

May 11, 2018

Mr. Stephen Judd, P.Eng. Manager of Infrastructure Engineering Services City of Port Moody 100 Newport Drive Port Moody, BC V3H 5C3

Dear Mr. Judd:

### Re: Soil, Sediment and Surface Water Analytical Results 1300 David Avenue, Port Moody, BC Project 13953

## 1. BACKGROUND

Keystone Environmental Ltd. (Keystone Environmental) is pleased to present this letter summarizing the analytical results of soil, sediment and surface water samples collected on City of Port Moody (the City) property and infrastructure located down-gradient from a residential septic field owned by Anmore Green Estates. The soil, sediment and surface water samples were collected on March 29, 2018.

A BC Ministry of Environment (MoE) letter titled *"Warning Letter, Permit, 4606"* dated March 7, 2017 (BC MoE, 2017a), reported non-compliance issues associated with the unconfirmed septic discharge associated with the Anmore Green Estates Strata Corporation LMS 3080 (Strata) septic field operating permit PE-04606, located within the Village of Anmore, adjacent to the northeast of Heritage Woods Secondary School, at 1300 David Avenue, Port Moody, BC. Associated Environmental Consultants Ltd. (Associated Environmental) developed an "action plan" for the Strata (AE, 2017a) to address the non-compliance issues. The action plan noted that soil and water sampling frequency would be determined by changes in precipitation, with monitoring during all seasons, and specifically, further communication with MoE if:

• *Escherichia coli* (*E. coli*) or fecal coliforms in water quality data from discharge points exceed the BC Approved Water Quality Guidelines for Primary-Contact Recreation and/or the Guidelines for Canadian Recreational Water Quality;

Suite 320 4400 Dominion Street Burnaby, British Columbia Canada V5G 4G3 Telephone: 604 430 0671 Facsimile: 604 430 0672 info@KeystoneEnviro.com KeystoneEnviro.com Environmental Consulting Engineering Solutions Assessment & Protection • Human-sourced viruses are detected in soil where septic breakthrough is believed to have occurred

In September 2017, Associated Environmental (AE, 2017b) collected eight soil samples from areas south and west of the wastewater treatment system for analysis of parameters outlined in their action plan, with the exception of human-source viruses. Analytical results reported four of the six Site sample locations tested positive for *Escherichia coli*. (*E. coli*) and fecal coliforms ranging from 2.3 to 160,000 MPN/g (230–16,000,000 MPN/100g), in addition to the two background sample locations northwest of the Site on a recreational trail. Provincial and federal human health protection values for the parameters analyzed are specific to water samples, and not applicable to the soil samples. Water samples were not collected by Associated Environmental during the September 2017 sampling event due to dry conditions.

The City of Port Moody noted that the soil samples collected by Associated Environmental focused on the embankment, located on the downgradient slope of the septic field on School District 43 (SD43) property. The City wishes to understand the extent and potential risk to public health from potential releases of septic waste into the City's storm water drainage system and watercourses, and whether the surface of the playing fields of the school have been affected.

As a result, Keystone Environmental personnel collected soil, sediment and surface water samples on City property, and from associated infrastructure (i.e., storm water collection basins), on March 29, 2018 and submitted these samples to Maxxam Analytical for analysis of *E. coli*, fecal coliforms, and total coliforms. Additionally, targeted field duplicate soil, sediment, and water samples were also submitted for analysis of total and human *Bacteroides*. This report provides the results of this investigation and describes visual observations of environment conditions encountered during the sampling program.

## 2. METHODS

For this sampling program, Keystone Environmental adopted a similar sampling approach as Associated Environmental (2017a; 2017b) but did not include analyses of nutrients that can be present naturally, and instead included parameters directly relevant to septic discharges. Keystone Environmental collected the following samples during the March 29, 2018 sampling event:

- Soil
  - Keystone Environmental collected 6 soil samples from 3 locations in proximity to the baseball playing field.
- Sediment
  - Keystone Environmental collected 8 sediment samples (7 characterization samples plus 1 field duplicate) from 4 creek locations (2 background and 2 possible exposure locations within Wilkes Creek to the west and Turner Creek to the east of the septic field), as well as 3 catch basin locations located downgradient from the playing field.
- Surface Water
  - Keystone Environmental collect 8 surface water samples (7 characterization samples plus 1 field duplicate) from 3 catch basin locations to characterize potential loading onto the property, and from 4 creek locations (2 background and 2 possible exposure locations from Wilkes and Turner Creeks).



Keystone Environmental staff arrived on Site at 11 am on March 29, 2018. Light rain was encountered throughout the sampling program. Based on a review of the Port Moody weather station [IBCPORTM8], 5.3 mm of precipitation fell on March 29, 2018 and 0.0 mm of precipitation fell on March 28, 2018. In the week leading up to the sampling event (i.e., March 22 to March 29), approximately 66.6 mm of precipitation was reported at the Port Moody weather station [IBCPORTM8] (www.wunderground.com).

Sample locations were logged using a global position system (GPS). Sample locations for soil and sediment samples are provided in **Figure 1 of Appendix A**. Surface water sampling locations are provided on **Figure 2 of Appendix A**. Visual observations at each sampling location were recorded in field note books to document environmental conditions, soil, sediment and/or water characteristics, and to record measurements of pH, temperature, dissolved oxygen, conductivity and turbidity of surface water samples using hand held meters.

Soil samples were collected from three locations in proximity to the playing fields at two depths (~3 cm and ~5 cm) per location, for a total of 6 samples. Soil samples were collected using a shovel. Surface organic material (e.g., grass) was first cleared from the surface soils, and then samples were collected from appropriate depths directly from the shovel. Soil characteristics were recorded in field note books prior to sample collection. Soil samples were labelled as follows:

- SS18-03 (0.3): soil sample located north of the baseball field at a depth of 3 cm below ground
- SS18-03 (0.5): soil sample located north of the baseball field at a depth of 5 cm below ground
- SS18-04 (0.3): soil sample located on the northern edge of the baseball field at a depth of 3 cm below ground
- SS18-04 (0.5): soil sample located on the northern edge of the baseball field at a depth of 5 cm below ground
- SS18-08 (0.3): soil sample collected from the base of the hill leading to the septic field on the north end of the school parking lot at a depth of 3 cm below ground
- SS18-08 (0.5): soil sample collected from the base of the hill leading to the septic field on the north end of the school parking lot at a depth of 5 cm below ground

Nine surficial sediment samples (~0-5 cm) were also collected from one depth at eight locations. Samples were labelled as follows:

- SED18-01: background sample collected from Wilkes Creek
- SED18-02: background sample collected from Turner Creek
- SED18-05: southern catch basin sample located west of the tennis courts
- SED18-06: catch basin sample located north of the soccer field
- SED18-07: catch basin sample located northwest of the baseball field
- SED18-09: Wilkes Creek sample located downgradient of the septic field



• SED18-10: Turner Creek sample located downgradient of the City's storm water collection system

Sufficial sediment samples collected from the creeks SED18-01, SED18-02, SED18-09 and SED18-10, as well as SED18-07 (the most upgradient catch basin location), were collected using a similar approach as the soil samples and were collected directly from the shovel. Due to the depth of the catch basins at SED18-05 and SED18-06, a ponar dredge sediment sampler was used to collect sufficial sediment samples at these locations. The samples were emptied into a stainless-steel pan, at which point sediment characteristics were noted and recorded prior to sample collection.

At approximately 1 pm, Keystone Environmental staff met with a member of the City of Port Moody Engineering Department to provide access to the catch basin sampling locations (SED18-05 to SED18-07). The employee was unable to access the large settling tank located below the tennis courts and, therefore, SED18-05 was collected from a catch basin located approximately 10 m west of the large settling tank location. The SED18-05 catch basin was located upgradient of the large settling tank and, thus, was considered to be representative of conditions that may occur downgradient in the larger settling tank.

Surface water sampling was also performed concurrently at each of the sediment sampling locations. Surface water (e.g., ponding or overland flow) was not observed at soil sampling locations, and thus were not collected. Surface water samples were collected in general accordance with the BC field sampling manual. Field measurements of pH, temperature, conductivity, dissolved oxygen, and turbidity were collected using hand held meters at each surface water sampling location.

