

Recommendations for Management of Noncytotoxic Vesicant Extravasations

Jennie Ong, PharmD • Ruth Van Gerpen, MS, RN-BC, APRN-CNS, AOCNS®

ABSTRACT

To prepare clinicians to treat extravasation of noncytotoxic vesicants with antidotes and thermal compresses, a literature review was performed to identify noncytotoxic vesicants and to create evidence and consensus-based recommendations. The stage of injury and vesicant's mechanism of tissue injury dictate treatment. For a vasopressor extravasation, warm compresses and administration of a vasodilator are recommended. For osmolarity, pH, absorption refractory, and cytotoxic concentration-dependent vesicants, warm compresses and administration of hyaluronidase are recommended. Compared with potentially catastrophic costs of undertreatment, the cost of overtreatment is minimal.

Key words: compartment syndrome, emergency treatment, extravasation, hyaluronidase, infiltration, irritant, noncytotoxic vesicant antidote, phentolamine, soft tissue injury, terbutaline, tissue or skin necrosis, warm or cold compress

Extravasation is a universal risk of intravenous (IV) vesicant administration. Appropriate precautions can reduce the risk but not eliminate it. The terms *extravasation* and *infiltration* are often used interchangeably in the literature, but the 2016 *Infusion Therapy Standards of Practice* (the *Standards*)¹ defines extravasation as “inadvertent infiltration of vesicant solution or medication into the surrounding tissue,”^{1(pS149)} whereas infiltration is the “inadvertent administration of a nonvesicant solution or medication into surrounding tissues.”^{1(pS150)} A vesicant is an agent capable of causing tissue damage when it escapes from the intended vascular pathway into surrounding

tissue. Vesicant identification and consensus treatment recommendations existed as early as 1979² for cytotoxic vesicants, but it is only in recent years that extravasation injury from noncytotoxic vesicants has begun to receive similar attention. In 2017, the Infusion Nurses Society (INS) Vesicant Task Force published an evidence-based list identifying noncytotoxic vesicants,³ but treatment modalities were outside the scope of that review. This article summarizes the evidence supporting treatments for noncytotoxic vesicant extravasations and contains recommendations to aid clinicians in making timely, evidence-based treatment decisions.

BARRIERS TO APPROPRIATE AND TIMELY TREATMENT

A calcium extravasation was the impetus for seeking evidence-based treatment recommendations for noncytotoxic vesicant extravasations. Every source reviewed had different lists of noncytotoxic vesicants and varying recommendations and reasoning for proposed treatments. The lack of consensus in the literature contributes both to undertreatment and inappropriate treatment of extravasations. In the case of extravasation injury, the cost of overtreatment is minimal. The antidotes are relatively inexpensive, easy to prepare, and cause minor if any discomfort to the patient. In contrast, the cost of undertreatment can be hospitalization, surgical intervention, and permanent cosmetic and functional defects. Identified barriers to receiving appropriate,

Authors' Affiliation: Bryan Medical Center, Lincoln, Nebraska (Dr Ong and Ms Van Gerpen).

Jennie Ong, PharmD, is a clinical pharmacist at Bryan Medical Center in Lincoln, Nebraska, with 10 years of hospital pharmacy experience. Her specialties include formulary management and patient safety-oriented process improvements. **Ruth Van Gerpen MS, RN-BC, APRN-CNS, AOCNS®**, is a clinical nurse specialist at Bryan Medical Center in Lincoln, Nebraska, with 36 years of oncology experience. Her subspecialties include pain management and infusion therapy.

The authors of this article have no conflicts of interest to disclose.

This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Corresponding Author: Jennie Ong, PharmD, Bryan Medical Center, 1600 S. 48th St., Lincoln, NE 68506 (jennie.ong@bryan-health.org).

DOI: 10.1097/NAN.0000000000000392

timely treatment include delayed recognition, unknown treatment options, lack of or inconsistent evidence supporting a treatment approach, and unknown or uncertain vesicant status of the medication.

Delayed Recognition

Despite recommendations in the *Standards*¹ to assess vascular access devices for signs and/or symptoms of infiltration and extravasation before each infusion and on a regular basis, some extravasations are recognized after irreversible damage has occurred, limiting the efficacy of intervention options. Loth and Eversmann⁴ proposed the concept of a necrosis interval to guide therapy decisions. The necrosis interval is the time from extravasation until irreversible tissue damage occurs, and it varies for each vesicant. If the offending agent is counteracted or removed within the time interval, then necrosis is prevented. The necrosis interval has not been determined for every vesicant. However, because the interval can be quite short, for example 4 to 6 hours for vasopressors, extravasation should be considered a medical emergency requiring time-sensitive treatment. Recognition of extravasation outside the necrosis interval may preclude treatments that would prevent or reduce damage and instead necessitate damage control.

Unknown Treatment Options

To date there are no published articles that comprehensively describe the evidence supporting or refuting a wide variety of treatments for noncytotoxic extravasations. Even for treatment methods with documentation of efficacy, when confronted with drug shortages affecting first-line options such as phentolamine, second-line treatment options have not been as well documented.

Uncertainty as to Appropriateness of Treatment Options

Every extravasation is unique. No two will have identical factors influencing the extent of injury or treatment decisions. Both patient- and incident-specific factors influence treatment decisions. These factors can include age, weight, comorbidities, communication barriers, skin integrity, site of extravasation, anatomic anomalies, care setting, response to current treatment, timing of recognition, and, perhaps most importantly, the identity, amount, and concentration of the vesicant. With so many factors influencing treatment and prognosis, it may seem difficult or even impractical to apply evidence from one extravasation to another.

Despite the unique nature of each extravasation, all are identical in that the patient's tissues have inadvertently been exposed to a toxic substance. Tissue damage will occur unless the vesicant's toxicity and mechanism of tissue injury can be eliminated or reversed. Thus, extravasation treatments are generalizable when the mechanism of tissue injury and method to eliminate or reverse it are considered. Large, multicenter, randomized, double-blind, head-to-head, or placebo-controlled trials are the gold

standard in evidence-based medicine. This level of evidence will never exist for extravasation, because ethical considerations preclude purposefully inflicting an extravasation on humans to test potential remedies. Perhaps, as a result of the varied nature of extravasations, the published recommendations and reported management of extravasations are just as varied. Conflicting recommendations and case reports describing a variety of treatment options have sometimes resulted in inappropriate therapy choices, such as applying cold compresses (instead of dry, warm compresses) to a norepinephrine extravasation. Applying reported or recommended treatments from one case to another should be done in the context of the mechanism of tissue injury and whether the proposed treatment is likely to eliminate or reverse the toxic potential of the vesicant.

Unknown or Uncertain Vesicant Status

Noncytotoxic vesicants are reported in many disparate lists. Even with all the evidence available, however, there are some medications that likely are vesicants but lack confirming evidence. These could include lower potency vesicants, those with a weaker propensity for causing tissue injury, or recently developed medications. Medications reviewed and ultimately excluded as noncytotoxic vesicants by the INS Vesicant Task Force³ (mostly because of insufficient evidence), as well as infiltration of any medication not yet identified as a vesicant, may warrant extravasation treatment if symptoms of extravasation occur. A medication may be a vesicant although it is not yet recognized as one.

LITERATURE SEARCH AND METHODOLOGY FOR EXTRAVASATION MANAGEMENT IDENTIFICATION

The *Standards* recommends that "each facility should reach a consensus on what medication is considered to be a vesicant and irritant based on their internal formularies" and "identify the vesicant nature of antineoplastic and noncytotoxic medications prior to administration and be prepared to use the correct antidote treatment for each medication."^{1(pS98)}

From published lists, 86 purported vesicants were identified. Inclusion in the vesicant list required either: (1) reports in literature or from manufacturer of tissue injury upon extravasation or (2) adverse effects or warnings in secondary drug information sources such as Micromedex or Lexicomp consistent with a vesicant along with a valid proposed mechanism of tissue injury.

Of the 86 purported vesicants vetted, 45 drugs met our inclusion criteria. This review, which began in August 2016, was bolstered in January 2017 when the INS Vesicant Task Force published an evidence-based list of noncytotoxic vesicants.³ Of 42 purported vesicants vetted by Gorski et al,³ 25 were classified as vesicants. The other 17 were

Downloaded from http://journals.lww.com/journalofinfusionnursing by H88SRLEF1XWZCEZ2bFUCaSmTDGqH4
kydLLZBBY8rHYD1V031w6INRQWYFQ0uID4E9Y6Z450CUIZ41F0ddGm3Y125hUnek1o2mWgmPYZZRgDlbf+
Y1vn4OGpFGbICFASW1231poCWgaTd+o6IdD6ys= on 09/19/2024

indeterminate at the time due to inconclusive or conflicting data. Based on more recent case reports, 3 of the 17 indeterminate vesicants (gentamicin, immune globulin, and propofol) met our first vesicant criteria. Of the remaining 14 indeterminate vesicants, another 8 met our first inclusion criteria (aminophylline, amphotericin, ampicillin, doxycycline, lorazepam, metronidazole, penicillin, and valproate). We had reviewed 44 purported vesicants that the INS Vesicant Task Force did not review, 4 of which met our first criteria (digoxin, lipids, methylene blue, and phosphate salts) and 5 of which met our second criteria (conivaptan, dantrolene, diazepam, esmolol, and etomidate). Radiographic contrast material is excluded from our recommendations due to the unique nature and scope of contrast extravasations. Other exclusions included antineoplastic vesicants, products not recommended for IV use, extravasations not involving skin or soft tissue (eg, intraperitoneal), and products not available in the United States.

Aside from the US Food and Drug Administration (FDA)-approved antidote phentolamine, literature support for antidotes and thermal compresses is largely limited to case reports or case series. More than 1800 cases were included in this review, but for treatments short on published evidence, published articles with recommendations have also been included to support a consensus recommendation. Previously published recommendations represent the authors' opinion, description of local practice, or institutional guidelines.

The US National Library of Medicine holdings in the PubMed database were searched without beginning date restriction through February 15, 2020, using the search string "Extravasation of Diagnostic and Therapeutic Materials" [Mesh] OR "Extravasation" [Title] OR (Infiltration OR extravasa*) AND drug AND "Humans" [Mesh]. Of the 23,159 titles or abstracts screened manually, the full text of 257 articles was sought. Because of the inability to translate or unavailability of the full text, 14 articles were excluded from full-text review. On bibliography screening, the full text of an additional 226 articles was reviewed. Of the 469 articles reviewed, 331 articles were ultimately included in the review, and 138 were excluded for the following reasons:

1. Chemotherapy extravasations (6 articles)
2. Vesicant not identified (25 articles)
3. Infiltration of irritant, not of a known vesicant (15 articles)
4. Adverse effect without extravasation, that is, a systemic effect (21 articles)
5. Norepinephrine untreated or phentolamine monotherapy, already FDA approved (15 articles)
6. Vesicant unavailable in the United States (mezlocillin/sulbactam, metiamide, toluidine blue) (3 articles)
7. Noncontributory in either recommendations or cases (45 articles)
8. Anatomically distinct extravasation (eg, intra-arterial or neonatal umbilical vein) (8 articles). For a thorough list

of other articles describing anatomically distinct extravasations, see Appendix 2 of Corbett et al.⁵

In conversation with J. Friedman, MD (July 11, 2019), and written communication with J. L. Thigpen, RNC, MN (November 1, 2019), V. Boyar, MD (November 2, 2019), and J. C. Schie, MS, OTR/L (November 18, 2019), we were able to clarify which drug extravasated in several previously published cases.⁶⁻⁹ Medication prescribing information was also used. Pertinent landmark animal case studies were also reviewed.

MECHANISMS OF TISSUE INJURY

Vesicant classification by mechanism of tissue injury is useful in determining appropriate management to eliminate or reverse the toxic potential of an extravasate. The 4 widely recognized mechanisms of tissue injury are pH, osmolarity, vasoconstriction, and cytotoxicity. We propose absorption refractory as a new fifth mechanism of tissue injury.

Nonphysiologic pH

Physiologic pH is 7.35–7.45. Extreme pH exposure, typically defined as a pH of <5 or >9,¹⁰ can damage venous endothelium and increase risk of vessel rupture. When agents with extreme pH extravasate, they can cause damage in the same manner.^{11,12} Tissue destruction and vasoconstriction with resulting inflammatory response, edema, sloughing, and ulceration may result. Neutralization of extreme pH should not be attempted because of the potential for exothermic or gas-producing reactions that may exacerbate the injury.¹³

Osmolarity

Physiologic osmolarity is approximately 310 mOsm/L.¹⁴ Both hypotonic and hypertonic solutions can cause tissue damage by forcing fluid shifts into or out of cells. Hypotonicity causes fluid shift into cells, which can result in cell rupture. The generally accepted cutoff for hypotonicity and risk of hemolysis with IV infusion is <112 mOsm/L,¹⁵ whereas for hypertonicity and risk of being a vesicant the cutoff is >900 mOsm/L.¹⁶ Hypertonicity disrupts cellular ion transport and causes fluid shift from cells to the interstitial space, which may lead to swelling and compartment syndrome. Histopathology confirms, for example, that extravasation of 10% calcium chloride, which has an osmolarity of 2050 mOsm/L, is "a true 'subcutaneous burn' and as such involves both the overlying skin and the underlying fascia and skeletal muscle."^{17(p155)} High osmolarity vesicants, including phosphate and calcium, also present a precipitation risk. Calcium precipitation in soft tissues is called *calcinosis cutis* and can occur with or without accompanying necrosis.^{1,18-20}

Vasoconstriction

Localized vasoconstriction attributed to extravasation can result in ischemia and necrosis by reducing blood flow.

Vesicant exposed tissues are at risk from both chemically induced and mechanically induced vasoconstriction. Electrolyte solutions such as calcium and sodium, along with pharmacologic vasopressors such as dopamine and epinephrine, can chemically induce vasoconstriction. Large volume or anatomically trapped extravasations can mechanically induce vasoconstriction when the interstitial pressure is raised enough to overcome the venous pressure, blocking blood flow and even causing compartment syndrome.

Cytotoxicity

Cytotoxicity is primarily associated with antineoplastic drugs. For antineoplastic drugs, extravasation injury occurs as a result of the vesicant binding to the nucleic acids in the DNA of healthy cells in the tissue causing cell death. Other cytotoxic vesicants damage cells or tissues on coming into direct contact with them. The identity and treatment of antineoplastic vesicants have been published elsewhere and are outside the scope of this review.²¹

Absorption Refractory

Absorption refractory is a newly proposed mechanism of tissue injury whereby drugs with insolubilities or limited ability to be absorbed into the bloodstream persist in the extravasated space. The prolonged presence of lipids in

the interstitial space has led to deep tissue necrosis, and propofol, often contained in a lipid carrier solution, seems particularly prone to causing necrosis and compartment syndrome because of its limited ability to disperse in the tissues and be absorbed into the bloodstream.²²⁻³⁶

MANAGEMENT RECOMMENDATIONS

Effective extravasation management requires early recognition of signs and symptoms and prompt implementation of appropriate clinical management strategies.¹ Extravasation is unique, but the patterns of initial symptoms are commonly staged as first proposed by Millam in 1988³⁷ and validated in a pediatric population by Flemmer and Chan in 1993.³⁸ Published iterations of this staging are used widely for patients of all ages. Strengths from multiple versions have been refined, and the resulting proposed staging recommendations can be found in Table 1.^{4-6,37-40} Extravasations of small volume with less potent vesicants are likely to fit into stages 1 or 2, whereas more potent vesicants and/or larger extravasated volumes will tend to reach stages 3 or 4. Once the necrosis interval has passed (which can be as short as 4 to 6 hours for vasopressors), the staging table is no longer relevant, and assessments should be made based on whether the injury is receding or advancing.

