Comparison of Blood Collection Methods in the Early Stages of Abstract: 146 (B) Anthrax Bacteremia in Mice

Z.N. LLEWELLYN, J.A. Boydston, J.E. Tromblev and L.E. Bowen

SOUTHERN RESEARCH

Legendary Discoveries. Leading Innovation

2000 Ninth Avenue South Birmingham, AL 35023 Phone: 205,581,2791 Fax: 205 581 2536 llewellyn@southernresearch.org

Abstract

Bacillus anthracis is the eausative agent of the disease anthrax in humans and animals. The route of blood collection could affect bacterial counts from potential ocular and dermal contamination during a nose-only inhalation exposure. The comparison of different blood collection routes to determine load during the early stages of bacteremia has not been well described

In this study we compared the retro-orbital sinus and intra-cardiac puncture blood collection routes during the early stages of the onset of bacteronia. Seven groups of BALBs mice, consisting of five males and five females, were exposed by nose-only inhalation to an average acrossol concentration of 3.24-96 CPU of B. authrusts Ames spores. Mice were cultinarized at T=0 3.2E+00 CFU of B. ammacas Ames spores. Mice were cultianized at 1-U (immediately following exposure), 6, 12, 18, and 24 hours post-exposure. Blood samples were collected via retro-orbital sinus or intra-cardiac puncture at 0 and 6 hours and by retro-orbital bleeding at subsequent time collections. Blood was serially diluted, plated, and incubated. The plates were counted to determine the number of CFU/mL.

For retro-orbital blood collection, bacteremia was observed in 100% of mice at T=0 and 50% at 6 hours. The average bacterial counts were 1.1E+03 CFU/mL and 77 CFU/mL, respectively. For cardiac puncture blood collection bacteremia was observed in 90% of mice at T=0 and 40% at 6 hours. The average bacterial counts were 5.9E+03 CFU/mL and 323 CFU/mL, respectively. By 12 hours, 100% of mice were negative for B. anthracis in the blood followed by 100% and 50% at 18 and 24 hours, respectively.

These results suggest there was there was a difference in the percent of bacteremia at T=0 and 6 hours post exposure between collection methods, but the differences were insignificant. Initial bacterial concentrations ranged between 7.7E+01 to 5.9E+03 at 0 to 6 hours. The bacterial load from different routes of collections was insignificant.

Introduction

Blood can be collected from animals using different techniques with differences in handling, restraining, anesthesia, invasiveness and the volume taken. Blood collection via the retro-orbital sinus or intra-cardiac puncture are standard techniques to obtain blood samples from mice. Comparisons of blood collection techniques and their effects on blood analyses have been documented. [1-5] However, there is a lack of information on collection methods for bacterial samples from mice in an acrosol challenge model

The onset of bacteremia of Bacillus anthracis Ames (BAA) during the first 24 hours in mice is crucial to determining the optimal time for antibiotic or post therapeutic treatment. Accurately determining the presence of bacteremia is critical to eliminate the possibility of ocular and dermal contamination.

In development of the inhalational anthrax murine model, an initial study on the onset to bacteremia resulted in mice challenged with an aerosol concentration of 2.0E+06. (4) Blood samples were collected at 0, 6, 12, and 24 hours post challenge for bacterial load. All mice euthanized immediately after aerosol challenge had bacteremia. Colony counts ranged from 1.3E+02 CFU/mL to greater than 3.0E+05 CFU/mL. The onset to bacteremia started 12 hours earlier and the bacterial counts were higher than published data. (5) It was not clear if the retro-orbital blood samples were contaminated from the aerosol challenge route at 0 and 6 hours. Therefore, we proposed to compare blood collection via intra-cardiac puncture and retro orbital sinus immediately after exposure and six hours after aerosol challenge.

Methods

Challenge Material Preparation:

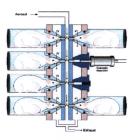
BAA spores were prepared by growing cultures in Difco Sporulation Media to nutrient exhaustion. The sporulated culture was harvested by centrifugation. Spores were then purified over Renografin-60 density gradients to remove vegetative cells and debris. The purified spores were resuspended in USP-Water For Injection (WFI). The spores were observed using phase contrast microscopy to quantify the number of phase bright spores, phase dark spores, vegetative cells, and other debris. Endotoxin levels were tested using the limulus amebocyte lysate assay to ensure that the spore preparation contained less than 1 endotoxin unit per milliliter. Spore number was quantified by serial dilution in USP-WFI and plating on TSA with 5% sheep's blood plates and determining the number of CFU/mL. Prior to exposure, the spore suspension was serially diluted in USP-WFI to achieve a concentration of 5.0E+09.

