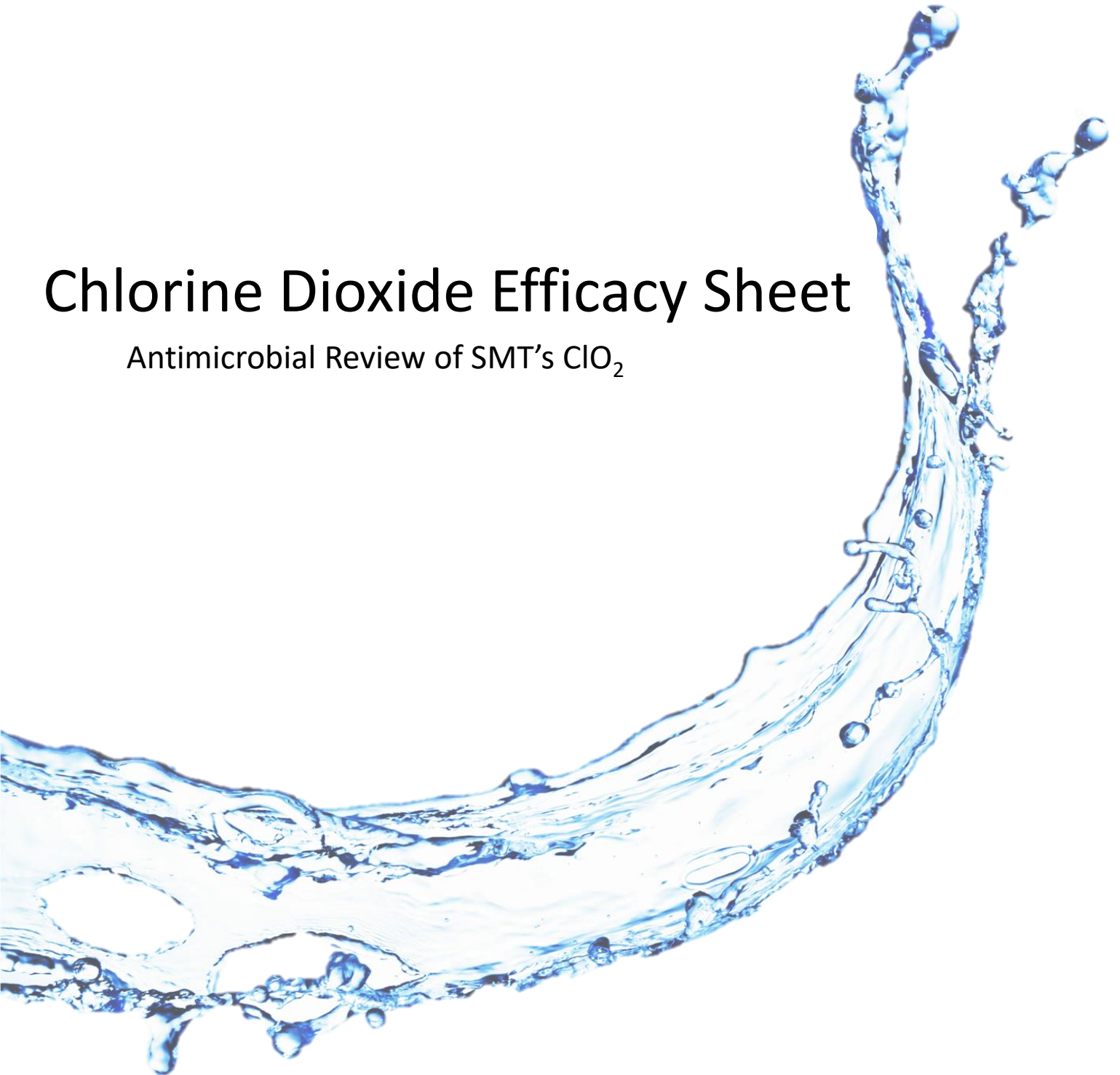




Chlorine Dioxide Efficacy Sheet

Antimicrobial Review of SMT's ClO_2



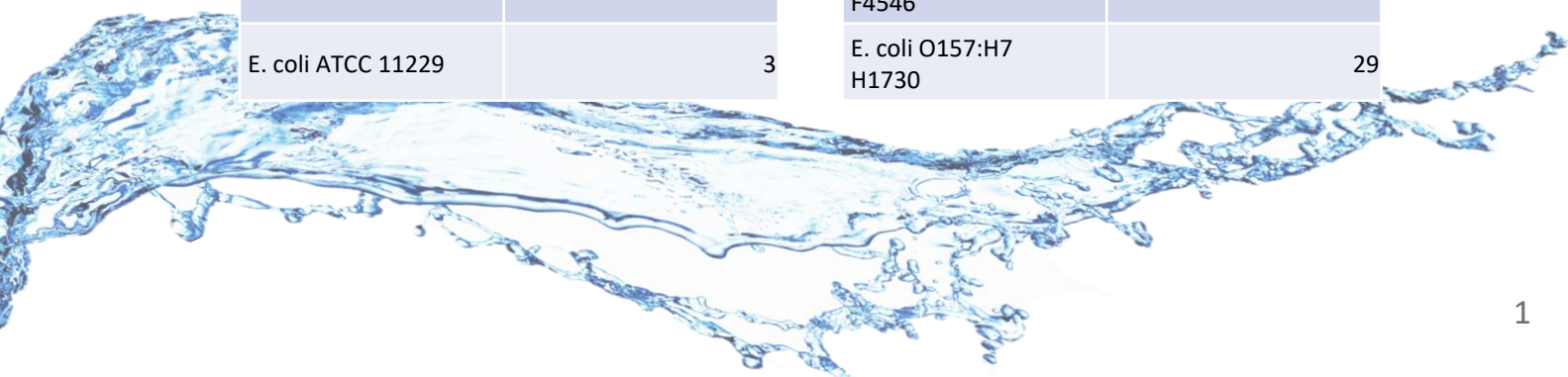


Selective Micro Technologies

Chlorine Dioxide Efficacy Sheet

Chlorine Dioxide has proven to be effective against a range of microbial pathogens. The following is a list of studies demonstrating the antimicrobial capabilities of chlorine dioxide against a range of organisms. These entries are sorted by name and pathogen classification (bacteria, virus, algae/fungi/mold/yeast, bacterial spores, dangerous chemicals, beta lactams, protozoa, or microsporidia). Please note that although no organism has ever proven to be resistant to chlorine dioxide, this list is not a comprehensive reference of chlorine dioxide’s full pathogenic capabilities. For further information chlorine dioxide’s biocidal efficacy against organisms not listed here, please reach out to us at inquiry@selectivemicro.com or 855.256.8299.

Bacteria	Reference No.	Bacteria	Reference No.
Blakeslea trispora	2	E. coli ATCC 51739	38