TABLE 1
Extravasation Staging

Stage	Assessment	Treatment options
1	• Painful infusion site • No erythema • Localized swelling (1%-10% of extremity above or below site)	• Remove cannula • Elevate extremity • Warm/cold compresses
2	• Painful infusion site • Slight swelling at site (up to 25% of extremity above or below site) • Slight erythema (localized to the central area of extravasation) • Good pulse below site • Brisk (1-2 s) capillary refill below site	• Remove cannula • Elevate extremity • Warm/cold compresses • Consider antidote
3	• Painful infusion site • Moderate swelling at site (25%-50% of extremity above or below site) • Marked erythema (extends beyond central area of extravasation) • Blanching (for vasopressor extravasation only) • Good pulse below site • Brisk (1-2 s) capillary refill below site • Skin cool to touch	• Leave cannula in place; using a 1 mL syringe, aspirate as much fluid as possible • Remove cannula unless it is needed for antidote administration • Elevate extremity • Warm/cold compresses • Consider antidote
4	• Painful infusion site • Severe swelling at site (>50% of extremity above or below site) • Very marked erythema (extends beyond borders of swelling) • Blanching (nonvasopressor extravasation) • Decreased or absent pulse • Prolonged capillary refill >4 s • Skin cool to touch • Skin breakdown including blistering or necrosis	• Leave cannula in place; using a 1-mL syringe, aspirate as much fluid as possible • Remove cannula unless it is needed for antidote administration • Elevate extremity • Warm/cold compresses • Consider antidote • If swelling of the site is tense and skin is blanched, obtain surgical consult

Data from Loth and Eversmann,⁴ Corbett et al,⁵ Friedman,⁶ Millam,³⁷ Flemmer and Chan,³⁸ Baharestani,³⁹ and Odom et al.⁴⁰

Where treatment options have limited evidence supporting efficacy, clinicians must consider the potential benefits of the treatment and initiate those therapies where the benefits outweigh the risks. More invasive extravasation treatments are associated with greater risks, but they are also associated with faster onset and greater impact. Recommended initial treatment options ranked in order from least to greatest impact and risk are as follows: elevation, thermal therapy, antidote administration, saline flush out, and open incision with irrigation. Extravasation management for stage 1 and 2 injuries should begin with elevation and thermal therapy and advance to antidote administration at 30 minutes if the injury is not resolving. Because stage 3 and 4 injuries are unlikely to downstage with the impact of elevation and thermal compresses alone, antidote therapy should be initiated immediately.³⁸ If the injury has not downstaged or is not resolving by 30 to 60 minutes after the initial antidote, consider repeating antidote administration or switching to alternative antidote until symptoms begin to resolve. If at any point it appears that antidote administration will not be sufficient to prevent extensive injury, consider immediately advancing to a saline flush out or open incision and irrigation.^{41,42} The level of treatment should be proportional to the level of injury and should err on the side of overtreatment when it comes to antidote administration, because the costs of antidote overtreatment are minimal when compared with the potential costs of undertreatment. The impact of elevation and thermal compress therapy on symptom resolution is small when compared with the impact of antidote administration or saline flush out.

For pH-mediated, osmolarity-mediated, and absorption refractory extravasations, the goal is to dilute the vesicant through absorption and dispersal, so the appropriate antidote is hyaluronidase. Cytotoxic vesicants with concentration-dependent toxicity can also be treated with hyaluronidase. Vasodilators, such as phentolamine, terbutaline, or nitroglycerin, should be used to counteract vasoconstriction, whether caused by a vasoconstrictor, chemical vasoconstriction, or mechanical vasoconstriction. Table 2⁴³⁻²⁹⁷ contains antidote recommendations for each noncytotoxic vesicant. Additional antidote and treatment evidence not definitively linked to a particular vesicant is presented in Table 3.²⁹⁸⁻³⁰⁴

The clinical practice setting will influence which treatments are available. Initiate as much treatment as is indicated and possible. If the injury has not downstaged to level 1 with the maximum level of care the current setting can accommodate, clinicians should consider transferring the patient to a hospital with plastic surgery capabilities while there is still potential to prevent the damage rather than transferring the patient for debridement and skin grafting after the damage has occurred.

Overview of Treatments

The toxicity of vasoconstrictive vesicants can be reversed by vasodilator administration, but pH, osmolarity, and even

to some extent cytotoxic vesicants have concentration-dependent toxicity. To dilute the vesicant to eliminate its toxicity, it must either be absorbed into the bloodstream or dispersed among interstitial fluids. Hyaluronidase facilitates both. An FDA-approved indication of hyaluronidase is as an adjuvant to increase the absorption and dispersion of other injected drugs.³⁰⁵ This can be used, for example, as a planned enhancement to localized analgesia, but with extravasation it is used as an unplanned rescue attempt to facilitate dispersion and absorption into the bloodstream from the extravascular space. The use of hyaluronidase for extravasation injury is not FDA approved and is therefore considered “off-label.”

Hyaluronidase facilitates absorption and dispersal of drugs and fluids by dissolving hyaluronic acid, one of the binders that holds soft tissue cell layers together and forms the dermal barrier. By loosening the layers from each other, fluid is able to flow freely between sheets of tissue. “The rate and extent of dispersion and absorption is proportionate to the amount of hyaluronidase and the volume of solution.”^{305p6} A 150-unit dose of hyaluronidase will facilitate the absorption of 1 L or more of subcutaneously administered fluid.³⁰⁵ In studying hyaluronidase for hypodermoclysis, Hallman et al³⁰⁶ used each pediatric patient as their own control, receiving subcutaneous fluids in both legs but hyaluronidase in only one leg. Without regard to bolus volume (range, 20–40 mL) or fluid type (saline or dextrose), hyaluronidase accelerated absorption in every measurable case (anatomy precluded measurement in 2 patients). On average, hyaluronidase accelerated absorption time to 84 minutes versus 214 minutes without hyaluronidase. In 2 cases in particular, skin tension decreased within only 2 minutes of hyaluronidase administration.³⁰⁶ The dermal barrier does not reform in full until approximately 48 hours later in a dose-dependent manner.³⁰⁵

Hyaluronidase is an effective mainstay of extravasation treatment. In 1950, Haire³⁰⁷ reported that the pain, swelling, and induration caused by extravasation of neoarsphenamine in one case and mapharsen in another resolved completely within 24 hours after treatment with 250 units of hyaluronidase. He reported that the amounts extravasated in these cases would normally have caused a painful induration lasting for weeks. Acceptance of hyaluronidase to treat extravasation injuries grew and was mentioned in surgical textbooks as early as 1953.³⁰⁸ Although ethical considerations preclude the study of hyaluronidase versus placebo for extravasation injury in humans, Zimmet³⁰⁹ demonstrated in rats that, after injection with 23.4% sodium chloride, hyaluronidase-treated rats had a decreased incidence of ulceration (50%) versus normal saline or water (80%). The average ulceration size in the normal saline and water groups was 2 to 3 times as large as in the hyaluronidase groups. Similarly, Laurie et al³¹⁰ demonstrated in rabbits that after parenteral nutrition injection, rabbits receiving no treatment had ulcers on average 13 times as large as hyaluronidase-treated rabbits. After injection with calcium

TABLE 2

Antidote Recommendations for Noncytotoxic Vesicants

Vesicant	Antidote recommendations	Antidote cases	Other treatments
Osmolarity mediated tissue injury			
Aminophylline: 170 mOsm/L ⁴³	Hyaluronidase ^{13,44-49}	Hyaluronidase: at least 1 case of injury prevention ^{44,45}	
Ampicillin 50 mg/mL: 566 mOsm/kg ¹⁰	Hyaluronidase ^{13,48-50}	Hyaluronidase: 1 co-extravasation with cefotaxime in dextrose 10% and 0.225% saline necrosed. ⁵¹	Conservative: 1 co-extravasation with chloramphenicol necrosed with PFD. ⁵²
Calcium salts, including calcium disodium edetate (EDTA) Calcium chloride 10%: 2040 mOsm/L ⁴³ Precipitation is an additional mechanism of injury ⁵³	Early treatment: Hyaluronidase ^{13,44,49,54-56} Alternative: sodium thiosulfate ^{57,58}	Hyaluronidase: 4 of 10 cases necrosed, 2 of which had delayed administration; the severest case had surgery. ^{6,44,59-61} Hyaluronidase, flush out and aspiration: 1 case without injury. ⁵⁹	Saline flush out: 1 calcinosis cutis case necrosed and required surgery (a co-extravasation with parenteral nutrition). ⁶² Conservative: all 43 cases necrosed, including 2 with calcinosis cutis and 25 cases requiring surgery of which 9 had PFD. ^{17,44,53,63-80}
Calcium salts, including calcium disodium edetate (EDTA) Calcinosis cutis without necrosis	Monitor superficial calcifications closely as many resolve spontaneously. ¹³ Alternative: sodium thiosulfate ^{13,58}		Prednisone: calcium deposits persisted at 1 y ⁸¹ Conservative: 6 cases of which 2 resolved over months, 1 required surgery and 3 had no reported outcome. ^{53,74,82,83}
Calcium gluconate or calcium gluceptate: 680 mOsm/L ⁴³ Precipitation is an additional mechanism of injury ⁵³	Early treatment: Hyaluronidase ^{13,44,46,48,49,54,56} Alternative: sodium thiosulfate ^{57,58}	Hyaluronidase: 1 case of necrosis where treatment mitigated damage. ⁸⁴ Hyaluronidase, flush out, and aspiration: 3 cases without necrosis ⁸⁵	Flush out and aspiration: 9 cases necrosed. ⁸⁶ Conservative: 39 cases necrosed, including 28 cases that required surgery with 8 cases of PFD. One nonsurgical case had PFD. ^{17,23,57,63,64,72,78,87-101}
Calcium gluconate or calcium gluceptate Calcinosis cutis without necrosis	Monitor superficial calcifications closely as many resolve spontaneously. ¹³ Alternative: sodium thiosulfate ^{13,58}	Sodium thiosulfate: 1 case where treatment started day 83 after inadequate response to warm compresses; improvement began day 100 with eventual full resolution. ¹⁰²	Conservative: 33 cases healed without sequelae. ^{87-89,103-116} Five cases healed with permanent cosmetic deficit ¹⁰⁸ and 1 case had PFD. ¹⁰⁵
Dextrose ≥10% 10% solution: 505 mOsm/L ³	Hyaluronidase ^{13,44,49,54,56,58}	Hyaluronidase: 1 case necrosed, but 7 had no injury including 1 case treated with incisions for fluid outlet and 1 co-extravasation with calcium ^{7,44,45,51,60,61,117-119}	Flush out: 1 case necrosed ⁹⁹ Flush out and aspiration: 1 case necrosed ¹²⁰ Conservative: 30 of 40 cases necrosed, including 14 that required surgery with 8 cases of PFD. One nonsurgical case had PFD. ^{42,63-65,67,68,70,79,80,85,99,121-126}
Diazepam: >2000 mOsm/kg ¹⁰	Hyaluronidase ¹³		Conservative: 2 necrosed and had PFD. ⁶⁴
Digoxin Cytotoxicity and vasoconstriction are additional mechanisms of tissue injury ¹²⁷	Hyaluronidase ¹³		
Etomidate: 4965 mOsm/L ¹²⁸	Hyaluronidase ¹³		
Lorazepam: >2000 mOsm/kg ¹⁰	Hyaluronidase ¹³		
Mannitol ≥20% 20%: 1100 mOsm/L ⁴³	Hyaluronidase ^{13,47,48,56,58}	Hyaluronidase: 2 cases had no necrosis, including 1 case treated with cold packs. ^{23,129}	Compartment syndrome: 4 cases ¹³⁰⁻¹³³ Conservative: 1 bullae case required surgery. ¹³⁴

(continues)

TABLE 2

Antidote Recommendations for Noncytotoxic Vesicants (Continued)

Vesicant	Antidote recommendations	Antidote cases	Other treatments
Nafcillin 40 mg/mL: 402 mOsm/kg ⁴³	Hyaluronidase ^{13,44,46,48,49,55}	Hyaluronidase: 2 cases of injury prevention. ¹³⁵	Conservative: 5 cases necrosed including 1 that required surgery. ^{135,137}
Parenteral nutrition: >900 mOsm/L	Hyaluronidase ^{13,44,49,55,56,58}	Hyaluronidase: 8 of 29 cases necrosed of which 2 had surgery. ^{6,9,60,117,138,139,139b} Hyaluronidase, flush out, and aspiration: 4 of 27 cases necrosed. ^{44,45,85,139b,140-142} Chondroitinsulfatase (like hyaluronidase, used in France): 2 cases had no necrosis. ¹⁴³	Incisions for fluid effluent: 9 of 12 cases necrosed and required surgery. ¹⁴⁴ Another unspecified number had no necrosis. ¹⁴⁵ Open incision and irrigation: 4 of 26 cases necrosed. ⁴² Surgical drainage: 1 case had no necrosis. ¹⁴⁶ Flush out and aspiration: 8 of 27 cases necrosed, 4 of which required surgery. ^{99,147} Compartment syndrome: 2 cases. ^{42,148} Nitroglycerin patch: 1 case without injury where the patch had been, but a necrotic lesion formed next to the patch site. ¹⁴⁹ Conservative: 314 cases of which 273 had necrosis. The severest were 38 cases that required surgery with 7 cases of PFD. One death occurred. ^{6,8,40,42,63-65,68,70,76,92,99,121,122,139a,149-165}
Potassium chloride 20 mEq/100 mL: 400 mOsm/L 40 mEq/100 mL: 799 mOsm/L ³	Hyaluronidase ^{13,44,49,55,56}	Hyaluronidase: at least 1 case of injury prevention. ^{44,45}	Conservative: 17 of 18 cases necrosed (1 each a co-extravasation with calcium gluconate, 5% dextrose, diazepam, hypertonic saline and 2 cases with 5% dextrose in 0.45% saline). One case had compartment syndrome (a co-extravasation with 5% dextrose). Six of 11 severest surgery cases had PFD. One nonsurgical case had PFD. ^{40,64,80,92,121,166-170} Lidocaine nerve block in 1 case failed to prevent necrosis and required surgery. ¹⁷¹
Sodium bicarbonate ≥8.4% 4.2%: 1000 mOsm/L 8.4%: 2000 mOsm/L ⁴³	Hyaluronidase ^{13,45-47,49,56} Hyaluronidase or lidocaine recommended by manufacturer ¹⁷²	Hyaluronidase and flush out: 1 of 2 cases necrosed. ^{41,173}	Conservative: 23 of 25 cases necrosed, including 16 surgery cases of which 2 had PFD. Two nonsurgical cases had PFD. ^{23,41,70,80,85,123,174-177} Incision and drains: 1 case necrosed. ¹⁶⁷
Sodium chloride ≥3% 0.9% 308 mOsm/L ⁴³ 3% 839 mOsm/L ³	Hyaluronidase ^{13,45,48-50}		Conservative: 3 of 18 cases had injury (1 was a co-extravasation with propofol), 2 had no injury, and 13 cases didn't specify injury. ^{85,178-180}
Sodium phosphate: 7000 mOsm/L ⁴³	Hyaluronidase ¹³		Conservative: 3 cases of persistent plaques, 1 of which had ulceration. ¹⁸¹
pH mediated tissue injury			
Acyclovir: pH 11 ⁴³	Hyaluronidase ^{13,49}	Hyaluronidase, flush out, and aspiration: 7 cases had no injury. ⁸⁵	Conservative: 5 of 10 cases necrosed, including 1 that required surgery; 4 cases vesiculated and 1 had no injury. ¹⁸²⁻¹⁸⁹
Amiodarone: pH 4.08 ⁴³	Hyaluronidase ^{13,56}	Hyaluronidase: no injury in 2 cases where dry heat was inadequate. ¹⁹⁰	Conservative: 2 cases necrosed, including 1 that required surgery and had PFD. ^{191,192}

(continues)

TABLE 2

Antidote Recommendations for Noncytotoxic Vesicants (Continued)

