

Fagron

genomics

Gene Comprehensive Nutrigenomic Report

Accession Number: #####

Specimen Collected: ###/###/####

Specimen Received: ###/###/####

Report Generated: February 27, 2025

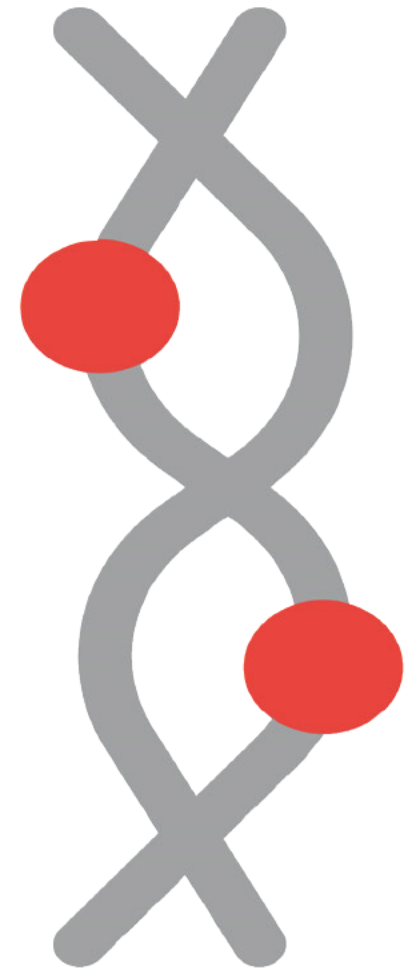
Specimen Type: Buccal Swab

Provider: #####

Patient Name: #####

Patient DOB: ###/###/####

Patient Gender: Female



Do not make any decisions about your health solely based on the information contained in this report.
Always consult with a licensed and experienced health practitioner when you receive this report.

– 18 – Female

(-/-) Normal Risk

(-/+) Medium Risk

(+/+) High Risk

rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
DEVELOPMENTAL							
Methylation							
	FOLR2	A/G (+/-)	Methyltetrahydrofolate (5-MTHF)	L-5-MTHF OR Homocysteine Supreme™			Complete Blood Count Serum and RBC Folate
	MTHFR 1298	G/T (+/-)					
	MTHFR 677	G/A (+/-)					
	TCN2	C/G (+/-)	Methylcobalamin, Adenosylcobalamin		Tricobalamin™ OR Vitamin B12		Serum Vitamin B12

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rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
Neurotransmitters							
	COMT	G/G (-/-)	Taurine, Choline, Trimethylglycine (TMG), Dimethylglycine (DMG), Methionine, SAMe, Inositol	NeuroLink™ if Focus Issues Present			Consider Neurotransmitter Metabolite Testing Consider PGx Testing
	MAOA	G/T (+/-)	Riboflavin (B2), Taurine, Choline, Trimethylglycine (TMG), Dimethylglycine (DMG), Methionine, SAMe, Inositol, L-Methionine		NeuroLink™ if Anxiety, Depression, or Focus Issues Present NeuroCalm™ if Chronic Anxiety or Depression Present		
	MAOB	T/C (+/-)					
	TPH2	G/G (-/-)	L-5-Methyl THF, Niacinamide, 5-HTP				
	HTR2	G/G (+/+)	5-HTP (Hydroxytryptophan)		May Benefit from NeuroCalm™ if Anxiety or Depression Present		
	SLC6A4	C/C (-/-)					

– 18 – Female

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rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
Neurotransmitters							
	DBH	G/G (-/-)	Vitamin C, Copper				
	GAD1	C/C (-/-)	Prescription Amantadine, Ketamine, Glycine, N-Acetyl-Cysteine (NAC), Zinc, Magnesium, Elderberry, L-Theanine, Melatonin	Liposomal NeuroCalm™ May Benefit from Melatonin if Sleep Is an Issue		May Need to Avoid Monosodium Glutamate (MSG) Be Cautious with Glutamine Supplementation	Consider Neurotransmitter Metabolite Testing
	GAD1	C/C (+/+)					
Sugar Sensitivity and Mood							
	ADRA2A	C/C (-/-)	Increased Risk of Hyperactivity From Sugar and Anti-Psychotic Induced Weight Gain				
Neurotrophic Factors							
	BDNF	C/C (-/-)	Curcumin, Lithium Orotate, D-Chiro-Inositol, Catechins, Resveratrol, Exercise				
	NGF	G/G (-/-)	Curcumin, Lithium Orotate, D-Chiro-Inositol, Catechins, Resveratrol, Exercise				
	SYN1	A/G (+/-)	Ginsenoside Rg3, Nicotinamide Riboside, Ginseng				

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(-/-) Normal Risk (-/+) Medium Risk (+/+) High Risk

rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
Neuro-Inflammation							
	C3	C/C (-/-)	Anti-Inflammatory Therapy: Curcumin, Omega-3 Fatty Acids, Resveratrol, Quercetin, Low Dose Naltrexone (LDN), CBD Oil	SPM Supreme™ C3 Curcumin Complex OmegaAvail™ Liquid		Consider Low Inflammatory Diet	
	CD14	G/G (-/-)					
	IL5	A/G (+/-)					
	IL13	C/T (+/-)					
	STAT4	C/G (+/-)					
	IL6	C/G (+/-)					
	TNF	G/G (-/-)					
	IDO1	C/T (+/-)					
	CTLA4	A/A (-/-)	Increased Efficacy of Naltrexone				
	DRD2	C/C (-/-)					

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(+/+) High Risk

rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
Autophagy							
	ATG5	T/T (-/-)	Curcumin, Lithium Orotate, D-Chiro-Inositol, Catechins, Resveratrol, Caffeine, 12+ Hour Fasting		Inositol OR Inositol Powder	12-15 Hour Fasting if Appropriate for Age	Routine Blood Sugar, Insulin, and HbA1c
	ATG12	C/C (+/+)					
	ATG16L1	C/T (+/-)					
Detoxification							
	CTH	G/G (-/-)	N-Acetyl Cysteine (NAC), Glutathione				
	GSTP1	A/A (-/-)	N-Acetyl Cysteine (NAC), Glutathione				

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rsID	Gene	Genetic Result	Therapeutics Associated With Positive Result	Highly Recommended Therapeutics	Provider Discretion: As Needed Formula Recommendations	Lifestyle Recommendations	Laboratory Recommendations
Environmental Inflammation							
	AOC1	C/T (+/-)	Poor Ability to Break Down Histamine in Foods		HistaGest-DAO™ if Histamine Response Presents after Eating	May Have Difficulty Digesting Foods Containing Histamine	
	HNMT	C/C (-/-)					
	FUT2	A/G (+/-)	Prebiotics and Probiotics Needed		ProBioMed™ Kids IgGI Shield™ if Intestinal Inflammation Is an Issue	Consider Consumption of Prebiotic and Probiotic Foods	Microbiome Testing, such as GI Spotlight , if Digestive Disorders or GI Inflammation Present
	HLA DQA1	C/C (-/-)	High Risk of Gluten and Casein Sensitivity, Broad Spectrum Enzyme				
	HLA DQB1	T/T (-/-)					

Summary for Developmental

Highly Recommended Therapeutics

Provider Discretion: As Needed Formula Recommendations

Lifestyle Recommendations

Laboratory Recommendations

Methylation

- L-5-MTHF OR Homocysteine Supreme™

- Tricobalamin™ OR Vitamin B12

- Complete Blood Count
- Serum and RBC Folate
- Serum Vitamin B12

Neurotransmitters

- Neurolink™ if Focus Issues Present

- Neurolink™ if Anxiety, Depression, or Focus Issues Present
- NeuroCalm™ if Chronic Anxiety or Depression Present
- May Benefit from NeuroCalm™ if Anxiety or Depression Present

- Consider Neurotransmitter Metabolite Testing
- Consider PGx Testing

- Liposomal NeuroCalm™
- May Benefit from Melatonin if Sleep Is an Issue

- May Need to Avoid Monosodium Glutamate (MSG)
- Be Cautious with Glutamine Supplementation

Neurotrophic Factors

- May Benefit from Synapsin Nasal Spray

Neuro-Inflammation

- SPM Supreme™
- C3 Curcumin Complex
- OmegaAvail™ Liquid

- Consider Low Inflammatory Diet

Autophagy

- Inositol OR Inositol Powder

- 12-15 Hour Fasting if Appropriate for Age

- Routine Blood Sugar, Insulin, and HbA1c

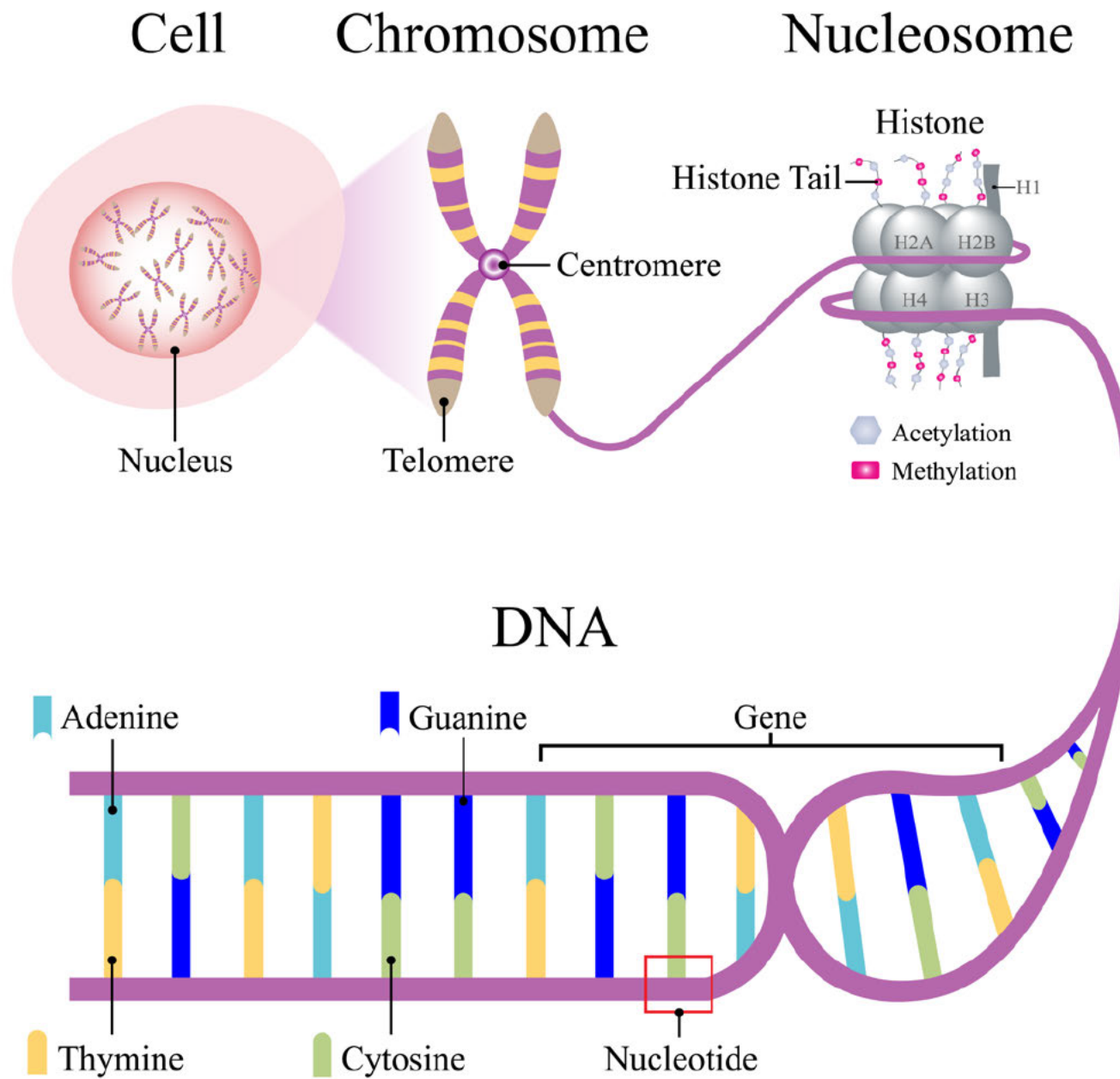
Detoxification

Environmental Inflammation

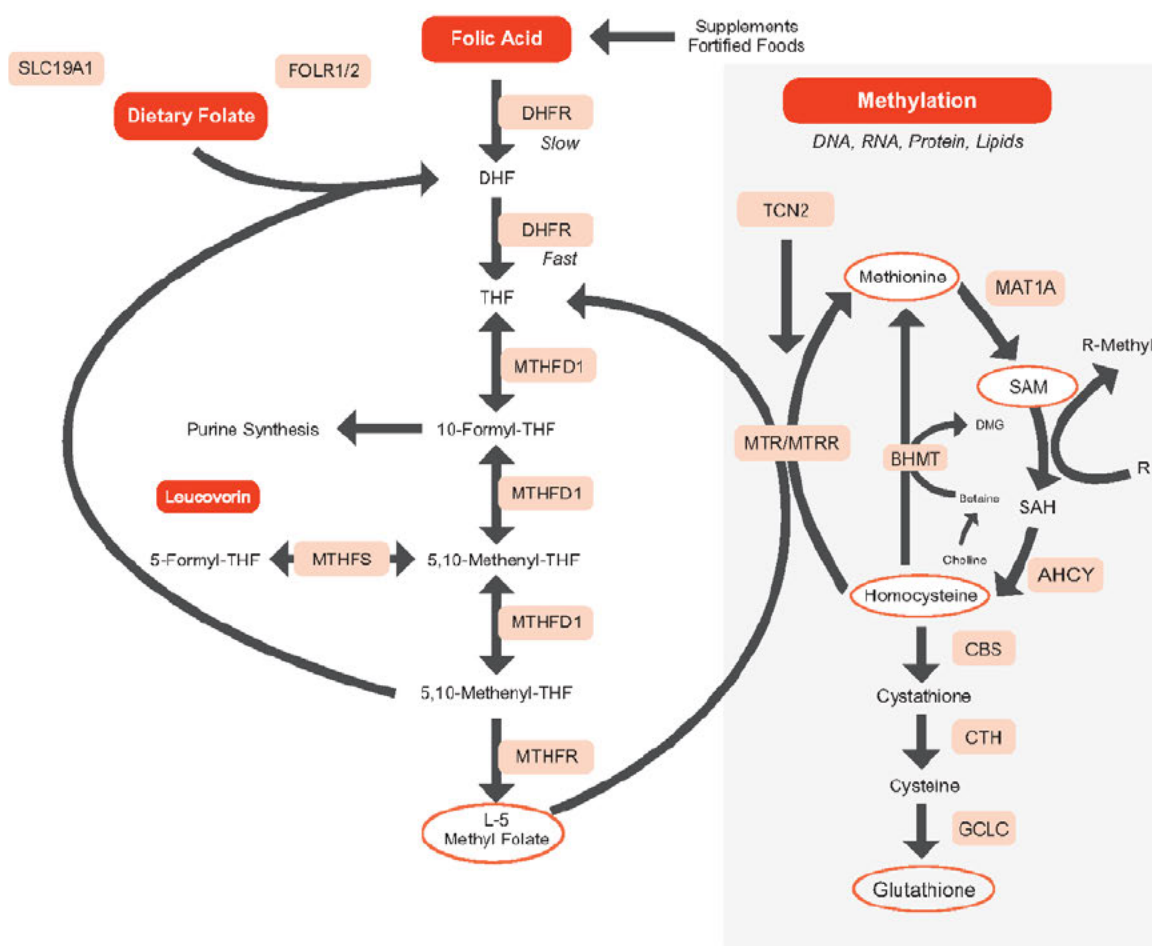
- HistaGest-DAOTM if Histamine Response Presents after Eating
- ProBioMed™ Kids
- IgGI Shield™ if Intestinal Inflammation Is an Issue

