The Journal of Animal & Plant Sciences, 22(2): 2012, Page: 283-288 ISSN: 1018-7081

CHEMICAL EVALUATION OF WEED SEEDS MIXED WITH WHEAT GRAINS AT HARVEST

M. N. Abbas, S. A. Rana, M. Shahid^{*}, N. Rana, M. Mahmood-ul-Hassan, M. Hussain^{*}

Department of Zoology and Fisheries, University of Agriculture, Faisalabad – Pakistan.
*Department of Chemistry and Biochemistry, University of Agriculture, Faisalabad, Pakistan Corresponding author's email: drmmhassan1@gmail.com

ABSTRACT

Phytochemical potential of fifteen weeds associated with wheat crop was evaluated. Qualitative and quantitative phytochemical analysis of weed seed extracts was made. Alkaloids, saponins, glycosides, terpenoids, anthraquinine, steroids, flavonoids and tannins were detected from the weed seeds. Tannins and alkaloids were in high concentration. Tannins ranged from 7.97 to 24.17%, alkaloids 0.88 to 4.00%, saponins 0.54 to 1.29% and flavonoids 3.91 to 15.55%. Wheat weeds care medicianlly important but their phytochemical potential needs to be further investigated.

Key words: Alkaloids, Biodiversity, Laxative, Tannins, Predator-Prey.

INTRODUCTION

Field of ethno-medicine is still unexplored in Pakistan. The country has a wealth of 5,700 species of medicinal plants of which 372 are endemic and 456 are in active trade for the manufacture almost 350 classical formulations to treat various ailments. Medicinal weeds play a key role in traditional health care system for human and animals. Most of allopathic drugs also comprise extracts taken from medicinal plants. The traditional and indigenous medical knowledge of plants, both oral and codified, is however eroding rapidly from our system (Rashid and Arshad, 2002; Ahmad *et al.*, 2003; Hussain *et al.*, 2006; Baquar1989; Mujtaba and Khan, 2007).

In spite of their ethno-botanical importance, weeds associated with wheat are exterminated from the crop to increase per hectare yield. A wide variety of weeds is associated with wheat in our country that not only adds phytomorphic diversity to the crop but also offers feeding, sheltering, refuge and breeding niches to beneficial arthropods. These weeds have been extensively used in herbal medicine (Table 1) but no such study is available from Pakistan.

Keeping in mind this dearth of knowledge, the present study was designed to investigate the scientific bases for the use of some weeds associated with wheat by defining and quantifying their percentage of crude phytochemical constituents. We focused our effort towards the characterization of Vicia sativa, Galium aparine, Rumex dentatus, Avena fatua, Lathyrus aphaca, Phalaris minor, Carthamus oxycantha, Convolvulus arvensis, Chenopodium album, Coronopus didymus, Melilots indica, Cichorium intybus, Anagallis arvensis, Chenopodium murale and Euphorbia sp.

MATERIALS AND METHODS

Seeds of wheat weeds (Vicia sativa, Galium aparine, Rumex dentatus, Avena fatua, Lathyrus aphaca, Phalaris minor, Carthamus oxycantha, Convolvulus arvensis, Chenopodium album, Coronopus didymus, Melilots indica, Cichorium intybus, Anagallis arvensis, Chenopodium murale and Euphorbia sp.) that had already been taxonomically identified and stored in the "Seed Bank, Ayuab Agriculture Research Institute, Faisalabad Pakistan" were collected from and were packed in sterilized polythene bags.

Preparation of extracts. The seeds were crushed into fine powder using a mill grinder and 25 g of each weed seed powder was dissolved separately in 100 ml of commercially available pure ethanol. The solution was kept at room temperature for seven days to allow the extraction of compounds from seeds. The solution for each weed variety was stirred after every 24 h using sterile glass rod. After seven days, the solution was filtered through Whatman filter paper no. 1 and a greenish filtrate was obtained. The solvent was evaporated and a black sticky substance was obtained that was stored in the refrigerator and suspended in 10% dimethyl sulfoxide prior to use.