Vesicant	Antidote recommendations	Antidote cases	Other treatments
Arginine: pH 5.6 ³ Hypertonicity ¹⁹³ and local hyperkalemia ¹⁹⁴ are additional mechanisms of tissue injury	Hyaluronidase for acid/base vesicants ¹³		Conservative: 4 cases necrosed including 2 that required surgery. ¹⁹³⁻¹⁹⁶
Conivaptan: pH 3.4-3.8 ⁴³	Hyaluronidase ¹³		
Dantrolene: pH 9.5-10.3 ⁴³	Hyaluronidase for acid/base vesicants ¹³		
Doxycycline: pH 1.8-3.3 ⁴³	Hyaluronidase ¹³		
Esmolol: pH 4.5-6.5 ⁴³	Hyaluronidase for acid/base vesicants ¹³	Hyaluronidase, flush out, and aspiration: Necrosis from unspecified "beta blocker." ⁵⁹	
Gentamicin: pH 3-5.5 ⁴³	Hyaluronidase ^{13,48-50}		Conservative: 3 cases necrosed; 2 were co-extravasations with penicillin. ^{23,72,99}
Immune globulin: pH 4-7.2 ⁴³	Hyaluronidase for osmolarity and acid/base vesicants ¹³	Hyaluronidase: 1 case with no necrosis. ⁶⁰ Hyaluronidase before or with subcutaneous infusion: 2 cases necrosed with small scarring. ¹⁹⁷	Conservative: 3 of 5 cases necrosed. ^{23,85,99,198}
Pentamidine: pH 4.5-7.5 ⁴³	Hyaluronidase ¹³		Conservative: 1 case necrosed with PFD. ¹⁹⁹
Pentobarbital: pH 9-10.5 ⁴³	Hyaluronidase for acid/base vesicants ¹³		
Phenobarbital: pH 9.2-10.2 ⁴³	Hyaluronidase ^{13,49}		Conservative: 1 case had bullae and 1 had necrosis and surgery. ^{200,201}
Phenytoin: pH 10-12.3 ⁴³ Precipitation an additional mechanism of injury. ¹²⁷	Hyaluronidase ^{13,46-49,118} Alternative: nitroglycerin discussed, but not explicitly recommended. ^{13,46}	Hyaluronidase: 1 case had no necrosis. ²⁰² Hyaluronidase, flush out, and cold pack: 1 case had no necrosis. ²⁰³ Nitroglycerin (patch) and dry heat: 1 case of fasciotomy prevention. ²⁰⁴	Flush out with lactated Ringers: 1 case had no necrosis. ²⁰⁵ Conservative: 56 of 63 cases necrosed, including 18 that required surgery with 10 cases of PFD and 1 nonsurgical PFD; 4 cases were treated with local heparin, heparinoid, or steroid therapy. ^{5,80,99,206-220}
Promethazine: pH 4.0-5.5 ⁴³ Cytotoxicity an additional mechanism of injury. ¹²⁷	Hyaluronidase ¹³		Papaverine (oral), stellate ganglion block, systemic steroids: 1 case necrosed with persistent cold intolerance. ²²¹
Vancomycin: pH 2.5-4.5 ⁴³	Hyaluronidase ^{13,46-49,55,56}		Conservative: 4 cases, of which 2 had bullous lesions and 2 had necrosis, including a co-extravasation with parenteral nutrition and Zosyn that required surgery. ²²²⁻²²⁵
Absorption refractory mediated tissue injury			
Lipids: pH 8.1 ¹⁰ 356 mOsm/kg ¹⁰	Authors recommend: Hyaluronidase and consider flush out.		Conservative: 1 case of necrosis required surgery. ²³

(continues)

TABLE 2

Antidote Recommendations for Noncytotoxic Vesicants (Continued)

Vesicant	Antidote recommendations	Antidote cases	Other treatments
Propofol: pH 6.0-8.5 ²⁶ Isotonic ²⁶	Authors recommend: hyaluronidase and consider flush out.	Hyaluronidase, flush out, and drains: 2 cases without necrosis including a co-extravasation with analgesics and Hartmann's solution and a case with inadequate response to hyaluronidase. ^{25,36}	Compartment syndrome: 4 cases with surgery. ^{32,33,35} Conservative: 6 of 7 cases necrosed including 4 that required surgery. ^{24,26,27,29-31,34}
Unknown mechanism of tissue injury			
Amphotericin: pH 5-7 ⁴³ 100 mg/L in dextrose 5%: 265 mOsm/kg ¹⁰	Authors recommend: hyaluronidase and for liposomal amphotericin consider flush out.		Aspirin: ice for 24 h then heat: 1 case without injury. ²²⁷ Oxygen therapy, heat, and 20% dextrose dressings: 1 stage IV case with co-extravasation of dopamine, amphotericin, famotidine, fentanyl, amikacin, and meropenem. Complete recovery by day 12. ²²⁸
Metronidazole: pH 5.5 (range 4.5-7.0) 310 mOsm/L ⁴³	Authors recommend: Hyaluronidase		Conservative: 1 case without injury. ²²⁹ Aspirin, pentoxifylline, and nifedipine: 1 case of necrosis required surgery. ²³⁰
Penicillin: pH 5-8.5 ⁴³ Iso-osmotic ⁴³	Hyaluronidase ^{46,49,50,55,56,118}		Conservative: 2 co-extravasations with gentamicin necrosed. ^{23,72}
Valproate: pH 7.6 ⁴³ Variable osmolality ⁴³ Toxicity to skin structures is a proposed mechanism of injury ²³²	Authors recommend: consider hyaluronidase with flush out.		Conservative: 1 case required surgery. ²³²
Vasoconstriction mediated tissue injury			
Dobutamine Cytotoxicity is an additional mechanism of injury ^{3,127}	First line: phenolamine ^{13,46-49,56,58,233,236} Second line: terbutaline or nitroglycerin ^{13,46,58}	Terbutaline: 1 case without damage (a co-extravasation with dopamine). ²³³	Conservative: 9 of 10 cases necrosed including 4 surgery cases with 1 PFD. ^{41,99,122,234,235}
Dopamine	First line: phenolamine ^{13,45-50,56,58,230,236} Second line: terbutaline ^{13,231,233} or nitroglycerin ^{13,46,58,231,233}	Phentolamine: 1 of 20 cases necrosed. ^{237,241} Phentolamine with nitroglycerin: 4 cases without necrosis, including 1 case of administration of nitroglycerin after inadequate response to phenolamine. ^{242,243} Phentolamine and papaverine: 1 case with functional deficit. ²⁴⁴ Nitroglycerin topical: 5 cases of which 4 were without necrosis. The necrosis case was an epinephrine co-extravasation that had surgery. ^{242,245-247}	Hyaluronidase, flush out, and aspiration: 1 of 7 cases necrosed, likely due to treatment delay of 6 h. ⁵⁹ Flush out with aspiration: 2 cases without injury. ⁵⁹ Conservative: 9 of 20 cases necrosed and required surgery, including 4 with PFD. ^{64,246,248-251}

(continues)

TABLE 2

Antidote Recommendations for Noncytotoxic Vesicants (Continued)

Vesicant	Antidote recommendations	Antidote cases	Other treatments
Epinephrine	First line: phentolamine ^{13,45-50,56,58,231,233,236} Second line: terbutaline ^{13,231} or nitroglycerin ^{13,46,58}	Lidocaine with epinephrine in digits: phentolamine (n = 54) vs saline (n = 54) had faster return of color (85 vs 319 min) and sensation (120 vs 549 min); no necrosis developed. ²⁵² Phentolamine administered after amyl nitrate inhalations were inadequate in a finger laceration exposed to topical epinephrine (1 case) ²⁵³ Autoinjector exposure with full symptom resolution (# of cases): • Phentolamine (12)²⁵⁴⁻²⁶³ • Phentolamine and oral nifedipine (1)²⁶⁴ • Phentolamine with lidocaine nerve block (2)^{265,266} • Nitroglycerin: 6 cases including 1 sublingual^{256,258,267,268} • Nitroglycerin and terbutaline (3)^{233,258} • Nitroglycerin and lidocaine nerve block (1)²⁵⁷ Phentolamine rescue therapy after failed therapy with: • Nitroglycerin (2)^{258,269} • Nitroglycerin with lidocaine nerve block (1)²⁷⁰ • Nitroglycerin, topical, with nifedipine sublingual (1)²⁷¹ • Terbutaline (1)²³³	Hyaluronidase, flush out, and aspiration: a co-extravasation with calcium gluconate, 50% dextrose, albumin, and sodium bicarbonate had no necrosis. ¹⁰⁰ Pentoxifylline: 1 case necrosed. ²⁷² Conservative: 5 cases necrosed, including 1 co-extravasation with aramine and 1 with sodium bicarbonate. Of the 5 cases, 4 required surgery with 2 cases of PFD. ^{64,151,167,177} Epinephrine (with or without lidocaine) in digits with conservative treatment: 8 cases with eventual full resolution; 1 case with 10 wk of impaired sensation and pain. ²⁷³ Autoinjector Exposure, symptoms resolved: • Iloprost intravenous with Stellate ganglion block in 1 case.²⁷⁴ • Nifedipine oral in 1 case.²⁵⁷ • Conservative: 9 cases.^{256-258,273}
Methylene blue: pH 3-4. ⁵⁴³ Cytotoxicity ¹²⁷ and pH ⁴³ are additional mechanisms of injury.	First line: nitroglycerin ^{13,58} Second line: phentolamine or terbutaline (for adrenergic antagonist) ^{13,58}		Conservative: 3 cases necrosed, including 2 surgery cases with 1 PFD. ²⁷⁵⁻²⁷⁷ Lymph mapping (1 bone marking): • Conservative: 21 of 47 cases necrosed, including 9 that required surgery.²⁷⁸⁻²⁸⁴
Norepinephrine	First line: phentolamine FDA approved ²⁸⁵ Second line: terbutaline ^{13,231,233} or nitroglycerin ^{13,46,58}	Phentolamine and nitroglycerin: 16 cases without injury. ²⁴³ Phentolamine and hyaluronidase: 15 cases without injury. ²⁸⁶ Axillary block and surgical vein decompression mitigated damage after inadequate phentolamine response. ²⁸⁷	Stellate ganglion block at 48 h and vein decompression at 72 h mitigated the damage. ²⁸⁷ Flush out: fully recovered at 4 wk. ¹⁶⁷ Conservative: 4 cases at stage 1 or 2 without injury. ²⁸⁸
Phenylephrine	First line: phentolamine ^{13,45-47,49,50,56,58,231,236} Second line: nitroglycerin ^{13,46,58}		Conservative: 2 of 16 cases necrosed and required surgery. ²⁸⁸⁻²⁹¹
Vasopressin	First line: nitroglycerin ^{13,48} Second line: phentolamine ^{13,50} or terbutaline ¹³		Stellate ganglion block and heat: 1 norepinephrine co-extravasation without injury. ²⁹² Conservative: 5 cases necrosed and required surgery, with 1 case of PFD. ²⁹³⁻²⁹⁷

Abbreviations: FDA: US Food and Drug Administration, PFD: permanent functional deficit.

chloride, ulcer size in normal saline-treated and untreated rabbits was on average more than twice as large as hyaluronidase-treated rabbits. When hyaluronidase treatments in calcium chloride injected rabbits were delayed by 1 hour, the benefits remained statistically significant, but at 30 minutes, 3 hours, 6 hours, and 12 hours, hyaluronidase trended toward benefit but did not reach significance ($P = .1$, $P = .3$, $P = .5$, and $P = .2$, respectively). The authors theorized that the acute inflammatory phase at 30 minutes postextravasation inhibited the benefit of hyaluronidase and that the positive impact then began to wear off after the 1-hour mark. Timely administration of hyaluronidase can promote complete absorption of a vesicant before the necrosis interval is reached, thus preventing permanent damage.

The effect of hyaluronidase in the treatment of accidental extravasation is rapid and marked. The effect of hyaluronidase has been described as "immediate blanching of the overlying erythematous skin and dramatic decrease in swelling,"^{202(p246)} that "within 5 minutes of hyaluronidase administration, the infant's foot developed a pinker hue and began to soften."^{51(p187)} "Within 30 minutes, the patient's pain lessened, erythema abated, and soft tissue swelling was noted to improve."^{119(pp257,e1)} More objectively, in 1 patient after only 15 minutes, the dorsalis pedis pulse that had been nonpalpable was again palpable,⁵¹ and in a second case after a repeat administration of hyaluronidase, the radial and superficial palmar pulses, which had been absent to auscultation, were once again present.²⁰² Others have described the effect as "swelling and redness markedly decreased" over the 5-hour observation period and the patient's limb was normal sized again at 24 hours.^{118(pp886,e4)} In the author's experience, a patient's intense itching and burning sensation from amiodarone extravasation completely resolved within 90 minutes of hyaluronidase administration without any adverse effects. Hyaluronidase is a highly effective, inexpensive therapy that can be administered with minimal discomfort.

Pharmacologic Antidotes **Hyaluronidase**

For adult and pediatric patients, 15 units (up to 150 units) is typically administered subcutaneously in 5 divided doses of 0.2 mL each on the periphery of swelling like the points of a star. Repeat dosing every 30 to 60 minutes until desired effect is achieved. Total doses up to 450 units have been used without adverse effect.^{45,59}

Sodium Thiosulfate

Calcinosis cutis treatment for adults is 12.5 g IV over 30 minutes and may increase gradually to 25 g 3 times per week.^{13,102} Sodium thiosulfate combines with calcium to create calcium thiosulfate, which has a solubility between 250 and 100,000 times greater than other calcium salts.³¹¹

For acute extravasation treatment and calcinosis cutis prevention, mice treated with 0.1 mL of 25% sodium thiosulfate intradermally after injection with 0.12 mL of

calcium gluconate had reduced incidence of calcification versus untreated mice (13% versus 53%) and 100% resolution of all ulcerations by day 21 versus only 73% resolution in untreated mice.³¹¹

Phentolamine ($\alpha 1$ Antagonist)

For adult and pediatric patients 5 to 10 mg in 10 to 20 mL of normal saline is typically administered intradermally in 5 divided doses on the periphery of blanching like the points of a star. This may be repeated every 30 to 60 minutes until desired effect or patient experiences an intolerably low blood pressure. Half-life in the bloodstream is 19 minutes.²⁸⁵ Doses as low as 0.5 mg have been used initially in adults, and Zenk et al⁴⁵ recommend 0.5 mg as the starting dose in neonates.²⁵⁷

Terbutaline ($\beta 2$ Agonist)

For adult and pediatric patients, terbutaline 1 mg in 10 mL of normal saline is typically administered intradermally in 5 divided doses on the periphery of blanching like the points of a star.²³³ This may be repeated every 30 to 60 minutes until desired effect or patient experiences intolerable systemic effects, such as tachycardia or irregular blood pressure. The drug's half-life is 2.9 hours.³¹²

Topical Nitroglycerin (Peripheral Vasodilator)

Apply 1 inch topically for adults and 4 mm/kg for neonates to affected area every 8 hours as needed until symptoms resolve.^{242,245}

Nonpharmacologic and Supportive Therapies

Other than thermal therapy with warm or cold compresses, the use of supportive therapies is outside the scope of this review. They are presented merely to illustrate which therapies could be considered without a recommendation for or against their use.

Elevation

Yucha et al³¹³ tested intentional 5-mL infiltrations of 0.45% saline and 3% saline when the infiltrated arm was kept at the level of the heart versus elevated by 4 inches. They found no difference in pain scores, induration size, or infiltrate volume. This could have been attributed to the low volume of infiltrate or the small difference in elevation angle. Elevation may have a greater impact with a larger volume of infiltrate, for example 50 mL, or with a steeper angle of elevation, which has been used in neonates.³⁰⁰

Warm and Cold Compresses

Compresses can be applied at 15-minute intervals 4 times daily, although continuous use of up to 72 minutes on initial extravasation has been reported.³¹⁴ Cold compresses should be used when the goal is to localize and limit the spread of the vesicant to mitigate the damage, such as with cytotoxic vesicants.⁵⁰ Cold compresses should be used to treat extravasations of valproate, because the proposed

TABLE 3**Case Series Not Linked to a Specific Vesicant**

Extravasates (# of cases)	Total number of cases	Treatments	Outcomes
Calcium, dextrose (high concentration), inotropes, parenteral nutrition, potassium, sodium bicarbonate ²⁹⁸	56	Surgical saline flush out	No skin or soft tissue damage reported and no patient required reconstructive surgery.
Blood (1), calcium (20), chemotherapy (39), contrast (1), dextrose 10%-20% (6), dobutamine (1), flucloxacillin (2), heroin (2), parenteral nutrition (14), potassium (3), sodium bicarbonate (4), thiopentone (3) ⁴¹	96	37 patients treated with surgical saline flush out, 1 with liposuction and 6 with both	39 patients with no skin damage and 5 patients with only minor skin blistering or delayed healing, compared with 52 patients initially treated conservatively of which 26 required surgical intervention.
Calcium gluconate (4), doxorubicin (4), parenteral nutrition (7) ²⁹⁹	15	10 treated with surgical saline flush out (with hyaluronidase in 2 cases). Five managed conservatively.	7 flush out patients with no symptoms. 2 flush out patients healed without need for skin graft. 1 flush out patient and 5 conservative management (late referral) patients treated with artificial skin with no functional deficits and acceptable cosmetic defects.
Albumin (1), antibiotics with dextrose 5% in 0.22% sodium chloride (7 cases), blood (2), chemotherapy (5), dextrose 5% with potassium chloride (30 mEq/L) in 0.45% sodium chloride (1), dopamine (1), heparin (2), intralipid (1), and parenteral nutrition: no additives (13), with antibiotics (6), with contrast (1). Antibiotics included nafcillin, chloramphenicol, oxacillin, ampicillin, carbenicillin, cephalothin and gentamicin. ³⁰⁰	34	Conservative management and wound care (silver sulfadiazine, povidone iodine, saline dressings)	No notable difference between treatment groups. No skin grafts required.
Calcium chloride (2), cloxacillin (1), epinephrine (2), dextrose 10% (5), parenteral nutrition without lipids (8), phenytoin (1), potassium chloride (11), sodium bicarbonate (1), vancomycin (1). Some patients had more than one drug extravasate. ⁴⁷	34	Hyaluronidase in 14 patients (including inappropriately in 1 epinephrine case)	No adverse effects of hyaluronidase. Outcomes not reported. Authors' note: reported that 34 extravasations should have been treated with an antidote according to published protocol, but upon reviewing list of extravasates and protocol, only 32 cases appear to have been eligible for antidote administration.
Aminophylline and insulin (1), blood (5), calcium and 10% dextrose (5), dextrose 10% and magnesium (1), flucloxacillin and dextrose 10% (1), parenteral nutrition (14), platelets (1), sodium bicarbonate (2), tromethamine (2). (4 cases had co-extravasation of 2 solutions). ³⁰¹	28	Hyaluronidase and saline irrigation (10). Saline irrigation without hyaluronidase (2).	Survey inclusion required that necrosis had developed.
Contrast (4), ciprofloxacin (3) dextrose 10% (4), dopamine (5), fentanyl (3), parenteral nutrition (38), potassium chloride (4). Other agents less than 3 cases each (29). ³⁰²	90		Parenteral nutrition: 31 cases treated with hyaluronidase. 46 of 90 cases fully recovered, 10 required wound care referral, 44 outcomes not documented.
Not reported ³⁰³	115	Hyaluronidase in 110 peripheral cases (3.9% of peripheral lines placed) and 5 midline cases (1.2% of midlines placed).	Outcomes not reported; 24 significant midline catheter infiltrations not treated with hyaluronidase. Original author unable to provide additional information.