- May Have Difficulty Digesting Foods Containing Histamine
- Consider Consumption of Prebiotic and Probiotic Foods

- Microbiome Testing, such as GI Spotlight, if Digestive Disorders or GI Inflammation Present



METHYLATION



Methylation

- Involves the addition of a methyl group (CH₃)
- Regulates gene expression and repression
- Reduces or removes toxins that eliminate essential nutrients
- Provides nutrients needed for processes such as detoxification, immune regulation, gut health

Methionine

- Used in protein formation and stabilization
- Elevated levels are associated with risk for coronary heart disease, stroke & neurological diseases

Glutathione

- Important for chemical detoxification & proper mitochondrial functioning
- Genes relevant for production include: AHCY, CTH, CGTP1, GSTM1, GSTM3, GSR, MTRR & MTR

5-Methyl Folate

- Important for dopamine and serotonin formation, detoxification and mitochondrial strength
- Genes relevant for production include: DHFR, FOLR1/2, MTHFD1, MTHFR, MTHFS

Homocysteine

- Elevated levels are associated with risk for coronary heart disease, stroke, neurological diseases
- Variants in the methylation pathway can be associated with increased/decreased levels

FOLATE

FOOD SOURCES



Eggs



Citrus Fruits



Leafy Greens



Legumes



Broccoli



Brussel Sprouts



Breads, Cereal,
Pastas, Rice



**ABSORBABLE
FOLATE**
NATURAL B9



**5-METHYL
FOLATE**
ACTIVE FORM

FUNCTIONS (OR BENEFITS AS YOU AGE)

- Maintains structure & function of proteins
- Maintains structure & function of DNA
- Facilitates DNA replication, neurotransmitter production & detoxification

DEFICIENCY CAUSES

- Neural tube defects
- Cardiovascular disease
- Memory problems
- Depression
- Insomnia
- Irritability

VITAMIN B12

FOOD SOURCES



Eggs



Seafood – clams, trout, salmon, tuna



Meats – liver, beef, ham, chicken



Fortified Foods



Swiss cheese



Nori/Seaweed



Nutritional Yeast



**ABSORBABLE
VITAMIN B12**



**METHYLCO-
BALAMIN**
ACTIVE FORM

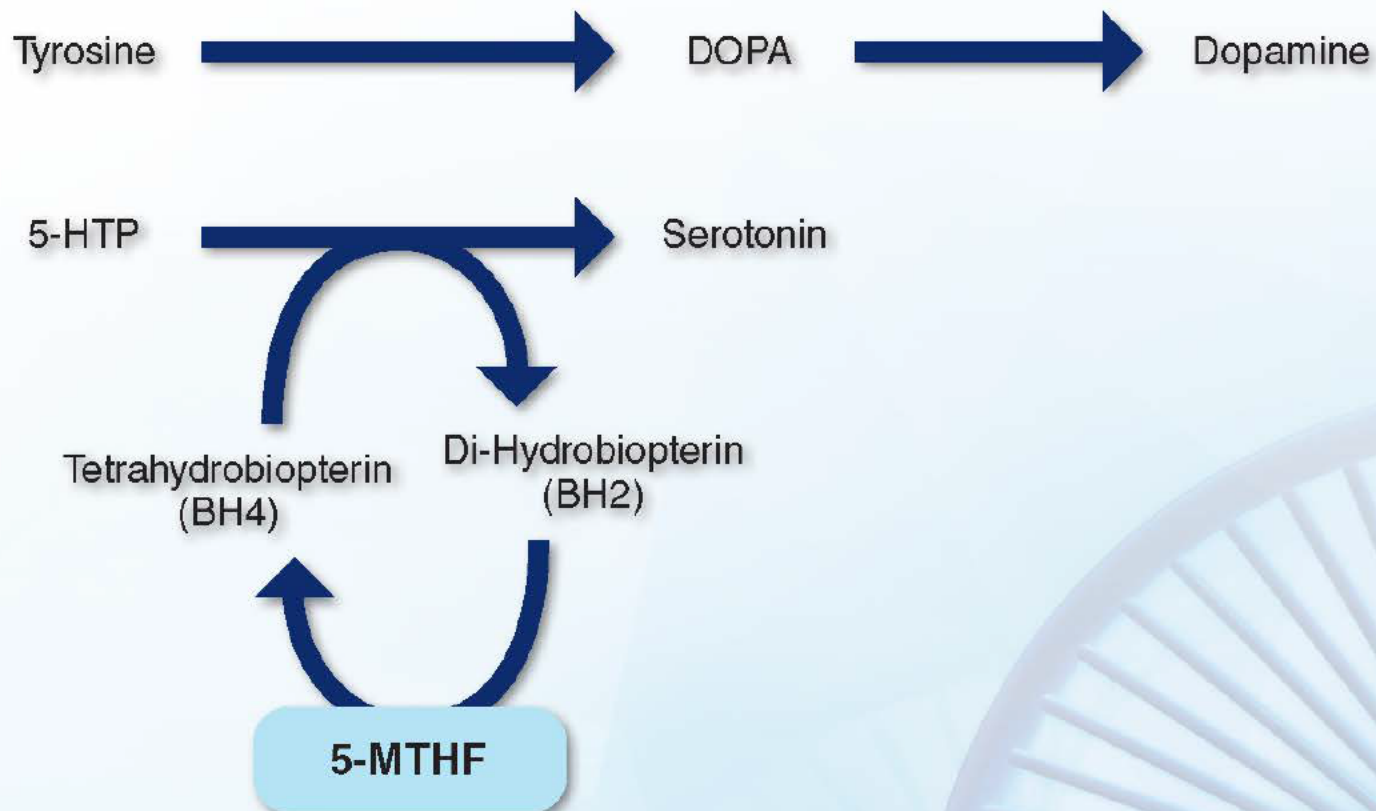
FUNCTIONS (OR BENEFITS AS YOU AGE)

- Formation and maintenance of red blood cells (RBCs)
- Facilitates DNA synthesis
- Regulates homocysteine levels (decreases)
- Facilitates neurological functioning

DEFICIENCY & PATHWAY ALTERATIONS

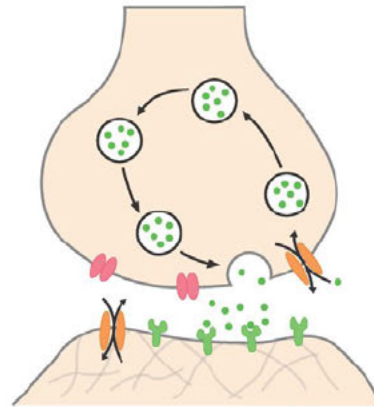
- Increased production of homocysteine
- Decreased breakdown of homocysteine
- Circadian Rhythm Problems
- Cancers
- Memory-related disorders
- Cardiovascular diseases
- Fatigue
- Poor balance
- Mood disorders

5-MTHF & Neurotransmitter Production



NEUROTRANSMITTERS & PATHWAY

TRANSMIT INFORMATION FOR ESSENTIAL PROCESSES SUCH AS DIGESTION, BREATHING, HEARTBEAT, MOVEMENT, PAIN REGULATION ETC.



RELEVANT GENES

- **HTR2, TPH2, SLC6A4, MAO-A** genes are important in the synthesis, breakdown, transport and/or functioning of serotonin
- **COMT, MAO-A, MAO-B** genes are important for the breakdown of serotonin, norepinephrine and/or dopamine
- The **DBH** gene is important for norepinephrine synthesis
- The **GAD1** gene is important for GABA synthesis
- Variants in **COMT, MAO-A, MAO-B** and **GAD1** genes have been associated with mood, anxiety and focus issues

WAYS TO INCREASE LEVELS



Aerobic Exercise



Dietary Factors



Mediation/Yoga



Increase Sun Exposure

NEURO-INFLAMMATION

WHAT IS IT?

- Inflammation of the brain and spinal cord
- Main causes: disease, injury, infection, acute/chronic stress
- Variants have been associated with increased inflammatory aggression and the inability to “shut down” neuro-inflammation
 - Interleukins (IL-1B, IL5, IL6, IL13) stimulate the immune response
 - C3 & STAT4 activate, form and/or differentiate T-cells
 - CTLA4 & CD14 are involved in the suppression of T-cells
 - TNF triggers inflammation
 - DRD2 suppresses neuroinflammation

WAYS TO DECREASE NEUROINFLAMMATION


Meditation & breathing exercises

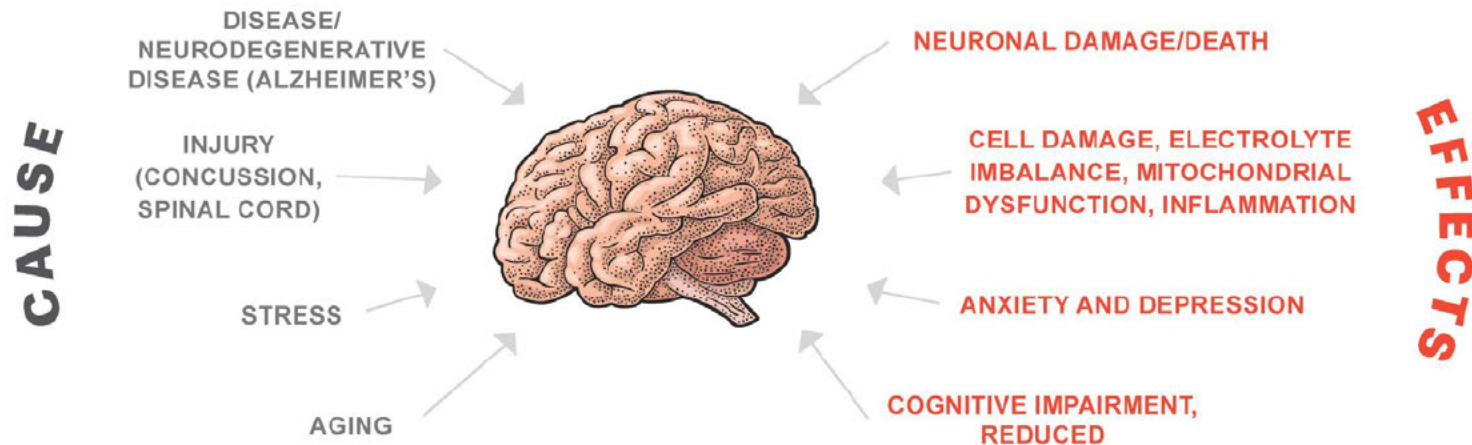

Therapeutic massages with herbalized oils (ex. Sesame oil)


Mediterranean Diet


Yoga


Curcumin, Bacopa herb


Anti-inflammatory Diet



NEUROTROPHIC FACTORS

VARIANTS IN THE SYN1, NGF & BDNF GENES CAUSE
DECREASED NEURON SYNTHESIS



Promote growth, development, survival, synaptic plasticity (strengthening) and repair of neurons



Regulate the development of the peripheral and central nervous systems



Regulate the formation of long-term memories

LOW LEVELS ARE CORRELATED WITH

- Neurodegenerative disorders
- Aging
- Chronic stress
- Mood disorders

WAYS TO INCREASE LEVELS



Exercise (physical or cognitive)



