 24 plates
- XB and XB high foam (HF) provided by sponsor/manufacturer – prepared at 0.5 oz/gallon and 2.0 oz/gallon
- Peroxigard (perox) disinfectant as positive control
- ASFV – working concentration of 100HAD₅₀. Pigs die of ASFV at lower than 100 HAD₅₀ infection.

Treatments:

- | | |
|-----------------------|---------|
| (1) 0.5oz/gallon XB | + ASFV |
| (2) 2oz/gallon XB | + ASFV |
| (3) 0.5oz/gallon XBHF | + ASFV |
| (4) 2oz/gallon XBHF | + ASFV |
| (5) Peroxigard | + ASFV |
| (6) PBS | +ASFV |
| (7) PBS | no ASFV |

Methodology:

1. 2 ml pure culture 100HAD₅₀ ASFV was added in petri dishes #1 to 6. Negative control is PBS.
2. 1 ml of XB was added and mixed thoroughly. This was repeated two more times to have a final volume of ~5ml of ASFV/XB or PBS mixture. Adequate mixing was done per XB or PBS addition with 10 minutes of contact time.
3. To determine the working concentration without killing the cells, different concentrations of mixtures were prepared by serial dilution (dilution 1:10, 1:20, 1:40 and 1:80) in 96-well plate and with cell culture media as diluent.