(continues)

TABLE 3

Case Series Not Linked to a Specific Vesicant (*Continued*)

Extravasates (# of cases)	Total number of cases	Treatments	Outcomes
Cyclophosphamide (1), dextrose 5% in 0.45% sodium chloride with 20 mEq potassium chloride/liter (2), lipids and parenteral nutrition (2), unreported (142). ⁴⁰	147	Heat (32), elevation (9), heat and elevation (28), cold (3), hyaluronidase (25), phentolamine (1), no treatment (49).	Only grade 3 or 4 injuries included. No surgical intervention required, no compartment syndrome. Severe stage 4 injuries from reported extravasates treated by wound care team with healing over 17 d on average. Original author unable to provide additional information.
Not reported. ³⁰⁴	4	Active leptospermum honey and dehydrated amniotic membrane allograft.	Healing in 27, 34, 21, and 41 d, respectively.
Included amino acids and calcium. ¹⁵⁷	61	Initial treatment not reported.	30 barely perceptible scars from fluid infiltration or extravasation. 31 noticeable scars including 4 significant cosmetic or functional scars. 1 of the 4 significant scars was due to parenteral nutrition. Also 3 patients with areas of alopecia from infiltration. 100 NICU graduates assessed for scars.
Antibiotics ⁸⁵	13	Conservative treatment	Healed without necrosis
Drugs ¹¹⁷	10	Hyaluronidase for stage 3 or 4 injuries	3 cases were stage IV and received hyaluronidase. No patients required debridement or skin grafting.
Calcium chloride and/or dextrose 10% ¹²⁶	15	Conservative management	Only stage 3 or 4 injuries included. Scarring in every patient. 4 patients required surgery.

Abbreviation NICU, neonatal intensive care unit.

mechanism of tissue injury is toxicity to skin structures.²³² Warm compresses in conjunction with vasodilatory antidotes (phentolamine, terbutaline, and nitroglycerin) should be used for extravasation of vasoconstrictive vesicants where the goal is to increase circulation. For pH-mediated, osmolarity-mediated, absorption refractory, and even some cytotoxic concentration-dependent vesicants, the goal is to disperse and absorb the vesicant, so warm compresses should be used along with the antidote hyaluronidase.^{13,50} Vesicants containing fats, including 3-in-1 parenteral nutrition and the absorption refractory vesicants propofol and lipids, will experience improved absorption and distribution with warm compresses because of improved solubility and decreased viscosity of fats with increased temperature. "Use of dry heat in conjunction with hyaluronidase works synergistically to increase blood flow and disperse the extravasated drug."^{1(p5100)} Hastings-Tolsma et al³¹⁴ demonstrated that warm compresses are effective at aiding in dispersion of infiltrated fluids, whereas cold compresses tended to keep the fluids trapped. Eighteen patients had 5 mL purposefully infiltrated; 6 with 0.45% saline, 6 with 0.9% saline, and 6 with 3% saline. At 12, 42, and 72 minutes postinfiltration, the 9 patients with warm compresses had statistically significantly lower infiltrate volumes remaining

($P < .001$) versus the 9 patients with cold compresses (3 in each concentration group). Thermal compress therapy is not without risks and should be performed carefully.^{236,315} See Table 4 for thermal compress recommendations.³¹⁶⁻³³²

Physical Massage or Compression Therapy

To manually aid in fluid dispersion and reduce tissue pressure, physical massage can be used, or an inflatable splint can be placed on a neonate and inflated for a portion of every hour to physically disperse infiltrated fluid that is anatomically trapped.^{333,334}

Localized Steroid Therapy

Topical or locally injected steroids have been reported with mixed results. Ahn et al³³⁵ found in rabbits that triamcinolone therapy seemed to reduce the reaction but was unable to prevent calcinosis cutis. Compañía et al³¹¹ conversely found that, in mice, triamcinolone therapy actually worsened the extent of the reaction.

Antimicrobial Therapy

Topical and systemic preventive antimicrobial therapy have been reported. Silver sulfadiazine cream can be used, because extravasation is a chemical burn. Secondary

TABLE 4

Thermal Compress Recommendations for Noncytotoxic Vesicants

	Thermal compress recommendations		Supportive literature
Vesicant	Primary	Secondary	
Osmolarity mediated tissue injury			
Aminophylline	Warm ^{13,50}	Warm or cold ^{13,58} Cold ⁴⁶	None available
Ampicillin	Warm ⁵⁰	Warm or cold ¹³ Cold ⁵⁶	Co-extravasation with cefotaxime, dextrose 10% in 0.225% sodium chloride. Warm packs inadequate, so hyaluronidase used resulting in no necrosis. ⁵¹
Calcium chloride	Warm ^{50,58}	Warm or cold ¹³ Cold ⁵⁶	Calcinosis cutis and fat necrosis resolved over 4 weeks with warm wet compresses. ³¹⁶
Calcium disodium edetate (EDTA)	Warm ³¹⁷		Calcinosis cutis that required surgical excision despite use of warm soaks. ³¹⁷
Calcium gluconate	Warm ^{50,58}	Warm or cold ¹³	1 case of inadequate response to dry heat that required sodium thiosulfate. ¹⁰² 3 cases of necrosis despite use of warm soaks. ^{87,88,318} 11 cases of necrosis despite use of cold compresses (with saline flush out in 9 cases). ^{86,319,320}
Dextrose	Warm ⁵⁰	Warm or cold ¹³ Cold ^{46,56,58}	3 cases used cold compresses of which 2 cases with hyaluronidase had no necrosis, whereas the case without hyaluronidase required fasciotomies. ^{118,119,321} One case with warm compresses had bullae formation, but necrosis was prevented. ³²²
Diazepam	Warm ⁵⁸	Warm or cold ¹³ Cold ⁴⁶	Diazepam and phenytoin co-extravasation: necrosis and surgery despite alternating hot and cold compresses for 3 days. ³²³
Digoxin	Warm ^a	Warm or cold ¹³ Cold ⁵⁸	None available
Etomidate	Warm ^a	Warm or Cold ¹³	None available
Immune globulin	Warm ²³		Necrosis despite warm packs ²³
Lorazepam	Warm ^a	Warm or cold ¹³ Cold ⁴⁶	None available
Mannitol	Warm ⁵⁰	Warm or cold ¹³ Cold ^{56,58}	No necrosis in 1 case of warm compress with hyaluronidase and 1 case of cold compress with hyaluronidase. ^{129,324}
Nafcillin	Warm ⁵⁰	Warm or cold ¹³ Cold ⁴⁶	Necrosis despite warm compresses. ¹³⁵
Parenteral nutrition	Warm ⁵⁰	Warm or cold ^{13,58} Cold ^{46,56}	Necrosis in 4 cases despite cold compresses in 2 cases, and warm compresses for co-extravasations with lipids in 2 cases. ^{155,167,325}
Phosphate salts	Warm ⁵⁰	Warm or cold ¹³	None available
Potassium chloride	Warm ^{50,58}	Warm or cold ¹³ Cold ^{46,56}	20 mEq/L in 5% dextrose and 0.225% sodium chloride: required fasciotomy despite improved edema and discoloration at periphery from use of warm compresses. ³²⁶
Sodium bicarbonate	Warm ^{50,58,172}	Warm or cold ¹³ Cold ^{46,56}	None available
Sodium chloride 3%	Warm ⁵⁰	Warm or cold ¹³ Cold ⁵⁸	18 patients had 5 mL purposefully infiltrated: 6 with half normal saline, 6 with normal saline, and 6 with 3% saline. 3 patients in each group were treated with cold compresses and 3 with warm compresses. At 12, 42, and 72 min postinfiltration, the 9 warm compress patients had statistically significantly lower infiltrate volumes versus the 9 cold compress patients. ³¹⁴
pH mediated tissue injury			
Acyclovir	Warm ¹³	Cold ⁵⁸	Cold compresses with gradual recovery over 1 mo. ³²⁷
Amiodarone	Warm ¹³	Warm or cold ⁵⁸ Cold ⁵⁶	2 cases (1 observed by an author of this article) with inadequate response to warm compresses where use of hyaluronidase prevented necrosis. ¹⁹⁰ One case of necrosis despite cold compresses ³²⁸
Arginine	Warm ¹³		2 cases of necrosis despite cold compresses ^{329,330}
Conivaptan	Warm ¹³		None available
Dantrolene	Warm ¹³		None available
Doxycycline	Warm ¹³	Cold ⁴⁶	None available

(continues)

TABLE 4

Thermal Compress Recommendations for Noncytotoxic Vesicants (Continued)

Vesicant	Thermal compress recommendations		Supportive literature
	Primary	Secondary	
Esmolol	Warm ¹³		None available
Gentamicin	Warm ^{13,48,49,58}		None available
Pentamidine	Warm ¹³		None available
Pentobarbital	Warm ¹³		None available
Phenobarbital	Warm ¹³	Cold ⁵⁸	None available
Phenytoin	Warm ^{13,55}		Fasciotomy prevented with use of dry heat and nitroglycerin patch. ²⁰⁴ Resolution over 3 wk with dry heat and local steroid injection. ²²⁰ Necrosis prevented with cold pack, hyaluronidase and saline flush out. ²⁰³ Necrosis despite cold compresses and heparinoid cream. ²¹⁵
Promethazine	Warm ¹³	Cold ⁴⁶	Fasciotomy required despite warm compresses ¹²
Vancomycin	Warm ^{13,50}	Cold ⁵⁶	None available
Absorption refractory mediated tissue injury			
Lipids	Warm ^a		Injury responded to warm compresses, but inadequate to prevent necrosis. ²²
Propofol	Warm ^a	Cold ⁵⁸	Cool packs, no tissue injury. Residual swelling and stiffness for 41 d required physiotherapy, resolution day 62. ²⁸
Unknown mechanism of tissue injury			
Amphotericin	Warm ^a	Warm or cold ⁵⁸	Ice on day 1 then heat and aspirin from day 2 resulting in no necrosis. ²²⁷ Necrosis despite local heat, hypertonic wet dressings and oxygen therapy in a co-extravasation with amikacin, dopamine, famotidine, fentanyl, and meropenem. ²²⁸
Metronidazole	Warm ^{50,58}		None available
Penicillin	Warm ⁵⁰	Cold ⁵⁶	None available
Valproate	Cold ^a		None available
Vasoconstriction mediated tissue injury			
Dobutamine	Warm ^{13,46,50,56,58}		Necrosis complicated by cellulitis responded to warm soaks. ³³¹
Dopamine	Warm ^{13,46,50,56,58}		2 cases of necrosis despite warm compresses and 1 case of phentolamine with warm compresses without necrosis. ^{80,241} Warm compresses in 1 co-extravasation with epinephrine resulted in no injury. ²⁴⁷
Epinephrine	Warm ^{13,46,50,56,58}		Incidental autoinjector hand exposures with warm compresses: 9 required antidote, ^{233,254,256,257,261,266,268,271} and 17 cases resolved without antidote administration. ²⁵⁸ Epinephrine applied to skin tear required antidote despite warm soaks. ²⁵³
Methylene blue	Warm ^{13,58}		None available
Norepinephrine	Warm ^{13,46,50,56,58}		"Therapeutic application of heat... has not been helpful." ^{332(p218)} Stellate ganglion block and warm blankets prevented necrosis in a co-extravasation with vasopressin. ²⁹²
Phenylephrine	Warm ^{13,50,58}		None available
Vasopressin	Warm ^{13,50}		Necrosis despite warm compresses. ²⁹⁷

^aAuthors' recommendation.

bacterial infections of damaged tissue have been reported and treated accordingly with topical or systemic antimicrobial agents as indicated.

Saline Flush Out/Irrigation

A highly effective method of physically removing the vesicant is through multiple surgical incisions along with

placement of a catheter to administer a physiologic saline flush out and perform aspiration that is similar to liposuction.⁴¹ Saline flush out has become so common in London that advance practice nurses are performing the procedure as opposed to a plastic surgeon.³³⁶ Other variations from this method include open incision and irrigation, incisions and drain insertion, incisions with mechanical

pressure from gentle massage, or saline flush out without aspiration.^{13,42,139a,142,167}

Once a wound develops, consultation with a wound care clinician is advisable, because a multitude of different wound care strategies have been reported.^{6,9,139a,173,228,323,333,337-340}

DISCUSSION

In the recognition and treatment of an extravasation injury, there are 4 possible outcomes:

- Treatment indicated—treated
- Treatment indicated—untreated: error
- Treatment not indicated—treated: error
- Treatment not indicated—untreated

The potential errors are either overtreatment, if treated without indication, or undertreatment, a lack of treatment when indicated. In the balance of how aggressively an institution manages extravasation injuries, they will tend to overtreat or undertreat extravasations. Because of the unique nature of each extravasation, determining the appropriate level of treatment is challenging. Whether extravasation treatment is indicated is difficult to assess initially. In most reported cases, the extent of injury was far greater than it first appeared. Some common reasons for this are as follows: (1) signs and symptoms occur internally, not just externally where visible; (2) physical signs of damage may be masked by other symptoms; (3) uncertainties in timing and volume of the extravasation; and (4) delayed presentation of symptoms (eg, calcium deposition). Imaging such as x-ray, magnetic resonance imaging, ultrasound, and gallium uptake bone scan (in cases of calcinosis cutis) can be used to estimate the volume of extravasation or the extent of internal injury, but in assessing whether to treat in the absence of compelling evidence either way, the cost of overtreatment or undertreatment must be considered.^{61,104,138,160}

Overtreatment of extravasation injury incurs minimal cost, because the antidotes are inexpensive and are well tolerated. Initial undertreatment of extravasation has caused prolonged hospitalization of up to 55 days.⁷² The extravasation injury is often more severe than the primary diagnosis. Morbidity and mortality on the magnitude of debridement of necrotic tissue, skin grafting, limb amputation, and even death from fungal meningitis following a parenteral nutrition extravasation have all been reported.^{64,162,167,171} Extravasations that appear to be managed appropriately but are undertreated may present later with unforeseen consequences, such as secondary necrotizing fasciitis, limb deformations from prematurely fused growth plates or fibrin deposition, scalp skin grafting with a residual bald spot, an ossified mass inhibiting foot flexion, or retinal detachment from secondary infection.^{64,68,341-344} Morbidity can also be accompanied by litigation.²¹⁸ Not every extravasation injury requires treatment, but in the

decision between potentially committing an overtreatment error versus an undertreatment error, generally the safer decision for the patient and for the clinician is to treat.