Social interactions

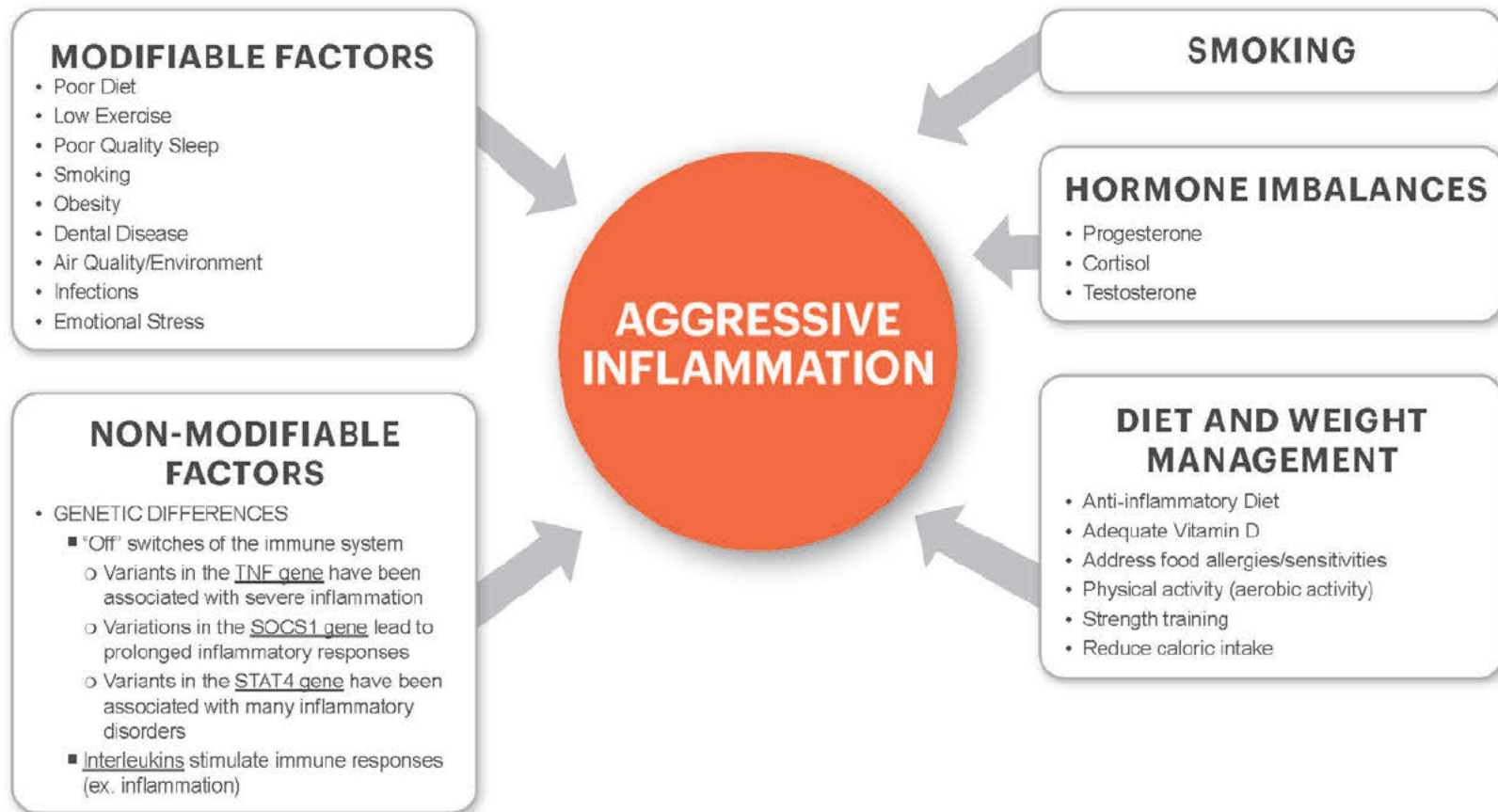


Reduce stress via breathing exercises and/or meditation

ANTI-INFLAMMATORY

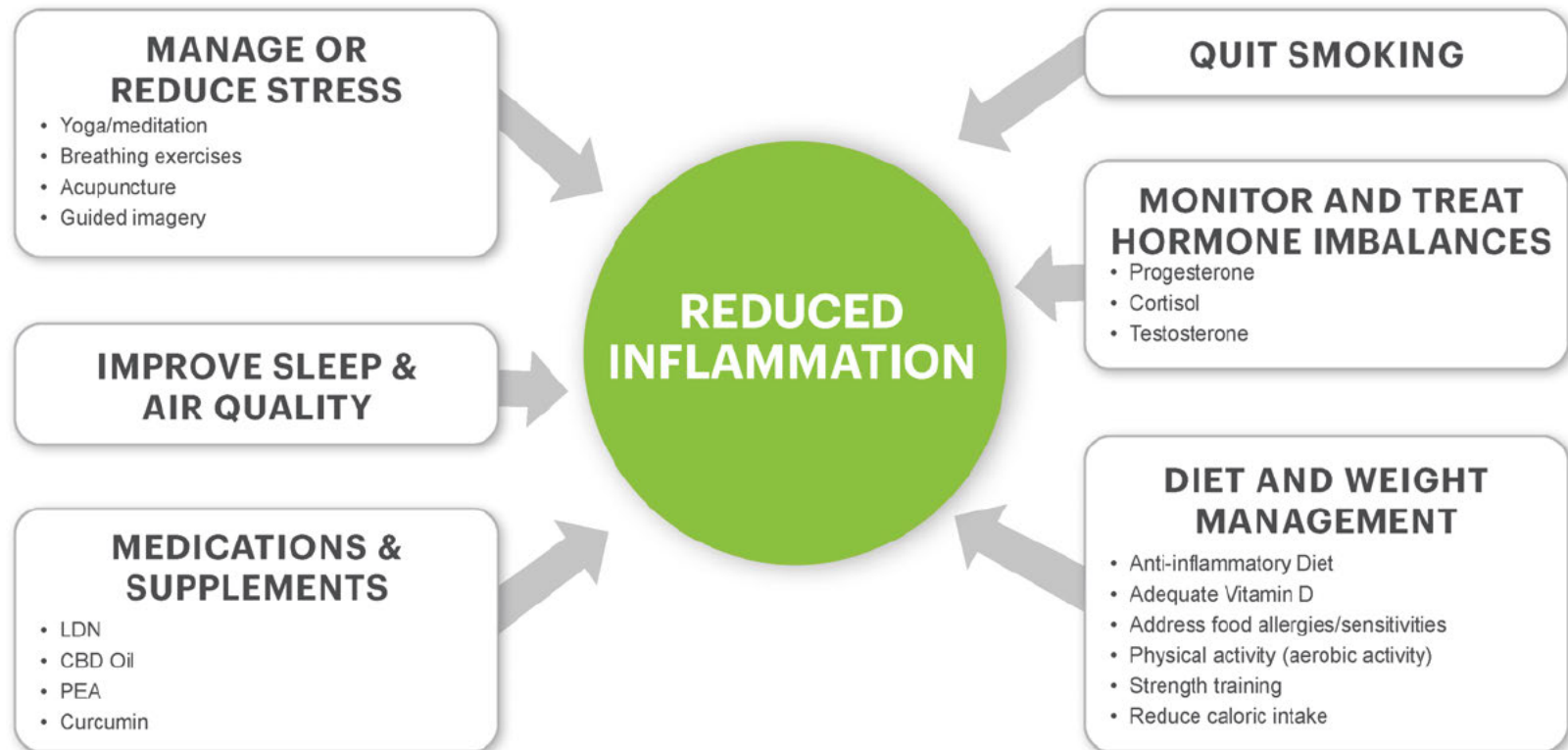
AN IMMUNE SYSTEM RESPONSE TRIGGERED BY HARMFUL STIMULI
(EX. PATHOGENS, DAMAGED CELLS, TOXIC COMPOUNDS, IRRADIATION)

DRIVERS OF INFLAMMATION



ANTI-INFLAMMATORY

WAYS TO REDUCE INFLAMMATION



THE IMMUNE SYSTEM & AUTOIMMUNITY

WHAT DOES THE IMMUNE SYSTEM DO?

Prevent or limit infections by distinguishing between healthy and unhealthy cells

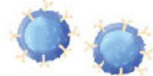
KEY PLAYERS & RELEVANT GENES



CYTOKINES

(ex. IL family, TNF- α)

- Helps with immune cell growth, activation, and function
- Interleukins (IL2, IL4, IL5, IL6, IL13, IL23R, IL2RA) stimulate the immune response
- SOCS1 & TNF are involved in cytokine signaling for the inflammatory response



LYMPHOCYTES

(ex. B, T & Natural Killer cells)

- Identify & kill infected cells
- Produces antibodies to fight future infections
- IDO1, CTLA4 & CD14 are involved in the suppression of T-cells
- C3, STAT4 & TRAF1 activate, form and/or differentiate T-cells

IMMUNE AGGRESSION

The immune system begins to attack healthy tissue

COMMON SYMPTOMS



Fatigue



Hair loss



Achy muscles



Inflammation



Skin rashes



Pain



Low-grade fever



Numbness and tingling in hands and feet



Trouble concentrating

MALFUNCTIONS LEAD TO

- Chronic inflammation
- Allergic reactions
- Immune aggressive diseases (Inflammatory bowel disease, skin & neurological disorders)

LOW-INFLAMMATORY

FOODS TO EAT



Fruits: strawberries, blueberries, cherries, oranges



Fatty fish: salmon, mackerel, tuna, sardines



Spices - turmeric, ginger



Green leafy vegetables & tomatoes



Dark chocolate



Olive oil



LOW-INFLAMMATORY DIET

FOODS TO AVOID



Soda & other sugar-sweetened drinks



Dairy products



Fried foods



Red & Processed meats (hotdogs, sausage)



Refined carbohydrates: white bread, pastries



Margarine, shortening, lard

BENEFITS



Reduces inflammation



Reduces risk for cardiovascular disease & Type II diabetes

DETOXIFICATION

GLUTATHIONE IN DETOXIFICATION

Relevant genes for production are AHCY, CTH, GSTP1, GSTM1, GSTM3, GSR, MTRR & MTR

WHY IS IT IMPORTANT?



Maintains health by protecting the body from toxins



Regulates cell production and programmed cell death



Critical role in chemical detoxification



Vital for proper mitochondrial function

DEFICIENCY CAUSES

- Auto-immune diseases
- Cardiovascular diseases
- Neurodegenerative diseases
- Cell death
- Poor mitochondrial function



WAYS TO INCREASE GLUTATHIONE

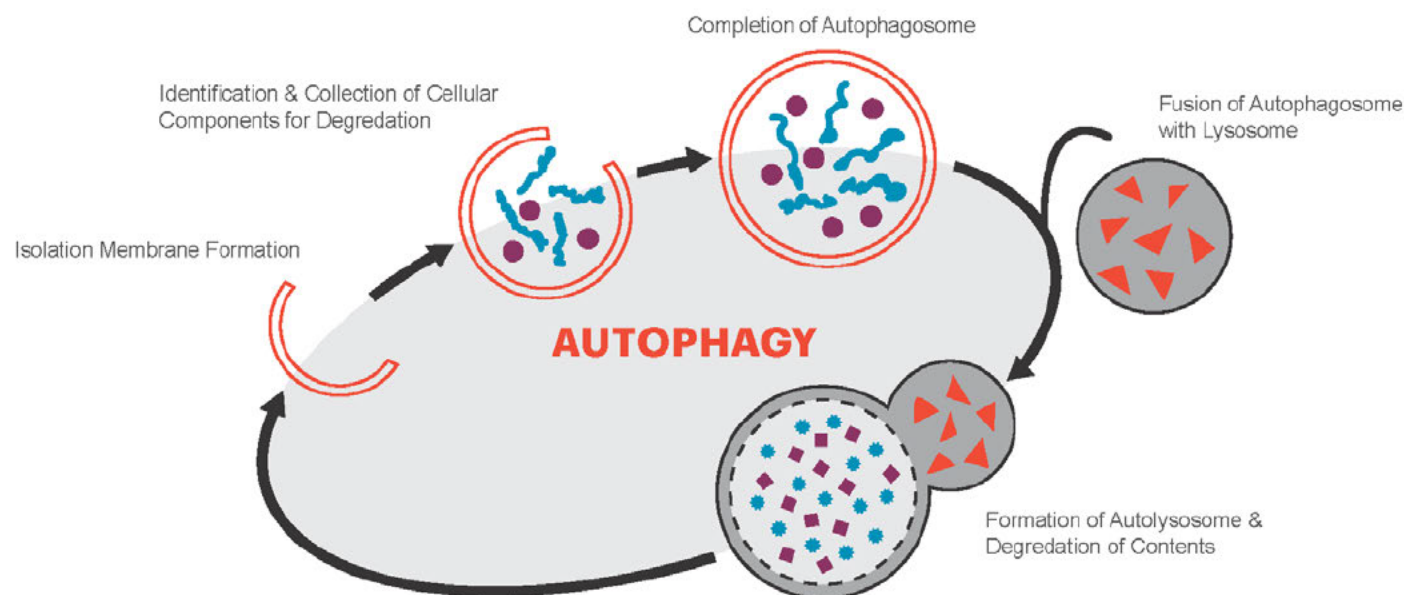
- Limit alcohol intake
- N-acetyl-cysteine (NAC)
- Glutathione therapies
- (ie. IV Glutathione, Glutathione suppository, Liposomal Glutathione)
- Include whey in diet, unless allergic or intolerant
- Methylation Support - if necessary

SUPEROXIDES & ANTIOXIDANTS

- SOD1, SOD2, SOD3 genes are important to transform superoxides to protect against mitochondrial damage
- Reactive Oxygen Species (ROS) can damage mitochondria and cause cell death.
- Antioxidants such as Vitamin A, Vitamin C and Vitamin E act as a defense against ROS

AUTOPHAGY: Cellular Housekeeping

VARIANTS IN THE ATG GENES HAVE BEEN ASSOCIATED WITH
CELLULAR BLOCKAGE



DEFECTS LEAD TO:

- Neurodegenerative Diseases
- Aging
- Heart Disease
- Developmental Disorders
- Type II Diabetes
- Insulin Resistance
- Fatty Liver
- Cancers

WAYS TO INCREASE

-  Intermittent fasting or low-calorie diet
-  Routine Exercise
-  Ketogenic diets (high fat, low carbs)
-  Medications & Supplements
D-Chiro Inositol (B8)
Metformin

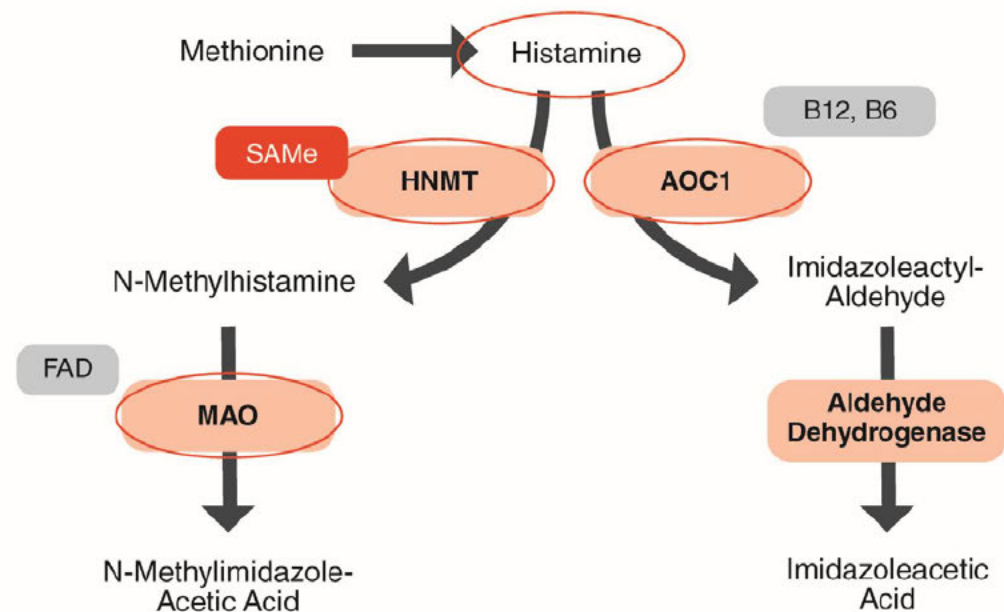
HISTAMINE

HISTAMINE

- Natural substance found in various foods

IMPLICATIONS

- Metabolic Enzymes: amine oxidases (ex. AOC1, MAO, DAO) & HNMT
- High histamine & low amine oxidase activity is associated with:
 - Diarrhea
 - Headaches
 - Nose congestion
 - Asthma
 - Hypotension
 - Arrhythmia
 - Flushing
 - Urticaria (hives)
 - Pruritus (itchy skin)
- Dietary histamine can be rapidly detoxified by amine oxidases, whereas persons with low amine oxidase activity are at risk of histamine toxicity



AOCI & HNMT POLYMORPHISM HISTAMINE

LOW HISTAMINE LEVEL FOODS



Meats & Fish
fresh meat (ex. chicken,
turkey, pork and red
meat), fresh fish (ex. hake,
trout, plaice)



Milk substitutes
(Coconut milk, rice milk)



Cream cheese, butter



Most cooking oils



**Most leafy greens
and herbs**



Beverages
(non-citric fruit juices,
herbal teas)



Fresh fruits
(with the exception of
strawberries)



Fresh vegetables



Grains



HIGH HISTAMINE LEVEL FOODS



Egg whites



**Processed, cured,
smoked and fermented
meats/fish** (lunch meat,
bacon, sausage, pepperoni,
canned tuna)



Leftover meat
(After meat is cooked, the
histamine levels increase due to
microbial action as the meat sits)