Bordetella bronchiseptica	8	E. coli K12	38
Brucella suis	30	E. coli K12 W3110	44
Burkholderia mallei	36	E. coli O157:H7 13B88	38
Burkholderia pseudomallei	36	E. coli O157:H7 204P	38
Campylobacter jejuni	39	E. coli O157:H7 43895	38
Clostridium botulinum	32	E. coli O157:H7 EDL933	13
Corynebacterium bovis	8	E. coli O157:H7 G5303	38
Coxiella burneti	35	E. coli O157:H7 F4546	29
E. coli ATCC 11229	3	E. coli O157:H7 H1730	29



Bacteria	Reference No.
<i>E. coli</i> O157:H7 E0018	29
<i>E. coli</i> O157:H7 C7927	38
<i>Erwina carotovora</i>	21
<i>Franscicella tularensis</i>	30
<i>Fusarium sambucinum</i>	21
<i>Fusarium solani</i> var. <i>coeruleum</i>	21
<i>Helicobacter pylori</i>	2
<i>Helminthosporium solani</i>	21
<i>Klebsiella pneumoniae</i>	3
<i>Lactobacillus acidophilus</i> NRRM B1910	38
<i>Lactobacillus brevis</i>	38
<i>Lactobacillus buchneri</i>	38
<i>Lactobacillus plantarum</i>	5
<i>Legionella</i>	2
<i>Legionella pneumophila</i>	42
<i>Leuconostoc mesenteroides</i>	5
<i>Listeria innocua</i> ATCC 33090	38
<i>Listeria monocytogenes</i> F4248	38