Phytochemical screening. Chemical tests were carried out both on the ethanolic extract and on the powdered specimens using standard procedures to identify the constituents as described by Harborne (1973), Trease and Evans (1989) and Sofowara (1993). The specific procedure involved for the evaulation of a particular group of chemicals is mention below.

1. **Tannins.** One ml of water and 1-2 drops of ferric chloride solution were added in 0.5 ml of extracted solution. Blue colour was observed for gallic tannins

- and green black for catecholic tannins (Iyengar, 1995).
- 2. **Saponins.** Foam test: Small amount of extract was shaken with little quantity of water. If foam produced persists for ten minutes it indicates the presence of saponins (Roopashree, *et al.*, 2008).
- 3. **Flavonoids (Alkaline Reagent Test).** Extracts were treated with few drops of sodium hydroxide solution. Formation of intense yellow colour, which becomes colourless on addition of dilute acid, indicates the presence of flavonoids (Roopashree, *et al.*, 2008).
- 4. Steriods. Two ml of acetic anhydride was added to 0.5 g ethanolic extract of each sample with 2 ml H₂SO₄. The colour changed from violet to blue or green in some samples indicating the presence of steroids.
- 5. Terpenoids (Salkowski test). Five ml of each extract was mixed in 2 ml of chloroform, and concentrated H₂SO₄ (3 ml) was carefully added to form a layer. A reddish brown colouration of the interface was formed to show the presence of terpenoids.
- 6. Cardiac glycosides (Keller-Killani test): Five ml of each extracts was treated with 2 ml of glacial acetic acid containing one drop of ferric chloride solution. This was underlayed with 1 ml of concentrated sulphuric acid. A brown ring of the interface indicates a deoxysugar characteristic of cardenolides. A violet ring may appear below the brown ring, while in the acetic acid layer, a greenish ring may form just gradually throughout thin layer.
- Alkaloids. Alkaloids are basic nitrogenous compounds with definite physiological and pharmacological activity. Alkaloid solution produces white yellowish precipitate when a few drops of Mayer's reagents are added (Siddiqui and Ali, 1997).
- 8. **Anthraquinones** Borntrager's test was used for detecting the presence of anthraquinones. In this case 0.5 g of the plant extract was shaken with benzene layer separated and half of its own volume of 10% ammonia solution added. A pink, red or violet colouration in the ammoniacal phase indicated the presence of anthraquinone.

Quantitative determination of the chemical constituency

Preparation of fat free sample: Two g of the sample were defatted with 100 ml of diethyl ether using a soxhlet apparatus for 2 h.

1. **Alkaloid.** Harborne (1973) method was used to determine alkaloid. Five gram of the sample was weighed into a 250 ml beaker and 200 ml of 10% acetic acid in ethanol was added and covered and allowed to stand for 4 h. This was filtered and the extract was concentrated on a water bath to one-quarter of the original volume. Concentrated

- ammonium hydroxide was added drop wise to the extract until the precipitation was complete. The whole solution was allowed to settle and the precipitated was collected and washed with dilute ammonium hydroxide and then filtered. The residue is the alkaloid, which was dried and weighed.
- 2. **Tannin.** Van-Burden and Robinson (1981) method was used to determine Tannin. For this 500 mg of the sample was weighed into a 50 ml plastic bottle. Fifty ml of distilled water was added and shaken for 1 h in a mechanical shaker. This was filtered into a 50 ml volumetric flask and made up to the mark. Then 5 ml of the filtered was pipetted out into a test tube and mixed with 2 ml of 0.1 M FeCl₃ in 0.1 N HCl and 0.008 M potassium ferrocyanide. The absorbance was measured at 605 nm within 10 min.
- Saponin. Obadoni and Ochuko (2001) method was used to determine Saponin. The samples were ground and 20 g of each were put into a conical flask and 100 Cm³ of 20% aqueous ethanol were added. The samples were heated over a hot water bath for 4 h with continuous stirring at about 55°C. The mixture was filtered and the residue re-extracted with another 200 ml 20% ethanol. The combined extracts were reduced to 40 ml over water bath at about 90°C. The concentrate was transferred into a 250 ml separatory funnel and 20 ml of diethyl ether was added and shaken vigorously. The aqueous layer was recovered while the ether layer was discarded. The purification process was repeated and 60 ml of nbutanol was added. The combined n-butanol extracts were washed twice with 10 ml of 5% aqueous sodium chloride. The remaining solution was heated in a water bath. After evaporation the samples were dried in the oven to a constant weight; the saponin content was calculated as percentage.
- 4. **Flavonoids.** Bohm and Kocipai-Abyazan (1994) method was used to determine flavonoid. For This Ten g of the plant sample was extracted repeatedly with 100 ml of 80% aqueous methanol at room temperature. The whole solution was filtered through Whatman filter paper No 42 (125 mm). The filtrate was later transferred into a crucible and evaporated into dryness over a water bath and weighed to a constant weight.