Dairy products: All fermented
milk products (ex. aged cheeses,
yogurt, buttermilk, kefir)



Chocolate, cocoa



Bone broth



Fruits (oranges, grapefruit,
lemons, lime, berries, dried fruit)



Vegetables (spinach,
tomatoes, eggplant)



**Artificial food colors and
preservatives**



**Fermented & vinegar-
containing foods**
(sauerkraut, kombucha, pickles,
relishes, ketchup, prepared
mustard)



Spices (cinnamon, chili
powder, cloves, nutmeg, curry
powder, cayenne)



Beverages (Black Tea, alcohol)

Gene Information Key

rsID	Gene	"_" variant	"+" variant
	ADRA2A	C	G
	AOC1	C	T
	ATG12	T	C
	ATG16L1	C	T
	ATG5	T	C
	BDNF	C	T
	C3	C	T
	CD14	G	A
	COMT	G	A
	CTH	G	T
	CTLA4	A	G
	DBH	G	A
	DRD2	C	A
	FOLR2	A	G
	FUT2	A	G
	GAD1	C	T
	GAD1	G	C
	GSTP1	A	G
	HLA-DQA1	C	T
	HLA-DQB1	T	C

rsID	Gene	"_" variant	"+" variant
	HNMT	C	T
	HTR2	A	G
	IDO1	T	C
	IL13	C	T
	IL5	A	G
	IL6	C	G
	MAOA	T	G
	MAOB	C	T
	MTHFR:	T	G
	MTHFR:	G	A
	NGF	G	A
	SLC6A4	C	A
	STAT4	C	G
	SYN1	A	G
	TCN2	C	G
	TNF	G	A
	TPH2	G	T

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Disclaimers

TESTING:

Testing Performed By: AC

METHODOLOGY AND LIMITATIONS DISCLAIMER:

Testing for genetic variation/mutation on listed genes was performed using ProFlex PCR and Real-Time PCR with TaqMan® allele-specific probes on the QuantStudio 12K Flex. All genetic testing is performed by GX Sciences, LLC d/b/a Fagron Genomics US ("Fagron Genomics US") (807 Las Cimas Pkwy, Suite 145, Austin, TX. 78746). This test will not detect all the known alleles that result in altered or inactive tested genes. This test does not account for all individual variations in the individual tested. Test results do not rule out the possibility that this individual could be a carrier of other mutations/variations not detected by this gene mutation/variation panel. Rare mutations surrounding these alleles may also affect our detection of genetic variations. Thus, the interpretation is given as a probability. Therefore, this genetic information shall be interpreted in conjunction with other clinical findings and familial history for the administration of specific nutrients. Patients should receive appropriate genetic counseling to explain the implications of these test results. Details of assay performance and algorithms leading to clinical recommendations are available upon request. The analytical and performance characteristics of this laboratory developed test (LDT) were determined by Fagron Genomics US's laboratory (Laboratory Director: James Jacobson, PhD) pursuant to Clinical Laboratory Improvement Amendments (CLIA) requirements (CLIA #: 45D2144988).

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If you have received the result variant Undetermined (UND) this indicates that we were not able to determine your carrier status based on your raw data. You may request your sample to be run again by emailing info@fagrongenomicsus.com

Fagron Genomics US SNP References

FOR2

• Henderson, G. B. Folate-binding proteins. *Annu. Rev. Nutr.* (1990). doi:10.1146/annurev.nutr.10.1.319 • Ragoussis, J., Senger, G., Townsland, J. & Campbell, I. G. Genomic organization of the human folate receptor genes on chromosome 11q13. *Genomics* (1992). doi:10.1016/S0888-7543(05)80236-8 • Freisheim, J. H., Price, E. M. & Ratnam, M. Folate coenzyme and antifolate transport proteins in normal and neoplastic cells. *Adv. Enzyme Regul.* (1989). doi:10.1016/0085-2571(89)90091-5 • Ratnam, M., Marquardt, H., Duhring, J. L. & Freisheim, J. H. Homologous membrane folate binding proteins in human placenta: cloning and sequence of a cDNA. *Biochemistry* (1989). • Shen, F., Ross, J. F., Wang, X. & Ratnam, M. Identification of a novel folate receptor, a truncated receptor, and receptor type beta in hematopoietic cells: cDNA cloning, expression, immunoreactivity, and tissue specificity. *Biochemistry* (1994). doi:10.1021/bi00171a021 • Page, S. T., Owen, W. C., Price, K. & Elwood, P. C. Expression of the Human Placental Folate Receptor Transcript is Regulated in Human Tissues. *Journal of Molecular Biology* 229, 1175–1183 (1993). • Suzuki, Y., Yoshitomo-Nakagawa, K., Maruyama, K., Suyama, A. & Sugano, S. Construction and characterization of a full length-enriched and a 5'-end-enriched cDNA library. *Gene* (1997). doi:10.1016/S0378-1119(97)00411-3 • Nakashima-Matsushita, N. et al. Selective expression of folate receptor beta and its possible role in methotrexate transport in synovial macrophages from patients with rheumatoid arthritis. *Arthritis Rheum.* (1999). doi:10.1002/1528-0131(199908)42:8<1608::AID-ANR7>3.0.CO;2-L • Strausberg, R. L. et al. Generation and initial analysis of more than 15,000 full-length human and mouse cDNA sequences. *Proc. Natl. Acad. Sci. U. S. A.* (2002). doi:10.1073/pnas.242803899 • Link, R. 15 Healthy Foods That Are High in Folate (Folic Acid). *Healthline* (2020). Available at: <https://www.healthline.com/nutrition/foods-high-in-folate-folic-acid#8.-Citrus-fruits>. • Office of Dietary Supplements - Folate. NIH Office of Dietary Supplements (2020). Available at: <https://ods.od.nih.gov/factsheets/Folate-HealthProfessional/> • 5-Methyl-tetrahydrofolate. National Center for Biotechnology Information. PubChem Compound Database (2004). Available at: <https://pubchem.ncbi.nlm.nih.gov/compound/135483968>. • Scaglione, F. & Panzavolta, G. Folate, folic acid and 5-methyltetrahydrofolate are not the same thing. *Xenobiotica* (2014). doi:10.3109/00498254.2013.845705 • Moore, L. D., Le, T. & Fan, G. DNA methylation and its basic function. *Neuropsychopharmacology* (2013). doi:10.1038/npp.2012.112 • Tanaka, T. et al. Genome-wide Association Study of Vitamin B6, Vitamin B12, Folate, and Homocysteine Blood Concentrations. *Am. J. Hum. Genet.* (2009). doi:10.1016/j.ajhg.2009.02.011 • Abu Seman, N., Wan Mohamad, W. N., Ostenson, C. G., Brismar, K. & Gu, H. F. Increased dna methylation of the slc30a8 gene promoter is associated with type 2 diabetes in a malay population. *Clin. Epigenetics* (2015). doi:10.1186/s13148-015-0049-5 • Ducker, G. S. & Rabinowitz, J. D. One-Carbon Metabolism in Health and Disease. *Cell Metabolism* (2017). doi:10.1016/j.cmet.2016.08.009 • Voutilainen, S., Rissanen, T. H., Virtanen, J., Lakka, T. A. & Salonen, J. T. Low dietary folate intake is associated with an excess incidence of acute coronary events: The Kuopio Ischemic Heart Disease Risk Factor Study. *Circulation* (2001). doi:10.1161/01.CIR.103.22.2674

MTHFR

• Donnelly, J. G. (2000). The 1298(A→C) mutation of methylenetetrahydrofolate reductase should be designated to the 1289 position of the gene. *American Journal of Human Genetics*, 66(2), 744–745. <https://doi.org/10.1086/302784> • Ducker, G. S., & Rabinowitz, J. D. (2017). One-Carbon Metabolism in Health and Disease. *Cell Metabolism*, 25(1), 27–42. <https://doi.org/10.1016/j.cmet.2016.08.009> • Frosst, P., Blom, H. J., Milos, R., Goyette, P., Sheppard, C. A., Matthews, R. G., Boers, G. J., den Heijer, M., Kluijtmans, L. A., & van den Heuvel, L. P. (1995). A candidate genetic risk factor for vascular disease: A common mutation in methylenetetrahydrofolate reductase. *Nature Genetics*, 10(1), 111–113. <https://doi.org/10.1038/ng0595-111> • Ogino, S., & Wilson, R. B. (2003). Genotype and haplotype distributions of MTHFR87C>T and 1298A>C single nucleotide polymorphisms: A meta-analysis. *Journal of Human Genetics*, 48(1), 1–7. <https://doi.org/10.1007/s100380300000> • Shen, O., Liu, R., Wu, W., Yu, L., & Wang, X. (2012). Association of the methylenetetrahydrofolate reductase gene A1298C polymorphism with male infertility: A meta-analysis. *Annals of Human Genetics*, 76(1), 25–32. <https://doi.org/10.1111/j.1469-1808.2011.00691.x> • van der Put, N. M., Gabreëls, F., Stevens, E. M., Smeitink, J. A., Trijbels, F. J., Eskes, T. K., van den Heuvel, L. P., & Blom, H. J. (1998). A second common mutation in the methylenetetrahydrofolate reductase gene: An additional risk factor for neural-tube defects? *American Journal of Human Genetics*, 62(5), 1044–1051. <https://doi.org/10.1086/301825> • Wei, L. K., Au, A., Menon, S., Griffiths, L. R., Kooi, C. W., Irene, L., Zhao, J., Lee, C., Alekseeva, A. M., Hassan, M. R. A., & Aziz, Z. A. (2017). Polymorphisms of MTHFR, eNOS, ACE, AGT, ApoE, PON1, PDE4D, and Ischemic Stroke: Meta-Analysis. *Journal of Stroke and Cerebrovascular Diseases: The Official Journal of National Stroke Association*, 26(11), 2482–2493. <https://doi.org/10.1016/j.jstrokecerebrovasdis.2017.05.048> • Weisberg, I. S., Jacques, P. F., Selhub, J., Bostom, A. G., Chen, Z., Curtis Ellison, R., Eckfeldt, J. H., & Rozen, R. (2001). The 1298A→C polymorphism in methylenetetrahydrofolate reductase (MTHFR): In vitro expression and association with homocysteine. *Atherosclerosis*, 156(2), 409–415. [https://doi.org/10.1016/S0021-9150\(00\)00671-7](https://doi.org/10.1016/S0021-9150(00)00671-7)

MTHFR

• Christensen, B., Arbour, L., Tran, P., Leclerc, D., Sabbaghian, N., Platt, R., Gilfix, B. M., Rosenblatt, D. S., Gravel, R. A., Forbes, P., & Rozen, R. (1999). Genetic polymorphisms in methylenetetrahydrofolate reductase and methionine synthase, folate levels in red blood cells, and risk of neural tube defects. *American Journal of Medical Genetics*, 84(2), 151–157. [https://doi.org/10.1002/\(sici\)1096-8628\(19990521\)84:2151::aid-ajmg12>3.0.co;2-t](https://doi.org/10.1002/(sici)1096-8628(19990521)84:2151::aid-ajmg12>3.0.co;2-t) • Frosst, P., Blom, H. J., Milos, R., Goyette, P., Sheppard, C. A., Matthews, R. G., Boers, G. J., den Heijer, M., Kluijtmans, L. A., & van den Heuvel, L. P. (1995). A candidate genetic risk factor for vascular disease: A common mutation in methylenetetrahydrofolate reductase. *Nature Genetics*, 10(1), 111–113. <https://doi.org/10.1038/ng0595-111> • Hazra, A., Kraft, P., Lazarus, R., Chen, C., Chanock, S. J., Jacques, P., Selhub, J., & Hunter, D. J. (2009). Genome-wide significant predictors of metabolites in the one-carbon metabolism pathway. *Human Molecular Genetics*, 18(23), 4677–4687. <https://doi.org/10.1093/hmg/ddp428> • Hong, H.-H., Hu, Y., Yu, X.-Q., Zhou, L., Lv, M.-Q., Sun, Y., Ren, W.-J., & Zhou, D.-X. (2017). Associations of C677T polymorphism in methylenetetrahydrofolate reductase (MTHFR) gene with male infertility risk: A meta-analysis. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*, 212, 101–109. <https://doi.org/10.1016/j.ejogrb.2017.03.004> • Kowa, H., Yasui, K., Takeshima, T., Urakami, K., Sakai, F., & Nakashima, K. (2000). The homozygous C677T mutation in the methylenetetrahydrofolate reductase gene is a genetic risk factor for migraine. *American Journal of Medical Genetics*, 96(6), 762–764. [https://doi.org/10.1002/1096-8628\(20001204\)96:6762::aid-ajmg12>3.0.co;2-x](https://doi.org/10.1002/1096-8628(20001204)96:6762::aid-ajmg12>3.0.co;2-x) • Ogino, S., & Wilson, R. B. (2003). Genotype and haplotype distributions of MTHFR87C>T and 1298A>C single nucleotide polymorphisms: A meta-analysis. *Journal of Human Genetics*, 48(1), 1–7. <https://doi.org/10.1007/s100380300000> • Shadrina, M., Bondarenko, E. A., & Slominsky, P. A. (2018). Genetics Factors in Major Depression Disease. *Frontiers in Psychiatry*, 9, 334. <https://doi.org/10.3389/fpsyt.2018.00334> • van der Put, N. M., Steegers-Theunissen, R. P., Frosst, P., Trijbels, F. J., Eskes, T. K., van den Heuvel, L. P., Mariman, E. C., den Heyer, M., Rozen, R., & Blom, H. J. (1995). Mutated methylenetetrahydrofolate reductase as a risk factor for spina bifida. *Lancet (London, England)*, 346(8982), 1070–1071. [https://doi.org/10.1016/s0140-6736\(95\)91743-8](https://doi.org/10.1016/s0140-6736(95)91743-8) • Wei, L. K., Au, A., Menon, S., Griffiths, L. R., Kooi, C. W., Irene, L., Zhao, J., Lee, C., Alekseeva, A. M., Hassan, M. R. A., & Aziz, Z. A. (2017). Polymorphisms of MTHFR, eNOS, ACE, AGT, ApoE, PON1, PDE4D, and Ischemic Stroke: Meta-Analysis. *Journal of Stroke and Cerebrovascular Diseases: The Official Journal of National Stroke Association*, 26(11), 2482–2493. <https://doi.org/10.1016/j.jstrokecerebrovasdis.2017.05.048> • Weisberg, I. S., Jacques, P. F., Selhub, J., Bostom, A. G., Chen, Z., Curtis Ellison, R., Eckfeldt, J. H., & Rozen, R. (2001). The 1298A→C polymorphism in methylenetetrahydrofolate reductase (MTHFR): In vitro expression and association with homocysteine. *Atherosclerosis*, 156(2), 409–415. [https://doi.org/10.1016/S0021-9150\(00\)00671-7](https://doi.org/10.1016/S0021-9150(00)00671-7)