Bacteria	Reference No.
<i>Listeria monocytogenes</i> serotype 4b, F8027	29
<i>Listeria monocytogenes</i> serotype 1/2b F8255	29
<i>Listeria monocytogenes</i> serotype 1/2a, F8369	29
<i>Listeria monocytogenes</i> serotype 4b, G1091	29
<i>Listeria monocytogenes</i> serotype 1/2a, H0222	29
<i>Listeria monocytogenes</i> F5069	19
<i>Listeria monocytogenes</i> LCDC-81-861	38
<i>Listeria monocytogenes</i> LCDC-81-886	19
<i>Listeria monocytogenes</i> Scott A	38
Methicillin-resistant <i>Staphylococcus aureus</i> (MRSA)	3
Multiple Drug-Resistant <i>Salmonella</i> (MDRS)	3
<i>Mycobacterium bovis</i>	28
<i>Mycobacterium fortuitum</i>	42
<i>Pediococcus acidilactici</i>	38
<i>Pediococcus acidilactici</i> AB1	44
<i>Pseudomonas aeruginosa</i>	3
<i>Pseudomonas fluorescens</i>	44

Bacteria	Reference No.
Salmonella spp.	16
Salmonella Agona	38
Salmonella Anatum Group E	38
Salmonella Choleraesins ATCC 13076	38
Salmonella choleraesuis	8
Salmonella Enterica (PT30) BAA-1045	38
Salmonella Gaminara	29
Salmonella Michigan	29
Salmonella Enterica Baildon	29
Salmonella Enterica S. Enteritidis	13
Salmonella Enterica S. Javiana	13
Salmonella Enterica S. Montevideo	13
Salmonella Enteritidis E190-88	38
Salmonella Gaminarum F2712-OJ	44
Salmonella Javiana	38

Bacteria	Reference No.
Salmonella Newport	4
Salmonella Stanley	44
Salmonella Typhimurium C133117	38
Salmonella Anatum Group E	38
Shigella	2
Staphylococcus aureus	23
Staphylococcus aureus ATCC 25923	38
Staphylococcus faecalis ATCC 344	38
Tuberculosis	3
Vancomycin-resistant Enterococcus faecalis	3
Vibrio strain Da-2	37
Vibrio strain Sr-3	37
Yersina enterocolitica	40
Yersina pestis	30
Yersinia ruckerii ATCC 29473	31



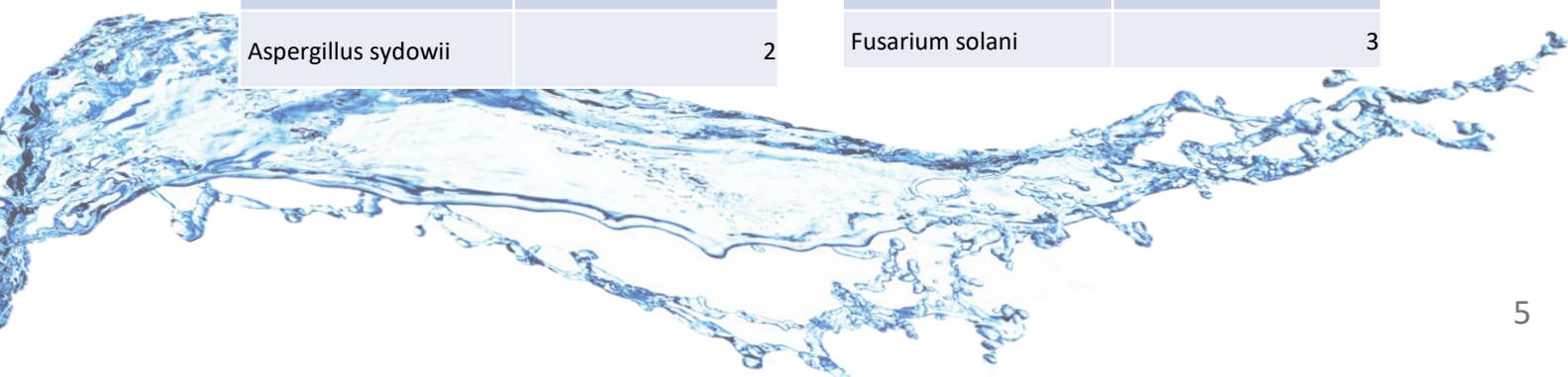
Virus	Reference No.
Adenovirus Type 40	6
Calciavirus	42
Canine Parvovirus	1
Coronavirus	3
Feline Calci Virus	3
Foot and Mouth disease	8
Hantavirus	8
Hepititus A Virus	3
Hepititus B Virus	8
Hepititus C Virus	8
Human coronavirus	1
Human Immunodeficiency Virus	3
Human Rotavirus type 2	15
Influenza A	22
Minute Virus of Mouse (Parvovirus) (MVM-i)	8