Statistical analysis. All values have been expressed as mean \pm standard deviation.

RESULTS AND DISCUSSION

Self growing plants (weeds) in association with wheat crop plants have ethno-botanical importance. They can be used as medicinal plants assessing the need to evaluate the constituents of these weed plants. The present study carried out on the seeds of weeds viz. *Vicia*

sativa, Galium aparine, Rumex dentatus, Avena fatua, Lathyrus aphaca, Phalaris minor, Carthamus oxycantha, Convolvulus arvensis, Chenopodium album, Coronopus didymus, Melilots indica, Cichorium intybus, Anagallis arvensis and Chenopodium murale. The chemical analysis revealed the presence of medicinally active constituents. The phytochemical screening and quantitative estimation of the crude yields of chemical constituents of the plant seeds studied showed that the seeds were rich in alkaloids, flavonoids, tannins, cardiac glycosides and saponins. They were known to show medicinal activity as well as exhibiting physiological activity (Sofowara, 1993; Ajayi, et al., 2011).

The phytochemical characteristics of the fifteen wheat weeds have been summarized in the Table 2. Flavinoids, tannins and alkaloids were present in all the weed plants. Anthraquinones and terpenoids were absent

in both the R. dentatus and C. murale whearse steroids and glycosides were absent in G. aparine. Flavinoids was found in high concentration in all seeds of weeds occurring in wheat crop may be responsible for their use in herbal medicine for cure of various ailments like capillary and vascular weakness (varicose veins, dysfunction, blood cloting and oedema) (Olivere- Bever, 1986; Rogar, 2002). Flavinoids were also useful for sexual stimulation (Benson, et al., 2008). Flavonoids had the record of being powerful water soluble free radical scavengers and wonderful antioxidants which prevented oxidative cell damage, had potent anticancer activity and inhibited tumor growth (Stauth, 1993; Sharma, et al., 2007). A study showed that flavonoids could inhibit the development of fluids that resulted in diarrhea by targeting the intestinal cystic fibrosis trans-membrane conductance regulator (Schuier, et al., 2005).

Table 1. Brief Review of the Various Selected Weeds Used For the Present Study of Wheat Crop