TCN2

• Afman, L. A., Lievers, K. J. A., van der Put, N. M. J., Trijbels, F. J. M., & Blom, H. J. (2002). Single nucleotide polymorphisms in the transcobalamin gene: Relationship with transcobalamin concentrations and risk for neural tube defects. *European Journal of Human Genetics: EJHG*, 10(7), 433–438. <https://doi.org/10.1038/sj.ejhg.5200830> • Castro, R., Barroso, M., Rocha, M., Esse, R., Ramos, R., Ravasco, P., Rivera, I., & de Almeida, I. T. (2010). The TCN2 778CNG polymorphism correlates with vitamin B12 cellular delivery in healthy adult populations. *Clinical Biochemistry*, 43(7–8), 645–649. <https://doi.org/10.1016/j.clinbiochem.2010.01.015> • Guéant, J.-L., Chabi, N. W., Guéant-Rodriguez, R.-M., Mutchinick, O. M., Debard, R., Payet, C., Lu, X., Guillaume, C., Bronowicki, J.-P., Quadros, E. V., Sanni, A., Amouzou, E., Xia, B., Chen, M., Anello, G., Bosco, P., Romano, C., Arrieta, H. R., Sánchez, B. E., ... Namour, F. (2007). Environmental influence on the worldwide prevalence of a 778C>G variant in the transcobalamin gene (TCN2). *Journal of Medical Genetics*, 44(6), 363–367. <https://doi.org/10.1136/jmg.2006.048041> • Lievers, K. J. A., Afman, L. A., Kluijtmans, L. A., Boers, G. H. J., Verhoeve, P., den Heijer, M., Trijbels, F. J. M., & Blom, H. J. (2002). Polymorphisms in the transcobalamin gene: Association with plasma homocysteine in healthy individuals and vascular disease patients. *Clinical Chemistry*, 48(9), 1383–1389. • Miller, J. W., Ramos, M. I., Garrod, M. G., Flynn, M. A., & Green, R. (2002). Transcobalamin II 775G>C polymorphism and indices of vitamin B12 status in healthy older adults. *Blood*, 100(2), 718–720. <https://doi.org/10.1182/blood-2002-01-0209> • Oussalah, A., Levy, J., Filhine-Trésarrieu, P., Namour, F., & Guéant, J.-L. (2017). Association of TCN2 rs1801198 c.776G>C polymorphism with markers of one-carbon metabolism and related diseases: A systematic review and meta-analysis of genetic association studies. *The American Journal of Clinical Nutrition*, 106(4), 1142–1156. <https://doi.org/10.3945/ajcn.117.156349> • von Castel-Dunwoody, K. M., Kauwell, G. P. A., Shelnutt, K. P., Vaughn, J. D., Griffin, E. R., Maneval, D. R., Theriaque, D. W., & Bailey, L. B. (2005). Transcobalamin 776C>G polymorphism negatively affects vitamin B-12 metabolism. *The American Journal of Clinical Nutrition*, 81(6), 1436–1441. <https://doi.org/10.1093/ajcn.81.6.1436>

COMT

• Dawling, S., Roodi, N., Mernaugh, R. L., Wang, X., & Parl, F. F. (2001). Catechol-O-methyltransferase (COMT)-mediated metabolism of catechol estrogens: Comparison of wild-type and variant COMT isoforms. *Cancer Research*, 61(18), 6716–6722. • Eriksson, A.-L., Suuriniemi, M., Mahonen, A., Cheng, S., & Ohlsson, C. (2005). The COMT val158met polymorphism is associated with early pubertal development, height and cortical bone mass in girls. *Pediatric Research*, 58(1), 71–77. <https://doi.org/10.1203/01.PDR.0000163383.49747.B5> • Kumar, P., & Rai, V. (2020). Catechol-O-methyltransferase gene Val158Met polymorphism and obsessive compulsive disorder susceptibility: A meta-analysis. *Metabolic Brain Disease*, 35(2), 241–251. <https://doi.org/10.1007/s11011-019-00495-0> • Lachman, H. M., Papolios, D. F., Saito, T., Yu, Y. M., Szumlanski, C. L., & Weinshilboum, R. M. (1996). Human catechol-O-methyltransferase pharmacogenetics: Description of a functional polymorphism and its potential application to neuropsychiatric disorders. *Pharmacogenetics*, 6(3), 243–250. <https://doi.org/10.1097/00008571-199606000-00007> • Sardahae, F. S., Holmen, T. L., Micali, N., & Kvale, K. (2017). Effects of single genetic variants and polygenic obesity risk scores on disordered eating in adolescents—The HUNT study. *Appetite*, 118, 8–16. <https://doi.org/10.1016/j.appet.2017.07.003> • Scanlon, P. D., Raymond, F. A., & Weinshilboum, R. M. (1979). Catechol-O-methyltransferase: Thermolabile enzyme in erythrocytes of subjects homozygous for allele for low activity. *Science (New York, N.Y.)*, 203(4375), 83–85. <https://doi.org/10.1126/science.758679> • Stein, D. J., Newman, T. K., Savitz, J., & Ramesar, R. (2008). Warriors versus worriers: The role of COMT gene variants. *CNS Spectrums*, 11(10), 745–748. <https://doi.org/10.1017/S1092852900014883> • Stein, M. B., Fallin, M. D., Schork, N. J., & Gelernter, J. (2005). COMT polymorphisms and anxiety-related personality traits. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 30(11), 2092–2102. <https://doi.org/10.1038/sj.npp.1300787> • Tunbridge, E. M., Harrison, P. J., Warden, D. R., Johnston, C., Refsum, H., & Smith, A. D. (2008). Polymorphisms in the catechol-O-methyltransferase (COMT) gene influence plasma total homocysteine levels. *American Journal of Medical Genetics. Part B: Neuropsychiatric Genetics: The Official Publication of the International Society of Psychiatric Genetics*, 147B(6), 996–999. <https://doi.org/10.1002/ajmg.b.30700> • Tunbridge, E. M., Narajos, M., Harrison, C. H., Beresford, C., Cipriani, A., & Harrison, P. J. (2019). Which Dopamine Polymorphisms Are Functional? Systematic Review and Meta-analysis of COMT, DAT, DBH, DDC, DRD1-5, MAOA, MAOB, TH, VMAT1, and VMAT2. *Biological Psychiatry*, 88(8), 608–620. <https://doi.org/10.1016/j.biopsych.2019.05.014> • Wichers, M., Aguilera, M., Kenis, G., Krabbendam, L., Myin-Germeyns, I., Jacobs, N., Peeters, F., Derom, C., Vlietinck, R., Mengelers, R., Delespaul, P., & van Os, J. (2008). The catechol-O-methyl transferase Val158Met polymorphism and experience of reward in the flow of daily life. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 33(13), 3030–3036. <https://doi.org/10.1038/sj.npp.1301520>

MAO-A

• Hotamisligil, G. S., & Breakefield, X. O. (1991). Human monoamine oxidase A gene determines levels of enzyme activity. *American Journal of Human Genetics*, 49(2), 383–392. • Hwang, I. W., Lim, M. H., Kwon, H. J., & Jin, H. J. (2018). Association of Monoamine Oxidase A (MAOA) Gene uVNTR and rs6323 Polymorphisms with Attention Deficit and Hyperactivity Disorder in Korean Children. *Medicina (Kaunas, Lithuania)*, 54(3), 32. <https://doi.org/10.3390/medicina54030032> • Kolla, N. J., & Bortolato, M. (2020). The role of monoamine oxidase A in the neurobiology of aggressive, antisocial, and violent behavior: A tale of mice and men. *Progress in Neurobiology*, 194, 101875. <https://doi.org/10.1016/j.pneurobio.2020.101875> • Larson, C. L., Taubitz, L. E., & Robinson, J. S. (2010). MAOA T941G polymorphism and the time course of emotional recovery following unpleasant pictures. *Psychophysiology*, 47(5), 857–862. <https://doi.org/10.1111/j.1469-8988.2010.01005.x> • Larson, C. L., Taubitz, L. E., & Robinson, J. S. (2010). MAOA T941G polymorphism and the time course of emotional recovery following unpleasant pictures. *Psychophysiology*, 47(5), 857–862. <https://doi.org/10.1111/j.1469-8988.2010.01005.x> • Leuchter, A. F., McCracken, J. T., Hunter, A. M., Cook, I. A., & Alpert, J. E. (2009). Monoamine oxidase a and catechol-o-methyltransferase functional polymorphisms and the placebo response in major depressive disorder. *Journal of Clinical Psychopharmacology*, 29(4), 372–377. <https://doi.org/10.1097/jcp.0b013e3181a04aaf> • M. B., & Jo, S. (2011). Behavioral outcomes of monoamine oxidase deficiency: Preclinical and clinical evidence. *International Review of Neurobiology*, 100. <https://doi.org/10.1016/B978-0-12-388467-3.00002-9> • Nordquist, N., & Orelund, L. (2006). Monoallelic expression of MAOA in skin fibroblasts. *Biochemical and Biophysical Research Communications*, 348(2), 763–767. <https://doi.org/10.1016/j.bbrc.2006.07.131> • Wang, M., Li, H., Deater-Deckard, K., & Zhang, W. (2018). Interacting Effect of Catechol-O-Methyltransferase (COMT) and Monoamine Oxidase A (MAOA) Gene Polymorphisms, and Stressful Life Events on Aggressive Behavior in Chinese Male Adolescents. *Frontiers in Psychology*, 9, 1079. <https://doi.org/10.3389/fpsyg.2018.01079>

MAO-B

• Babi? Leko, M., Nikolac Perkovi?, M., Nedi? Erjavec, G., Klepac, N., Švob Štrac, D., Borove?ki, F., Pivac, N., Hof, P. R., & Šimi?, G. (2021). Association of the MAOB rs1799836 Single Nucleotide Polymorphism and APOE ?4 Allele in Alzheimer’s Disease. *Current Alzheimer Research*, 18(7), 585–594. <https://doi.org/10.2174/156720501866210917162843> • Bortolato, M., & Shih, J. C. (2011). Behavioral outcomes of monoamine oxidase deficiency: Preclinical and clinical evidence. *International Review of Neurobiology*, 100, 13–42. <https://doi.org/10.1016/B978-0-12-388467-3.00002-9> • Dlugos, A. M., Palmer, A. A., & de Wit, H. (2009). Negative emotionality: Monoamine oxidase B gene variants modulate personality traits in healthy humans. *Journal of Neural Transmission (Vienna, Austria?)*, 116(10), 1323–1334. <https://doi.org/10.1007/s00702-009-0281-2> • Löhle, M., Mangone, G., Hermann, W., Hausbrand, D., Wolz, M., Mende, J., Reichmann, H., Hermann, A., Corvol, J.-C., & Storch, A. (2022). Functional MAOB Gene Intron 13 Polymorphism Predicts Dyskinesia in Parkinson’s Disease. *Parkinson’s Disease*, 2022, 5597503. <https://doi.org/10.1155/2022/5597503> • Löhle, M., Mangone, G., Wolz, M., Beuthien-Baumann, B., Oehme, L., van den Hoff, J., Kotzerke, J., Reichmann, H., Corvol, J.-C., & Storch, A. (2018). Functional monoamine oxidase B gene intron 13 polymorphism predicts putaminal dopamine turnover in de novo Parkinson’s disease. *Movement Disorders: Official Journal of the Movement Disorder Society*, 33(9), 1496–1501. <https://doi.org/10.1002/mds.27466>

TPH2

• Gao, J., Pan, Z., Jiao, Z., Li, F., Zhao, G., Wei, Q., Pan, F., & Evangelou, E. (2012). TPH2 Gene Polymorphisms and Major Depression – A Meta-Analysis. *PLoS ONE*, 7(5), e36721. <https://doi.org/10.1371/journal.pone.0036721> • Inoue, H., Yamasue, H., Tochigi, M., Takei, K., Suga, M., Abe, O., Yamada, H., Rogers, M. A., Aoki, S., Sasaki, T., & Kasai, K. (2010). Effect of tryptophan hydroxylase-2 gene variants on amygdalar and hippocampal volumes. *Brain Research*, 1331, 51–57. <https://doi.org/10.1016/j.brainres.2010.03.057> • Popova, N. K., & Kulikov, A. V. (2010). Targeting tryptophan hydroxylase 2 in affective disorder. *Expert Opinion on Therapeutic Targets*, 14(11), 1259–1271. <https://doi.org/10.1517/14728222.2010.524208>

HTR2

• Clinical Annotation for rs8313 (HTR2A); paroxetine; Depression (level 3 Toxicity). (n.d.). PharmGKB. Retrieved November 21, 2023, from <https://www.pharmgkb.org/clinicalAnnotation/1183618859> • Numata, S., Umehara, H., Ohmori, T., & Hashimoto, R. (2018). Clozapine Pharmacogenetic Studies in Schizophrenia: Efficacy and Adgranulocytosis. *Frontiers in Pharmacology*, 9, 1049. <https://doi.org/10.3389/fphar.2018.01049> • Smith, R. M., Papp, A. C., Webb, A., Ruble, C. L., Munsie, L. M., Nisenbaum, L. K., Kleinman, J. E., Lipska, B. K., & Sadee, W. (2013). Multiple regulatory variants modulate expression of 5-hydroxytryptamine 2A receptors in human cortex. *Biological Psychiatry*, 73(6), 546–554. <https://doi.org/10.1016/j.biopsych.2012.09.028>

SLC6A4

• Rs1042173. (n.d.). PharmGKB. Retrieved November 20, 2023, from <https://www.pharmgkb.org/variant/PA166155172/clinicalAnnotation> • Ait-Daoud, N., Seneviratne, C., Smith, J. B., Roache, J. D., Dawes, M. A., Liu, L., Wang, X.-Q., & Johnson, B. A. (2012). Preliminary Evidence for cue-induced Alcohol Craving Modulated by Serotonin Transporter Gene Polymorphism rs1042173. *Frontiers in Psychiatry*, 3, 6. <https://doi.org/10.3389/fpsy.2012.00006> • Bauer, I. E., Graham, D. F., Soares, J. C., & Nielsen, D. A. (2015). Serotonergic gene variation in substance use pharmacotherapy: A systematic review. *Pharmacogenomics*, 16(11), 1–8. <https://doi.org/10.2217/pgs.15.72> • Johnson, B. A., Ait-Daoud, N., Seneviratne, C., Roache, J. D., Javors, M. A., Wang, X.-Q., Liu, L., Penberthy, J. K., DiClemente, C. C., & Li, M. D. (2011). Pharmacogenetic approach at the serotonin transporter gene as a method of reducing the severity of alcohol drinking. *The American Journal of Psychiatry*, 168(3), 265–275. <https://doi.org/10.1176/appi.ajp.2010.10050755> • Seneviratne, C., Huang, W., Ait-Daoud, N., Li, M. D., & Johnson, Bankole. A. (2009). Characterization of a functional polymorphism in the 3’UTR of SLC6A4 and its association with drinking intensity. *Alcoholism, Clinical and Experimental Research*, 33(2), 332–339. <https://doi.org/10.1111/j.1530-0277.2008.00837.x>