Soil, water and sediment samples were submitted to Maxxam Analytical for analysis of *E. coli*, fecal coliforms, and total coliforms. Additional soil/sediment and water samples were collected concurrently at each location for possible future analysis of total *Bacteroides* and human *Bacteroides*. These additional samples were collected to help to differentiate between human and animal sources of bacteria. Following review of the *E. coli*, fecal coliforms, and total coliforms analytical results, five sediment samples (SED18-05, SED18-06, SED18-07, SED18-09, and SED18-10), one soil sample (SS18-08 (0.5)), and two surface water samples (SW18-05 and SW18-06) were selected to be analyzed for total and human *Bacteriodes*.

## 3. APPROACH

For this report, Keystone Environmental assumed that the public, mainly school children and park users may be exposed to fecal bacteria in two ways:

- To fecal bacteria in surface water (resulting from groundwater seepages from the septic field) through dermal skin contact, and incidental ingestion of surface water from splashes and hand mouth-transfer
- To fecal bacteria in soil/sediment through dermal skin contact and incidental ingestion of soil via hand mouth-transfer



To gauge the potential for human health risks, Keystone Environmental compared surface water *E. coli* results to human health based provincial and federal benchmarks; the BC MoE Approved Water Quality Guidelines for Primary-Contact Recreation (BC MoE 2017b) and Health Canada Guidelines for Canadian Recreational Water Quality (Health Canada, 2012). The 2017 BC MoE guidelines involved adopting the Health Canada guidelines, and supersede the 2001 BC MoE Water Quality Criteria for Microbiological Indicators Overview Report (BC MoE, 2001). The PCR criterion is used as the safety limit to trigger beach and public area closures in Metro Vancouver (Vancouver Coastal Health, 2018).

With respect to soil and sediment quality, a human health benchmark for fecal coliforms in soil and sediment has not been formally established in Canada. The BC Organic Matter Recycling Regulation specifies a limit of 1,000 MPN/100g, which is consistent with the Council of Ministers of the Environment (CCME) guideline for compost quality, but this value is specific to biosolids and compost, and is not applicable to soils alone (OMRR, 2018). CCME indicates the "use of *E. coli* content as a direct indicator of pathogen levels is not yet supported by all regulatory agencies in Canada, but it may be used to help verify the reason for the high fecal coliform levels" (CCME, 2005). As such, soil results in this report were used to infer presence/absence of fecal coliforms and human *Bacteroids*. Soil samples submitted for analysis of these parameters were presented as quantity detected, either Most Probable Number (MPN/100g) or Cell Equivalents (CEs/g).

For the purposes of this report, data supporting a potential septic discharge was inferred if human *Bacteroides* were detected in soil, sediment, or surface water down-gradient to the septic field, as total coliforms can occur naturally in the environment, and the presence of *E. coli* can be associated with both human and animal feces (Health Canada, 2012).

## 4. RESULTS

Soil and sediment sample results are provided in **Appendix B**. Surface water sample results are provided in **Appendix C**. Laboratory analytical reports presented in **Appendix D**.

## 4.1 Physical Conditions

During the March 29, 2018 sampling event, groundwater seeps were not observed to be leaching from the hill side of the Strata's septic field onto City property. Additionally, pooling of surface water on the playing fields located south of the Strata's septic field was not observed.

Table 1 describes soil and sediment characteristics observed during the sampling program.



Sample	Characteristics	Odour
SED18-01	Dark brown to black, predominately fines, with some organic matter	No odour
SED18-02	Brown to black sandy silt, with some organics	No odour
SS18-03 (0.3)	Brown to black sandy soil with some fines and clumps of light gray moist fines	No odour
SS18-03 (0.5)	Brown and grey sand, moist with minimal organics	No odour
SS18-04 (0.3)	Brown to black sand with some organics	No odour
SS18-04 (0.5)	Brown to grey sand, moist, with minimal organics	No odour
SED18-05	Predominately black fines (silt and clay)	organic/decomposition like odour
SED18-06	Grey to black sandy sediments with some organics	Slight organic/decomposition like odour
SED18-07	Brown to black sandy sediment with some fines	Slight organic/decomposition like odour
SS18-08 (0.3)	Dark brown/black sand, with some organics	No odour
SS18-08 (0.5)	Brown and grey sandy soil with some organics	No odour
SED18-09	Reddish brown sandy sediments	No odour
SED18-10	Reddish brown sandy sediments	No odour

## Table 1 Soil and Sediment Characteristics of Collected Samples

Table 2 describes water quality encountered during the sampling program.

Table 2Water Quality Characteristics of Collected Surface Water Samples
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Sample	Characteristics	рН	Dissolved Oxygen (mg/L)	Conductivity (µS/cm)	Turbidity (NTU)	Temperature (°C)
SW18-01	Clear, no odour	6.7	10.4	8.6	4.20	6.5
SW18-02	Clear, no odour	7.5	11.6	8.4	4.20	7.5
SW18-05	Turbid, organic odour	6.7	10.8	245	34.9	10.0
SW18-06	Clear, no odour	7.5	10.4	128	32.4	8.5
SW18-07	Clear, no odour	7.0	10.4	68	32.2	10.0
SW18-09	Clear, no odour	7.6	11.8	71	4.84	6.0
SW18-10	Clear, no odour	7.1	11.6	82	5.91	7.0

## 4.2 Surface Water

Analytical results reported detectable *E. coli* and fecal coliforms in two of the three surface water samples (SW18-05 and SW18-06) collected from the catch basin locations. *E. coli* concentrations were less than provincial and federal PCR guidelines, with exception of SW18-06, which had an *E. coli* concentration of 940 CFU/100mL (CFU = Colony Forming Units). Notably, measurements of *E. coli*, total coliforms, and fecal coliforms in surface water



collected at SW18-05, the catch basin sample located downgradient of SW18-06, had an *E. coli* concentration of 100 CFU/100mL, which was less than provincial and federal PCR guidelines (**Appendix C**). Total *Bacteriodes* analyses on SW18-06 and SW18-05 resulted in measured concentrations of non-detect and 27,526 CEs/100 mL, respectively. Notably, human *Bacteriodes* analyses were non-detect in both cases.

Surface water samples collected from background locations in Wilkes and Turner Creeks had *E. coli* concentrations less than provincial and federal PCR guidelines, and creek samples collected downgradient of the septic field (SW18-09 and SW18-10) had non-detectable total coliform and *E. coli* concentrations.

## 4.3 Soil

Total coliform concentrations were detected in one of the three soil sampling locations, SS18-08 (at both depths SS18-08 (0.3) and SS18-08 (0.5)), however *E. coli* concentrations were less than the detection limit at this location. Additionally, results of the human *Bacteriode* analysis at SS18-08 (0.5) were also non-detectable.

Regulatory criteria for *E. coli*, total coliforms or human *Bacteroides* are not available for soil and sediment samples.

## 4.4 Sediment

Total coliform concentrations were detected in five of the seven sediment sampling locations: SED18-05, SED18-06, SED18-07, SED18-09, and SED18-10. Additionally, sediment *E. coli* concentrations were measured at detectable concentrations in samples collected downgradient of the septic field in Wilkes and Turner Creeks (SED18-09 and SED18-10) at 210 and 45 MPN/100g, respectively. The results of the human *Bacteriodes* analyses for SED18-05, SED18-06, SED18-07, SED18-09, and SED18-10 were determined to be non-detectable.

Regulatory criteria for *E. coli*, total coliforms or human *Bacteroides* are not available for soil and sediment samples.

## 5. CONCLUSION AND RECOMMENDATIONS

The findings of the soil, sediment and surface water sampling program did not provide evidence that the septic field was discharging beyond the septic field limits during the sampling program, as soil, sediment, and surface water samples collected from locations downgradient of the septic field with detectable total coliforms, fecal coliforms, and/or *E. coli* concentrations did not contain detectable human *Bacteroides*. These results suggest that the coliforms and *E. coli* concentrations measured in the downgradient samples may have been from animal sources.

Surface water samples that were co-located with sediment sampling locations had concentrations of *E. coli* at concentrations less than provincial and federal PCR guidelines, with the exception of SW18-06. This water sample, as well as the co-located sediment sample (SED18-06), were tested for human *Bacteriodes* and, the results were non-detect, suggesting that the elevated *E. coli* concentrations measured at this location were likely of animal origin.



## 6. CLOSURE

This letter has been prepared solely for the internal use of the City of Port Moody pursuant to the agreement between Keystone Environmental Ltd. and the City of Port Moody. By using this report, the City of Port Moody agrees that they will review and use the letter in its entirety. Any use which other parties make of this letter, or any reliance on or decisions made based on it, are the responsibility of such parties. Keystone Environmental Ltd. accepts no responsibility for damages, if any, suffered by other parties as a result of decisions made or actions based on this letter.