For treatment of large-volume infiltrations of medications that are not known vesicants, hyaluronidase should be considered. It has the potential to prevent tissue injury or compartment syndrome as “it can reduce pressure necrosis from mechanical compression of tissues by large amounts of IV fluids trapped in a limited tissue space.”^{45(p295)} Infiltrations of nonvesicant medications that are large in volume compared with the size of the anatomic infusion site can cause vesicant-like damage. In very low birth weight infants, the line between irritant and vesicant blurs, because most infusion volumes are large compared with their very small size, so large-volume extravasations of any fluid in these patients should be treated with hyaluronidase.³⁴⁵

Treatment bias exists in the extravasation body of literature. Less severe extravasations are less likely to receive an antidote but are also more likely to resolve on their own without necrosis, making it appear that antidotes are unnecessary to facilitate healing without necrosis. More severe extravasations are more likely to receive an antidote but are also more likely to result in necrosis or permanent functional deficit, making the antidotes appear ineffective because they were unable to prevent necrosis. The efficacy of an antidote is demonstrated when antidote administration mitigated the catastrophic damage that could have resulted or altogether prevented damage that would have occurred. Because there is no controlled comparator, the damage that might have occurred if not treated can only be presumed based on cases where no antidote was administered. Without standardized severity reporting, appropriately comparing results of antidote-treated cases with untreated cases is difficult if not impossible.

Reporting and observation bias are also seen in extravasation literature. Cases that resolved spontaneously or are similar to previously reported cases are generally not considered worthy of reporting. Alternatively, case series targeting the safety of a peripheral infusion of a vesicant are watching for extravasations to happen and so are likely to catch them earlier, resulting in a low incidence of mild extravasation injuries. This may misrepresent the true incidence and severity of extravasations.

CONCLUSION

Extravasation injury is an established risk of IV vesicant administration. Preventative measures can decrease the risk but not eliminate it, so clinicians should be prepared to treat extravasation injuries in a time-sensitive, evidence-based manner. Treatments are based on the vesicant's mechanism of tissue injury. Cytotoxic vesicants with concentration-dependent toxicity can be treated with warm compresses and hyaluronidase. If the potency of the cytotoxic vesicant cannot be mitigated through dilution, the

most appropriate therapy might be cold compresses with strong consideration of saline flush out. The antidotes for vasopressor extravasations are phentolamine, terbutaline, or nitroglycerin. For pH-mediated, osmolarity-mediated, and absorption refractory extravasations, the antidote is hyaluronidase. Warm compresses should be used for vasopressor, pH-mediated, osmolarity-mediated, and absorption refractory vesicants. Future study regarding absorption refractory vesicants to better characterize this newly proposed mechanism of tissue injury would help clinicians better understand this pattern of tissue damage and could add support to treatment recommendations.

Because the body of extravasation literature is subject to multiple biases, more evidence must be gathered in a standardized way before extravasation recommendations can rely solely on evidence without the need for supporting consensus recommendations. In scenarios similar to extravasation injury where ethical considerations preclude clinical trials, like fetal drug exposure, evidence has been gathered in registries. Registries can be managed by a health system-based research organization, as part of a clinical trial, by a drug manufacturer or joint manufacturer effort potentially as part of a Risk Evaluation and Mitigation Strategy, by a nonprofit research organization or by a professional organization (<https://www.fda.gov/science-research/womens-health-research/list-pregnancy-exposure-registries>). The American College of Radiology has a registry for IV contrast extravasation, but no registry covers any other noncytotoxic extravasations (<https://www.acr.org/-/media/ACR/Files/Registries/NRDROverview.pdf>). According to poison control operatives, poison control advises patients and clinicians on accidental epinephrine autoinjector exposure management and advises clinicians for noncytotoxic extravasations on a regular basis (personal communication, December 3, 2019). When reports of extravasation accumulate, the evidence-based recommendations will become even more robust. Outside of a registry, adverse effects can and should be reported within the United States to the FDA MedWatch. Because the scope of MedWatch contains any serious adverse effect, the required details are not tailored specifically to extravasation injury like a registry reporting form could be. Registry data collected and published by the task force of a professional organization have the greatest chance of producing the requisite case information in the volume necessary to support evidence-based recommendations. The authors hope that as clinicians follow the recommendations contained in this document, the registry could support or refute and refine these recommendations to provide a more robust set of evidence-based noncytotoxic extravasation management recommendations.

ACKNOWLEDGMENTS

The authors would like to thank Jan Rice, MLS, AHIP, and Terri Raburn, MLIS, for their expert assistance in the literature search and

citations and Lisa Gorski, MS, RN, HHCNS-BC, CRNI®, FAAN, for her encouragement, support, and assistance in manuscript preparation.

REFERENCES

- Gorski L, Hadaway L, Hagle ME, McGoldrick M, Orr M, Doellman D. Infusion therapy standards of practice. *J Infus Nurs*. 2016;39(suppl 1):S1–S159.
- Swartz AJ. Chemotherapy extravasation management: part I—doxorubicin (Adriamycin). *Cancer Nurs*. 1979;2(5):405–407.
- Gorski L, Stranz M, Cook LS, et al. Development of an evidence-based list of noncytotoxic vesicant medications and solutions. *J Infus Nurs*. 2017;40(1):26–40. doi:10.1097/NAN.0000000000000202
- Loth TS, Eversmann Jr WW. Extravasation injuries in the upper extremity. *Clin Orthop Relat Res*. 1991;(272):248–254.
- Corbett M, Marshall D, Harden M, Oddie S, Phillips R, McGuire W. Treatment of extravasation injuries in infants and young children: a scoping review and survey. *Health Technol Assess*. 2018;22(46):1–112. doi:10.3310/hta22460.
- Friedman J. Plastic surgical problems in the neonatal intensive care unit. *Clin Plast Surg*. 1998;25(4):599–617.
- Thigpen JL. Peripheral intravenous extravasation: nursing procedure for initial treatment. *Neonatal Netw*. 2007;26(6):379–384. doi:10.1891/0730-0832.26.6.379
- Boyar V, Handa D, Clemens K, Shimborske D. Clinical experience with Leptospermum honey use for treatment of hard to heal neonatal wounds: case series. *J Perinatol* 2014;34(2):161–163. doi:10.1038/jp.2013.158
- Schie JC, Goodman KL. Treatment of neonatal extravasation injuries using non-contact, low-frequency ultrasound: development of a new treatment protocol. *Newborn Infant Nurs Rev*. 2013;13(1):42–47.
- Clark E, Giambra BK, Hingl J, Doellman D, Tofani B, Johnson N. Reducing risk of harm from extravasation: a 3-tiered evidence-based list of pediatric peripheral intravenous infusates. *J Infus Nurs*. 2013;36(1):37–45. doi:10.1097/NAN.0b013e3182798844
- Doellman D, Hadaway L, Bowe-Geddes LA, et al. Infiltration and extravasation: update on prevention and management. *J Infus Nurs*. 2009;34(9):203–211.
- Hadaway L. Infiltration and extravasation: preventing a complication of IV catheterization. *Am J Nurs*. 2007;107(8):64–72.
- Reynolds PM, MacLaren R, Mueller SW, Fish DN, Kiser TH. Management of extravasation injuries: a focused evaluation of noncytotoxic medications. *Pharmacotherapy*. 2014; 34(6):617–32. doi:10.1002/phar.1396
- Perucca R. Peripheral venous access devices. In: Alexander M, Corrigan A, Gorski L, Hankins J, Perucca R, eds. *Infusion Nursing: An Evidence-Based Approach*. 3rd ed. Saunders; 2010:456–479.
- Sterile water. Package insert*. Hospira; 2010.
- Stranz M, Kastango ES. A review of pH and osmolarity. *Int J Pharm Compd*. 2002;6(3):216–220.
- Heckler FR, McCraw JB. Calcium-related cutaneous necrosis. *Surg Forum*. 1976;27(6):553–555.
- Sty JR, Starshak RJ, Hubbard AM. GA-67 scintigraphy: calcium gluconate extravasation. *Clin Nucl Med*. 1982;7(8):377.
- Orellana P, Velasquez C, Meneses L, et al. Tc-99m MDP uptake secondary to soft tissue extravasation of calcium gluconate in a newborn thought to have osteomyelitis. *Clin Nucl Med*. 2002;27(9):653–655.
- De Beuckeleer LH, Vanhoenacker FM, Spinhoven M, Dieltjens A. Images in clinical radiology: calcinosis cutis following calcium gluconate extravasation. *JBR-BTR*. 2003;86(5):305.
- Olsen M, LeFebvre K, Brassil K, eds. *Chemotherapy and Immunotherapy Guidelines and Recommendations for Practice*. Oncology Nursing Society; 2019.

22. Graber R, Laufman H. Massive subcutaneous fat necrosis after extravasation of intravenous infusion of fat emulsion. *Am J Surg.* 1964;107(6): 878–880. doi:10.1016/0002-9610(64)90181-3
23. Kumar RJ, Pegg SP, Kimble RM. Management of extravasation injuries. *ANZ J Surg.* 2001;71(5):285–289.
24. Tokumine J, Sugahara K, Tomori T, Nagasawa Y, Takaesu Y, Hokama A. Tissue necrosis caused by extravasated propofol. *J Anesth.* 2002;16(4):358–359. doi:10.1007/s005400200057
25. Mahajan R, Gupta R, Sharma A. Extravasation injury caused by propofol. *Anesth Analg.* 2006;102(2):648. doi:10.1213/01.ANE.0000190745.29994.58
26. Roth W, Eschertzhuber S, Gardetto A, Keller C. Extravasation of propofol is associated with tissue necrosis in small children. *Paediatr Anaesth.* 2006;16(8):887–889. doi:10.1111/j.1460-9592.2006.01873.x
27. Abdelmalak BB, Bashour CA, Yared JP. Skin infection and necrosis after subcutaneous infiltration of propofol in the intensive care unit. *Can J Anaesth.* 2008;55(7):471–473. doi:10.1007/BF03016315
28. Huijbers EJM, Baars JW, Schutte PFE, Schellens JHM, Beijnen JH. Propofol extravasation in a breast cancer patient. *J Oncol Pharm Pract.* 2008;14(4):195–198. doi:10.1177/1078155208094454
29. Basak P, Poste J, Jesmajian S. Propofol extravasation and tissue necrosis. *Indian J Dermatol.* 2012;57(1):78–79. doi:10.4103/0019-5154.92692
30. Sharma R, Yoshikawa H, Abisaab J. Chemical burn secondary to propofol extravasation. *The West J Emerg Med.* 2012;13(1):121–122. doi:10.5811/westjem.2011.6.6813
31. LeBlanc JM, Lalonde D, Cameron K, Mowatt JA. Tissue necrosis after propofol extravasation. *Intensive Care Med.* 2014;40(1):129–130. doi:10.1007/s00134-013-3137-z.
32. Kalraiya AJ, Madanipour S, Coloco H, Cobiella C. Propofol extravasation: a rare cause of compartment syndrome. *BMJ Case Rep.* 2015;2015. doi:10.1136/bcr-2015-209360
33. Varacallo M, Shirey L, Kavuri V, Harding S. Acute compartment syndrome of the hand secondary to propofol extravasation. *J Clin Anesth.* 2018;47:1–2. doi:10.1016/j.jclinane.2018.01.020
34. Riley RH, Westhoff GP. Extravasation of propofol. *Anaesth Intensive Care.* 1993;21(5):720–721.
35. Bebawy JF, Gupta DK, Koht A. Compartment syndrome caused by a properly functioning infusion pump. *J Clin Anesth.* 2011;23(2):134–136. doi:10.1016/j.jclinane.2009.11.006
36. Rodger D, Hammerschlag J. Extravasation injury leading to acute compartment syndrome in a child: the vital role of pulse oximetry in early detection and management. *J Perioper Pract.* 2018;28(4):95–98. doi:10.1177/1750458918762324
37. Millam DA. Managing complications of I.V. therapy. *Nursing.* 1988;18(3):34–43. doi:10.1097/00152193-198803000-00020
38. Flemmer L, Chan JS. A pediatric protocol for management of extravasation injuries. *Pediatr Nurs.* 1993;19(4):355–358, 424.
39. Baharestani MM. An overview of neonatal and pediatric wound care knowledge and considerations. *Ostomy Wound Manage.* 2007;53(6):34–36,38,40.
40. Odom B, Lowe L, Yates C. Peripheral infiltration and extravasation injury methodology: a retrospective study. *J Infus Nurs.* 2018;41(4):247–252. doi:10.1097/NAN.0000000000000287
41. Gault DT. Extravasation injuries. *Br J Plast Surg.* 1993;46(2):91–96. doi:10.1016/0007-1226(93)90137-z
42. Kostoglou N, Demiri E, Tsimponis A, et al. Severe extravasation injuries in neonates: a report of 34 cases. *Pediatr Dermatol.* 2015;32(6):830–835. doi:10.1111/pde.12664
43. McEvoy GK, Snow EK, Miller J, Kohoutek LM, eds. *Handbook on Injectable Drugs.* 20th ed. American Society of Health-System Pharmacists; 2018.
44. MacCara ME. Extravasation: a hazard of intravenous therapy. *Drug Intell Clin Pharm.* 1983;17(10):713–717. doi:10.1177/106002808301701002
45. Zenk KE, Sills J, Koeppel R. Guidelines for management of intravenous extravasations: appendix G. In Zenk KE, Sills J, Koeppel R, eds. *Neonatal Medications and Nutrition: A Comprehensive Guide.* 3rd ed. NICU INK Book Publishers; 2003:627–629.
46. Hurst S, McMillan M. Innovative solutions in critical care units: extravasation guidelines. *Dimens Crit Care Nurs.* 2004;23(3):125–128.
47. Paquette V, McGloin R, Northway T, Dezorzi P, Singh A, Carr R. Describing intravenous extravasation in children (DIVE Study). *Can J Hosp Pharm.* 2011;54(5):340–345. doi:10.4212/cjhp.v64i5.1069
48. Hanrahan KM. Hyaluronidase for treatment of intravenous extravasations. *Series on Evidence-Based Practice for Children and Families.* University of Iowa College of Nursing, Office for Nursing Research; 2012. https://works.bepress.com/kirsten_hanrahan/21/
49. Desarno J, Sandate I, Green K, Chavez P. When in doubt, pull the catheter out: implementation of an evidence-based protocol in the prevention and management of peripheral intravenous infiltration/extravasation in neonates. *Neonatal Netw.* 2018;37(6):372–377. doi:10.1891/0730-0832.37.6.372
50. Martin SM. Extravasation management of nonchemotherapeutic medications. *J Infus Nurs.* 2013;36(6):392–396. doi:10.1097/NAN.0000000000000010
51. Kuensting LL. Treatment of intravenous infiltration in a neonate. *J Pediatr Health Care.* 2010;24(3):184–188. doi:10.1016/j.pedhc.2010.02.001
52. Borman H, Tuncali E, Apak A, Kostakoğlu N. Progressive gangrene of the hand following extravasation of antibiotics associated with hereditary resistance to activated protein C. *Ann Plast Surg.* 1998;41(2):94–200. doi:10.1097/0000637-199808000-00014
53. Jucglà A, Sais G, Curco N, Marcoval J, Moreno A, Peyri J. Calcinosis cutis following liver transplantation: a complication of intravenous calcium administration. *Br J Dermatol.* 1995;132(2):275–278. doi:10.1111/j.1365-2133.1995.tb05026.x
54. Lund C. Prevention and management of infant skin breakdown. *Nurs Clin North Am.* 1999;34(4):907–920, vii.
55. Montgomery LA, Hanrahan K, Kottman K, Otto A, Barrett T, Hermiston B. Guideline for I.V. infiltrations in pediatric patients. *Pediatr Nurs.* 1999;25(2):167–169, 173–180.
56. Maly C, Fan KL, Rogers GF, et al. A primer on the acute management of intravenous extravasation injuries for the plastic surgeon. *Plast Reconstr Surg Glob Open.* 2018;6(4):e1743. doi:10.1097/GOX.00000000000001743
57. Compañía P, Javier F, Míguez JM, de Toro Santos FJ. Lesions associated with calcium gluconate extravasation: presentation of 5 clinical cases and analysis of cases published. *Ann Plast Surg.* 2017;79(5):444–449. doi:10.1097/SAP.0000000000001110
58. David V, Christou N, Etienne P, et al. Extravasation of noncytotoxic drugs. *Ann Pharmacother.* 2020;54(8):804–814. doi:10.1177/1060028020903406
59. Casanova D, Bardot J, Magalon G. Emergency treatment of accidental infusion leakage in the newborn: report of 14 cases. *Br J Plast Surg.* 2001;54(5):396–399. doi:10.1054/bjps.2001.3593
60. Yan YM, Fan QL, Li AQ, Chen JL, Dong FF, Gong M. Treatment of cutaneous injuries of neonates induced by drug extravasation with hyaluronidase and hirudoid. *Iran J Pediatr.* 2014;24(4):352–358.
61. Boyar V, Galiczewski C, Kurepa D. Point-of-care ultrasound use in neonatal peripheral intravenous extravasation injuries: a case series. *J Wound Ostomy Continence Nurs.* 2018;45(6):503–509. doi:10.1097/WON.0000000000000475
62. Cizmeci MN, Kanburoglu MK, Tatli MM. Extravasation and extra vigilance. *J Vasc Access.* 2013;14(2):201. doi:10.5301/jva.5000120