DBH

• Barrie, E. S., Weinschenker, D., Verma, A., Pendergrass, S. A., Lange, L. A., Ritchie, M. D., Wilson, J. G., Kuivaniemi, H., Tromp, G., Carey, D. J., Gerhard, G. S., Brilliant, M. H., Hebbring, S. J., Cubells, J. F., Pinsonneault, J. K., Norman, G. J., & Sadee, W. (2014). Regulatory polymorphisms in human DBH affect peripheral gene expression and sympathetic activity. *Circulation Research*, 115(12), 1017–1025. <https://doi.org/10.1161/CIRCRESAHA.116.304398> • Cubells, J. F., & Zabetian, C. P. (2004). Human genetics of plasma dopamine beta-hydroxylase activity: Applications to research in psychiatry and neurology. *Psychopharmacology*, 174(4), 463–476. <https://doi.org/10.1007/s00213-004-1840-8> • Köhnke, M. D., Kolb, W., Köhnke, A. M., Lutz, U., Schick, S., & Batra, A. (2006). DBH*444G/A polymorphism of the dopamine-beta-hydroxylase gene is associated with alcoholism but not with severe alcohol withdrawal symptoms. *Journal of Neural Transmission (Vienna, Austria: 1996)*, 113(7), 869–878. <https://doi.org/10.1007/s00702-005-0365-6> • Wood, J. G., Joyce, P. R., Miller, A. L., Mulder, R. T., & Kennedy, M. A. (2002). A polymorphism in the dopamine beta-hydroxylase gene is associated with “paranoid ideation” in patients with major depression. *Biological Psychiatry*, 51(5), 365–369. [https://doi.org/10.1016/s0006-3223\(01\)01367-1](https://doi.org/10.1016/s0006-3223(01)01367-1)

GAD1

• Lim, S. W. et al. Genetic Prediction of Antidepressant Drug Response and Nonresponse in Korean Patients. *PLoS One* 9, e107098 (2014). • Darrah, S. D. et al. Genetic Variability in Glutamic Acid Decarboxylase Genes: Associations with Post-traumatic Seizures after Severe TBI. *Epilepsy Res* 103, 180–194 (2013).

GAD1

• Ulge, S. et al. A population-based association study of candidate genes for depression and sleep disturbance. *Am J Med Genet B Neuropsychiatr Genet* 153B, 468–476 (2010). • Hettema, J. M. et al. Association between glutamic acid decarboxylase genes and anxiety disorders, major depression, and neuroticism. *Mol Psychiatry* 11, 752–762 (2006). • Barakat, A. K. et al. Citalopram-induced pathways regulation and tentative treatment-outcome-predicting biomarkers in lymphoblastoid cell lines from depression patients. *Transl Psychiatry* 10, 210 (2020). • Weber, H. et al. Gender Differences in Associations of Glutamate Decarboxylase 1 Gene (GAD1) Variants with Panic Disorder. *PLoS One* 7, e37851 (2012).

ADRA2A

• da Silva, T. L. et al. Adrenergic ?2A receptor gene and response to methylphenidate in attention-deficit/hyperactivity disorder-predominantly inattentive type. *J Neural Transm* 115, 341–345 (2008). • Mäestu, J. et al. Human adrenergic ?2A receptor C-1291G polymorphism leads to higher consumption of sweet food products. *Mol Psychiatry* 12, 520–521 (2007). • Lario, S. et al. MspI identifies a biallelic polymorphism in the promoter region of the alpha 2A-adrenergic receptor gene. *Clin Genet* 51, 129–130 (1997). • Myer, N. M., Boland, J. R. & Faraone, S. V. Pharmacogenetics predictors of methylphenidate efficacy in childhood ADHD. *Mol Psychiatry* 23, 1929–1936 (2018). • Zhang, X. & Sun, Y. The Predictive Role of ADRA2A rs1800544 and HTR3B rs3758987 Polymorphisms in Motion Sickness Susceptibility. *Int J Environ Res Public Health* 18, 13163 (2021).

BDNF

• Losenkov, I. S. et al. Association Between BDNF Gene Variant Rs6285 and the Severity of Depression in Antidepressant Treatment-Free Depressed Patients. *Frontiers in Psychiatry* 11, (2020). • Szarowicz, C. A., Steece-Collier, K. & Caulfield, M. E. New Frontiers in Neurodegeneration and Regeneration Associated with Brain-Derived Neurotrophic Factor and the rs6285 Single Nucleotide Polymorphism. *International Journal of Molecular Sciences* 23, 8011 (2022). • Chen, Z.-Y. et al. Variant Brain-Derived Neurotrophic Factor (BDNF) (Met68) Alters the Intracellular Trafficking and Activity-Dependent Secretion of Wild-Type BDNF in Neurosecretory Cells and Cortical Neurons. *J. Neurosci.* 24, 4401–4411 (2004). • Colucci-D’Amato, L., Speranza, L. & Volpielli, F. Neurotrophic Factor BDNF, Physiological Functions and Therapeutic Potential in Depression, Neurodegeneration and Brain Cancer. *Int J Mol Sci* 21, 7777 (2020).

NGF

• Coskun, S., Varol, S., Ozdemir, H. H., Agacayak, E., Aydzın, B., Kapan, O., Camkurt, M. A., Tunc, S., & Cevik, M. U. (2016). Association of brain-derived neurotrophic factor and nerve growth factor gene polymorphisms with susceptibility to migraine. *Neuropsychiatric Disease and Treatment*, 12, 1779–1785. <https://doi.org/10.2147/NDT.S108814>

• Lang, U. E., Hellweg, R., Bajbouj, M., Gaus, V., Sander, T., & Gallinat, J. (2008). Gender-dependent association of a functional NGF polymorphism with anxiety-related personality traits. *Pharmacopsychiatry*, 41(5), 196–199. <https://doi.org/10.1055/s-0028-1082070>

• Lester, K. J., Hudson, J. L., Tropeano, M., Creswell, C., Collier, D. A., Farmer, A., Lyneham, H. J., Rapee, R. M., & Eley, T. C. (2012). Neurotrophic gene polymorphisms and response to psychological therapy. *Translational Psychiatry*, 2(5), e108. <https://doi.org/10.1038/tp.2012.33>

• Zakharyan, R., Atshemyan, S., Gevorgyan, A., & Boyajyan, A. (2014). Nerve growth factor and its receptor in schizophrenia. *BBA Clinical*, 1, 24–29. <https://doi.org/10.1016/j.bbaci.2014.05.001>

SYN1

• Santos, M., Lima, L., Carvalho, S., Mota-Pereira, J., Pimentel, P., Maia, D., Correia, D., Barroso, M. F., Gomes, S., Cruz, A., & Medeiros, R. (2023). The Impact of BDNF, NTRK2, NGFR, CREB1, GSK3B, AKT, MAPK1, MTOR, PTEN, ARC, and SYN1 Genetic Polymorphisms in Antidepressant Treatment Response Phenotypes. *International Journal of Molecular Sciences*, 24(7), 6758. <https://doi.org/10.3390/ijms24076758>

• Yan, P., Liu, H., Zhou, T., Sun, P., Wang, Y., Wang, X., Zhang, L., Wang, T., Dong, J., Zhu, J., Lv, L., Li, W., Qi, S., Liang, Y., & Kong, E. (2022). Crosstalk of Synapsin1 palmitoylation and phosphorylation controls the dynamics of synaptic vesicles in neurons. *Cell Death & Disease*, 13(9), 1–15. <https://doi.org/10.1038/s41419-022-05235-4>

C3

• Asteris, P. G., Gavrilaki, E., Touloumenidou, T., Koravou, E.-E., Koutra, M., Papayanni, P. G., Poulères, A., Karali, V., Lemonis, M. E., Mamou, A. D., Papalexandri, A., Varelas, C., Chatzopoulou, F., Chatzidimitriou, M., Chatzidimitriou, D., Veleni, A., Rapti, E., Kioumis, I., ... Anagnostopoulos, A. (2022). Genetic prediction of ICU hospitalization and mortality in COVID-19 patients using artificial neural networks. *Journal of Cellular and Molecular Medicine*, 26(5), 1445–1455. <https://doi.org/10.1111/jcmm.17008>

• Phillips, C. M., Goumidi, L., Bertrais, S., Ferguson, J. F., Field, M. R., Kelly, E. D., Peloso, G. M., Cupples, L. A., Shen, J., Ordovas, J. M., McManus, R., Hercberg, S., Portugal, H., Lairon, D., Planells, R., & Roche, H. M. (2009). Complement component 3 polymorphisms interact with polyunsaturated fatty acids to modulate risk of metabolic syndrome. *The American Journal of Clinical Nutrition*, 90(6), 1665–1673. <https://doi.org/10.3945/ajcn.2009.28101>

CD14

• Kamel, M. A., Selim, E. S., Tantawy, E. A., Elgendy, A., Abdulmageed, A., & Anis, R. H. (2023). Association of serum CD14 level and functional polymorphism C-159T in the promoter region of CD14 gene with allergic rhinitis. *Clinical and Experimental Medicine*, 23(8), 4861–4869. <https://doi.org/10.1007/s10238-023-01097-y>

• Mertens, J., Bregadze, R., Mansur, A., Askar, E., Bickeböller, H., Ramadori, G., & Mihm, S. (2009). Functional impact of endotoxin receptor CD14 polymorphisms on transcriptional activity. *Journal of Molecular Medicine (Berlin, Germany)*, 87(8), 815–824. <https://doi.org/10.1007/s00109-009-0479-7>

• Wang, Z., Hu, J., Fan, R., Zhou, J., & Zhong, J. (2012). Association between CD14 gene C-260T polymorphism and inflammatory bowel disease: A meta-analysis. *PLoS One*, 7(9), e45144. <https://doi.org/10.1371/journal.pone.0045144>

• Williams, L. K., McPhee, R. A., Ownby, D. R., Peterson, E. L., James, M., Zoratti, E. M., & Johnson, C. C. (2006). Gene-environment interactions with CD14 C-260T and their relationship to total serum IgE levels in adults. *The Journal of Allergy and Clinical Immunology*, 118(4), 851–857. <https://doi.org/10.1016/j.jaci.2006.07.007>

• Xu, J.-J., Liu, K.-Q., Ying, Z.-M., Zhu, X.-W., Xu, X.-J., Zhao, P.-P., Bai, W.-Y., Qiu, M.-C., Zhang, X.-W., & Zheng, H.-F. (2019). Effect of CD14 polymorphisms on the risk of cardiovascular disease: Evidence from a meta-analysis. *Lipids in Health and Disease*, 18(1), 74. <https://doi.org/10.1186/s12944-019-1018-3>

IL5

• Ganesh, B. B., Bhattacharya, P., Gopisetty, A., & Prabhakar, B. S. (2011). Role of Cytokines in the Pathogenesis and Suppression of Thyroid Autoimmunity. *Journal of Interferon & Cytokine Research*, 31(10), 721–731. <https://doi.org/10.1089/jir.2011.0049>

• Ishigaki, K., Akiyama, M., Kanai, M., Takahashi, A., Kawakami, E., Sugishita, H., Sakae, S., Matoba, N., Low, S.-K., Okada, Y., Terao, C., Amariuta, T., Gazal, S., Kochi, Y., Horikoshi, M., Suzuki, K., Ito, K., Koyama, S., Ozaki, K., ... Kamatani, Y. (2020). Large-scale genome-wide association study in a Japanese population identifies novel susceptibility loci across different diseases. *Nature Genetics*, 52(7), 669–679. <https://doi.org/10.1038/s41588-020-0640-3>

• Kabisch, M., Depner, M., Dahmen, I., Welland, S. K., Vogelberg, C., Niggemann, B., Lau, S., Illig, T., Klopp, N., Wahn, U., Reinhardt, D., von Mutius, E., & Nickel, R. (2007). Polymorphisms in eosinophil pathway genes, asthma and atopy. *Allergy*, 62(4), 423–428. <https://doi.org/10.1111/j.1365-9995.2006.01300.x>

• Mestiri, S., Zaafer, I., Inoubli, O., Abid, N., Omrani, A., Nejehi, H., & Marmouch, H. (2020). Association of cytokine Th2 gene polymorphisms with autoimmune thyroid diseases in Tunisian population. *International Journal of Immunogenetics*, 47(3), 294–308. <https://doi.org/10.1111/iji.12472>

• Principe, S., Porsberg, C., Bolm Litlev, S., Kjærsgaard Klein, D., Golebski, K., Dyhre-Petersen, N., van Dijk, Y. E., van Bragt, J. J. M. H., Dankelman, L. L. H., Dahlen, S., Brightling, C. E., Vijverberg, S. J. H., & Maitland-van der Zee, A. H. (2021). Treating severe asthma: Targeting the IL-5 pathway. *Clinical and Experimental Allergy*, 51(8), 992–1005. <https://doi.org/10.1111/cea.13885>

• Zhu, W., Liu, N., Zhao, Y., Jia, H., Cui, B., & Ning, G. (2010). Association analysis of polymorphisms in IL-3, IL-4, IL-5, IL-6, and IL-13 with Graves' disease. *Journal of Endocrinological Investigation*, 33(10), 751–755. <https://doi.org/10.1007/BF03346862>

IL-13

• Cameron, L., Webster, R. B., Strempel, J. M., Kiesler, P., Kabisch, M., Ramachandran, H., Yu, L., Stern, D. A., Graves, P. E., Lohman, I. C., Wright, A. L., Halonen, M., Klimecki, W. T., & Vercelli, D. (2006). Th2 cell-selective enhancement of human IL13 transcription by IL13-1112C>T, a polymorphism associated with allergic inflammation. *Journal of Immunology (Baltimore, Md.: 1950)*, 177(12), 8633–8642. <https://doi.org/10.4049/jimmunol.177.12.8633>

• Dimberg, J., Rubén, M., Skarstedt, M., Andersson, M., & Andersson, R. E. (2020). Genetic polymorphism patterns suggest a genetic driven inflammatory response as pathogenesis in appendicitis. *International Journal of Colorectal Disease*, 35(2), 277–284. <https://doi.org/10.1007/s00384-019-03473-1>

• Liao, N., Zhao, H., Chen, M.-L., & Xie, Z.-F. (2017). Association of the IL-13 polymorphisms rs1800925 and rs20541 with chronic obstructive pulmonary disease risk: An updated meta-analysis. *Medicine*, 96(47), e8558. <https://doi.org/10.1097/MD.0000000000008556>

• Omraninava, M., Eslami, M. M., Aslani, S., Razi, B., Imani, D., & Feyziniya, S. (2022). Interleukin 13 gene polymorphism and susceptibility to asthma: A meta-regression and meta-analysis. *European Annals of Allergy and Clinical Immunology*, 54(4), 150–167. <https://doi.org/10.23822/EurAnnACI.1764-1489.180>