If you have any questions, please do not hesitate to contact the undersigned.

Sincerely,

## Keystone Environmental Ltd.

Brett Lucas, M.Sc., R.P.Bio. Risk Assessor / Toxicologist

Craig Patterson, R.P.Bio. Project Manager

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### ATTACHMENTS:

- Appendix A: Figures
- Appendix B: Soil and Sediment Analytical Results
- Appendix C: Surface Water Analytical Results
- Appendix D: Analytical Laboratory Reports

### REFERENCES

- Associated Environmental Ltd. (AE). 2017a. Action Plan for the Anmore Green Estates (Permit PE 4606). Associated Environmental Ltd. October 11, 2017.
- AE. 2017b. Technical Memorandum Re: Results of September 2017 Soil Testing. Associated Environmental Ltd. October 16, 2017.
- BC Ministry of Environment (BC MoE). 2001. Water Quality. Water Quality Criteria for Microbiological Indicators Overview Report. BC Ministry of Environment. August 7, 2001.
- BC MoE. 2013. Field Sampling Manual. BC Ministry of Environment. 2013 edition.
- BC MoE. 2017a. Warning Letter, Permit, 4606. BC Ministry of Environment. March 7, 2017.



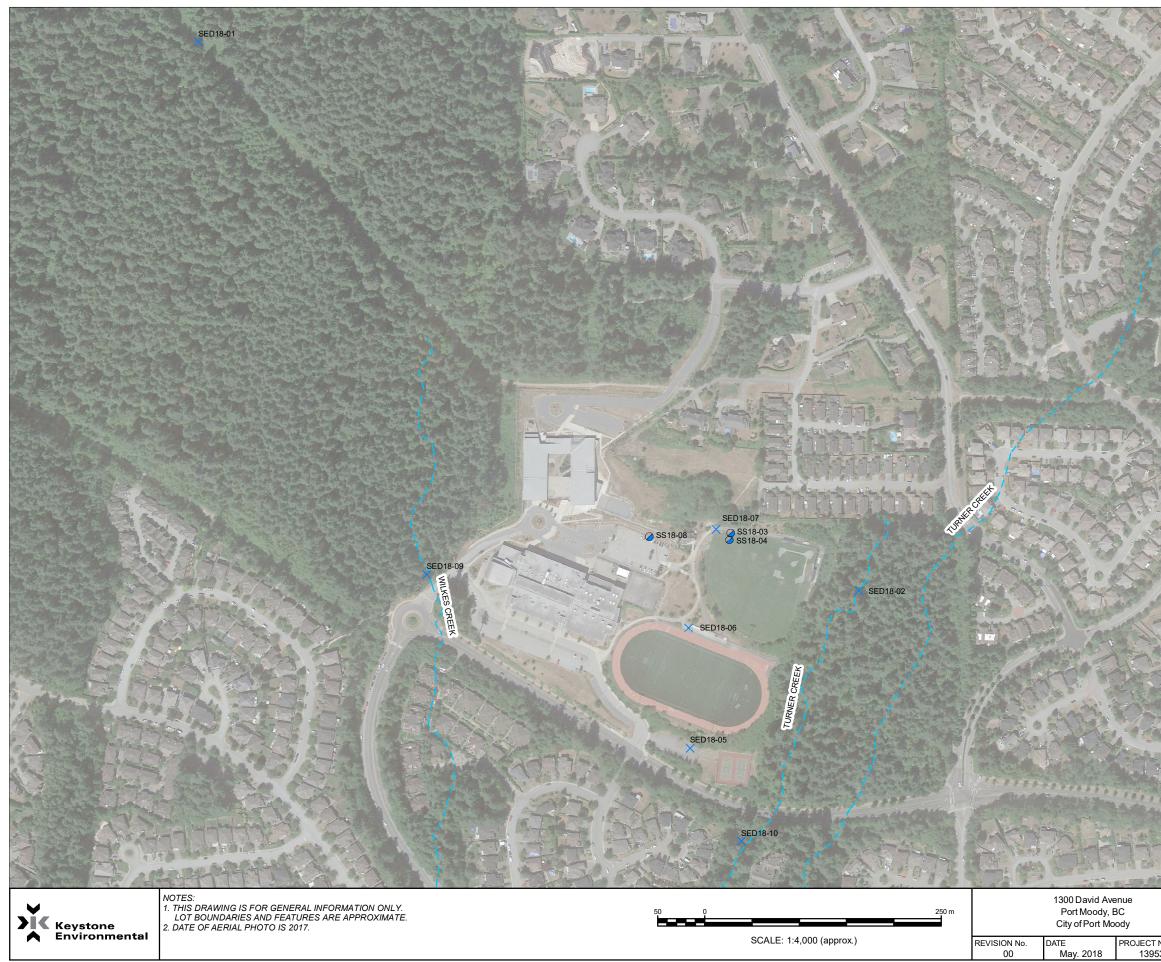
- BC MoE. 2017b. Recreational Water Quality Guidelines. Guideline Summary. Ministry of Environment, Water Protection and Sustainability Branch. December 2017.
- Canadian Council Ministers of the Environment (CCME). 2005. Guidelines for Compost Quality. Canadian Council Ministers of the Environment. PN 1340. 2005.
- Health Canada. 2012. Guidelines for Canadian Recreational Water Quality. Third Edition. Prepared by the Federal-Provincial-Territorial Working Group on Recreational Water Quality of the Federal-Provincial-Territorial Committee on Health and the Environment. April 2012.
- Organic Matter Recycling Regulation (OMRR). 2018. Organic Matter Recycling Regulation. Includes amendments up to BC Reg. 243/2016, November 1, 2017 to February 13, 2018.
- Vancouver Coastal Health. 2018. Beach Water Quality Reports. http://www.vch.ca/publichealth/environmental-health-inspections/pools-beaches/beach-water-quality-reports (Accessed February 15, 2018).



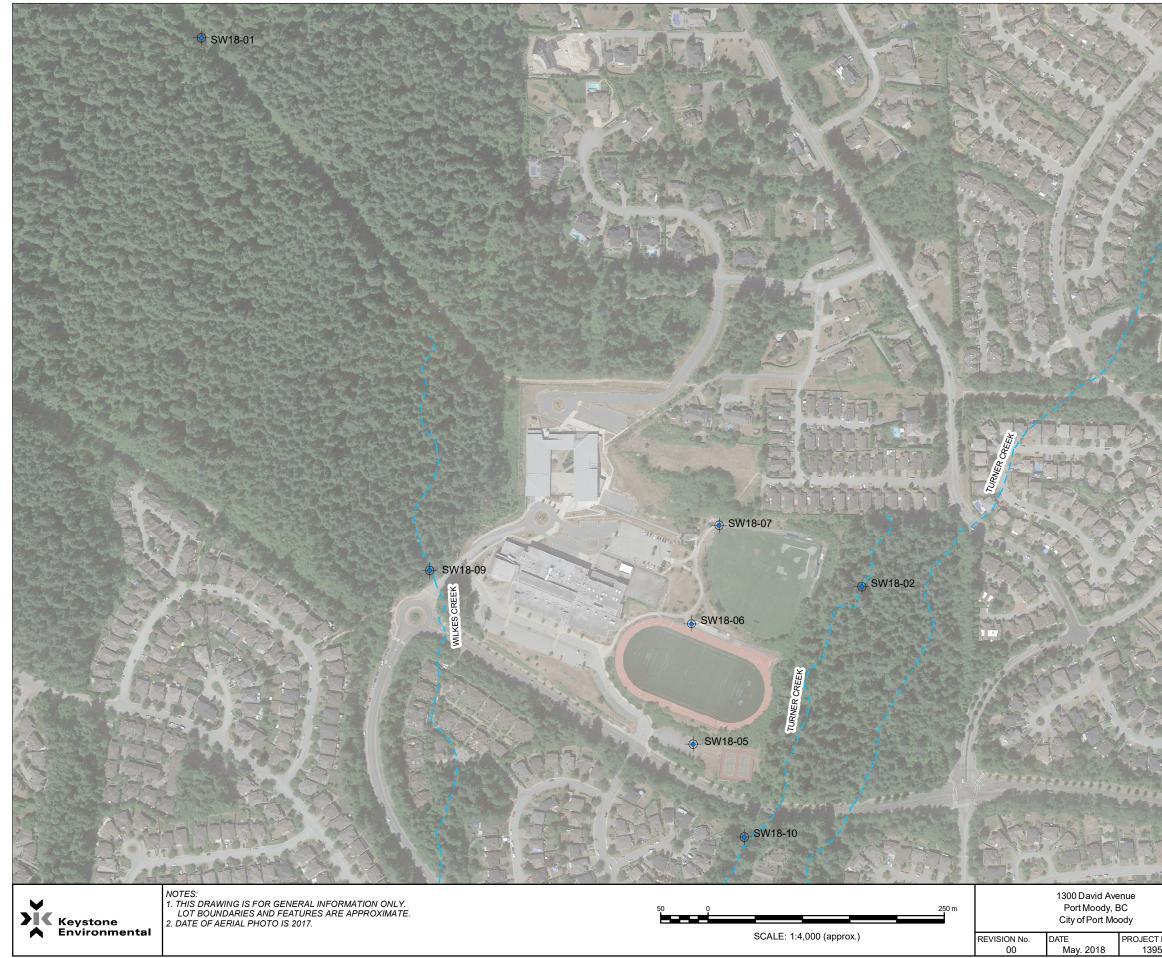
APPENDIX A

FIGURES





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	Figure 1
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No. 53-103	