63. Yosowitz P, Ekland DA, Shaw RC, Parsons RW. Peripheral intravenous infiltration necrosis. *Ann Surg.* 1975;182(5):553–556. doi:10.1097/0000658-197511000-00003
64. Upton J, Mulliken JB, Murray JE. Major intravenous extravasation injuries. *Am J Surg.* 1979;137(4):497–506. doi:10.1016/0002-9610(79)90121-1
65. Wales JKH, Hall MA, Walton P. Neonatal infusion site complications. *Br J Parenteral Ther.* 1985: 6–8.
66. Sanchez V, Segedin ER, Moser M, Pallares VS. Role of lumbar sympathectomy in the pediatric intensive care unit. *Anesth Analg.* 1988;67(8):794–797.
67. Sanpera I, Fixsen JA, Hill RA. Injuries to the physis by extravasation: a rare cause of growth plate arrest. *J Bone Joint Surg Br.* 1994;76(2):278–280.
68. Fullilove S, Fixsen J. Major limb deformities as complications of vascular access in neonates. *Paediatr Anaesth.* 1997;7(3):247–250. doi:10.1111/pan.1997.7.3.247
69. Wada A, Fujii T, Takamura K, Yanagida H, Matsuura A, Katayama A. Physeal arrest of the ankle secondary to extravasation in a neonate and its treatment by the gruca operation: a modern application of an old technique. *J Pediatr Orthop B.* 2003;12(2):129–132. doi:10.1097/01.bpb.0000049576.53117.ae
70. Sawatzky-Dickson D, Bodnaryk K. Neonatal intravenous extravasation injuries: evaluation of a wound care protocol. *Neonatal Netw.* 2006;25(1):13–19. doi:10.1891/0730-0832.25.1.13
71. Lin CY, Hsieh KC, Yeh MC, Sheen-Chen SM, Chou FF. Skin necrosis after intravenous calcium chloride administration as a complication of parathyroidectomy for secondary hyperparathyroidism: report of four cases. *Surg Today.* 2007;37(9):778–781. doi:10.1007/s00595-006-3426-z
72. Rose REC, Felix R, Crawford-Sykes A, Venugopal R, Wharfe G, Arscott G. Extravasation injuries. *West Indian Med J.* 2008;57(1):40–47.
73. Younus M. A medication error by calcium ampule led to an ADR. *Drug Saf.* 2010;33(10):943–944.
74. Pugashetti R, Shinkai K, Ruben BS, Grossman ME, Maldonado J, Fox LP. Calcium may preferentially deposit in areas of elastic tissue damage. *J Am Acad Dermatol.* 2011;64(2):296–301. doi:10.1016/j.jaad.2010.01.046
75. Xu C, Turner A, Yeoh TM, Carney B. Management of severe calcium chloride extravasation injury: a case report. *ANZ J Surg.* 2016;86(5):421–422. doi:10.1111/ans.13437
76. de Blécourt RA, Bloem JJ. Treatment of extravasation of intravenously administered agents [article in Dutch]. *Ned Tijdschr Geneesk.* 1996;140(8):409–411.
77. Semple P, Booth C. Calcium chloride; a reminder. *Anaesthesia.* 1996;51(1):93.
78. Girard P, Plancq MC, Tourneux P, Deroussen F, Gouron R, Klein C. Extravasation of calcium solution in the child: value of negative-pressure wound therapy. *Arch Pediatr.* 2019;26(7):407–410. doi:10.1016/j.arcped.2019.09.011
79. Guy RL, Holland JP, Shaw DG, Fixsen JA. Limb shortening secondary to complications of vascular cannulae in the neonatal period. *Skeletal Radiol.* 1990;19(6):423–425. doi:10.1007/bf00241795
80. Firat C, Erbatur S, Aytekin AH. Management of extravasation injuries: a retrospective study. *J Plast Surg Hand Surg.* 2013;47(1):60–65. doi:10.3109/2000656X.2012.741065
81. Domínguez-Fernández I, Goñiz R, Pérez-Gala S, Fraga J, Fernández-Herrera J. Calcinosis cutis following extravasation of calcium salts. *J Eur Acad Dermatol Venereol.* 2008;22(4):505–506. doi:10.1111/j.1468-3083.2007.02369.x
82. Kagen MH, Bansal MG, Grossman M. Calcinosis cutis following the administration of intravenous calcium therapy. *Cutis.* 2000;65(4):193–194.
83. Tuncer S, Aydin A, Erer M. Extravasation of calcium solution leading to calcinosis cutis surrounding the dorsal cutaneous branch of the ulnar nerve. *J Hand Surg Br.* 2006;31(3):288–289. doi:10.1016/j.jhsb.2005.12.001
84. Nickel B. Peripheral intravenous access: applying infusion therapy standards of practice to improve patient safety. *Crit Care Nurse.* 2019;39(1):61–71. doi:10.4037/ccn2019790
85. Ghanem AM, Mansour A, Exton R, et al. Childhood extravasation injuries: improved outcome following the introduction of hospital-wide guidelines. *J Plast Reconstr Aesthet Surg.* 2015;68(4):505–518. doi:10.1016/j.bjps.2014.12.029
86. Sivrioğlu N, Irkören S. Versajet hydrosurgery system in the debridement of skin necrosis after Ca gluconate extravasation: report of 9 infantile cases. *Acta Orthop Traumatol Turc.* 2014;48(1):6–9. doi:10.3944/AOTT.2014.2941
87. Berger PE, Heidelberger KP, Poznanski AK. Extravasation of calcium gluconate as a cause of soft tissue calcification in infancy. *Am J Roentgenol Radium Ther Nucl Med.* 1974;121(1):109–117. doi:10.2214/ajr.121.1.109
88. Roberts JR. Cutaneous and subcutaneous complications of calcium infusions. *JACEP.* 1977;6(1):16–20.
89. Lee FA, Gwinn JL. Roentgen patterns of extravasation of calcium gluconate in the tissues of the neonate. *J Pediatr.* 1975;86(4):598–601. doi:10.1016/s0022-3476(75)80161-2
90. Weiss Y, Ackerman C, Shmilovitz L. Localized necrosis of scalp in neonates due to calcium gluconate infusions: a cautionary note. *Pediatrics.* 1975;56(6):1084–1086.
91. Wolfe MS, North ER. Extravasation of injected calcium solution leading to calcifications in the upper extremity of the neonate. Report of a case. *J Bone Joint Surg Am.* 1983;65(4):558–559.
92. Falcone PA, Barrall DT, Jeyarajah DR, Grossman JA. Nonoperative management of full-thickness intravenous extravasation injuries in premature neonates using enzymatic debridement. *Ann Plast Surg.* 1989;22(2):146–149. doi:10.1097/0000637-198902000-00010
93. Morrison W, Hurley JV, Ahmad TS, Webster HR. Scar formation after skin injury to the human foetus in utero or the premature neonate. *Br J Plast Surg.* 1999;52(1):6–11. doi:10.1054/bjps.1998.3009
94. Caksen H, Odaş D. An infant with gigantic subcutaneous calcium deposition following extravasation of calcium gluconate. *Pediatr Dermatol.* 2002;19(3):277–279. doi:10.1046/j.1525-1470.2002.00663.x
95. Ruiz Genao D, Fernández AT, Zambrano AZ. Cutaneous necrosis due to calcium gluconate extravasation in a newborn [in Spanish]. *Actas Dermo-Sifiliográficas.* 2003;94(1):123–124. doi:10.1016/S0001-7310(03)79241-2
96. Tiras U, Erdevi O, Karabulut AA, Dallar Y, Eksioğlu HM. Debridement via Collagenase application in two neonates. *Pediatr Dermatol.* 2005;22(5):472–475. doi:10.1111/j.1525-1470.2005.00119.x
97. Puvabanditsin S, Garrow E, Titapiwatanakun R, Getachew R, Patel JB. Severe calcinosis cutis in an infant. *Pediatr Radiol.* 2005;35(5):539–542. doi:10.1007/s00247-004-1363-9
98. Domizio S, Puglielli C, Barbante E, et al. Calcinosis cutis in a newborn caused by minimal calcium gluconate extravasation. *Int J Dermatol.* 2006;45(12):1439–1440. doi:10.1111/j.1365-4632.2006.02693.x
99. Murphy AD, Gilmour RF, Coombs CJ. Extravasation injury in a paediatric population. *ANZ J Surg.* 2019;89(4):E122–E126. doi:10.1111/ans.14104
100. Martin PH, Carver N, Petros AJ. Use of liposuction and saline wash-out for the treatment of extensive subcutaneous extravasation of corrosive drugs. *Br J Anaesth.* 1994;72(6):702–704. doi:10.1093/bja/72.6.702
101. Lee TG, Chung S, Chung YK. A retrospective review of iatrogenic skin and soft tissue injuries. *Arch Plast Surg.* 2012;39(4):412–416. doi:10.5999/aps.2012.39.4.412

102. Raffaella C, Annapaola C, Tullio I, Angelo R, Giuseppe L, Simone C. Successful treatment of severe iatrogenic calcinosis cutis with intravenous sodium thiosulfate in a child affected by T-acute lymphoblastic leukemia. *Pediatr Dermat.* 2009;26(3):311–315. doi:10.1111/j.1525-1470.2008.00776.x
103. Ravenel SD. Cellulitis from extravasation of calcium gluconate simulating osteomyelitis. *Am J Dis Child.* 1983;137(4):402–403. doi:10.1001/archpedi.1983.02140300080024
104. Balsam D, Goldfarb CR, Stringer B, Farruggia S. Bone scintigraphy for neonatal osteomyelitis: simulation by extravasation of intravenous calcium. *Radiology.* 1980;135(1):185–186. doi:10.1148/radiology.135.1.7360958
105. Leape LL. Calcification of the leg after calcium infusion. *J Pediatr Surg.* 1975;10(5):831–833. doi:10.1016/0022-3468(75)90394-2
106. Packer JE, Naidech HJ, Young LW. Radiological case of the month: soft-tissue calcification caused by calcium gluconate extravasation. *Am J Dis Child.* 1984;138(5):505–506.
107. Lakhani JK. Calcium gluconate-its unusual complication. *Indian Pediatr.* 1996;33(6):510–512.
108. Mu SC, Lin CH, Sung TC. Calcinosis cutis following extravasation of calcium gluconate in neonates. *Acta Paediatr Taiwan.* 1999;40(1):34–35.
109. Garcia-Alvarez F, Bello ML, Albareda J, Seral F. Neonatal tumoration in the secondary to calcium extravasation. *Rev Esp Pediatr.* 1999;55(1):279–280.
110. Millard TP, Harris AJ, MacDonald DM. Calcinosis cutis following intravenous infusion of calcium gluconate. *Br J Dermatol.* 1999;140(1):184–186. doi:10.1046/j.1365-2133.1999.02641.x
111. Soon SL, Chen S, Warshaw E, Caughman SW. Calcinosis cutis as a complication of parenteral calcium gluconate therapy. *J Pediatr.* 2001;138(5):778. doi:10.1067/mpd.2001.112060
112. Arora A, Agarwal A, Kumar S, Gupta SK. Iatrogenic calcinosis cutis—a rare differential diagnosis of soft-tissue infection in a neonate: a case report. *J Orthop Surg (Hong Kong).* 2005;13(2):195–198. doi:10.1177/230949900501300218
113. Moss J, Syrengelas A, Antaya R, Lazova R. Calcinosis cutis: a complication of intravenous administration of calcium gluconate. *J Cutan Pathol.* 2006;33(Suppl 2):60–62. doi:10.1111/j.1600-0560.2006.00519.x
114. Sonohata M, Akiyama T, Fujita I, Asami A, Mawatari M, Hotokebuchi T. Neonate with calcinosis cutis following extravasation of calcium gluconate. *J Orthop Sci.* 2008;13(3):269–272. doi:10.1007/s00776-007-1217-z
115. Vaccari S, Ismaili A, Barisani A, Neri I, Patrizi A. Solitary erythematous, tender plaque of the heel in a young infant. *Dermatol Online J.* 2013;19(9):19617.
116. Ching DLH, Wong KY, Milroy C. Iatrogenic calcinosis cutis following a neonatal extravasation injury. *Br J Hosp Med (Lond).* 2014;75(5):295. doi:10.12968/hmed.2014.75.5.295
117. Chowdhury M, Wong LH, Horn V, Eaton E, Pierro A. Extravasation injury in surgical neonates and children. *Proc Nutr Soc.* 2004;63(suppl 1):23A.
118. Lawson SL, Brady W, Mahmoud A. Identification of highly concentrated dextrose solution (50% dextrose) extravasation and treatment—a clinical report. *Am J Emerg Med.* 2013;31(5):886.e3–5. doi:10.1016/j.ajem.2012.12.010
119. Wiegand R, Brown J. Hyaluronidase for the management of dextrose extravasation. *Am J Emerg Med.* 2010;28(2):257.e1–2. doi:10.1016/j.ajem.2009.06.010
120. Aribit F, Laville J, Baron J. Extravasation injuries and subcutaneous aspiration in children [article in French]. *Rev Chir Orthop Reparatrice Appar Mot.* 2000;86(1):87–88.
121. Pasquesoone L, Aljudaihi N, Ellart J, Guerreschi P, Duquennoy-Martinot V. Emergency management of extravasation in children [in French]. *Ann Chir Plast Esthet.* 2016;61(5):598–604. doi:10.1016/j.anplas.2016.07.016
122. Meszes A, Tólosi G, Máder K, Orvos H, Kemény L, Csoma ZR. Lesions requiring wound management in a central tertiary neonatal intensive care unit. *World J Pediatr.* 2017;13(2):165–172. doi:10.1007/s12519-016-0070-6
123. Milcheski DA, Mota WM, Lobato RC, Monteiro Jr AA, Gemperli R. Surgical treatment of extravasation injuries: experience of the Hospital das Clínicas, Faculty of Medicine, University of São Paulo. *Rev Col Bras Cir.* 2018;45(4):e1912. doi:10.1590/0100-6991e-20181912
124. Levy SB, Rosh AJ. Images in emergency medicine: dextrose extravasation causing skin necrosis. *Ann Emerg Med.* 2006;48(3):236, 239. doi:10.1016/j.annemergmed.2006.01.004
125. Pantelides NM, Shah AK. Extravasation injury: a simple technique to maintain limb elevation within a neonatal intensive care unit. *J Neonatal Nurs.* 2013;19(5):243–245. doi:10.1016/j.jnn.2013.04.001
126. Nandiolo-Anelone KR, Allah KC, Cissé L, Bankolé SR, Oulaï M, Aké AYL. Extravasation injuries in newborns: our experience about 15 cases [in French]. *Chir Main.* 2014;33(1):44–50. doi:10.1016/j.main.2013.11.007
127. Buter J, Steele KT, Chung KC, Elzinga K. Extravasation injury from chemotherapy and other non-antineoplastic vesicants. Savarese DMF, Collins KA, eds. UpToDate.com website. Accessed December 5, 2019. <https://www.uptodate.com/contents/extravasation-injury-from-chemotherapy-and-other-non-antineoplastic-vesicants>
128. Niedermirtl F, Eberhardt M, Namer B, et al. Etomidate and propylene glycol activate nociceptive TRP ion channels. *Mol Pain.* 2018;14:1744806918811699. doi:10.1177/1744806918811699
129. Vacca VM. Vesicant extravasation. *Nursing.* 2013;43(9):21–22. doi:10.1097/01.NURSE.0000432917.59376.55
130. Stahl S, Lerner A. Compartment syndrome of the forearm following extravasation of mannitol in an unconscious patient. *Acta Neurochir (Wien).* 2000;142(8):945–946. doi:10.1007/s007010070083
131. Edwards JJ, Samuels D, Fu ES. Forearm compartment syndrome from intravenous mannitol extravasation during general anesthesia. *Anesth Analg.* 2003;96(1):245–246. doi:10.1097/00000539-200301000-00049
132. Eroglu A, Uzunlar H. Forearm compartment syndrome after intravenous mannitol extravasation in a carbosulfan poisoning patient. *J Toxicol Clin Toxicol.* 2004;42(5):649–652. doi:10.1081/clt-200026975
133. Erickson BA, Yap RL, Pazona JF, Hartigan BJ, Smith ND. Mannitol extravasation during partial nephrectomy leading to forearm compartment syndrome. *Int Braz J Urol.* 2007;33(1):68–71. doi:10.1590/s1677-55382007000100010
134. Chang KA, Jawan B, Luk HN, Fung ST, Lee JH. Bullous eruptions caused by extravasation of mannitol—a case report. *Acta Anaesthesiol Sin.* 2001;39(4):195–198.
135. Zenk KE, Dungy CI, Greene GR. Nafcillin extravasation injury: use of hyaluronidase as an antidote. *Am J Dis Child.* 1981;135(12):1113–1114. doi:10.1001/archpedi.1981.02130360021008
136. Tilden SJ, Craft JC, Cano R, Daum RS. Cutaneous necrosis associated with intravenous nafcillin therapy. *Am J Dis Child.* 1980;134(11):1046–1048. doi:10.1001/archpedi.1980.02130230026008
137. Moore RA, Terry BE. Nafcillin necrosis. *NITA.* 1984;7(1):61–62.
138. Yan Y-M, Gong M, Chen J-L, et al. Incidence, risk factors and treatment outcomes of drug extravasation in pediatric patients in china. *Turk J Pediatr.* 2017;59(2):1621–1168. doi:10.24953/turk-jped.2017.02.008
139. Khan I, Rizvi S-SA, Malik I, Weinberger B, Puvabanditsin S, Hegyi T. Extravasation of intravenous fluid in a preterm neonate. *Consultant.* 2014;54(3):188–190.
- 139a. Boyar V, Galiczewski C. Efficacy of dehydrated human amniotic membrane allograft for the treatment of severe extravasation injuries in preterm neonates. *Wounds.* 2018;30(8):224–228.