STAT4

• Jiang, Y., Zhang, R., Zheng, J., Liu, P., Tang, G., Lv, H., Zhang, L., Shang, Z., Zhan, Y., Lv, W., Shi, M., & Zhang, R. (2012). Meta-analysis of 125 rheumatoid arthritis-related single nucleotide polymorphisms studied in the past two decades. *PLoS One*, 7(12), e51571. <https://doi.org/10.1371/journal.pone.0051571>

• Lee, H.-S., Park, H., Yang, S., Kim, D., & Park, Y. (2008). STAT4 polymorphism is associated with early-onset type 1 diabetes, but not with late-onset type 1 diabetes. *Annals of the New York Academy of Sciences*, 1150, 93–98. <https://doi.org/10.1196/annals.1447.013>

• Lee, H.-S., Remmers, E. F., Le, J. M., Kastner, D. L., Bae, S.-C., & Gregersen, P. K. (2007). Association of STAT4 with rheumatoid arthritis in the Korean population. *Molecular Medicine (Cambridge, Mass.)*, 13(9–10), 455–460. <https://doi.org/10.2119/2007-00072Lee>

• Namjou, B., Sestak, A. L., Armstrong, D. L., Zidovetzki, R., Kelly, J. A., Jacob, N., Ciobanu, V., Kaufman, K. M., Ojwang, J. O., Ziegler, J., Quismorio, F., Reiff, A., Myones, B. L., Guthridge, J. M., Nath, S. K., Bruner, G. R., Mehrian-Shai, R., Silverman, E., Klein-Gitelman, M., ... Jacob, C. O. (2009). High density genotyping of STAT4 gene reveals multiple haplotypic associations with Systemic Lupus Erythematosus in different racial groups. *Arthritis and Rheumatism*, 60(4), 1085–1095. <https://doi.org/10.1002/art.24387>

• Sigurdsson, S., Nordmark, G., Garnier, S., Grundberg, E., Kwan, T., Nilsson, O., Eloranta, M.-L., Gunnarsson, I., Svenungsson, E., Sturfelt, G., Bengtsson, A. A., Jönsson, A., Truedsson, L., Rantapää-Dahlqvist, S., Eriksson, C., Alm, G., Göring, H. H. H., Pastinen, T., Sjöström, A.-C., & Rönnblom, L. (2008). A risk haplotype of STAT4 for systemic lupus erythematosus is over-expressed, correlates with anti-dsDNA and shows additive effects with two risk alleles of IRF5. *Human Molecular Genetics*, 17(18), 2868–2876. <https://doi.org/10.1093/hmg/ddn184>

• Yan, N., Meng, S., Zhou, J., Xu, J., Muhal, F. S., Jiang, W., Shi, L., Shi, X., & Zhang, J. (2014). Association between STAT4 gene polymorphisms and autoimmune thyroid diseases in a Chinese population. *International Journal of Molecular Sciences*, 15(7), 12280–12293. <https://doi.org/10.3390/ijms150712280>

IL6

• Zhu, S., Wang, B., Jia, Q., & Duan, L. (2019). Candidate single nucleotide polymorphisms of irritable bowel syndrome: A systemic review and meta-analysis. *BMC Gastroenterology*, 19(1), 165. <https://doi.org/10.1186/s12876-019-1084-z>

• Carini, M., Fredi, M., Cavazzana, I., Bresciani, R., Ferrari, F., Monti, E., Franceschini, F., & Biasotto, G. (2023). Frequency Evaluation of the Interleukin-6 ?174G>C Polymorphism and Homeostatic Iron Regulator (HFE) Mutations as Disease Modifiers in Patients Affected by Systemic Lupus Erythematosus and Rheumatoid Arthritis. *International Journal of Molecular Sciences*, 24(22), 16300. <https://doi.org/10.3390/ijms242216300>

• Chen, L., Zhang, Z., Huang, J., & Jin, M. (2018). Association between rs1800795 polymorphism in the interleukin-6 gene and the risk of polycystic ovary syndrome: A meta-analysis. *Medicine*, 97(29), e11558. <https://doi.org/10.1097/MD.00000000000011558>

• Cheng, H., Zhu, W., Zhu, M., Sun, Y., Sun, X., Jia, D., Yang, C., Yu, H., & Zhang, C. (2021). Meta-analysis: Interleukin 6 gene -174G/C polymorphism associated with type 2 diabetes mellitus and interleukin 6 changes. *Journal of Cellular and Molecular Medicine*, 25(12), 5628–5639. <https://doi.org/10.1111/jcmm.16575>

• Fishman, D., Faulds, G., Jeffery, R., Mohamed-Ali, V., Yudkin, J. S., Humphries, S., & Woo, P. (1998). The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. *Journal of Clinical Investigation*, 102(7), 1369–1376. <https://doi.org/10.1172/JCI63575>

• Nie, G., Xie, C. L., Cao, Y. J., Xu, M. M., Shi, X., Zou, A. L., & Qi, J. H. (2016). Meta-analysis of IL-6 -174G/C polymorphism and psoriasis risk. *Genetics and Molecular Research: GMR*, 15(2), 15028255. <https://doi.org/10.4238/gmr.15028255>

• Tanaka, T., Narazaki, M., & Kishimoto, T. (2014). IL-6 in Inflammation, Immunity, and Disease. *Cold Spring Harbor Perspectives in Biology*, 6(10), a018295. <https://doi.org/10.1101/cshperspect.a018295>

• Wang, X., Yan, Z., & Ye, Q. (2019). Interleukin-6 gene polymorphisms and susceptibility to liver diseases: A meta-analysis. *Medicine*, 98(50), e18408. <https://doi.org/10.1097/MD.00000000000018408>

• Wu, W., Clark, E. A. S., Stoddard, G. J., Watkins, W. S., Esplin, M. S., Manuck, T. A., Xing, J., Vamer, M. W., & Jorde, L. B. (2013). Effect of interleukin-6 polymorphism on risk of preterm birth within population strata: A meta-analysis. *BMC Genetics*, 14, 30. <https://doi.org/10.1186/1471-2156-14-30>

TNF-?

• Chen, L., Huang, Z., Liao, Y., Yang, B., & Zhang, J. (2019). Association between tumor necrosis factor polymorphisms and rheumatoid arthritis as well as systemic lupus erythematosus: A meta-analysis. *Brazilian Journal of Medical and Biological Research = Revista Brasileira De Pesquisas Medicas E Biologicas*, 52(3), e7927. <https://doi.org/10.1590/1414-431X20187927>

• de Luis, D. A., Aller, R., Izaola, O., Gonzalez Sagrado, M., & Conde, R. (2013). Role of G308 promoter variant of tumor necrosis factor alpha gene on weight loss and metabolic parameters after a high monounsaturated versus a high polyunsaturated fat hypocaloric diets. *Medicina Clínica*, 141(5), 189–193. <https://doi.org/10.1016/j.medcli.2012.12.021>

• Kilpeläinen, T. O., Laaksonen, D. E., Lakka, T. A., Herder, C., Koenig, W., Lindström, J., Eriksson, J. G., Uusitupa, M., Kolb, H., Laakso, M., Tuomilehto, J., & Finnish Diabetes Prevention Study. (2010). The rs1800629 polymorphism in the TNF gene interacts with physical

activity on the changes in C-reactive protein levels in the Finnish Diabetes Prevention Study. *Experimental and Clinical Endocrinology & Diabetes: Official Journal, German Society of Endocrinology [and] German Diabetes Association*, 118(10), 757–759. <https://doi.org/10.1055/s-0030-1249686> • Kroeger, K. M., Carville, K. S., & Abraham, L. J. (1997). The -308 tumor necrosis factor-alpha promoter polymorphism effects transcription. *Molecular Immunology*, 34(5), 391–399. [https://doi.org/10.1016/s0161-5890\(97\)00052-7](https://doi.org/10.1016/s0161-5890(97)00052-7) • Loures, M. A. R., Alves, H. V., de Moraes, A. G., Santos, T. da S., Lara, F. F., Neves, J. S. F., Macedo, L. C., Teixeira, J. J. V., Sell, A. M., & Visentainer, J. E. L. (2019). Association of TNF, IL12, and IL23 gene polymorphisms and psoriatic arthritis: Meta-analysis. *Expert Review of Clinical Immunology*, 15(3), 303–313. <https://doi.org/10.1080/1744666X.2019.1564039> • Song, G. G., Seo, Y. H., Kim, J.-H., Choi, S. J., Ji, J. D., & Lee, Y. H. (2015). Association between TNF-? (-308 A/G, -238 A/G, -857 C/T) polymorphisms and responsiveness to TNF-? blockers in spondyloarthritis, psoriasis and Crohn's disease: A meta-analysis. *Pharmacogenomics*, 16(12), 1427–1437. <https://doi.org/10.2217/pgs.15.90> • Tu, Y., Fan, G., Zeng, T., Cai, X., & Kong, W. (2018). Association of TNF-? promoter polymorphism and Graves' disease: An updated systematic review and meta-analysis. *Bioscience Reports*, 38(2), BSR20180143. <https://doi.org/10.1042/BSR20180143> • Yang, G., Chen, J., Xu, F., Bao, Z., Yao, Y., & Zhou, J. (2014). Association between Tumor Necrosis Factor-? rs1800629 Polymorphism and Risk of Asthma: A Meta-Analysis. *PLoS ONE*, 9(6), e99962. <https://doi.org/10.1371/journal.pone.0099962>

IDO1

• Barnes, J., Mondelli, V., & Pariante, C. M. (2017). Genetic Contributions of Inflammation to Depression. *Neuropsychopharmacology*, 42(1), 81–98. <https://doi.org/10.1038/npp.2016.169> • Golimbet, V. E., Korova?seva, G. I., Gabaeva, M. V., Velikaia, N. V., Snegireva, A. A., Kasparov, S. V., Kolesina, N. I., Ganisheva, T. K., & Save'eva, T. M. (2014). [A study of IL-17 and IDO gene polymorphisms in patients with schizophrenia]. *Zhurnal Nevrologii i Psikiatrii Imeni S.S. Korsakova*, 114(5), 46–49. • Smith, A. K., Simon, J. S., Gustafson, E. L., Novello, S., Cubellis, J. F., Epstein, M. P., Devlin, D. J., Qiu, P., Albrecht, J. K., Brass, C. A., Sulkowski, M. S., McHutchinson, J. G., & Miller, A. H. (2012). Association of a polymorphism in the indoleamine- 2,3-dioxygenase gene and interferon-?induced depression in patients with chronic hepatitis C. *Molecular Psychiatry*, 17(8), 781–789. <https://doi.org/10.1038/mp.2011.67>

CTLA4

• Chen, M., & Li, S. (2019). Associations between cytotoxic T-lymphocyte-associated antigen 4 gene polymorphisms and diabetes mellitus: A meta-analysis of 76 case-control studies. *Bioscience Reports*, 39(5), BSR20190309. <https://doi.org/10.1042/BSR20190309> • Chen, Y., Chen, S., Gu, Y., Feng, Y., Shi, Y., Fu, Q., Wang, Z., Cai, Y., Dai, H., Zheng, S., Sun, M., Zhang, M., Xu, X., Chen, H., Xu, K., & Yang, T. (2018). CTLA-4 +49 G/A, a functional T1D risk SNP, affects CTLA-4 level in Treg subsets and IA-2A positivity, but not beta-cell function. *Scientific Reports*, 8(1), 10074. <https://doi.org/10.1038/s41598-018-28423-9> • Fathima, N., Narne, P., & Ishaq, M. (2019). Association and gene-gene interaction analyses for polymorphic variants in CTLA-4 and FOXP3 genes: Role in susceptibility to autoimmune thyroid disease. *Endocrine*, 64(3), 591–604. <https://doi.org/10.1007/s12020-019-01859-3> • Mäurer, M., Loserth, S., Kolb-Mäurer, A., Ponath, A., Wiese, S., Kruse, N., & Rieckmann, P. (2002). A polymorphism in the human cytotoxic T-lymphocyte antigen 4 (CTLA4) gene (exon 1 +49) alters T-cell activation. *Immunogenetics*, 54(1), 1–8. <https://doi.org/10.1007/s00251-002-0429-9> • Mousavi, M. J., Shayesteh, M. R. H., Jamalzehi, S., Alimohammadi, R., Rahimi, A., Aslani, S., & Rezaei, N. (2021). Association of the genetic polymorphisms in inhibiting and activating molecules of immune system with rheumatoid arthritis: A systematic review and meta-analysis. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, 26, 22. https://doi.org/10.4103/jrms.JRMS_567_20 • Patel, H., Mansuri, M. S., Singh, M., Begum, R., Shastri, M., & Misra, A. (2016). Association of Cytotoxic T-Lymphocyte Antigen 4 (CTLA4) and Thyroglobulin (TG) Genetic Variants with Autoimmune Hypothyroidism. *PLoS One*, 11(3), e0149441. <https://doi.org/10.1371/journal.pone.0149441>

DRD2

• Clarke, T.-K., Weiss, A. R. D., Ferraro, T. N., Kampman, K. M., Dackis, C. A., Pettinati, H. M., O'brien, C. P., Oslin, D. W., Lohoff, F. W., & Berrettini, W. H. (2014). The dopamine receptor D2 (DRD2) SNP rs1076560 is associated with opioid addiction. *Annals of Human Genetics*, 78(1), 33–39. <https://doi.org/10.1111/ahg.12046> • Gluskin, B. S., & Mickey, B. J. (2016). Genetic variation and dopamine D2 receptor availability: A systematic review and meta-analysis of human in vivo molecular imaging studies. *Translational Psychiatry*, 6(3), e747. <https://doi.org/10.1038/tp.2016.22> • Sasabe, T., Furukawa, A., Matsusita, S., Higuchi, S., & Ishiura, S. (2007). Association analysis of the dopamine receptor D2 (DRD2) SNP rs1076560 in alcoholic patients. *Neuroscience Letters*, 412(2), 139–142. <https://doi.org/10.1016/j.neulet.2006.10.064> • Tunbridge, E. M., Narajos, M., Harrison, C. H., Beresford, C., Cipriani, A., & Harrison, P. J. (2019). Which Dopamine Polymorphisms Are Functional? Systematic Review and Meta-analysis of COMT, DAT, DBH, DDC, DRD1-5, MAOA, MAOB, TH, VMAT1, and VMAT2. *Biological Psychiatry*, 86(8), 608–620. <https://doi.org/10.1016/j.biopsych.2019.05.014> • Zheng, C., Shen, Y., & Xu, Q. (2012). Rs1076560, a functional variant of the dopamine D2 receptor gene, confers risk of schizophrenia in Han Chinese. *Neuroscience Letters*, 518(1), 41–44. <https://doi.org/10.1016/j.neulet.2012.04.052>