Family	Botanical Name	Local Name	Traditional Uses	Reference
Fabaceae	Vicia sativa	Rivari	Antiseptic	Dwivedi, 2008; Jabeen et al., 2009
Fabaceae	Lathyrus aphaca	Jangali matar	Famine food.	Dwivedi, 2008. Jabeen <i>et al.</i> , 2009; Jabeen <i>et al.</i> , 2009
Rubiaceae	Galium aparine	Betistraw	Diuretic, Dropsies, Gravel and Calculi, Tonic Lymphatic alterative, Anti-inflammatory, Astringent, Anti-neoplastic	Proving Report 2006;
Polygonaceae	Rumex dentatus	Jangli palak	Treatment for pneumonia, Cough, Bscesses, stomach-ache and smallpox	Hussain <i>et al.</i> , 2006; Kumar, 2010
Poaceae	Avena fatua	Jangli jai	Seeds are nerve tonic, stimulant and laxative.	Islam et al., 2006;
Poaceae	Phalaris minor	Dumbi setti	The seed can be ground into a flour and used in making bread, cakes etc.	Moerman, 1998.
Asteraceaae	Carthamus oxycantha	Pohli	Antihyperlipidemic, Antihyperglycemic, Antinephrolithiatic and Hepatoprotective.	Bukhsh <i>et al.</i> , 2007; Jabeen <i>et al.</i> , 2009
Convolvulaceae	Convolvulus arvensis	Lehli	Roots are purgative and possess blood coagulating properties due to presence of vitamin K like substances, Used as animal feed.	Khan, 2004; Gorsi and Rizwan, 2002
Chenopodiaceae	Chenopodium album	Batho	Antipruritic and Antinociceptic.	Yadav et al., 2007; Jabeen et al., 2009; Qureshi, et al., 2009
Chenopodiaceae	Chenopodium murale	Krund	The leaves are used in salads.	Yadav et al., 2007;
Brassicaceae	Coronopus didymus	Jangli halon	Antiallergic, Antipyretic, Hypoglycemic and hepatoprotective.	Qureshi et al., 2009; Busnardo, et al., 2009
Leguminosae	Melilots indica	Zaed sangi	The leaves and flowers are added to bathwater for the external treatment for warts, Rheumatism, and wounds. Ocular infections are also treated.	Shinwari,2000; Jabeen <i>et al.</i> , 2009
Asteraceae	Cichorium intybus	Kasni	liver tonic, cardiotonic, diuretic, stomachic, cholagogue, depurative, emmenagogue, hepatomegaly, cephalalgia, inflammations, anorexia, dyspepsia, flatulence, colic, jaundice, splenomegaly, amenorrhea dysmenorrhea, and asthma, etc.	Sala, 1994; Jabeen et al., 2009
Primulaceae	Anagallis arvensis	Billi boti	Skin diseases: infected wounds.	Akerreta et al., 2007; Jabeen, et al., 2009; Kumar, 2010
Euphorbiaceae	Euphorbia sp.	Chatri dohdhk	Stem pound and mother liquor used (Mtakalang'onyo) to expel retained Placenta.	Komwihangilo <i>et al.</i> , 1995; Guèye,2002

Table 2. Qualitative phytochemical components of seeds of selected weed plants relevant to wheat crop.

Weed Plants	Flavinoids	Saponins	Tannins	Steroids	Glycosides	Alkaloids	Anthrequinones	Terpenoids
Vicia sativa	+	+	+	+	+	+	+	+
Galium aparine	+	+	+	-	-	+	-	+
Rumex dentatus	+	+	+	+	-	+	-	-
Avena fatua	+	+	+	-	+	+	-	+
Lathyrus aphaca	+	-	+	-	+	+	+	+
Phalaris minor	+	+	+	-	+	+	+	+
Carthamus								
oxycantha	+	+	+	-	+	+	+	+
Convolvulus arvensis	+	+	+	-	+	+	+	+
Chenopodium album	+	+	+	+	+	+	-	+
coronopus didymus	+	+	+	+	+	+	+	+
Melilots indica	+	-	+	+	+	+	+	+
Cichorium intybus	+	-	+	+	+	+	+	+
Anagallis arvensis	+	+	+	+	+	+	+	-
Chenopodium murale	+	+	+	+	+	+	-	-
Euphorbia sp.	+	-	+	+	+	+	-	+

Table 3. Qualitative (percent) Phytochemical components of seeds of selected weed plants