KEYSTONE SURFACE WATER SAMPLE (2018)
<u>LEGEND</u>
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**APPENDIX B** 

SOIL AND SEDIMENT ANALYTICAL RESULTS



# SOIL AND SEDIMENT ANALYTICAL RESULTS Microbiological

1300 David Avenue, Port Moody, BC Project #: 13953 April 2018

Standards	Г	SAMPLE ID	Units	SED18-01	SED18-A	RPD or MS	SED18-02	SED18-05	SED18-06	SED18-07	SED18-09	SED18-10	SS18-03 (0.3)
		DATE SAMPLED		29-Mar-18	29-Mar-18	for	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18
		LAB CERTIFICATE		B823570	B823570	SED18-01	B823570	B823570	B823570	B823570	B823570	B823570	B823570
		LAB SAMPLE ID		TE7252	TE8792	and	TE7253	TE7258	TE7259	TE7260	TE7263	TE7264	TE7254
		SAMPLE DEPTH (mbg)		~0-0.05	Duplicate of	SED18-A	~0-0.05	~0-0.05	~0-0.05	~0-0.05	~0-0.05	~0-0.05	0.03
		SOIL DESCRIPTION		Brown Fines	SED18-01		Black Sandy Silt	Black Fines	Black Sand	Black Sand	Redish Sand	Redish Sand	Brown Sand
Microbiology													
n/s		E. coli	MPN/100g	<20	<20		<20	<20	<20	<20	210	45	<20
n/s		Total Coliform	MPN/100g	<20	<20		<20	240,000	45	3,300	210	140	<20
n/s		Human Bacteriodes	CEs/g					ND	ND	ND	ND	ND	
n/s		Total Bacteriodes	CEs/g					902,216	25,780	5,749	ND	ND	

Notes:

MPN/100g Most Probable Number per 100 grams	MPN/100g	Most Probable Number per 100 grams
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CEs/100g	Cell Equivalents per 100 grams
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n/s No standards

RPD Relative Percent Difference

MS Maximum Spread

ND Non Detect

mbg Metres below ground

# SOIL AND SEDIMENT ANALYTICAL RESULTS Microbiological

1300 David Avenue, Port Moody, BC Project #: 13953 April 2018

Standards	]	SAMPLE ID	Units	SS18-03 (0.5)	SS18-04 (0.3)	SS18-04 (0.5)	SS18-08 (0.3)	SS18-08 (0.5)
		DATE SAMPLED		29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18
		LAB CERTIFICATE		B823570	B823570	B823570	B823570	B823570
		LAB SAMPLE ID		TE7255	TE7256	TE7257	TE7261	TE7262
		SAMPLE DEPTH (mbg)		0.05	0.03	0.05	0.03	0.05
		SOIL DESCRIPTION		Brown Sand				
		Microbiology						
n/s		E. coli	MPN/100g	<20	<20	<20	<20	<20
n/s		Total Coliform	MPN/100g	<20	<20	<20	3,500	7,000
n/s		Human Bacteriodes	CEs/g					ND
n/s		Total Bacteriodes	CEs/g					ND

Notes:

MPN/100g	Most Probable Number per 100 grams
CEs/100g	Cell Equivalents per 100 grams

n/s No standards

RPD Relative Percent Difference

MS Maximum Spread

ND Non Detect

mbg Metres below ground

APPENDIX C

SURFACE WATER ANALYTICAL RESULTS



# SURFACE WATER ANALYTICAL RESULTS Microbiology

1300 David Avenue, Port Moody, BC Project #: 13953 April 2018

BCWQG	Health Canada		SAMPLE ID	Units	SW18-01	SW18-A	RPD or MS	SW18-02	SW18-05	SW18-06	SW18-07	SW18-09	SW18-10
(2017)	(2012)		DATE SAMPLED		29-Mar-18	29-Mar-18	for	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18	29-Mar-18
			LAB CERTIFICATE		B823570	B823570	SW18-01	B823570	B823570	B823570	B823570	B823570	B823570
			LAB SAMPLE ID		TE7265	TE8793	and	TE7266	TE7267	TE7268	TE7269	TE7270	TE7271
Primary Contact	Primary Contact		TOP OF SCREEN (mbg)			Duplicate of	SW18-A						
Recreation	Recreation		BOTTOM OF SCREEN (mbg)			SW18-01							
	Microbiology												
n/g	n/g		Fecal Coliforms	CFU/100mL	<1	0	-	7	1	1	<1	<1	6
200/100 mL	200/100 mL		E. coli	CFU/100mL	4	4		67	100	<u>940</u>	0	0	0
n/g	n/g		Total Coliform	CFU/100mL	11	10		79	780	980	0	0	0
n/s	n/s		Human Bacteriodes	CEs/100mL					ND	ND			
n/s	n/s		Total Bacteriodes	CEs/100mL					27,526	ND			

#### Notes:

BC WQG BC Water Quality Guidelines

Health Canada Health Canada Guidelines for Canadian Recreational Water Quality

CFU Colony Forming Units

n/g No guideline

RPD Relative Percent Difference

MS Maximum Spread

# APPENDIX D

ANALYTICAL LABORATORY REPORTS



Maxia Stroup Company

Your Project #: 13953-103 Your C.O.C. #: K019946, K019947, K019948

#### **Attention: CRAIG PATTERSON**

KEYSTONE ENVIRONMENTAL LTD SUITE 320 4400 DOMINION STREET BURNABY, BC CANADA V5G 4G3

> **Report Date: 2018/05/10** Report #: R2552492 Version: 4 - Revision

## **CERTIFICATE OF ANALYSIS – REVISED REPORT**

MAXXAM JOB #: B823570 Received: 2018/03/29, 17:17

Sample Matrix: DRINKING WATER # Samples Received: 8

		Date	Date		
Analyses	Quantity	Extracted	Analyzed	Laboratory Method	Analytical Method
Fecal Coliform membrane filt - Potable W	7	N/A	2018/03/29	BBY4SOP-00001	SM 22 9222 m
Fecal Coliform membrane filt - Potable W	1	N/A	2018/04/02	BBY4SOP-00001	SM 22 9222 m
Tot Coliform/E.Coli by MF-Chromocult(PW)	7	N/A	2018/03/29	BBY4SOP-00143	SM 22 9222
Tot Coliform/E.Coli by MF-Chromocult(PW)	1	N/A	2018/04/02	BBY4SOP-00143	SM 22 9222
Human Bacteroides in Water Subcontract (1)	2	N/A	2018/05/08		
Total Bacteroides in Water Subcontract (1)	2	N/A	2018/05/08		

Sample Matrix: Sediment # Samples Received: 8

		Date	Date		
Analyses	Quantity	Extracted	Analyzed	Laboratory Method	Analytical Method
Total Coliforms (MTF) in Soil (2)	7	N/A	2018/03/29	COR1 SOP-00019	Health Can MFHPB-19
Total Coliforms (MTF) in Soil (2)	1	N/A	2018/04/02	COR1 SOP-00019	Health Can MFHPB-19
Escherichia Coli (MTF) in Soil (2)	7	N/A	2018/03/29	COR1 SOP-00019	Health Can MFHPB-19
Escherichia Coli (MTF) in Soil (2)	1	N/A	2018/04/02	COR1 SOP-00019	Health Can MFHPB-19
Moisture	7	2018/03/31	2018/04/02	BBY8SOP-00017	BCMOE BCLM Dec2000 m
Moisture	1	2018/04/03	2018/04/04	BBY8SOP-00017	BCMOE BCLM Dec2000 m
Human Bacteroides in Soil Subcontract (1)	4	N/A	2018/04/19		
Human Bacteroides in Soil Subcontract (1)	1	N/A	2018/05/08		
Total Bacteroides inSoil Subcontract (1)	4	N/A	2018/04/19		
Total Bacteroides inSoil Subcontract (1)	1	N/A	2018/05/08		