Virus	Reference No.
Minute Virus of Mouse (Parvovirus) (MVM-p)	8
Mouse Hepatitis Virus (MHV-A59)	8
Mouse Hepatitis Virus (MHV-JHM)	8
Mouse Parvovirus type 1 (MPV-1)	8
Murine Parainfluenza Virus Type 1 (Sendai)	8
Newcastle Disease Virus	8
Norwalk Virus	8
Poliovirus	20
Rotavirus	3
Severe Acute Respiratory Syndrome (SARS) Coronavirus	43
Sialodscryoadenitis Virus (Coronavirus) (SDAV)	8
Simian rotavirus SA-11	15
Theiler's Mouse Encephalomyelitis Virus (TMEV)	8
Vaccinia Virus	10



Algae/Fungi/Mold/Yeast	Reference No.
Alternaria alternate	26
Aspergillus aeneus	2
Aspergillus aurolatus	2
Aspergillus brunneo uniseriatus	2
Aspergillus caespitosus	2
Aspergillus cervinus	2
Aspergillus clavatonanicus	2
Aspergillus clavatus	2
Aspergillus egyptiacus	2
Aspergillus elongatus	2
Aspergillus fischeri	2
Aspergillus fumigatus	2
Aspergillus giganteus	2
Aspergillus longivesica	2
Aspergillus niger	12
Aspergillus ochraceus	2
Aspergillus parvathecicus	2
Aspergillus sydowii	2

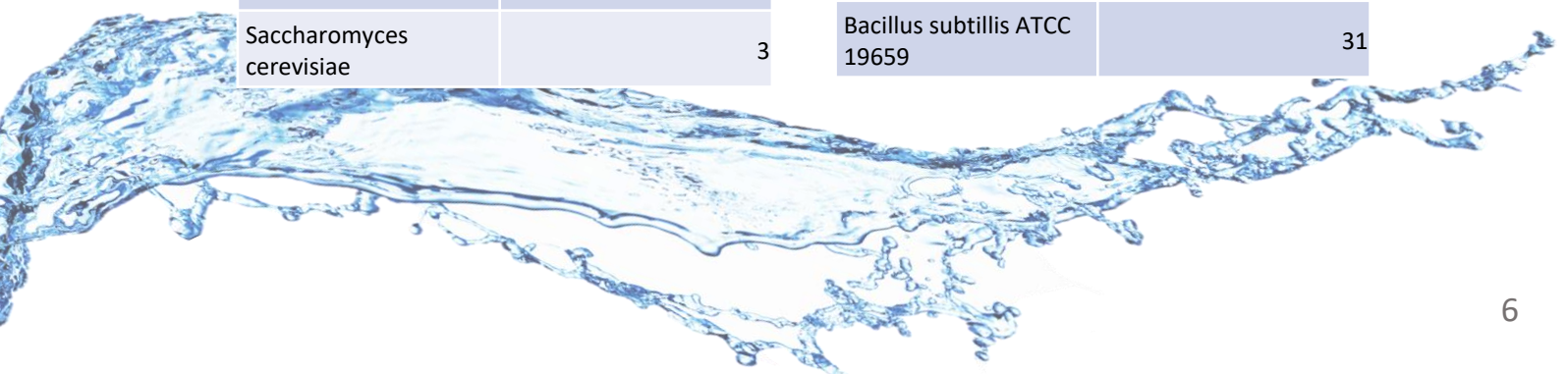
Algae/Fungi/Mold/Yeast	Reference No.
Aspergillus unguis	2
Aspergillus ustus	2
Aspergillus versicolor	2
Botrytis species	3
Candida spp.	5
Candida albicans	2
Candida dubliniensis	2
Candida maltose	2
Candida parapsilosis	2
Candida sake	2
Candida sojae	2
Candida tropicalis	2
Candida viswanathil	2
Chaetomium globosum	7
Cladosporium cladosporioides	7
Debaryomyces etchellsii	2
Eurotium spp.	5
Fusarium solani	3