Weed Plants	Alkaloids	Saponins	Flavinoids	Tannins
Vicia sativa	2.27±0.14	0.85±0.04	4.62±0.06	10.41±0.08
Galium aparine	2.76 ± 0.03	0.99 ± 0.03	6.36 ± 0.03	16.96 ± 0.01
Rumex dentatus	1.81 ± 0.07	0.56 ± 0.03	10.94 ± 0.05	15.18 ± 0.04
Avena fatua	2.19 ± 0.01	0.87 ± 0.06	3.91 ± 0.08	18.53 ± 0.05
Lathyrus aphaca	4.00 ± 0.04	0.00 ± 0.00	4.93 ± 0.07	15.56 ± 0.03
Phalaris minor	1.21 ± 0.03	0.70 ± 0.04	11.85 ± 0.11	7.97 ± 0.02
Carthamus oxycantha	2.61 ± 0.03	0.48 ± 0.06	4.14 ± 0.28	15.58 ± 0.03
Convolvulus arvensis	2.63 ± 0.07	0.76 ± 0.04	13.88 ± 0.11	20.26 ± 0.06
Chenopodium album	2.12 ± 0.02	0.76 ± 0.04	8.80 ± 0.04	16.09 ± 0.03
coronopus didymus	2.25 ± 0.02	0.74 ± 0.02	15.55 ± 0.13	15.17 ± 0.03
Melilots indica	2.62 ± 0.07	0.00 ± 0.00	13.52 ± 0.09	21.97 ± 0.02
Cichorium intybus	2.13 ± 0.01	0.00 ± 0.00	14.13 ± 0.13	14.53 ± 0.02
Anagallis arvensis	2.21 ± 0.02	1.29 ± 0.03	4.43 ± 0.10	9.56 ± 0.03
Chenopodium murale	0.88 ± 0.06	0.54 ± 0.01	3.92 ± 0.02	24.17 ± 0.02
Euphorbia sp.	2.46 ± 0.07	0.00 ± 0.00	9.20 ± 0.54	20.86 ± 0.05

The biological functions of flavonoiods include protection against inflammatory allergies, free radical scavenging, ulcers, microbes, hepatoxins, platelets aggregation, viruses and tumors (Okwu and Omodamiro, 2005; Okwu and Emenike, 2006). This accounts for the natural antioxidants by acting against oxidative stress related disease such infections as diabetics, cancer and coronary heart diseases (Burits and Bucar, 2002). So the people that are suffering from such infection can feed on wheat weed seeds as source of natural antioxidants. Tannins were reported by many researchers that the molecule has high molecular weight water soluble polycyclic aromatic compounds widely distributed through the plant kingdom and almost found in every plant part and is one of active component of seeds cotyledon (Haslam, 1989; Kaur and Arora, 2009; Okoli and Okere, 2010). Enzo (2007) reported that tannins were also responsible for different anti-diarrheal activities.

Quantitative estimation of percent crude chemical constituents in these wheat weeds determined has been given in Table 3. *C. murale* contained highest percentage of crude tannins (24.17%), *P. minor* contained the lowest yield of tannins (7.97%) but the highest yield of flavinoids was found in *C. didymus* (15.55%) and the lowest was found in *C. murale* (3.92%). *L. aphaca* was found to contain highest yield of alkaloids that is (4.00%) and the lowest yield was found in *C. murale* (0.88%). Alkaloids might be found in all plant parts but tended to accumulate in storage organs of plants. Alkaloids act as stimulators, inhibitors and growth terminators (Rowson, 1958; Waller, 1978; Nazrullaev, *et al.*, 2001)

Saponins were also obtained from the seeds of weeds but the yield recorded were minimal ranging from 1.29% in *A. arvensis* and 0.48% in the *C. oxycantha*. Saponin has been reported as antinutrient, found in different plant parts and in low quantitiy in seeds and

posses both beneficial (i.e. Cholesterol lowering) as well as deleterious properties and reveal structure dependent biological activity (Savage, 1993; Akubugwo *et al.*, 2007).

The analyzed seed cotyledons of weeds contained steroidal compounds which were of importance and interest in pharmacy due to their relationship with compounds as sex hormones (Okwu, 2001). Presence of terpenoids in the studied seeds is in line with with the results of Goto, *et al.* (2010), who stated that Terpenoids were present in many herbal plants, and several terpenoids had been shown to be available for pharmaceutical applications, for example, artemisinin and taxol as malaria and cancer medicines, respectively. Accordingly various terpenoids were available in many plants for not only herbal but also for dietary use.