Sample Matrix: Soil

# Samples Received: 6

		Date	Date		
Analyses	Quantity	Extracted	Analyzed	Laboratory Method	Analytical Method
Total Coliforms (MTF) in Soil (2)	6	N/A	2018/03/29	COR1 SOP-00019	Health Can MFHPB-19
Escherichia Coli (MTF) in Soil (2)	6	N/A	2018/03/29	COR1 SOP-00019	Health Can MFHPB-19
Moisture	6	2018/03/31	2018/04/02	BBY8SOP-00017	BCMOE BCLM Dec2000 m
Human Bacteroides in Soil Subcontract (1)	1	N/A	2018/04/19		



Your Project #: 13953-103 Your C.O.C. #: K019946, K019947, K019948

#### **Attention: CRAIG PATTERSON**

KEYSTONE ENVIRONMENTAL LTD SUITE 320 4400 DOMINION STREET BURNABY, BC CANADA V5G 4G3

> Report Date: 2018/05/10 Report #: R2552492 Version: 4 - Revision

### **CERTIFICATE OF ANALYSIS – REVISED REPORT**

#### MAXXAM JOB #: B823570 Received: 2018/03/29, 17:17

Sample Matrix: Soil # Samples Received: 6

		Date	Date		
Analyses	Quantity	y Extracted	Analyzed	Laboratory Method	Analytical Method
Total Bacteroides inSoil Subcontract (1)	1	N/A	2018/04/19	)	

#### Remarks:

Maxxam Analytics' laboratories are accredited to ISO/IEC 17025:2005 for specific parameters on scopes of accreditation. Unless otherwise noted, procedures used by Maxxam are based upon recognized Provincial, Federal or US method compendia such as CCME, MDDELCC, EPA, APHA.

All work recorded herein has been done in accordance with procedures and practices ordinarily exercised by professionals in Maxxam's profession using accepted testing methodologies, quality assurance and quality control procedures (except where otherwise agreed by the client and Maxxam in writing). All data is in statistical control and has met quality control and method performance criteria unless otherwise noted. All method blanks are reported; unless indicated otherwise, associated sample data are not blank corrected.

Maxxam Analytics' liability is limited to the actual cost of the requested analyses, unless otherwise agreed in writing. There is no other warranty expressed or implied. Maxxam has been retained to provide analysis of samples provided by the Client using the testing methodology referenced in this report. Interpretation and use of test results are the sole responsibility of the Client and are not within the scope of services provided by Maxxam, unless otherwise agreed in writing.

Solid sample results, except biota, are based on dry weight unless otherwise indicated. Organic analyses are not recovery corrected except for isotope dilution methods.

Results relate to samples tested.

This Certificate shall not be reproduced except in full, without the written approval of the laboratory.

Reference Method suffix "m" indicates test methods incorporate validated modifications from specific reference methods to improve performance.

\* RPDs calculated using raw data. The rounding of final results may result in the apparent difference.

(1) This test was performed by Sub Burnaby to EMSL (NJ)

(2) The matrix is non-food and is outside of the scope of the method. Sample(s) analyzed have not been subjected to Maxxam's standard validation process for the submitted matrix and is not an accredited method.

Maxia Stroup Company

Your Project #: 13953-103 Your C.O.C. #: K019946, K019947, K019948

#### **Attention: CRAIG PATTERSON**

KEYSTONE ENVIRONMENTAL LTD SUITE 320 4400 DOMINION STREET BURNABY, BC CANADA V5G 4G3

> Report Date: 2018/05/10 Report #: R2552492 Version: 4 - Revision

### **CERTIFICATE OF ANALYSIS – REVISED REPORT**

MAXXAM JOB #: B823570 Received: 2018/03/29, 17:17

**Encryption Key** 

Nancy Niklis Project Manager 10 May 2018 11:01:59

Please direct all questions regarding this Certificate of Analysis to your Project Manager. Nancy Niklis, Project Manager Email: NNiklis@maxxam.ca Phone# (604) 734 7276

Maxxam has procedures in place to guard against improper use of the electronic signature and have the required "signatories", as per section 5.10.2 of ISO/IEC 17025:2005(E), signing the reports. For Service Group specific validation please refer to the Validation Signature Page.



KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

Maxxam ID		TE7267	TE7268							
Sampling Date		2018/03/29 13:00	2018/03/29 13:30							
COC Number		K019948	K019948							
	UNITS	SW18-05	SW18-06	QC Batch						
External Sublet Analysis	External Sublet Analysis									
Subcontract Parameter	N/A	ATTACHED	ATTACHED	8982520						

### **RESULTS OF CHEMICAL ANALYSES OF DRINKING WATER**



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Maxxam Job #: B823570 Report Date: 2018/05/10

#### KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

# **MICROBIOLOGY (DRINKING WATER)**

Maxxam ID		TE7265	TE7266	TE7267	TE7268	TE7269	TE7270	TE7271		
Sampling Date		2018/03/29 11:30	2018/03/29 12:00	2018/03/29 13:00	2018/03/29 13:30	2018/03/29 14:00	2018/03/29 15:15	2018/03/29 15:45		
COC Number	-	K019948								
	UNITS	SW18-01	SW18-02	SW18-05	SW18-06	SW18-07	SW18-09	SW18-10	RDL	QC Batch
Microbiological Param.										
Fecal Coliforms	CFU/100mL	<1	7	1	1	<1	<1	6	1	8948217
Total Coliforms	CFU/100mL	11	79	780	980	0	0	0	N/A	8948211
E. coli	CFU/100mL	4.0	67	100	940	0	0	0	N/A	8948211
RDL = Reportable Detection	Limit									

N/A = Not Applicable

Maxxam ID		TE8793	
Sampling Date		2018/03/29	
		11:30	
COC Number		K019948	
	UNITS	SW18-A	QC Batch
Microbiological Param.			
Fecal Coliforms	CFU/100mL	0	8949386
Total Coliforms	CFU/100mL	10	8949384
E. coli	CFU/100mL	4.0	8949384



Report Date: 2018/05/10

KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

### **RESULTS OF CHEMICAL ANALYSES OF SEDIMENT**

Maxxam ID		TE7258	TE7259	TE7260	TE7263	TE7264	
Sampling Date		2018/03/29 13:00	2018/03/29 13:30	2018/03/29 14:00	2018/03/29 15:15	2018/03/29 15:45	
COC Number		K019946	K019946	K019946	K019947	K019947	
	UNITS	SED18-05	SED18-06	SED18-07	SED18-09	SED18-10	QC Batch
External Sublet Analysis							
Subcontract Parameter	N/A	ATTACHED	ATTACHED	ATTACHED	ATTACHED	ATTACHED	8964567



KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

# **PHYSICAL TESTING (SEDIMENT)**

Maxxam ID		TE7252	TE7252	TE7253	TE7258	TE7259	TE7260	TE7263		
Sampling Date		2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29		
Sampling Date		11:30	11:30	12:00	13:00	13:30	14:00	15:15		
COC Number		K019946	K019946	K019946	K019946	K019946	K019946	K019947		
	UNITS	SED18-01	SED18-01 Lab-Dup	SED18-02	SED18-05	SED18-06	SED18-07	SED18-09	RDL	QC Batch
Physical Properties										
Moisture	%	74	72	26	74	39	55	28	0.30	8948334

RDL = Reportable Detection Limit

Lab-Dup = Laboratory Initiated Duplicate

Maxxam ID		TE7264		TE8792					
Sampling Date		2018/03/29		2018/03/29					
Sampling Date		15:45		11:30					
COC Number		K019947		K019946					
	UNITS	SED18-10	QC Batch	SED18-A	RDL	QC Batch			
Physical Properties									
Moisture	%	21	8948334	70	0.30	8950379			
Moisture         %         21         8948334         70         0.30         8950379           RDL = Reportable Detection Limit									



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Maxxam Job #: B823570 Report Date: 2018/05/10

#### KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

# **MICROBIOLOGY (SEDIMENT)**

Maxxam ID		TE7252	TE7253	TE7258	TE7259	TE7260	TE7263	TE7264		
Sampling Date		2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29		
Sampling Date		11:30	12:00	13:00	13:30	14:00	15:15	15:45		
COC Number		K019946	K019946	K019946	K019946	K019946	K019947	K019947		
	UNITS	SED18-01	SED18-02	SED18-05	SED18-06	SED18-07	SED18-09	SED18-10	RDL	QC Batch
Microbiological Param.										
E. coli	MPN/100g	<20	<20	<20	<20	<20	210	45	20	8948187
Total Coliforms	MPN/100g	<20	<20	240000	45	3300	210	140	20	8948185
RDL = Reportable Detection L	imit									

Maxxam ID		TE8792						
Sampling Date		2018/03/29 11:30						
COC Number		K019946						
	UNITS	SED18-A	RDL	QC Batch				
Microbiological Param.								
Microbiological Param.								
<b>Microbiological Param.</b> E. coli	MPN/100g	<20	20	8949393				
	MPN/100g MPN/100g	<20 <20	20 20	8949393 8949389				



KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

### **RESULTS OF CHEMICAL ANALYSES OF SOIL**

Maxxam ID		TE7262						
Sampling Date		2018/03/29 14:45						
COC Number		K019947						
	UNITS	SS18-08 (0.5)	QC Batch					
External Sublet Analysis								
Subcontract Parameter	N/A	ATTACHED	8964567					



KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

# **PHYSICAL TESTING (SOIL)**

Maxxam ID		TE7254	TE7255	TE7256	TE7257	TE7261	TE7262			
Sampling Date		2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29	2018/03/29			
Sampling Date		12:30	12:35	12:45	12:50	14:30	14:45			
COC Number		K019946	K019946	K019946	K019946	K019946	K019947			
	UNITS	SS18-03 (0.3)	SS18-03 (0.5)	SS18-04 (0.3)	SS18-04 (0.5)	SS18-08 (0.3)	SS18-08 (0.5)	RDL	QC Batch	
Physical Properties										
Moisture	%	36	12	11	11	26	18	0.30	8948334	
RDL = Reportable Detection Limit										



KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

# **MICROBIOLOGY (SOIL)**

Maxxam ID		TE7254	TE7255	TE7256	TE7257	TE7261	TE7262			
Sampling Date		2018/03/29 12:30	2018/03/29 12:35	2018/03/29 12:45	2018/03/29 12:50	2018/03/29 14:30	2018/03/29 14:45			
COC Number		K019946	K019946	K019946	K019946	K019946	K019947			
	UNITS	SS18-03 (0.3)	SS18-03 (0.5)	SS18-04 (0.3)	SS18-04 (0.5)	SS18-08 (0.3)	SS18-08 (0.5)	RDL	QC Batch	
Microbiological Param.										
E. coli	MPN/100g	<20	<20	<20	<20	<20	<20	20	8948187	
Total Coliforms	MPN/100g	<20	<20	<20	<20	3500	7000	20	8948185	
RDL = Reportable Detection Limit										



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KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

### **GENERAL COMMENTS**

Each tempera	ature is the aver	age of up to thr	ee cooler temperatures taken at receipt
Pack	kage 1	11.3°C	
• .	est: Report to in pectively, as per		r total, fecal coliforms and E. coli on samples SED18-A and SW18-A, duplicates of samples SED18-01 and on 2018/04/02.
Sample(s) ana	alyzed past hold	time. Analysis p	performed with client's consent.
	port reissued to d SED18-10 as p		tracted results for bacteriodes and human bacteriodes on samples SED18-05, SED18-07, SS18-08 (0.5), 018/04/12
•		•	alyzed past recommended hold time for Total Coliforms (MTF) in Soil. Sample was analyzed past (MTF) in Soil. Sample was analyzed past recommended hold time for Fecal Coliforms (MTF) in Soil - Wet.
•	• •	•	yzed past method specific hold time for Fecal Coliform membrane filt - Potable W. Sample was analyzed rm/E.Coli by MF-Chromocult(PW).
Results relate	e only to the ite	ms tested.	



### **QUALITY ASSURANCE REPORT**

KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

			Method Bl	ank	RPD				
QC Batch	Parameter	Date	Value	UNITS	Value (%)	QC Limits			
8948334 Moisture 2018/04/02 <0.30 % 2.5 20									
8950379 Moisture 2018/04/04 <0.30 % 3.1 20									
Duplicate: Paired analysis of a separate portion of the same sample. Used to evaluate the variance in the measurement.									
Method Blank: A b	Method Blank: A blank matrix containing all reagents used in the analytical procedure. Used to identify laboratory contamination.								



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KEYSTONE ENVIRONMENTAL LTD Client Project #: 13953-103 Sampler Initials: BTL

#### VALIDATION SIGNATURE PAGE

The analytical data and all QC contained in this report were reviewed and validated by the following individual(s).

Bhana

Mandheraj Chana, Junior Project Manager

Rob Reinert, B.Sc., Scientific Specialist

Maxxam has procedures in place to guard against improper use of the electronic signature and have the required "signatories", as per section 5.10.2 of ISO/IEC 17025:2005(E), signing the reports. For Service Group specific validation please refer to the Validation Signature Page.



# CHAIN OF CUSTODY RECORD

K 019946

A Bureau V	eritas Group Company	Burnaby: 4606 Canad	a Way, Burnaby, BC \	/5G 1K5. Te	oll Free (	800) 665	5-8566								BBY FCD-00077/07 Page ) of 3
	Invoice Information		Report Information (	if differs fr	om invo	ice)			Proje	ct Informatio	n (where	e applicable)			Turnaround Time (TAT) Required
Company Name:	3763 - Keystone Environmen	tal Ltd. Company Nam	KEL	/				Quotati	on #:						Regular TAT 5 days (Most analyses)
Contact Name:	Crali Patterso	Contact Name:	Sun	Q.				P.O. #/ /	AFE#:	1395.	3-	103		PLEA	SE PROVIDE ADVANCE NOTICE FOR RUSH PROJECTS
Address:	#320 - 4400 Dominion Street	Address:	Sun	re				Project	ŧ:						Rush TAT (Surcharges will be applied)
	Burnaby, BC PC: V5G 4G3		-		PC:		(1):2-	Site Loca	tion:						Same Day 2 Days
Phone: (604) 43		Phone:	/					Site #:	-	2		1			🗌 1 Day 🔲 3 Days
Email: CSP	Herson@keyste	accause and Tu	·cu					Sampled	I By:	<u></u>	2+	KM	-	Date	Required:
	Regulatory Criteria		Special Instructions					· · · · ·	Analysis	Requested		0	_	Rush	Confirmation #:
BC CSR Soil	BC CSR Wa	iter										all's			LABORATORY USE ONLY
YK CSR Soil	YK CSR Wa	ter 🗌	Return Cooler		MTBE		perved.	erved		a 00 4		and the			CUSTODY SEAL
CCME (Specify	y) 🗌 Other (Spe	-16.0			Ha		F2 - F4	Pres d?	8	Alkalin	Ammo	25			Y / N COOLER TEMPERATURES
			Ship Sample Bottles (Please Specify)		VOC / BTEXS / VPH	ТЕРН/НЕРН/РАН			eserve			ese	8		Present Intact
Drinking Wate	er D BC Water (	Dunlitu			C/BT	HIHE	Filtered?	Filtered? Field Pre	Field Pr	BOI	- -	the	BMITT	YZE	NIA 11,10,13
						-	15H S	v u	u.	Fluorio	Nitrate	an	CRS SU	ANAL	
SAMPLES M	UST BE KEPT COOL ( < 10 °C ) FROM	TIME OF SAMPLING UNTIL	DELIVERY TO MAXXA	M	Hely I		1 Meta	d Merc tals	L	] j		EECO	TAIN	DO NOT ANALYZE	JUST SAMPLE
,	Sample Identification	Date Sa (YYYY/M		Matrix	BTEXS / VP BTEX / F1	PAH	EPH Dissolves	Dissolved Total Met	Total Me	TISS Chioride	Nitrite	10 to	# OF CONTAINERS SUBMITTED	HOLD - C	COOLING MEDIA PRESENT 0 / N COMMENTS
1 SED	18-01	20/8/2		selat							1 k	XX	3		DB Total gul
1	13-02	1	12:00	Alt	(				$\square$			XXX	3		Aunung berteinste
	8-03 (0.3)		12:30	50:1							1	XXX	3		sumples to be
	8-03 (0.5)		12:35	soil					П			XX	3		placed on holo).
10	8-04 (0.3)		2:45	soil								XX	3		
Visi 1 1996 COURSE	8-04 (0.5)		the second s	sil					T			- WW	3		
	218-05		13:00	adart	-								3		
	18-06			dist	-						T Ś	NVV	3		
	218-07		14:00	AL.	-						tκ	100	3		
	8-08 (0.3)		- A Management	soi/		+			+	++-	10		13		
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Solar.	- Bretthmus 201	3102/2017	001	1110	Iron		TAN		2.0	18/03/7	0	H:17			
-	Dicurans 201	0/05/29.1	1	μν	- CD		LHU		10	10100/	4	17317	-		NUL PROPERTY AND A SPACE AND A DECIDE
						_							T		
mess otherwise agreed to	o in writing, work submitted on this Chain of G	ustody is subject to Maxiam's stand	ard Terms and Conditions.	Signing of this	Chain of C	ustody doc	ument is ack	nowledgmen	t and accep	tance of our term	is which are	available for viewing	at w	<b>B</b> 8	823570_COC

COC-1027 Keystone



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### CHAIN OF CUSTODY RECORD

BBY FCD-00077/07

St ....