Algae/Fungi/Mold/Yeast	Reference No.
Lodderomyces elongisporus	2
Mucor circinelloides	2
Mucor flavus	2
Mucor indicus	2
Mucor mucedo	2
Mucor rademosus	2
Mucor ramosissimus	2
Mucor saturnus	2
Penicillium spp.	5
Penicillium chrysogenum	7
Penicillium digitartum	3
Penicillium herquei	2
Phormidium boneri	2
Pichia pastoris	3
Poitrasia circinans	2
Rhizopus oryzae	2
Roridin A	33
Saccharomyces cerevisiae	3

Algae/Fungi/Mold/Yeast	Reference No.
Stachybotrys chartarum	2
Trichophyton mentagrophytes	2
Verrucarin A	2

Bacterial Spores	Reference No.
Alicyclobacillus acidoterrestris	12
Bacillus anthracis	30
Bacillus anthracis Ames	14
Bacillus atrophaeus	31
Bacillus atrophaeus ATCC 49337	12
Bacillus cereus 232	44
Bacillus coagulans	12
Bacillus megaterium	12
Bacillus polymyxa	12
Bacillus pumilus ATCC 27142	12
Bacillus pumilus ATCC 27147	11
Bacillus subtilis (globigii) ATCC 9372	11
Bacillus subtilis ATCC 19659	31



Bacterial Spores	Reference No.
Bacillus subtilis 5230	12
Bacillus thuringiensis	18
Clostridium sporogenes ATCC 19404	12
Geobacillus stearothermophilus ATCC 12980	11
Geobacillus stearothermophilus ATCC 7953	31
Geobacillus stearothermophilus VHP	11

Chemical Agents	Reference No.
Ricin Toxin	10
dihydronicotinamide adenine dinucleotide	24
microcystin-LR (MC-LR)	25
cylindrospermopsin (CYN)	25

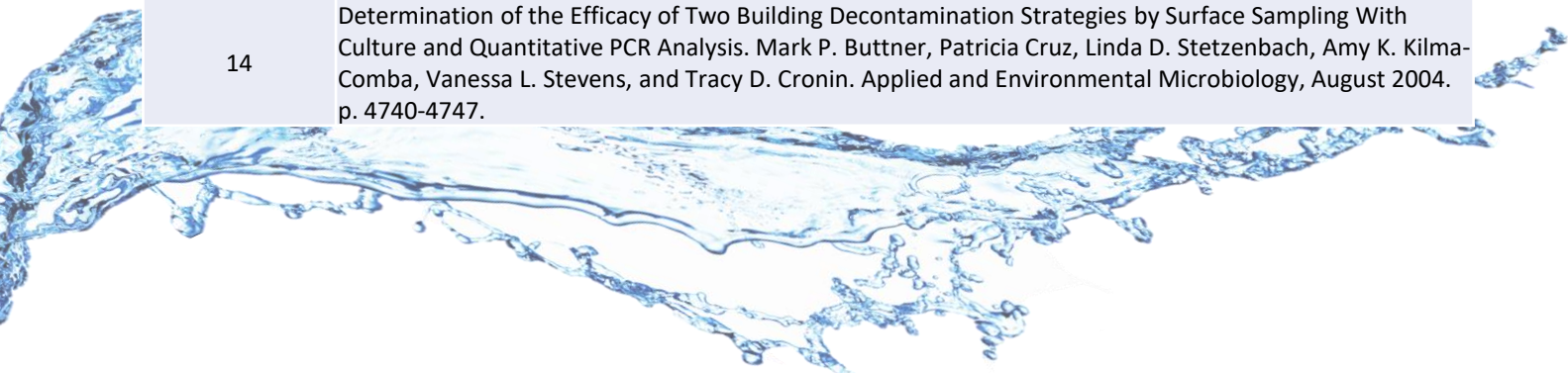
Beta Lactams	Reference No.
Amoxicillin	2
Ampicillin	2
Cefadroxil	2
Cefazolin	2
Cephalexin	2
Imipenem	2
Penicillin G	2
Penicillin V	2