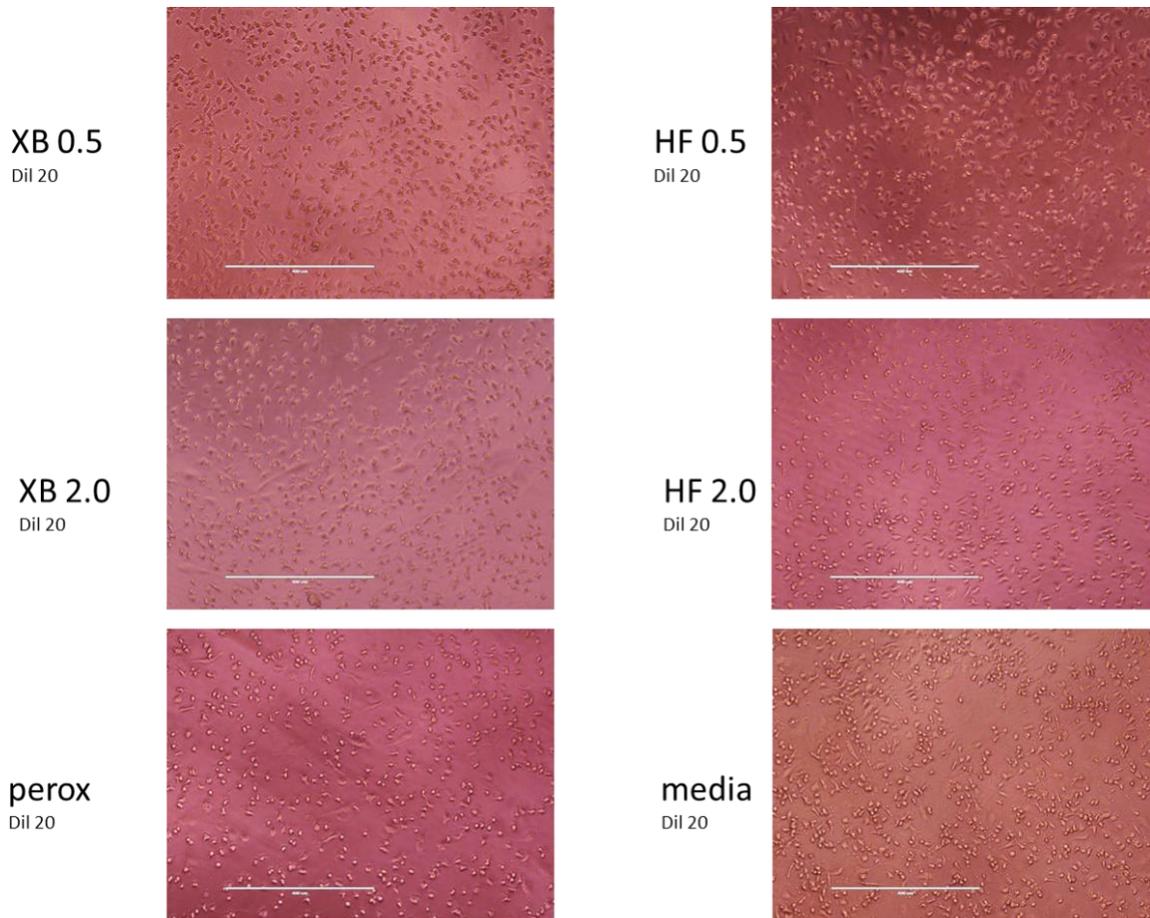


Figure 2. At Dilution 1: 20 Healthy, attached cells were observed in all treatments.

Figure 3. Day 3 Cell Toxicity Results. (✓) cell alive (✗) dead cells

	1	2	3	4	5	6	7	8	9	10	11	12	dilution	250ul vol
A	+/-	+/-	+/-	+/-	✗	✗	✗	✗	✓	✓	✓	✓	1:10	25ul + 225ul media
B	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:20	25ul dil A+ 225ul
C	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:40	25ul dil B + 225ul
D	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:80	25ul dil C + 225ul
E	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	1:10	25ul + 225ul media
F	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:20	25ul dil E+ 225ul
G	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:40	25ul dil F + 225ul
H	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	1:80	25ul dil G + 225ul
		0.5 XB			0.5 XBHF			Peroxigard					PBS + ASFV	
		2.0 XB			2.0 XBHF								PBS, no ASFV	

2022-11-16 XB study ASFv RT-PCR				
				HAD ₅₀ /ml
Sample	C_T	C_T Mean	C_T SD	Quantity
Standard	19.54	19.66	0.18	7,500,000
Standard	19.79			7,500,000
Standard	22.24	22.35	0.16	750,000
Standard	22.46			750,000
Standard	25.73	25.81	0.10	75,000
Standard	25.88			75,000
Standard	28.97	29.14	0.24	7,500
Standard	29.31			7,500
Standard	31.96	32.25	0.40	750
Standard	32.53			750
Standard	36.04	36.21	0.25	75
Standard	36.39			75
NTC	Undetermined			
NTC	Undetermined			
AFV Plasmid	26.19	26.18	0.01	62,146
AFV Plasmid	26.17			62,830

	P2		P3	
XB 0.5	Undetermined	Undetermined	Undetermined	Undetermined
XB 2.0	Undetermined	Undetermined	Undetermined	Undetermined
XBHF 0.5	Undetermined	Undetermined	Undetermined	Undetermined
XBHF 2.0	Undetermined	Undetermined	Undetermined	Undetermined
Perox	Undetermined	Undetermined	Undetermined	Undetermined
+ASFV	CT: 38.52; Qty: 11.6	Undetermined	Undetermined	Undetermined
media	Undetermined	Undetermined	Undetermined	Undetermined

Figure 4. ASFv RT-PCR results. ASFv was not detected in all disinfectant treatments.

Preliminary Observations/Future Experiments:

- Mixture dilution of 1:20 is the concentration that is not toxic to cells.
- ASFv was not detected by RT-PCR in all mixtures with disinfectants.
- However, ASFv was barely detected in only one of two duplicates in P2 ASFV and none on P3 ASFV.
- Suggested plan of action on future experiments (to be done after reverification – February 2023)
 - Continue passaging of cells. Samples were frozen and will be thawed for further passaging upon facility re-opening.
 - Increase cultivation from 3 to 5 days to allow ample virus amplification. In this preliminary experiment, passages were only every 3 days due to limited time before facility shutdown.
 - Increase number of passages to allow ample virus amplification (if there is any virus remaining after treatment), that would be detected by RT-PCR.
 - Possibly increase virus concentration at the start of experiment.

Prepared by RM 2022-11-17