Methods

Aerosol Challenge of BALB/c Mice

Seven groups consisting of 5 male and 5 female BALB/c mice were placed in nose only restraint tubes and connected to the exposure chamber using Positive Flow-ByTM nose cones (In-Tox Products, LLC; Albuquerque, NM) as shown in Figure 1 The Collison nebulizer was filled with a pre-mixed nebulizer stock suspension of 5.0E+09 BAA and operated at a constant pressure. The start of the exposure period (T = 0) began once the nebulizer was activated. The actual concentration of challenge material in the exposure atmosphere was determined by analysis of stainless steel impinger samples collected from the breathing zone of the animals during the exposure. Impinger samples were analyzed on the day of collection Aerosol concentration were reported as colony forming units per liter of air (CFU/L). Real-time aerosol concentration was monitored using a laser-based aerosol photometer. Each group was exposed for approximately 60 minutes

Figure 1. Positive Flow-By™ Nose Cones



Blood Collection Methods

Blood from the retro-orbital sinus was collected at 0, 6, 12, 18, and 24 hours. Two additional groups were bled via the intra-cardiac puncture at 0 and 6 hours. All blood collections were terminal bleeds. Blood was serially diluted, plated, and incubated. The plates were counted to determine the number of CFU/mL.

Intra Cardiac Puncture Blood Collection Method:

The mice were anesthetized prior to euthanasia and the thoracic cavity was wiped with 70% ethanol. The thoracic cavity was palpated to identify the bottom of the rib cage, the needle inserted at the base and under the rib cage to the heart. Blood was collected into the syringe and placed into the collection tube

Retro-orbital Sinus Rlood Collection Method:

The mice were anesthetized prior to euthanasia and the head immobilized. The neparinized microhematocrit capillary tube was inserted into the eye socket. The first drop of blood was discarded to flush out any benarin and then held so that the blood flows from the capillary tube into the collection tube.

Microbiological Analyses

Blood samples were serially diluted in USP-WFI, plated in triplicate on blood agar and incubated for colony formation. The plates were then counted to determine the number of CFU/mL.

Results

BALB/c mice were exposed by nose-only inhalation to a mean aeroso concentration of 3.2E+06 CFU/L of BAA spores. Individual aerosol concentrations with for each group are shown in Table 1. Using Guyton's formula (6) the calculated mean inhaled dose for male and female mice combined was 4.1E+06 CFU and corresponded to an inhaled dose of 44 LD₅₀ (1 LD₅₀ 9.4E+04 CFU based on prior data).(7)

Table 1. Summary of Aerosol Concentration Data

Time of Blood Collection (Hour)	Impinger Concentration (CFU/mL)	Aerosol Concentration (CFU/L)	Mean Inhaled Dose (CFU)
0	3.20E+07	2.60E+06	3.30E+06
6	5.70E+07	4.50E+06	5.90E+06
12	3.20E+07	2.60E+06	3.25E+06
18	5.70E+07	4.50E+06	5.85E+06
24	3.20E+07	2.40E+06	3.10E+06

Results

Retro-orbital Sinus vs. Intra-cardiac Puncture Blood Collection

The percent of bacteremia is shown in Table 2. The percent of bacteremia in mice was 100% and 90% at 0 hours in retro-orbital sinus and intra-cardiac puncture blood collections, respectively. At 6 hours 50% and 40% of mice had bacteremia in retro-orbital sinus and intra-cardiac puncture blood collections, respectively. Subsequent blood collections were only taken via the retro-orbital sinus. Figure 2 illustrates the percent of bacteremia from retro-orbital samples collected at all time points within 24 hours post exposure

Table 2. Percentage of Mice with Bacillus anthracis Ames Bacteremia

Route of Blood Collection*	Time of Blood Plating	Percent Bacteremia (%)	
	(Hour)		
RO	0	100	
IC	0	90	
RO	6	50	
IC	6	40	
RO	12	0	
RO	18	100	
RO	24	50	

Figure 2. Percentage of Mice with Bacillus anthracis Ames Bacteremia From Blood Collected Via the Retro-orbital Sinus

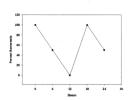


Table 3 summarizes the bacterial loads of BAA in the blood taken at T=0. 6. 12. 18, and 24 hours post exposure. Initial concentrations were 1.1E+03 CFU/mL and 5.9E+03 for retro-orbital sinus and intra-cardiac puncture blood collection respectively at T=0. At 6 hours the concentrations decreased to 77 CFU/mL and 323 CFU/mL for retro-orbital sinus and intra-cardiac puncture blood collections respectively. Bacteremia was characterized by an initial spike followed by a rapid decline with no detectable bacteria at 12 hours as shown in Figure 3. increase was identified with concentrations of 3.4E+02 at 18 hours and 3.4E+06 at