Protozoa	Reference No.
Chironomid larvae	27
Cryptosporidium	34
Cryptosporidium parvum Oocysts	9
Cyclospora cayentanesis oocysts	41
Giardia	34



References

Reference No	Reference
1	Selective Micro Technologies EPA Pesticide Label
2	Biological Efficacy of Chlorine Dioxide, ClorDisys Solutions, Inc., 2018.
3	Chlorine Dioxide, Part 1: A Versatile, High-Value Sterilant for the Biopharmaceutical Industry, Barry Wintner, Anthony Contino, and Gary O'Neal. BioProcess International December 2005.
4	Chlorine Dioxide Gas Decontamination of Large Animal Hospital Intensive and Neonatal Care Units, Henry S. Luftman, Michael A. Regits, Paul Lorcheim, Mark A. Czarneski, Thomas Boyle, Helen Aceto, Barbara Dallap, Donald Munro, and Kym Faylor. Applied Biosafety, 11(3) 144-154 ABSA 2006.
5	Efficacy of Chlorine Dioxide Gas as a Sanitizer for Tanks Used for Aseptic Juice Storage. Y. Han, A.M. Guentart, R.S. Smith, R.H. Linton, and P.E. Nelson. Food Microbiology, 16(1) 53-61. 1991.
6	Inactivation of Enteric Adenovirus and Feline Calcivirus by Chlorine Dioxide, Jeanette A. Thurston-Enriquez, Charles N. Haas, Joseph Jacangelo, and Charles P. Gerba. Applied and Environmental Microbiology, June 2005, p. 3100-3105.
7	Effect of Chlorine Dioxide Gas on Fungi and Mycotoxins Associated With Sick Building Syndrome, S.C. Wilson, C. Wu, L.A. Aandriychuck, J.M. Martin, T. L. Brasel, C.A. Jumper, and D.C. Strauss. Applied and Environmental Microbiology, Sept. 2005, p. 5399-5403.
8	BASF Aseptrol EPA Pesticide Label
9	Effects of Ozone, Chlorine Dioxide, Chlorine, and Monochloramine on Cryptosporidium parvum Oocyst Viability, D.G. Korich, J.R. Mead, M.S. Madore, N.A. Sinclair, and C.R. Sterling. Applied and Environmental Microbiology, May 1990, p. 1423-1428.
10	NHSRC's Systematic Decontamination Studies, Shawn P. Ryan, Joe Wood, G. Blair Martin and Vipin K. Rastogi (ECBC), Harry Stone (Batelle). 2007 Workshop on Decontamination, Cleanup, and Associated Issues for Sites Contaminated With Chemical, Biological, or Radiological Materials, Sheraton Imperial Hotel, Research Triangle Park, North Carolina, June 21, 2007.
11	Validation of Pharmaceutical Processes 3rd Edition, edited by James Agalloco and Frederick J. Carleton. Informa Healthcare USA, Inc., 2008, p. 267.
12	Chlorine Dioxide Gas Sterilization Under Square-Wave Conditions, D.K. Jeng, Applied Environmental Microbiology. 56:514-519. 1990.
13	Inactivation Kinetics of Inoculated Escherichia coli O157:H7 and Salmonella enterica on Lettuce by Chlorine Dioxide Gas, Barakat S.M. Mahound and R.H. Linton. Food Microbiology 25(2): 244-252. February 2008.
14	Determination of the Efficacy of Two Building Decontamination Strategies by Surface Sampling With Culture and Quantitative PCR Analysis. Mark P. Buttner, Patricia Cruz, Linda D. Stetzenbach, Amy K. Kilma-Comba, Vanessa L. Stevens, and Tracy D. Cronin. Applied and Environmental Microbiology, August 2004. p. 4740-4747.