Conclusion: Seeds from all the fifteen weed plants associated with wheat crops possessed a significant amount of phytochemicals. The presence of phytochemicals like alkaloids, tannins, saponins etc. in these weed plants might be responsible for their curative effects. Moreover, it revealed a strong hope for the development of more novel chemotherapeutic agents from such weed plants which in future may serve for the production of synthetically improved therapeutic agents. Further studies are recommended to explore other parts of weed frequently occurring in our cropland.

Acknowledgement: All the field and laboratory facilities were provided under the Research Project No. 20-813/R and D/HEC. The Authors highly acknowledge Higher Education Commission, Islamabad, Pakistan for funding the same to accomplish this work.

REFERENCES

- Ahmad, M., M. A. Khan and R. A. Qureshi (2003). Ethnobotanical study of some cultivated plants of chhuchh region (District Attock). J. Hamd. Medi. 3: 15-19.
- Ajayi, I. A., O. Ajibade and R. A. Oderinde (2011). Preliminary Phytochemical Analysis of some Plant Seeds. Res.J.Chem.Sci.1(3).
- Akubugwo, I. E., A. N. Obasi and S. Ginika (2007). Nutritional Potential of Leaves and Seeds of Black Nightshade *Solanum nigrum* L. Var *virginicum* from Afikpo-Nigeria. Pakistan J. Nutr. 6: 323-326.
- Aletor, V. A. (1993). Allelochemicals in plant food and feeding stuffs: I. National, biochemical and physiopathological aspects in animal production. Vet. Hum. Toxicol. 35:57-67.
- Baquar, S. R. (1989). Medicinal and Poisonous Plants of Pakistan. Printas Karachi, Pakistan. pp. 343-344.
- Benson, B. B., Y. A. Bekro, J. A. Mamyrbékova-Békro, W. K. Coulibaly and E. E. Ehilé. (2008).

- Assessment of Sexual Stimulant Potential of Total Flavonoids Extracted from Leaves of *Palisota Hirsuta* Thunb. K. Schum (Commelinaceae). European J. Scientific Res. 22: 533-538.
- Boham, B. A. and R. Kocipai-Abyazan (1994). Flavonoids and condensed tannin from leaves of Hawaiian *Vccinium vaticulatum* and *V. calycinium*. Pacific Sci. 48:458-463.
- Burits, M. and F. Bucar (2002). Antioxidant activity of *Chrysphylum albidum* essential oil. Phtother Res. 14:323-328.
- Enzo, A. P. (2007). Traditional plants and herbal remedies used in the treatment of diarrheal disease: Mode of action, quality, efficacy and safety considerations. Modern Phytomedicine Turning Medicinal Plants in to Drugs. WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim. pp. 248-260.
- Goto, T., N. Takahashi, S. Hirai and T. Kawada (2010).

 Various Terpenoids Derived from Herbal and
 Dietary Plants Function as PPAR Modulators
 and Regulate Carbohydrate and Lipid
 Metabolism. Hindawi Publishing Corporation
 PPAR Research. Article ID 483958, p. 9.
- Harborne, J. B. (1973). Phytochemical methods, London. Chapman and Hall, Ltd. pp. 49-188.
- Haslam, E. (1989). Plant Polyphenols, Vegetable Tannins Revisited, Cambridge Univ. Press, U.K.
- Hussain, F., M. Islam and A. Zaman (2006). Ethnobotanical profile of plants of Shawar Valley, District Swat, Pakistan. Int. J. Biol. Biotec. 3: 301-307.
- Iyengar, M.A. (1995). Study of Crude Drugs. 8th ed., Manipal Power Press.
- Kaur, G. J. and D. S. Arora (2009). Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare* and *Trachyspermum ammi*. BMC Complementary and alternative medicine. 9: 30.
- Marjan, N. and H. Hossein (2008). Review of pharmacological effects of *Glycyrrhiza sp.* and its bioactive compounds. Phytotherapy Res. 22:709-24.
- Mojab, F., M. Kamainejad, N. Ghaderi and H. R. Vahidipour (2003). Phytochemical screening of some species of Iranian plants. Iranian J. Pharm. Res. 77-82.
- Mujtaba, G. and M. A. Khan (2007). Check list of medicinal plants of siran valley mansehra-pakistan. Leaflet Quaid-I-Azam University, Islamabad-Pakistan. pp.15.
- Nazrullaev, S. S., I. A. Bessonova and K. S. Akhmedkhodzhaeva (2001). Estrogenic activity as a function of chemical structure in