Downloaded from https://journals.lww.com/journalofinfusionnursing by H88RLEF1WZCEZ2BDFUCaSkmtDQGH
kydLLZBBY8Y81YD1V031W6INRQWYFQ0UdIAE9V3Z450CUIUZ41F0d0Gm3Y1Z5hUnek102mWmgpYpYZZRGdJhF+
Y1v4D0GpFGBICFASW13T3poCWgaT4d06Id6gys= on 09/19/2024

140. Chen YY, Wang HZ, Pan CH, Hsieh KS. Treatment of extravasation of intravenous fluid: a case report. *Clinical Neonatology*. 1996;3:27–29.

141. Siu SL, Kwong KL, Poon SST, So KT. The use of hyaluronidase for treatment of extravasations in a premature infant. *HK J Paediatr (New Series)*. 2007;12:130–132.

142. Davies J, Gault D, Buchdahl R. Preventing the scars of neonatal intensive care. *Arch Dis Child Fetal Neonatal Ed*. 1994;70(1):F50–51.

143. Gil ME, Mateu J. Treatment of extravasation from parenteral nutrition solution. *Ann Pharmacother*. 1998;32(1):51–55. doi:10.1345/aph.16487

144. Sung KY, Lee SY. Nonoperative management of extravasation injuries associated with neonatal parenteral nutrition using multiple punctures and a hydrocolloid dressing. *Wounds*. 2016;28(5):145–151.

145. Chandavasu O, Garrow E, Valda V, Alsheikh S, Vega SD. A new method for the prevention of skin sloughs and necrosis secondary to intravenous infiltration. *Am J Perinatol*. 1986;3(1):4–5. doi:10.1055/s-2007-999813

146. Govind B, Tete PI, Thomas N. Percutaneous central line extravasation masquerading as an abscess. *Indian Pediatr*. 2014;51(4):309–310. doi:10.1007/s13312-014-0384-1

147. Onesti MG, Carella S, Maruccia M, et al. The use of hyalomatrix PA in the treatment of extravasation affecting premature neonates. *Plast Reconstr Surg*. 2012;129(1):219e–221e.

148. Park HJ, Kim KH, Lee HJ, Jeong EC, Kim KW, Suh DI. Compartment syndrome due to extravasation of peripheral parenteral nutrition: extravasation injury of parenteral nutrition. *Korean J Pediatr*. 2015;58(11):454–458. doi:10.3345/kjp.2015.58.11.454

149. O'Reilly C, McKay FM, Duffty P, Lloyd DJ. Glyceryl trinitrate in skin necrosis caused by extravasation of parenteral nutrition. *Lancet (London, England)*. 1988;2(8610):565–566.

150. Benalleg A, Mazouni M, Grangaud J, Benabdal J, Bagrich M. Subdural extravasation in newborns [article in French]. *Arch Fr Pediatr*. 1969;26(7):818.

151. Lynch DJ, Key JC, White RR. Management and prevention of infiltration and extravasation injury. *Surg Clin North Am*. 1979;59(5):939–949. doi:10.1016/s0039-6109(16)41940-7

152. Koo WW, Fong T, Gupta JM. Parenteral nutrition in infants. *Aust Paediatr J*. 1980;16(3):169–174.

153. Reynolds BC. Neonatal extravasation injury: case report. *Infant*. 2007;3(6):230–232.

154. Cho KY, Lee SJ, Burm JS, Park EA. Successful combined treatment with total parenteral nutrition fluid extravasation injuries in preterm infants. *J Korean Med Sci*. 2007;22(3):588–594. doi:10.3346/jkms.2007.22.3.588

155. Lehr VT, Lulic-Botica M, Lindblad WJ, Kazzi NJ, Aranda JV. Management of infiltration injury in neonates using duoderm hydroactive gel. *Am J Perinatol*. 2004;21(7):409–414. doi:10.1055/s-2004-835309

156. Tobin C. Managing an extravasation wound in a premature infant. *Wounds UK*. 2007;3(1):90–91.

157. Cartlidge PH, Fox PE, Rutter N. The scars of newborn intensive care. *Early Hum Dev*. 1990;21(1):1–10. doi:10.1016/0378-3782(90)90105-r

158. Bourneau-Martin D, Leboucher B, Le Bouedec S, et al. Risk of extravasation injury with PEDIAVEN and other parenteral nutrition solutions used with peripheral intravenous in French neonatal intensive care unit. *Drug Saf*. 2015;38(10):993–994.

159. Moon HS, Burm JS, Yang WY, Kang SY. Prognosis of full-thickness skin defects in premature infants. *Arch Plast Surg*. 2012;39(5):463–468. doi:10.5999/aps.2012.39.5.463

160. Kurepa D, Galiczewski C, Civala A, Boyar V. Point-of-care ultrasound as an adjunct in the diagnosis of neonatal and pediatric superficial soft tissue infection: a report of two cases. *Ostomy Wound Manage*. 2017;63(7):14–19.

161. Hirsch SD, Powers JM, Rhodes JL. Neonatal soft tissue reconstruction using a bioengineered skin substitute. *J Craniofac Surg*. 2017;28(2):489–491. doi:10.1097/SCS.0000000000003346

162. Rosales CM, Jackson MA, Zwick D. Malassezia furfur meningitis associated with total parenteral nutrition subdural effusion. *Pediatr Dev Pathol*. 2004;7(1):86–90. doi:10.1007/s10024-003-4030-5

163. Atay S, Sen S, Cukurlu D. Incidence of infiltration/extravasation in newborns using peripheral venous catheter and affecting factors. *Rev Esc Enferm USP*. 2018;52:e03360. doi:10.1590/S1980-220X2017040103360

164. Ozalp Gerceker G, Kahraman A, Yardimci F, et al. Infiltration and extravasation in pediatric patients: a prevalence study in a children's hospital. *J Vasc Access*. 2018;19(3):266–271.

165. Tomàs Tomàs MM, Pérez JE, Amorós Cerdá SM. Complications of peripheral parenteral nutrition: clinical observations of 2 cases [in Spanish]. *Enferm Intensiva*. 2014;25(1):30–34. doi: 10.1016/j.enfi.2013.11.006

166. Williams HP. Accidental subcutaneous infiltration of potassium chloride solution causing necrosis. *Br Med J (Clin Res Ed)*. 1984;289(6460):1742. doi:10.1136/bmj.289.6460.1742-b

167. Schummer W, Schummer C, Bayer O, Müller A, Bredle D, Karzai W. Extravasation injury in the perioperative setting. *Anesth Analg*. 2005;100(3):722–727. doi:10.1213/01.ANE.0000154442.30278.3C

168. Altmann S, Damert HG, Schneider W. Clinical manifestation of extravasation caused by infusion and its therapeutic management [in German]. *Zentral Chir*. 2014;139(1):83–88. doi:10.1055/s-0031-1271432

169. Onesti MG, Fino P, Ponzio I, Ruggieri M, Scuderi N. Non-surgical treatment of deep wounds triggered by harmful physical and chemical agents: a successful combined use of collagenase and hyaluronic acid. *Int Wound J*. 2016;13(1):22–26. doi:10.1111/iwj.12215

170. Richa FC, Chalhoub VR, El-Hage CF, Yazbeck PH. Severe skin extravasation injury following intravenous injection of potassium chloride. *Saudi J Anaesth*. 2019;13(4):397–398. doi:10.4103/sja.SJA_451_19

171. Steib A, Bing J, Bantzhauf P, Galani M, Otteni JC. Tissue necrosis after accidental perivenous injection of potassium chloride [in French]. *Ann Fr Anesth Reanim*. 1984;3(5):396–397. doi:10.1016/s0750-7658(84)80082-9

172. Sodium bicarbonate. Package insert. Hospira; 2005.

173. Lee H, Kwon S, Choi J, et al. The management of infantile extravasation injury using maternal platelet-rich plasma. *Pediatr Dermatol*. 2013;30(6):e114–117.

174. Hampton S. Wound care. Vacuum wrapped. *Nurs Times*. 1999;95(3):77–78.

175. Rustogi R, Mill J, Fraser JF, Kimble RM. The use of Acticoat in neonatal burns. *Burns*. 2005;31(7):878–882. doi:10.1016/j.burns.2005.04.030

176. Grabois FS, Voievda T, Aqcuvita A, Kizlansky V, Genez DS, Vidaurreta S. Use of sterile petrolatum for extravasation injury in a premature infant [in Spanish]. *Arch Argent Pediatr*. 2008;106(6):533–535. doi:10.1590/S0325-00752008000600011

177. Gaze NR. Tissue necrosis caused by commonly used intravenous infusions. *Lancet*. 1978;2(8086):417–419. doi:10.1016/s0140-6736(78)91879-2

178. Andira Perez C, Figueroa SA. Complication rates of 3% hypertonic saline infusion through peripheral intravenous access. *J Neurosci Nurs*. 2017;49(3):191–195. doi:10.1097/JNN.0000000000000286

179. Dillon RC, Merchan C, Altschuler D, Papadopoulos J. Incidence of adverse events during peripheral administration of sodium chloride 3. *J Intensive Care Med*. 2018;33(1):48–53. doi:10.1177/0885066617702590

180. Jones GM, Bode L, Riha H, et al. Safety of continuous peripheral infusion of 3% sodium chloride solution in neurocritical care patients. *Am J Crit Care*. 2016;26(1):37–42.

181. Verykiou S, Aljefri K, Gopee H, et al. Cutaneous manifestations of phosphate solution extravasation. *Clin Exp Dermatol*. 2018;43(1):42–45. doi:10.1111/ced.13233

182. Sylvester RK, Ogden WB, Draxler CA, Lewis FB. Vesicular eruption: a local complication of concentrated acyclovir infusions. *JAMA*. 1986;255(3):385–386. doi:10.1001/jama.255.3.385
183. Robbins MS, Stromquist C, Tan LH. Acyclovir pH—possible cause of extravasation tissue injury. *Ann Pharmacother*. 1993;27(2):238.
184. De Souza BA, Shibu M. Painless acyclovir extravasation injury in a diabetic. *Br J Plast Surg*. 2002;55(3):264. doi:10.1054/bjps.2001.3822
185. Sarica A. Bullous cutaneous eruption due to extravasation of acyclovir in an adolescent with acute lymphoblastic leukemia. *Turk J Haematol*. 2012;29(1):109–110. doi:10.5505/tjh.2012.93685
186. Neocleous C, Andonopoulou E, Adramerina A, et al. Tissue necrosis following extravasation of acyclovir in an adolescent: a case report. *Acta Med Acad*. 2017;46(1):55–58. doi:10.5644/ama2006-124.187
187. Laskin OL, Longstreth JA, Saral R, de Miranda P, Keeney R, Lietman PS. Pharmacokinetics and tolerance of acyclovir, a new anti-herpesvirus agent, in humans. *Antimicrob Agents Chemother*. 1982;21(3):393–398. doi:10.1128/aac.21.3.393
188. Buck ML, Vittone SB, Zaglul HF. Vesicular eruptions following acyclovir administration. *Ann Pharmacother*. 1993;27(12):1458–1459. doi:10.1177/106002809302701208
189. Armingaud P, Arsac P, Kerdraon R, Esteve E. Localized bullous eruption after intravenous injection of aciclovir: toxic or immunoallergic mechanism? *Ann Dermatol Venereol*. 2000;127(5):496–498.
190. Fox AN, Villanueva R, Miller JL. Management of amiodarone extravasation with intradermal hyaluronidase. *Am J Health Syst Pharm*. 2017;74(19):1545–1548. doi:10.2146/ajhp160737
191. Russell SJ, Saltissi S. Amiodarone induced skin necrosis. *Heart*. 2006;92(10):1395. doi:10.1136/hrt.2005.086157
192. Ledingham D, Cordato D. Focal myositis and contracture secondary to amiodarone extravasation from a peripheral cannula. *BMJ Case Rep*. 2019;12(1):bcr-2018-227725. doi:10.1136/bcr-2018-227725
193. Bowlby HA, Elanjian SI. Necrosis caused by extravasation of arginine hydrochloride. *Ann Pharmacother*. 1992;26(2):263–264. doi:10.1177/106002809202600226
194. Amano H, Nagai Y, Kowase T, Ishikawa O. Cutaneous necrosis induced by extravasation of arginine monohydrochloride. *Acta Derm Venereol*. 2008;88(3):310–311. doi:10.2340/00015555-0420
195. Salameh Y, Shoufani A. Full-thickness skin necrosis after arginine extravasation—a case report and review of literature. *J Pediatr Surg*. 2004;39(4):e9–11. doi:10.1016/j.jpedsurg.2003.12.030
196. Bassi E, Lonati D, Pandolfi R, Gatti M, Jolliffe VML, Del Forno C. Case of skin necrosis due to arginine monohydrochloride extravasation. *J Dermatol*. 2007;34(3):198–200. doi:10.1111/j.1346-8138.2007.00249.x
197. Carne E, Ponsford M, El-Shanawany T, Jolles S. Skin necrosis following subcutaneous immunoglobulin (SCIG). *J Clin Immunol*. 2017;37(1):27–28. doi:10.1007/s10875-016-0346-6
198. Datta R, Kuruvilla M, Gill M, de la Morena MT. Association of skin necrosis with subcutaneous immunoglobulin therapy. *Ann Allergy Asthma Immunol*. 2014;113(2):232–233.
199. Bolognia JL. Cutaneous ulceration: an unusual complication of intravenous pentamidine therapy. *Dermatologica*. 1991;183(3):221–224. doi:10.1159/000247675
200. Haroun M, Jakubovic HR, Nethercott JR. Localized subepidermal bullae after intravenous phenobarbital. *Cutis*. 1987;39(3):233–234.
201. Schafer T, Kukies S, Stokes TH, Levin LS, Donatucci CF, Erdmann D. The prepuce as a donor site for reconstruction of an extravasation injury to the foot in a newborn. *Ann Plast Surg*. 2005;54(6):664–666. doi:10.1007/s000637-200506000-00019
202. Sokol DK, Dahlmann A, Dunn DW. Hyaluronidase treatment for intravenous phenytoin extravasation. *J Child Neurol*. 1998;13(5):246–247. doi:10.1177/088307389801300512
203. Khan MS, Holmes JD. Reducing the morbidity from extravasation injuries. *Ann Plast Surg*. 2002;48(6):628–632. doi:10.1097/0000637-200206000-00011
204. Edwards JJ, Bosek V. Extravasation injury of the upper extremity by intravenous phenytoin. *Anesth Analg*. 2002;94(3):672–673. doi:10.1097/0000539-200203000-00035
205. von Heimburg D, Pallua N. Early and late treatment of iatrogenic injection damage [in German]. *Chirurg*. 1998;69(12):1378–1382. doi:10.1007/s001040050588
206. Comer JB. Extravasation from intravenous phenytoin. *Am J Intraven Ther Clin Nutr*. 1984;11(1):23–29.
207. Kilarski DJ, Buchanan C, Von Behren L. Soft-tissue damage associated with intravenous phenytoin. *N Engl J Med*. 1984;311(18):1186–1187.
208. Spengler RF, Arrowsmith JB, Kilarski DJ, Buchanan C, Von Behren L, Graham DR. Severe soft-tissue injury following intravenous infusion of phenytoin: patient and drug administration risk factors. *Arch Intern Med*. 1988;148(6):1329–1333.
209. Rao VK, Feldman PD, Dibble DG. Extravasation injury to the hand by intravenous phenytoin: report of three cases. *J Neurosurg*. 1988;68(6):967–969. doi:10.3171/jns.1988.68.6.0967
210. Hagan HJ, Hastings H. Extravasation of phenytoin in the hand. *J Hand Surg Am*. 1988;13(6):942–943. doi:10.1016/0363-5023(88)90276-6
211. Sharief N, Goonasekera C. Soft tissue injury associated with intravenous phenytoin in a neonate. *Acta Paediatr*. 1994;83(11):1218–1219. doi:10.1111/j.1651-2227.1994.tb18288.x
212. Appleton RE, Gill A. Adverse events associated with intravenous phenytoin in children: a prospective study. *Seizure*. 2003;12(6):369–372.
213. Twardowsky CA, De Paola L, Germiniani FMB, Werneck LC, Silgado C. Pearls & oysters: soft-tissue necrosis as a result of intravenous leakage of phenytoin. *Neurology*. 2009;73(19):e94–95. doi:10.1212/WNL.0b013e3181c0d401
214. Hasija N, Hazarika AJ, Sokhal N, Kumar S. Tissue necrosis of hand caused by phenytoin extravasation: an unusual occurrence. *Saudi J Anaesth*. 2014;8(2):309–310. doi:10.4103/1658-354X.130766
215. Yan YY, Tan TJ. Utility of magnetic resonance imaging in the surgical management of drug extravasation: a case report. *J Hand Surg Asian Pac Vol*. 2017;22(1):100–103. doi:10.1142/S0218810417720030
216. Okogbaa JI, Onor IO, Arije OA, Harris MB, Lillis RA. Phenytoin-induced purple glove syndrome: a case report and review of the literature. *Hosp Pharm*. 2015;50(5):391–395. doi:10.1310/hpj5005-391
217. Viale PH, Sanchez DY. Violaceous skin reaction of the hand. *Clin J Oncol Nurs*. 2002;6(5):310–312. doi:10.1188/02.CJON.310-312
218. von Muehlendahl KE. Complications due to infusions in infants and young children: lessons from six cases of litigation of the “Norddeutsche Schlichtungsstelle für Arzthaftpflichtfragen.” *Monatsschr Kinderheilkd*. 2012;160(10):988–991. doi:10.1007/s00112-012-2699-0
219. Hayes AG, Chesney TM. Necrosis of the hand after extravasation of intravenously administered phenytoin. *J Am Acad Dermatol*. 1993;28(2 Pt 2):360–363.
220. Sonohata M, Asami A, Tsunoda K, Hotokebuchi T. Purple glove syndrome associated with intravenous phenytoin administration in a patient with severe mental and motor retardation. *J Orthop Sci*. 2006;11(4):409–411. doi:10.1007/s00776-006-1024-y
221. Malesker MA, Malone PM, Cingle CM, Cochran RM. Extravasation of IV promethazine. *Am J Health Syst Pharm*. 1999;56(17):1742–1743.
222. Hoelen DWM, Tjan DHT, van Vugt R, van der Meer YG, van Zanten ARH. Severe local vancomycin induced skin necrosis. *Br J Clin Pharmacol*. 2007;64(4):553–554. doi:10.1111/j.1365-2125.2007.02897.x
223. Bohm NM, Wong JG. Bullous dermatosis associated with vancomycin extravasation. *Am J Med Sci*. 2012;343(2):177–179. doi:10.1097/MAJ.0b013e318237bb47
224. Nanjappa S, Snyder M, Greene JN. Vancomycin infiltrate-induced dermatitis mimicking bullous cellulitis. *J Drugs Dermatol*. 2017;16(11):1160–1163.