K 019947

Inpany Name:	pie Bottles pecify)			N	P.O. # Projec Site Lc Site #: Sampl	AFE#: / AFE#: t #: boation: Analysi Cpawasaul play Lpawasaul play		Conductivity Alkalinity			# OF CONTAINERS SUBMITTED	Date	Regular TAT     Rush TAT (Surchar)     Same Day     1 Day     te Required:     th Confirmation #:     LABORAT     Y / N     Present Intact	TORY USE ONLY
itact Name: iress: ires: if Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (YVY/IAM/OD)	nstructions ooler ple Bottles pecify) TO MAXXAM	ирн 🔲 voc/втехs/ирн 🗍 Мтве 🗌		N	P.O. # Projec Site Lc Site #: Sampl	/ AFE#: t #: ccation: led By: Analysi Cpausaud pials Cpausaud pials	Fluoride Sulphate Sulphate	sted	Sendining	ut tead celitains	IS SUBMITTED	Date	ASE PROVIDE ADVANCE I Rush TAT (Surchar, Same Day 1 Day te Required: th Confirmation #: LABORAT CUSTODY SEAL Y / N Present Intact	NOTICE FOR RUSH PROJE rges will be applied) 2 Days 3 Days TORY USE ONLY COOLER TEMPERATURES
Iress: me: Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (YVY/IMM/OD)	ple Bottles pecify) TO MAXXAM			N	Project Site Lc Site #: Sampl	tt #: bocation: led By: Analysi Cpanasald pial3 Cpanasald pial3	Fluoride Sulphate Sulphate	sted	Sendining	ut tead celitains	IS SUBMITTED	Date	Rush TAT (Surchar Same Day 1 Day te Required: th Confirmation #: LABORAT CUSTODY SEAL Y / N Present Intact	TORY USE ONLY COOLER TEMPERATURES
Ine: Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (Yvyy (AMA/OD)	ple Bottles pecify) TO MAXXAM			N	Site Lc	ed By: Analysi Charleseved	Fluoride Sulphate Sulphate	sted	Sendining	at tead celiptins	ISS SUBMITTED	Rush	Same Day          I Day         I Day         te Required:         th Confirmation #:         LABORAT         CUSTODY SEAL         Y / N         Present       Intact	COOLER TEMPERATURES
Special In: Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (VVV (MM/OD)	ple Bottles pecify) TO MAXXAM			N	Site #: Sampl	Interferenced Preserved?	Fluoride Sulphate	Conductivity   Akailinity   Application		ut tead celitains from brother of of the	RS SUBMITTED	Rush	1 Day te Required: th Confirmation #:     LABORAT     CUSTODY SEAL     Y / N Present Intact	3 Days TORY USE ONLY COOLER TEMPERATURES
Special In: Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (VVV (MM/OD)	ple Bottles pecify) TO MAXXAM			N	Sampl	Analysi	Fluoride Sulphate	Conductivity Alkalinity		ul Real colitains	RS SUBMITTED	Rush	CUSTODY SEAL Y / N Present Intact	TORY USE ONLY COOLER TEMPERATURES
Special In: Return Co Ship Samp (Please Sp UNG UNTIL DELIVERY Date Sampled (YVY/MM/OD)	ple Bottles pecify) TO MAXXAM			N	rcury Filtered?	Field Preserved?	Fluoride Sulphate	Conductivity Alkalinity		ul Real colitains	RS SUBMITTED	Rush	CUSTODY SEAL Y / N Present Intact	COOLER TEMPERATURES
Return Co     Ship Samp     (Please Sp     (Pl	ple Bottles pecify) TO MAXXAM			N	roury	Field Preserved?	Fluoride Sulphate	Conductivity Alkalinity		ul terred colitains	RS SUBMITTED		LABORAT CUSTODY SEAL Y / N Present Intact	COOLER TEMPERATURES
UNG UNTIL DELIVERY	ple Bottles pecify) TO MAXXAM Time			N	roury	Field Press Field Press				ul fead celitains	RS SUBMITTED	r Analyze	CUSTODY SEAL Y / N Present Intact	COOLER TEMPERATURES
2018/03/28	(HH:MM)	t is	BTEX/F1	EPH	Disolved Me	Total Metals Total Mercury	Chloride C		XXXE.co	XXX 10th 10	W W # OF CONTAINE		De Total a preteride.	Sent of Anna
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	) TIME: (НН:М ]7:0С	D) TIME: (HH:MM) REC 17:00	D) TIME: (HH:MM) RECEIVED	D) TIME: (HH:MM) RECEIVED BY: (Signa 17:00 Mu FORE	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print 17:00 MUL FORD TA	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) 17:00 MUL FORD TACK	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DA' 17:00 MUL PEORO TACK JU	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DATE: (YYY) 17:00 MUU FORO TACK LO18/	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DATE: (YYYY/MM/DD) 17:00 JULY FORD TACK LO18/03/29	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DATE: (YYYY/MM/DD) TIME 17:00 MUL FORD TACK 2018/03/29 F	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DATE: (YYYY/MM/DD) TIME: (HH:MM) 17:00 MUL FORD TACK 2018/03/29 F7>17	D)         TIME: (HH:MM)         RECEIVED BY: (Signature/Print)         DATE: (YYYY/MM/DD)         TIME: (HH:MM)	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) DATE: (YYYY/MM/DD) TIME: (HH:MM) 17:00 MUL EORO TACK D018/03/29 F2:17 axeem's standard Terms and Carolitons. Spring of this Chain of Custody document is acknowledgement and acceptance of our terms which are available for viewing at w	D) TIME: (HH:MM) RECEIVED BY: (Signature/Print) 17:00 MUL FORD TACK JO18/03/29 F=:17