- Haplophyllum quinoline alkaloids. Chemistry of Natural Compounds. 37:551-555.
- Obdoni, B. O. and P. O. Ochuko (2001). Phytochemical studies and comparative efficacy of the crude extracts of some Homostatic plants in Edo and Delta States of Nigeria. Global J. Pure Appl. Sci. 8b:203-208.
- Okoli, B. J. and O. S. Okere (2010). Antimicrobial activity of the phytochemical constituents of *Chrysophyllum albidum* G. Don_Holl (African Star apple). Journal of research in national development, 8(1).
- Okwu, D. E. (2001). Evaluation of the chemical composition of indigenous spices and flavouring agents. Global J. Pure Appl. Sci. 7: 455-459.
- Okwu, D. E. and I. N.Emenike (2006). Evaluation of the phyto-nutrients and vitamins content of the citrus fruit. International J. Molecular Med. Advan. Sci. 2:1-6.
- Okwu, D. E. and O. D. Omodamiro (2005). Effect of hexane extract and phytochemical content of *Xylopia aethiopica* and *Ocimum gratissium* on uterus of Guinea pig. Bio-Rese. 3: 40-44.
- Oliver-Bever, B. (1986). Medicinal plants in tropical West Africa. Cambridge University press, Cambridge. pp. 123-125.
- Rashid, A. and M. Arshad (2002). Medicinal plant diversity, threat imposition and interaction of a mountain people community. In Proceeding of Workshop on Curriculum Development in Applied Ethnobotany. Published by the Ethnobotany Project, WWF Pakistan, 34-D/2, Sahibzada Abdul Qayuum Road Peshawar, Pakistan. Pp. 84-90.
- Rogar, G. D. P. (2002). Encyclopedia of Medicinal plants (vol 1). Education and health. Library editorial safeliz S.L Spain. pp.153-154, 265-267.
- Roopashree T. S., R. Dang, R. H. S. Rani and C. Narendra (2008). Antibacterial activity of

- antipsoriatic herbs: *Cassia tora*, *Momordica charantia* and *Calendula officinalis*. International J. App. Res. Nat.1 Prod. 1: 20-28.
- Rowson, J. M. (1958). Alkaloids and plant taxonomt. Proceedings of the linnean society of London. 169: 212–216.
- Savage, G.P. (1993). Saponins. In: Encyclopedia of Food Science, Food Technology and Nutrition. R. Macrae, R.K. Robinson and M.J. Sadler (eds) Academic Press 24/28 Oval Road, London NW17DX. pp. 3998-4001.
- Schuier, M., H. Sies, B. Illek and H. Fischer (2005). "Cocoa-related flavonoids inhibit CFTR-mediated chloride transport across T84 human colon epithelia". J. Nutr. 135: 2320–5.
- Sharma, V. K., A. Bhattacharya, A. Kumar and H. K. Sharma (2007). Health Benefits of Tea Consumption. Tropical J. Pharmaceutical Res. 6: 785-792.
- Siddiqui, A. A. and M. Ali (1997). Practical Pharmaceutical chemistry. Ist ed., CBS Publishers and Distributors, New Delhi. Pp. 126-131.
- Sofowara, A. (1993). Medicinal plants and Traditional medicine in Africa. Spectrum Books Ltd, Ibadan, Nigeria. p. 289.
- Stauth, D. (1993). Studies force new view on biology of flavonoids", EurekAlert. Adapted from a news release issued by Oregon State University. URL accessed.
- Trease, G. E. and W.C. Evans (1989). Pharmacognsy. 11th edn. Brailliar Tiridel Can. Macmillian publishers.
- Van-Burden, T. P. and W.C. Robinson (1981). Formation of complexes between protein and tannin acid. J. Agric. Food Chem. 1:77.
- Waller, G. R. (1978). Alkaloid Biology and Metabolism in Plants Springer.