# 2X SuProbe qPCR Mastermix

| Catalog No | Size |
|---|---|
| • PCR01D0351 | 2 ml |
| • PCR01D0352 | 5 ml |
| • PCR01D0353 | 10 ml |

## INSTRUCTIONS FOR USE

### 1. PRODUCT NAME

2X SuProbe qPCR Mastermix

**Cat No** : PCR01D0351, PCR01D0352 and PCR01D0353
**Size** : 2 ml, 5 ml and 10 ml

### 2. MANUFACTURER

SuGenomik Biyoteknoloji, METU Technopolis, Yenimahalle, Ankara.

### 3. INTENDED USE

2X SuProbe qPCR Mastermix provides everything you need for probe-based (such as MBG and TaqMan) PCR amplification and detection in a convenient, single-tube format. It was optimized for broad range qPCR applications. It includes HotStart Taq DNA polymerase, dUTP/dTTP blend to enable UNG digestion to minimize the risk of contamination, and universal ROX dye in a standart concentration.

### 4. KIT CONTENT

| Content | Volume |
|---|---|
| 2X SuProbe qPCR Mastermix, PCR01D0351 | 1 ml x 2 |
| 2X SuProbe qPCR Mastermix, PCR01D0352 | 1 ml x 5 |
| 2X SuProbe qPCR Mastermix, PCR01D0353 | 1 ml x 10 |

### 5. STORAGE CONDITIONS

Store at -20±5°C away from light for 12 months.

### 6. REACTION SETUP

- It is recommended that thaw the 2X SuProbe qPCR Mastermix and mix thoroughly but gently to ensure even distribution of the components.
- Put the specimen's DNA on the PCR tubes or multi-well plate.
- For negative control sample do not add DNA please use nuclease-free water.
- Keep plate or PCR tubes on ice during reaction set up.

| Components | Volume |
|---|---|
| 2X SuProbe qPCR Mastermix | 10 µl |
| Forward primer (10 µM) | 0.5 µl |
| Reverse primer (10 µM) | 0.5 µl |
| Probe (10 µM) | 0.2 µl |
| Sample DNA (max 10 µl) | 2 µl |
| Nuclease-free water | X to complete 20 µl |
| TOTAL Volume | 20 µl |

### 7. RECOMMENDED qPCR PROTOCOL

- This kit was validated on ABI StepOne Plus system and compatible with standard RT-PCR machines such as ABI 7000, 7300,7500, Roche 480, MX3000P, MX3005P, Rotorgene TM6000, Icycler IQTM4/5, Bio-Rad CFX96 and LongGene Q2000B.
- (Optional) An additional melting curve step can be added when needed to ensure specific amplification and to detect possible primer dimers events.

| Steps | Temp. | Time | Cycles |
|---|---|---|---|
| **HotStart Activation** | 95 ℃ | 5 min | 1 |
| **Amplification** | 95 ℃ | 10 sec | 40 |
| | 57-60 ℃ | 45-90 sec (reading) | |

- For Applied Biosystems ABI (QS5, 7500 and StepOne etc) real-time PCR instruments, set to "passive reference" dye as "yes" which is a default setting.

### 8. EVALUATIONS of RESULTS

Analyze the amplification of the samples, positive and negative controls for any increase. If the fluorescence increases in Positive Control but doesn't in Negative Control, amplification reaction is proceeding properly. If any other situation occurs, however, amplification reaction may be proceeding in a wrong way. In such a case, re-test the samples from reagent preparation.

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