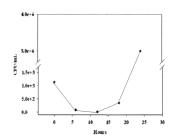
Subsequent analysis was performed using the criteria listed by the Food and Drug Administration. (8) Bacterial counts less than 25 CFU per plate were excluded from the additional bacterial count analyses as they were below the detectable limit of quantification (LOQ) for aerobic bacteria. One sample at 6 hours post challenge had a detectable colony count, all other samples were below the level of

Table 3. Average Concentration of Bacillus anthracis Ames in Blood

Time Collection (Hours)	Initial Data Analysis RO ¹ (CFU/mL)	Initial Data Analysis IC ² (CFU/mL)	Subsequent Data Analysis RO ¹ (CFU/mL)	Subsequent Data Analysis IC ² (CFU/mL)
0	1.12E+03	5.91E+03	2.87E+03	1.40E+04
6	7.67E+01	3.23E+02	BLOQ3	2.97E+03
12	0.00E+00		BLOQ3	
18	3.43E+02		BLOQ3	
24	3.39E+06		BLOQ3/TNTC4	

Results

Figure 3. Concentration of Bacillus anthracis Ames From Blood Collected via the Retro-Orbital Sinus



Conclusions

teremia was characterized by an initial spike followed by a rapid decline and then rapid increase in mice enthanized from 0 hours to 24 hours. These results suggest that blood collection from the retro-orbital sinus is an acceptable method following an inhalation exposure

References

- 1. Grouzmann E, Cavadas C, Grand D, Moratel M, Aubert JF, Brunner HR, Mazzolai L. Blood sampling methodology is crucial for precise holamines concentrations in mice. Pflugers Arch 2003 Nov-447(2):254-8
- 2. Hui YH, Huang NH, Ebbert L, Bina H, Chiang A, Maples C, Pritt M, Kern 1. mit 171, Huang N11, Ebbert 1., Dina 11, Chang A, Magiset V, Pritt M, Kern T, Patel N, Pharmacokinetic comparisons of tail-hieleding with cannula- or retro-orbital bleeding techniques in rate using six marketed drugs. J Pharmacol Toxicol Methoda; 2007 Sep-Oct;56(2):256-64
 Van Herck H, Bauman V, Brandt CJ, Borce HA, Hesp AP, van Lith HA, Schuriat M, Bevnen AC. Blood sampling from the retro-orbital plexus, the
- saphenous vein and the tail vein in rats: comparative effects on selected behavioural and blood variables. Lab Anim. 2001 Apr;35(2):131-9.
- behavioural and blood variables. Lab Anim. 2001 Apr3(2):131-9.

 Bowen LE, Deskins AR, Llewellyn ZN. (2008). Determination of the Pulmonary Mean Deposited Dose and Onset of Bacteronia in Mice Following Nasc-Only Inhalation of Bacillus authorices Sporse. Final report for Southern Research Institute study 11051.01.06 to the National Institute of Health daded September 5, 2008.

 Heine HS, Bassett J, Miller L, Hartings JM, Ivins BE, Pitt ML, Pritz D, Norris SI. Stress WP. Determination of Authbrists Efficiency assisted.
- Heine HS, Basselt J, Miller L, Hartings JM, Ivus HE, Pitt ML, Pritz D, Norris SL, Byme WR. Determination of Antibiotic Efficacy against Bacillus authracis in a Mouse Aerosol Challenge Model. Antimicrobial Agents and Chemotheraphy. 2007. Apr.51(4):1373-1379.
 Guyton, AC. Measurement of the respiratory volumes of laboratory animals (1947). American Journal of Physiology. 1947. 150: 70-77.
- Bowen LE, Deskins AR, Llewellyn ZN. (2008). Determination of the 50%
- Lethal Dose (LD₅₀) of Bacillus authracis in Mice Following Nose-Only Inhalation Exposure. Final report for Southern Research Institute study
- Institution Exposure. That report for Southern Research institute study 1105.10.105 to the National Institute of Health dated August 8, 2008.
 Bacteriological Analytical Manual January 2001. Chapter 3. Aerobic Plate Count. U.S. Food and Drug Administration.

Acknowledgements

This contract was funded by the National Institutes of Health, National Institute of Allergy and Infectious Disease, Contract N01-AI-30063.