COC-1027 Keystone



## CHAIN OF CUSTODY RECORD

BBY	FCD-00	0077/	07

K 019948

Invoice Information	Report Inform	nation (if differs fro	om invoice)	Project Informat	ion (where applicable)	Turnaround Time (TAT) Required
mpany Name: 3763 - Keystone Environmental Ltd.	Company Name:	SAM	E	Quotation #:		Regular TAT 5 days (Most analyses)
intact Name: Craig Putterson	Contact Name:			P.O. #/ AFE#:		PLEASE PROVIDE ADVANCE NOTICE FOR RUSH PROJECT
dress: #320 - 4400 Dominion Street	Address:			Project #: 139	153-103	Rush TAT (Surcharges will be applied)
Burnaby, BC PC: V5G 4G3			PC:	Site Location:		Same Day 🗌 2 Days
one: (604) 430-0671	Phone:		3 28	Site #:		🗌 1 Day 🔲 3 Days
nail: CSputterson@keydenlewinound	Email:			Sampled By: BT	L + EM	Date Required:
Regulatory Criteria	Special Instru	uctions		Analysis Requested	0	Rush Confirmation #:
BC CSR Soil DC CSR Water						LABORATORY USE ONLY
YK CSR Soil YK CSR Water	Return Coole	er	MTBE	ved?	No.	CUCTODA (CAL)
	1000		лен [] Н [] Н [] F2 - F4	Preser hate	Ammoni	CUSTODY SEAL Y / N COOLER
CCME (Specify) Other (Specify)	Ship Sample I	Bottles	HAV/	Ved?	A C A	Present Intact
	(Please Specif		BTEXS BTEX / HEPH/	Prese		
Drinking Water BC Water Quality			VOC / BTEXS / VPH VOC / BTEX / F1 LEPH/HEPH/РАН H	Fitered? Field Pre- Field Pre- oride [ 900	Late Late	\$ AVA 11,10,13
				Fluo	Nitrate	TAN
SAMPLES MUST BE KEPT COOL ( < 10 °C ) FROM TIME OF	SAMPLING UNTIL DELIVERY TO	MAXXAM	H N	1 Merr	Nuclue and Nuclear and Nuclear and Nuclear and Action a	TUT DU 10,15
Sample Identification	Date sampled	ime npled Matrix	BTEXS / VPH BTEX / F1 PAH EPH E	Dissolved M Total Metals Total Mercu Chloride		COOLING MEDIA PRESENT
The second se	(YYYY/MM/DD) (HH	(MM)	BTEX BTEX PAH EPH Disso	Diss Tota Chio Chio		
SW18-01	2018/03/29 11	:30 unter			XXX 3	D total and hear
5618-02	1 12	:00 miter			XXX 3	bicteriste samples
SW18-05		:00 unter			VXX 3	to be ducation
SW18-06	13	:30 mile			XXX 3	111
SW18-07		:00 water			XXX 3	199
5418-09	Sec. 22 (1997)		+++-	+++++	XXX 3	+ +
		s:15 water				
SW18-10	18:	:45 water			XXX 3	
4						
						<del>   </del>
RELINQUISHED BY: (Signature/Print) DATE: (YYYY/	MM/DD) TIME: (HH:MM)	RECEIV	/ED BY: (Signature/Print	) DATE: (YYYY/MIN	1/DD) TIME: (HH:MM)	
Blas / Biet/ Lucas TO13/03,	179 17:00	01.00	Dans			
Blas / Brett Lucas 2018/03,	0 11.00	m	HEORO TA	EK 2013/03/	19 F:F	

COC-1027 Keystone

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200 Route 130 North, Cinnaminson, NJ 08077 Phone/Fax: (800) 220-3675/ 786-0262 <u>http://www.emsl.com</u> E-mail: <u>Dnalab2@EMSL.com</u>



Client:	Maxxam Analytics	EMSL Order ID:	611800666
	4606 Canada Way	Date Received:	4/16/2018
	Burnaby, BC V5G 1K5	Date Analyzed:	4/19/2018
Attn:	Nancy Niklis	Date Reported:	4/19/2018
Project:	B823570	Date Amended:	

# Rapid Detection of Total Bacteroides by Quantitative PCR

EMSL Test: M095

Lab Sample Number	Client Sample ID	Description	Soil Received	Soil Sampled (g)	CEs */g Soil
0666-1	TE7258-SED18-05	N/A	Bulk	1.2313	902,216
0666-2	TE7260-SED18-07	N/A	Bulk	1.2613	5,749
0666-3	TE7262-SS18-08 (0.5)	N/A	Bulk	1.1335	None Detected
0666-4	TE7263-SED18-09	N/A	Bulk	1.214	None Detected
0666-5	TE7264-SED18-10	N/A	Bulk	1.2066	None Detected

CEs: Cell Equivalent, measured by PCR using genomic DNA standards.

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**Sergey Balashov, Ph.D.** PCR Laboratory Director

200 Route 130 North, Cinnaminson, NJ 08077 Phone/Fax: (800) 220-3675/ 786-0262 <u>http://www.emsl.com</u> E-mail: <u>Dnalab2@EMSL.com</u>



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Attn:	Nancy Niklis	Date Reported:	4/19/2018
Project:	B823570	Date Amended:	

# Rapid Detection of Human Bacteroides by Quantitative PCR

EMSL Test: M199

led (g) CEs* /g Soil
3 None Detected
3 None Detected
5 None Detected
None Detected
6 None Detected

Note: The qPCR assay for human *Bacteroides* is based on HF183 marker which was evaluated by EPA scientists (SAM, 33, 2010). The qPCR detects human specific *Bacteroides* predominantly.

CEs: Cells Equivalent, measured by PCR using genomic DNA standards.

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Client:	Maxxam Analytics	EMSL Order ID:	611800780
	4606 Canada Way	Date Received:	5/3/2018
	Burnaby, BC V5G 1KS	Date Analyzed:	5/4/2018
Attn:	Nancy Niklis	Date Reported:	5/8/2018
Project:	B823570	Date Amended:	5/10/2018

# Rapid Detection of Total Bacteroides by Quantitative PCR

EMSL Test: M095

Lab Sample Number	Client Sample ID	Description	Soil Received	Soil Sampled (g)	CEs */g Soil
0780-1	TE7259-SED18-06	N/A	Bulk	1.368	25,780

CEs: Cell Equivalent, measured by PCR using genomic DNA standards.

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**Sergey Balashov, Ph.D.** PCR Laboratory Director

Retort amendment date 05/10/2018 following customer's request0780-1TE7259-SED18-06Sample name is corrected

200 Route 130 North, Cinnaminson, NJ 08077 Phone/Fax: (800) 220-3675/ 786-0262 http://www.emsl.com E-mail: Dnalab2@EMSL.com



Client:	Maxxam Analytics	EMSL Order ID:	611800780
	4606 Canada Way	Date Received:	5/3/2018
	Burnaby, BC V5G 1KS	Date Analyzed:	5/4/2018
Attn:	Nancy Niklis	Date Reported:	5/8/2018
Project:	B823570	Date Amended:	5/10/2018

# Rapid Detection of Human *Bacteroides* by Quantitative PCR

EMSL Test: M199

Lab Sample Number	Client Sample ID	Description	Bulk Received	Bulk Sampled (g)	CEs* /g Soil
0780-1	TE7259-SED18-06	N/A	Bulk	1.368	None Detected

Note: The qPCR assay for human *Bacteroides* is based on HF183 marker which was evaluated by EPA scientists (SAM, 33, 2010). The qPCR detects human specific *Bacteroides* predominantly.

CEs: Cells Equivalent, measured by PCR using genomic DNA standards.

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Retort is amended 05/10/2018 following customer's request0780-1TE7259-SED18-06Sample name is corrected

**Sergey Balashov, Ph.D.** PCR Laboratory Director

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Client:	Maxxam Analytics 4606 Canada Way	EMSL Order ID: Date Received:	
	Burnaby, BC V5G 1KS	Date Analyzed:	5/4/2018
Attn.	Nancy Niklis	Date Reported:	5/8/2018
Project:	B823570	Date Amended:	5/9/2018

# Rapid Detection of Total *Bacteroides* by Quantitative PCR

EMSL Test: M095

Lab Sample Number	Client Sample ID	Description	Water Received (mL)	Water Sampled (mL)	CEs*/100 mL
0780-2	TE7267-SW18-05	N/A	200	200	27,526
0780-3	TE7268-SW18-06	N/A	200	200	None Detected

CEs: Cell Equivalent, measured by PCR using genomic DNA standards.

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Retort amendment date 05/09/2018 following customer's request

0780-3 TE7268-SW18-06 Sample name corrected

Sergey Balashov, Ph.D. PCR Laboratory Director

# EMSL Analytical, Inc. 200 Route 130 North, Cinnaminson, NJ 08077 Phone/Fax: (800) 220-3675/ 786-0262 http://www.emsl.com E-mail: Dnalab2@EMSL.com



Client:	Maxxam Analytics	EMSL Order ID:	611800780
	4606 Canada Way	Date Received:	5/3/2018
	Burnaby, BC V5G 1KS	Date Analyzed:	5/4/2018
Attn.	Nancy Niklis	Date Reported:	5/8/2018
Project:	B823570	Date Amended:	5/10/2018

# Rapid Detection of Human *Bacteroides* by Quantitative PCR

EMSL Test Code: M199

Lab Sample Number	Client Sample ID	Description	Water Received (mL)	Water Sampled (mL)	CEs /100 mL
0780-2	TE7267-SW18-05	N/A	200	200	None Detected
0780-3	TE7268-SW18-06	N/A	200	200	None Detected

Note: The qPCR assay for human *Bacteroides* is based on HF183 marker which was evaluated by EPA scientists (SAM, 33, 2010). The qPCR detects human specific *Bacteroides* predominantly.

CEs: Cells Equivalent, measured by PCR using genomic DNA standards.

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Sergey Balashov, Ph.D. PCR Laboratory Director

Retort amendment date 05/10/2018 following customer's request0780-3TE7268-SW18-06Sample name corrected