Reference No	Reference
15	Inactivation of Human and Simian Rotaviruses by Chlorine Dioxide. Yu-shiaw Chen and James M. Vaughn. Applied and Environmental Microbiology, May 1990, p. 1363-1366.
16	Decontamination of Produce Using Chlorine Dioxide Gas Treatment, Richard Linton, Philip Nelson, Bruce Applegate, Dave Gerrard, Yingchang Han, and Travis Selby, Purdue University.
17	Efficacy of Chlorine Dioxide Gas Against Alicyclobacillus acidoterrestris Spores on Apple Surfaces, Sun-Young Lee, Genesis Iris Dancer, Su-sen Chang, Min-Suk Rhee and Dong-Hyun Kang, International Journal of Food Microbiology 108(3): 364-386. May 2006.
18	Decontamination of Bacillus thuringiensis Spores on Selected Surfaces by Chlorine Dioxide Gas, Y. Han, B. Applegate, R.H. Linton, and P.E. Nelson, Journal of Environmental Health 66(4): 16-21. Nov. 2003.
19	Decontamination of Strawberries Using Batch and Continuous Chlorine Dioxide Treatments, Y. Han, T.L. Selby, K.K. Schultze, and P.E. Nelson, Journal of Environmental Health 67(12) 2004.
20	Mechanisms of Inactivation of Poliovirus by Chlorine Dioxide and Iodine, Maria E. Alvarez and R.T. O'Brian, Applied and Environmental Microbiology, Nov. 1982, p. 1064-1071.
21	The Use of Chlorine Dioxide in Potato Storage, Nora Olsen, Glae Kleinkopf, Gary Secor, Lynn Woodell, and Phil Nolte, University of Idaho, Bulletin 825
22	Protective Effect of Low-Concentration Chlorine Dioxide Gas Against Influenza A Virus Infection, Norio Ogata and Takashi Shibata, Journal of General Virology 89(Pt 1): 60-67. Jan 2008.
23	Preparation and Evaluation of Novel Solid Chlorine Dioxide-Based Disinfectant Powder in Single-Pack, M. Zhu, L.S. Zhang, X.F. Pei, and X. Xu, Biomedical Environmental Science, 21(2): 157-162. Apr. 2008
24	Chlorine Dioxide Oxidation of Dihyronicotinamide Adenine Dinucleotide (NADH), E.V. Bakhmutova-Albert, D.W. Margerum, J.G. Auer, and B.M. Applegate, Inorganic Chemistry 47(6):2205-2211. 16 Feb. 2008.
25	Oxidative Elimination of Cyanotoxins: Comparison of Ozone, Chlorine, Chlorine Dioxide, and Permanganate, E. Rodriguez, G.D. Onstad, T.P. Kull, J.S. Metcalf, J.L. Acero, and U. von Gunten, Water Research 41(15): 3381-3393. 20 June 2007.
26	Inhibition of Hyphal Growth of the Fungus Alternaria alternata by Chlorine Dioxide Gas at Very Low Concentrations, H. Morina, A. Matsubara, T. Fukuda, and T. Shibata, Yakugaku Zasshi 127(4): 773-777. Apr. 2007.
27	Inactivation of Chironomid Larvae With Chlorine Dioxide, X.B. Sun, F.Y. Cui, J.S. Zhang, F. Xu, and L.J. Liu, Journal of Hazardous Materials, 142(1-2): 348-353. 2 Apr. 2007.
28	William A. Rutala, Ph.D., M.P.H., Eugene C. Cole, D.P.H., Nancy S. Wannamaker, M.S., David J. Weber, M.D., M.P.H., Inactivation of Mycobacterium tuberculosis and Mycobacterium bovis by 14 Hospital Disinfectants Chapel, The University of North Carolina at Chapel Hill, The American journal of medicine, 91(3b): S267-S271. 1991.
29	Evaluation of Gaseous Chlorine Dioxide as a Sanitizer for Killing Salmonella, Escherichia coli O157:H7, Listeria monocytogenes, and Yeasts and Molds on Fresh and Fresh-Cut Produce, Kaye V. Sy, Melinda B. Murray, M. David Harrison, and Larry R. Beuchat, Journal of Food Protection, 68(6): 1176-1187. 2005.
30	Decontamination of Surfaces Contaminated With Biological Agents Using Fumigant Technologies, S. Ryan, and J. Wood, 2008 Workshop on Decontamination, Cleanup, and Associated Issues for Sites Contaminated with Chemical, Biological, or Radiological Materials, Sheraton Imperial Hotel, Research Triangle Park, North Carolina, September 24, 2008.