ATG5

• Grosjean, I., Roméo, B., Domdom, M.-A., Belaid, A., D'Andréa, G., Guillot, N., Gherardi, R. K., Gal, J., Milano, G., Marquette, C. H., Hung, R. J., Landi, M. T., Han, Y., Brest, P., Von Bergen, M., Klionsky, D. J., Amos, C. I., Hofman, P., & Mograbi, B. (n.d.). Autophagopathies: From autophagy gene polymorphisms to precision medicine for human diseases. *Autophagy*, 18(11), 2519–2536. <https://doi.org/10.1080/15548627.2022.2039994> • Martin, L. J., Gupta, J., Jyothula, S. S. S. K., Butsch Kovacic, M., Biagini Myers, J. M., Patterson, T. L., Erickson, M. B., He, H., Gibson, A. M., Baye, T. M., Amirsetty, S., Tsoras, A. M., Sha, Y., Eissa, N. T., & Hershey, G. K. K. (2012). Functional variant in the autophagy-related 5 gene promoter is associated with childhood asthma. *PLoS One*, 7(4), e33454. <https://doi.org/10.1371/journal.pone.0033454> • Shao, Y., Chen, F., Chen, Y., Zhang, W., Lin, Y., Cai, Y., Yin, Z., Tao, S., Liao, Q., Zhao, J., Mai, H., He, Y., He, J., & Cui, L. (2017). Association between genetic polymorphisms in the autophagy-related 5 gene promoter and the risk of sepsis. *Scientific Reports*, 7, 9399. <https://doi.org/10.1038/s41598-017-09978-5> • Tamargo-Gómez, I., Fernández, Á. F., & Mariño, G. (2020). Pathogenic Single Nucleotide Polymorphisms on Autophagy-Related Genes. *International Journal of Molecular Sciences*, 21(21), 8196. <https://doi.org/10.3390/ijms21218196>

ATG12

• Yuan, J. et al. Polymorphisms in autophagy related genes and the coal workers' pneumoconiosis in a Chinese population. *Gene* 632, 36–42 (2017). • Anton, R. F. et al. Pharmacogenomics. *Nat. Genet.* 16, 268–278 (2008). • Lindberg, S. Autophagy: Definition, Diet, Fasting, Cancer, Benefits, and More. *Healthline* (2014). Available at: <https://www.healthline.com/health/autophagy#bottom-line>. • Takagi, A., Kume, S., Maegawa, H. & Uzu, T. Emerging role of mammalian autophagy in ketogenesis to overcome starvation. *Autophagy* (2016). doi:10.1080/15548627.2016.1151597 • Antunes, F. et al. Autophagy and intermittent fasting: the connection for cancer therapy? *Clinics* (Sao Paulo, Brazil) (2018). doi:10.6061/clinics/2018/e814s • Levine, B. & Kroemer, G. Autophagy in the Pathogenesis of Disease. *Cell* (2008). doi:10.1016/j.cell.2007.12.018 • Smith, G. S., Walter, G. L. & Walker, R. M. Clinical Pathology in Non-Clinical Toxicology Testing. in *Haschek and Rousseaux's Handbook of Toxicologic Pathology* (2013). doi:10.1016/B978-0-12-415759-0.00018-2 • Mizushima, N. Autophagy: Process and function. *Genes and Development* (2007). doi:10.1101/gad.1599207

ATG16L1

• Levine, B., & Kroemer, G. (2008). Autophagy in the Pathogenesis of Disease. *Cell*, 132(1), 27–42. <https://doi.org/10.1016/j.cell.2007.12.018> • Wellcome Trust Case Control Consortium. (2007). Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*, 447(7145), 661–678. <https://doi.org/10.1038/nature05911>

CTH

• Wenstrom, K. D., Johanning, G. L., Owen, J., Johnston, K. E., Acton, S., & Tamura, T. (2000). Role of amniotic fluid homocysteine level and of fetal 5, 10-methylenetetrahydrofolate reductase genotype in the etiology of neural tube defects. *American Journal of Medical Genetics*, 90(1), 12–16. [https://doi.org/10.1002\(sic\)1096-8628\(20000103\)90:112::aid-ajmg3>3.0.co;2-h](https://doi.org/10.1002(sic)1096-8628(20000103)90:112::aid-ajmg3>3.0.co;2-h)

GSTP1

• Dai, X., Bui, D. S., & Lodge, C. (2021). Glutathione S-Transferase Gene Associations and Gene-Environment Interactions for Asthma. *Current Allergy and Asthma Reports*, 21(5), 31. <https://doi.org/10.1007/s11882-021-01005-y> • do Nascimento, M. R., Silva de Souza, R. O., Silva, A. L., Lima, E. S., Gonçalves, M. S., & de Moura Neto, J. P. (2021). GSTP1 rs1695 and rs1871042, and SOD2 rs4880 as molecular markers of lipid peroxidation in blood storage. *Blood Transfusion = Trasfusione Del Sangue*, 19(4), 309–316. <https://doi.org/10.2450/2020.0062-20> • Hollman, A. L., Tchounwou, P. B., & Huang, H.-C. (2016). The Association between Gene-Environment Interactions and Diseases Involving the Human GST Superfamily with SNP Variants. *International Journal of Environmental Research and Public Health*, 13(4), 379. <https://doi.org/10.3390/ijerph13040379> • Katsarou, M.-S., Giakoumaki, M., Papadimitriou, A., Demertzis, N., Androutsopoulos, V., & Drakoulis, N. (2018). Genetically driven antioxidant capacity in a Caucasian Southeastern European population. *Mechanisms of Ageing and Development*, 172, 1–5. <https://doi.org/10.1016/j.mad.2017.08.010> • Melén, E., Nyberg, F., Lindgren, C. M., Berglind, N., Zucchini, M., Nordling, E., Hailberg, J., Svartengren, M., Morgenstern, R., Kere, J., Bellander, T., Wickman, M., & Pershagen, G. (2008). Interactions between glutathione S-transferase P1, tumor necrosis factor, and traffic-related air pollution for development of childhood allergic disease. *Environmental Health Perspectives*, 116(8), 1077–1084. <https://doi.org/10.1289/ehp.11117> • Moyer, A. M., Salavaggione, O. E., Wu, T.-Y., Moon, I., Eckloff, B. W., Hildebrandt, M. A., T., Schaid, D. J., Wieben, E. D., & Weinshilboum, R. M. (2008). Glutathione S-Transferase P1: Gene Sequence Variation and Functional Genomic Studies. *Cancer Research*, 68(12), 4791–4801. <https://doi.org/10.1158/0008-5472.CAN-07-6724> • Mukhammadayeva, G. F., Bakirov, A. B., Karimov, D. O., Zlatindinov, M. M., Valova, A. V., Borisova, A. I., & Distanova, A. A. (2022). Analysis of the GSTP1 rs1695 polymorphism association with the development of asthma and phenotypic manifestations. *The Journal of Asthma: Official Journal of the Association for the Care of Asthma*, 59(6), 1065–1069. <https://doi.org/10.1080/02770903.2021.1910295> • Palmer, C. N. A., Doney, A. S. F., Lee, S. P., Murrie, I., Ismail, I., Macgregor, D. F., & Mukhopadhyay, S. (2006). Glutathione S-transferase M1 and P1 genotype, passive smoking, and peak expiratory flow in asthma. *Pediatrics*, 118(2), 710–716. <https://doi.org/10.1542/peds.2005-3030> • Scarfo, M., Sciardra, C., Ruberto, S., & Santovito, A. (2021). GSTT1, GSTP1 and XPC genes are associated with longevity in an Italian cohort. *Annals of Human Biology*, 48(5), 443–447. <https://doi.org/10.1080/03014460.2021.1985170> • Simeunovic, D., Odanovic, N., Pljesa-Ercegovac, M., Radic, T., Radovanovic, S., Coric, V., Milinkovic, I., Matic, M., Djukic, T., Ristic, A., Ristic, D., Seferovic, P., Simic, T., Simic, D., & Savic-Radojevic, A. (2019). Glutathione Transferase P1 Polymorphism Might Be a Risk Determinant in Heart Failure. *Disease Markers*, 2019, 6984845. <https://doi.org/10.1155/2019/6984845> • Tamer, L., Kaliko?lu, M., Ates, N. A., Yildirim, H., Ercan, B., Saritas, E., Unlü, A., & Atik, U. (2004). Glutathione-S-transferase gene polymorphisms (GSTT1, GSTM1, GSTP1) as increased risk factors for asthma. *Respirology* (Carlton, Vic.), 9(4), 493–498. <https://doi.org/10.1111/j.1440-1843.2004.00657.x>

AOC1

• Maintz, L., & Novak, N. (2007). Histamine and histamine intolerance. *The American Journal of Clinical Nutrition*, 85(5), 1185–1196. <https://doi.org/10.1093/ajcn/85.5.1185> • Maintz, L., Yu, C.-F., Rodríguez, E., Baurecht, H., Bieber, T., Illig, T., Weidinger, S., & Novak, N. (2011). Association of single nucleotide polymorphisms in the diamine oxidase gene with diamine oxidase serum activities. *Allergy*, 66(7), 893–902. <https://doi.org/10.1111/j.1398-9995.2011.02548.x>

HNMT

• Pang, Y. P., Zheng, X. E., & Weinshilboum, R. M. (2001). Theoretical 3D model of histamine N-methyltransferase: Insights into the effects of a genetic polymorphism on enzymatic activity and thermal stability. *Biochemical and Biophysical Research Communications*, 287(1), 204–208. <https://doi.org/10.1006/bbrc.2001.5570> • Preuss, C. V., Wood, T. C., Szumliński, C. L., Raftogiannis, R. B., Ottemess, D. M., Girard, B., Scott, M. C., & Weinshilboum, R. M. (1998). Human histamine N-methyltransferase pharmacogenetics: Common genetic polymorphisms that alter activity. *Molecular Pharmacology*, 53(4), 708–717. <https://doi.org/10.1124/mol.53.4.708> • Szczepankiewicz, A., Br?borowicz, A., Sobkowiak, P., & Popiel, A. (2010). Polymorphisms of two histamine-metabolizing enzymes genes and childhood allergic asthma: A case control study. *Clinical and Molecular Allergy: CMA*, 8, 14. <https://doi.org/10.1186/1476-7961-8-14> • Yan, L., Galinsky, R. E., Bernstein, J. A., Liggett, S. B., & Weinshilboum, R. M. (2000). Histamine N-methyltransferase pharmacogenetics: Association of a common functional polymorphism with asthma. *Pharmacogenetics*, 10(3), 261–266. <https://doi.org/10.1097/00008571-200004000-00007>

FUT2

• Hu, M., Zhang, X., Li, J., Chen, L., He, X., & Sui, T. (2022). Fucosyltransferase 2: A Genetic Risk Factor for Intestinal Diseases. *Frontiers in Microbiology*, 13, 940196. <https://doi.org/10.3389/fmicb.2022.940196> • Rausch, P., Rehman, A., Künzel, S., Häslér, R., Ott, S. J., Schreiber, S., Rosenstiel, P., Franke, A., & Baines, J. F. (2011). Colonic mucosa-associated microbiota is influenced by an interaction of Crohn disease and FUT2 (Secretor) genotype. *Proceedings of the National Academy of Sciences of the United States of America*, 108(47), 19030–19035. <https://doi.org/10.1073/pnas.1106408108> • Tong, M., McHardy, I., Ruegger, P., Goudarzi, M., Kashyap, P. C., Haritunians, T., Li, X., Graeber, T. G., Schwager, E., Huttenhower, C., Fornace, A. J., Sonnenburg, J. L., McGovern, D. P. B., Borneman, J., & Braun, J. (2014). Reprograming of gut microbiome energy metabolism by the FUT2 Crohn's disease risk polymorphism. *The ISME Journal*, 8(11), 2193–2206.

<https://doi.org/10.1038/ismej.2014.84>

HLA-DQA1

• de Bakker, P. I. W., McVean, G., Sabeti, P. C., Miretti, M. M., Green, T., Marchini, J., Ke, X., Monsuur, A. J., Whittaker, P., Delgado, M., Morrison, J., Richardson, A., Walsh, E. C., Gao, X., Galver, L., Hart, J., Hafler, D. A., Pericak-Vance, M., Todd, J. A., ... Rioux, J. D. (2006). A high-resolution HLA and SNP haplotype map for disease association studies in the extended human MHC. *Nature Genetics*, 38(10), 1166–1172. <https://doi.org/10.1038/ng1885> • Dubois, P. C. A., Trynka, G., Franke, L., Hunt, K. A., Romanos, J., Curtotti, A., Zhemakova, A., Heap, G. A. R., Adány, R., Aromaa, A., Bardella, M. T., van den Berg, L. H., Bockett, N. A., de la Concha, E. G., Dema, B., Fehrmann, R. S. N., Fernández-Arquero, M., Fialal, S., Grandone, E., ... van Heel, D. A. (2010). Multiple common variants for celiac disease influencing immune gene expression. *Nature Genetics*, 42(4), 295–302. <https://doi.org/10.1038/ng.543> • Monsuur, A. J., de Bakker, P. I. W., Zhemakova, A., Pinto, D., Verduijn, W., Romanos, J., Auricchio, R., Lopez, A., van Heel, D. A., Crusius, J. B. A., & Wijmenga, C. (2008). Effective detection of human leukocyte antigen risk alleles in celiac disease using tag single nucleotide polymorphisms. *PloS One*, 3(5), e2270. <https://doi.org/10.1371/journal.pone.0002270> • Sallése, M., Lopetuso, L. R., Eftymakis, K., & Neri, M. (2020). Beyond the HLA Genes in Gluten-Related Disorders. *Frontiers in Nutrition*, 7. <https://www.frontiersin.org/articles/10.3389/fnut.2020.575844> • Senapati, S., Sood, A., Midha, V., Sood, N., Sharma, S., Kumar, L., & Thelma, B. K. (2016). Shared and unique common genetic determinants between pediatric and adult celiac disease. *BMC Medical Genomics*, 9(1), 44. <https://doi.org/10.1186/s12920-016-0211-8> • van Heel, D. A., Franke, L., Hunt, K. A., Gwilliam, R., Zhemakova, A., Inouye, M., Wapenaar, M. C., Bamardo, M. C. N. M., Bethel, G., Holmes, G. K. T., Feighery, C., Jewell, D., Kelleher, D., Kumar, P., Travis, S., Walters, J. R. F., Sanders, D. S., Howdle, P., Swift, J., ... Wijmenga, C. (2007). A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. *Nature Genetics*, 39(7), 827–829. <https://doi.org/10.1038/ng2058>

HLA-DQB1

• Monsuur, A. J., de Bakker, P. I. W., Zhemakova, A., Pinto, D., Verduijn, W., Romanos, J., Auricchio, R., Lopez, A., van Heel, D. A., Crusius, J. B. A., & Wijmenga, C. (2008). Effective detection of human leukocyte antigen risk alleles in celiac disease using tag single nucleotide polymorphisms. *PloS One*, 3(5), e2270. <https://doi.org/10.1371/journal.pone.0002270>