Reference No	Reference
31	Sporicidal Action of CD and VHP Against Avirulent Bacillus Anthracis-Effect if Organic Bio-Burden and Titer Challenge Level, Vipin K. Rastogi, Lanie Wallace, and Lisa Smith, 2008 Workshop on Decontamination, Cleanup, and Associated Issues for Sites Contaminated with Chemical, Biogical, or Radiological Materials, Sheraton Imperial Hotel, Research Triangle Park, North Carolina September 25, 2008.
32	Clostridium Botulinum, ESR Ltd., May 2001.
33	Efficacy of Chlorine Dioxide as a Gas and in Solution in the Inactivation of Two Trichothecene Mycotoxins, S.C. Wilson, T.L. Brasel, J.M. Martin, C. Wu, L. Andrichuk, D.R. Douglas, L. Cobos, and D.C. Straus, International Journal of Toxicology, 24(3) 181-186. 3 May 2005.
34	Guidelines For Drinking-water Quality, World Health Organization, p. 40.
35	Division of Animal Resources Agent Summary Sheet, M. Huerkamp, 30 June 2003.
36	NRT Quick Reference Guide; Glanders and Melioidosis
37	Seasonal Occurance of the Pathogenic Vibrio sp. of the Disease of the Sea Urchoin Strongylocentrotus intermedius Occurring at Low Water Temperatures and the Prevention Method of the Disease, K. tajima, K. Takeuchi, M. Takahata, M. Hasegawa, S. Watanabe, M. Iqbal, and Y. Ezura. Nippon Suisan Gakkaishi. 66(5) 799-804. 2000.
38	Selecting Surrogate Microorganisms for Evaluation of Pathogens on Chlorine Dioxide Gas Treatment, Jeongmok Kim, Somi Koh, Arpan Bhagat, Arun K. Bunia, and Richard H. Linton. Purdue University Center for Food Safety 2007 Annual Meeting October 30-31, 2007 at Forestry Center, West Lafayette, IN.
39	Sensitivity of Listeria monocytogenes, Campylobacter jejuni, and Escherichia Coli Stec to Sublethal Bactericidal Treatments and Development of Increased Resistance After Repetitive Cycles of Inactivation, N. Smigic, A. Rajkovic, H. Medic, M. Uyttendaele, and F. Devlieghere, Oral Presentation. FoodMicro 2008, 1-4 September 2008, Aberdeen, Scotland.
40	Susceptibility of Chemostat-Grown Yersinia enterocolitica and Klebsiella pneumoniae to Chlorine Dioxide, M.S. Harakeh, J.D. Berg, J.C. Hoff, and A. Matin, Applied Environmental Microbiology 49(1): 69-72. Jan 1985.
41	Efficacy of Gaseous Chlorine Dioxide as a Sanitizer Against Cryptosporidium parvum, Cyclospora cayetanensis, and Encephalitozoon intestinalis on Produce, Y. Ortega, A. Mann, M. Torres, and V. Cama, Journal of Food Protection, 71(12): 2410-2414. Dec. 2008.
42	J. Jacangelo, Inactivation of Waterborne Emerging Pathogens by Selected Disinfectants, p. 23.
43	SARS Fact Sheet, National Agricultural Biosecurity Center, Kansas State University.