225. Boyar V. Pediatric extravasation injuries. *Ostomy Wound Manage.* 2019;65(2):14–15.
226. Propofol. Package insert. Hospira; 2018.
227. Olson C, McCoy LK. Amphotericin B extravasation: a case report. *NITA.* 1985;8(4):299–300.
228. Beytut D, Ozdamar N, Isler N, Turgut N. Abstract 594: Extravasation injury: Lokal oxigene therapy application within pediatric intensive care unit. *Pediatr Crit Care Med.* 2014;15(4 suppl):134–135. doi:10.1097/01.pcc.0000449320.67961.46
229. North AM, Yee JM. A case of metronidazole injection infiltration without sequelae. *Hosp Pharm.* 2017;52(8):559–563. doi:10.1177/0018578717722885
230. Bharani A, Chattopadhyay BP, Dani P, Bhargava KD. Metronidazole extravasation causing digital gangrene. *Indian J Physiol Pharmacol.* 1995;39(3):307–308.
231. Le A, Patel S. Extravasation of noncytotoxic drugs: a review of the literature. *Ann Pharmacother.* 2014;48(7):870–886. doi:10.1177/1060028014527820
232. Santivasi WL, Kulkarni S, Patton ML, et al. Infiltration of sodium valproate with compartment syndrome and bullous reaction: case report and literature review. *Burns.* 2011;37(7):e59–62. doi:10.1016/j.burns.2011.05.010
233. Stier PA, Bogner MP, Webster K, Leikin JB, Burda A. Use of subcutaneous terbutaline to reverse peripheral ischemia. *Am J Emerg Med.* 1999;17(1):91–94. doi:10.1016/s0735-6757(99)90028-1
234. Reed WP, Newman KA, Applefeld MM, Sutton FJ. Drug extravasation as a complication of venous access ports. *Ann Intern Med.* 1985;102(6):788–790. doi:10.7326/0003-4819-102-6-788
235. Applefeld MM, Newman KA, Sutton FJ, et al. Outpatient dobutamine and dopamine infusions in the management of chronic heart failure: clinical experience in 21 patients. *Am Heart J.* 1987;114(3):589–595.
236. Brandon D, Hill CM, Heimal L, et al. *Neonatal Skin Care, Evidence-based Clinical Practice Guideline.* 4th ed. Association of Women's Health, Obstetric and Neonatal Nurses; 2018.
237. Stetson JB, Reading GP. Avoidance of vascular complications associated with the use of dopamine. *Can Anaesth Soc J.* 1977;24(6):727–733. doi:10.1007/bf03006717
238. Siwy BK, Sadove AM. Acute management of dopamine infiltration injury with Regitine. *Plast Reconstr Surg.* 1987;80(4):610–612. doi:10.1097/00006534-198710000-00024
239. Dugger B. Peripheral dopamine infusions: are they worth the risk of infiltration? *J Intraven Nurs.* 1997;20(2):95–99.
240. Subhani M, Sridhar S, DeCristofaro JD. Phentolamine use in a neonate for the prevention of dermal necrosis caused by dopamine: a case report. *J Perinatol.* 2001;21(5):324–326. doi:10.1038/sj.jp.7200493
241. Chen JL, O'Shea M. Extravasation injury associated with low dose dopamine. *Ann Pharmacother.* 1998;32(5):545–548.
242. Wong AF, McCulloch LM, Sola A. Treatment of peripheral tissue ischemia with topical nitroglycerin ointment in neonates. *J Pediatr.* 1992;121(6):980–983. doi:10.1016/s0022-3476(05)80356-7
243. Cardenas-Garcia J, Schaub KF, Belchikov YG, Narasimhan M, Koenig SJ, Mayo PH. Safety of peripheral intravenous administration of vasoactive medication. *J Hosp Med.* 2015;10(9):581–585. doi:10.1002/jhm.2394
244. Boltax RS, Dineen JP, Scarpa FJ. Gangrene resulting from infiltrated dopamine solution. *N Engl J Med.* 1977;296(14):823. doi:10.1056/nejm197704072961426
245. Denkler KA, Cohen BE. Reversal of dopamine extravasation injury with topical nitroglycerin ointment. *Plast Reconstr Surg.* 1989;84(5):811–813. doi:10.1097/00006534-198911000-00017
246. Bhosale GP, Shah VR. Extravasation injury due to dopamine infusion leading to dermal necrosis and gangrene. *J Anaesthesiol Clin Pharmacol.* 2012;28(4):534–535. doi:10.4103/0970-9185.101954
247. Driscoll C, Langer M, Burke S, El Metwally D. Improving detection of IV infiltrates in neonates. *BMJ Qual Improv Rep.* 2015;4(1). doi:10.1136/bmjquality.u204253.w3874
248. Ebels T, van der Heide JN. Dopamine-induced ischaemia. *Lancet.* 1977;2(8041):762. doi:10.1016/s0140-6736(77)90264-1
249. Yamamoto Y, Igawa H, Minakawa H, et al. Reconstruction of iatrogenic dorsal skin defects of hands with reverse forearm flaps. *Eur J Plast Surg.* 1994;17(2):104–105.
250. Phillips RA, Andrades P, Grant JH, Ray PD. Deep dopamine extravasation injury: a case report. *J Plast Reconstr Aesthet Surg.* 2009;62(7):e222–224. doi:10.1016/j.bjps.2008.11.064
251. Turner DA, Kleinman ME. The use of vasoactive agents via peripheral intravenous access during transport of critically ill infants and children. *Pediatr Emerg Care.* 2010;26(8):563–566. doi:10.1097/PEC.0b013e3181ea71e1
252. Nodwell T, Lalonde D. How long does it take phentolamine to reverse adrenaline-induced vasoconstriction in the finger and hand? A prospective, randomized, blinded study: The Dalhousie project experimental phase. *Can J Plast Surg.* 2003;11(4):187–190. doi:10.1177/229255030301100408
253. Jordan LK. An unusual case of digital ischaemia. *N C Med J.* 1969;30(10):418–419.
254. Singh T, Randhawa S, Khanna R. The EpiPen and the ischaemic finger. *Eur J Emerg Med.* 2007;14(4):222–223. doi:10.1097/MEJ.0b013e3280b17eec
255. Sinclair MD, Bailey MA, McAree BJ, Dewhurst D, Kent PJ. Images in vascular medicine: rapid epinephrine “reversal” with phentolamine following accidental autoinjector inoculation. *Vasc Med.* 2011;16(3):215–216. doi:10.1177/1358863X11404281
256. Velissariou I, Cottrell S, Berry K, Wilson B. Management of adrenaline (epinephrine) induced digital ischaemia in children after accidental injection from an EpiPen. *Emerg Med J.* 2004;21(3):387–388.
257. El Maraghy MW, El Maraghy AW, Evans HB. Digital adrenaline injection injuries: a case series and review. *Can J Plast Surg.* 1998;6(4):196–200.
258. Mrvos R, Anderson BD, Krenzelok EP. Accidental injection of epinephrine from an autoinjector: invasive treatment not always required. *South Med J.* 2002;95(3):318–320.
259. Markovchick V, Burkhart KK. The reversal of the ischaemic effects of epinephrine on the finger with local injections of phentolamine. *J Emerg Med.* 1991;9(5):323–324.
260. Hinterberger JW, Kintzi HE. Phentolamine reversal of epinephrine induced digital vasospasm. *Arch Fam Med.* 1994;3(2):193–195.
261. Turner MJ, Purushotham AD. Accidental EpiPen injection into a digit – the value of a Google search. *Ann R Coll Surg Engl.* 2004;86(3):218–219.
262. Deshmukh N, Tolland JT. Treatment of accidental epinephrine injection in a finger. *J Emerg Med.* 1989;7(4):408.
263. Ahearn MA. A pulseless hand: accidental epinephrine injection. *Am Fam Phys.* 1998;57(6):1238.
264. Khairalla E. Epinephrine-induced digital ischemia relieved by phentolamine. *Plast Reconstr Surg.* 2001;108(6):1831–1832. doi:10.1097/00006534-200111000-00084
265. Maguire WM, Reisdorff EG, Smith D, Wiegenstein JG. Epinephrine-induced vasospasm reversed by phentolamine digital block. *Am J Emerg Med.* 1990;8(1):46–47.
266. Hardy SJ, Agostini DE. Accidental epinephrine autoinjector-induced digital ischaemia reversed by phentolamine digital block. *J Am Osteopath Assoc.* 1995;95(6):377–378.
267. Claudy FR. Pertinent medical intelligence: accidental digital injection of epinephrine. *Md Med J.* 1995;44(4):292–293.
268. Kaspersen J, Vedsted P. Accidental injection of adrenaline in a finger with EpiPen [in Danish]. *Ugeskr Laeger.* 1998;160(45):6531–6532.

269. Sellens C, Morrison L. Accidental injection of epinephrine by a child: a unique approach to treatment. *Can J Emerg Med*. 1999;1(1):34–36.
270. McCauley WA, Gerace RV, Scilley C. Treatment of accidental digital injection of epinephrine. *Ann Emerg Med*. 1991;20(6):665–668.
271. Mol CJ, Gaver J. A 39 year old nurse with accidental discharge of epinephrine autoinjector into left index finger. *J Emerg Nurs*. 1992;18(4):306–307.
272. Ozcan A, Baratali E, Meral O, et al. Bullous dermatitis and skin necrosis developing after adrenalin extravasation. *Eurasian J Med*. 2015;47(3):226–228. doi:10.5152/eurasianjmed.2015.58
273. Fitzcharles-Bowe C, Denkler K, Lalonde D. Finger injection with high-dose (1:1,000) epinephrine: does it cause finger necrosis and should it be treated? *Hand (NY)*. 2007;2(1):5–11. doi:10.1007/s11552-006-9012-4
274. Barkhorvarian AR, Wakelin SH, Paes TR. Accidental digital injection of adrenaline from an autoinjector device. *Br J Dermatol*. 2000;143(6):1359.
275. Ruhlen JL. Tissue necrosis. Cutaneous and subcutaneous damage following extravasation of methylene blue. *J Kans Med Soc*. 1982;83(5):236, 260.
276. Dumbarton TC, Gorman SK, Minor S, Loubani O, White F, Green R. Local cutaneous necrosis secondary to a prolonged peripheral infusion of methylene blue in vasodilatory shock. *Ann Pharmacother*. 2012;46(3):e6. doi:10.1345/aph.1Q560
277. Khokhar RS, Aqil M, Al-Zahrani T, Gelidan A, Al Khayal K. Novel management of methylene blue extravasation: a case report and review of literature. *Saudi J Anaesth*. 2015;9(2):211–213. doi:10.4103/1658-354X.152891
278. Perry PM, Meinhard E. Nectotic subcutaneous abscesses following injections of methylene blue. *Br J Clin Pract*. 1974;28(8):289–291.
279. Kim YH, Cho SI. Skin flap necrosis by bone marking with methylene blue in cochlear implantation. *J Audiol Otol*. 2015;19(2):108–110. doi:10.7874/jao.2015.19.2.108
280. Lee JH, Chang CH, Park CH, Kim JK. Methylene blue dye-induced skin necrosis in immediate breast reconstruction: evaluation and management. *Arch Plast Surg*. 2014;41(3):258–263. doi: 10.5999/aps.2014.41.3.258
281. Bleicher RJ, Kloth DD, Robinson D, Axelrod P. Inflammatory cutaneous adverse effects of methylene blue dye injection for lymphatic mapping/sentinel lymphadenectomy. *J Surg Oncol*. 2009;99(6):356–360.
282. Zakaria S, Hoskin TL, Degnim AC. Safety and technical success of methylene blue dye for lymphatic mapping in breast cancer. *Am J Surg*. 2008;196(2):228–233.
283. Salhab M, Al Sarakbi W, Mokbel K. Skin and fat necrosis of the breast following methylene blue dye injection for sentinel node biopsy in a patient with breast cancer. *Int Semin Surg Oncol*. 2005;2:26.
284. Stradling B, Aranha G, Gabram S. Adverse skin lesions after methylene blue injections for sentinel lymph node localization. *Am J Surg*. 2002;184(4):350–352.
285. Phentolamine. Package insert. West-Ward Pharmaceuticals; 2015.
286. Close AS, Frackelton WH, Kory RC. Cutaneous necrosis due to norepinephrine: II—mechanism and prevention. *Ann Surg*. 1958;147(1):44–50. doi:10.1097/00000658-195801000-00007
287. Weeks PM. Ischemia of the hand secondary to levarterenol bitartrate extravasation: methods of management. *JAMA*. 1966;196(3):288–290.
288. Lewis T, Merchan C, Altshuler D, Papadopoulos J. Safety of the peripheral administration of vasopressor agents. *J Intensive Care Med*. 2019 Jan;34(1):26–33. doi:10.1177/0885066616686035
289. Peirce EC, Polley VB. Necrosis following intravenous use of neo-synephrine. *N Engl J Med*. 1954;250(3):114–115. doi:10.1056/NEJM195401212500305
290. Datar S, Gutierrez E, Schertz A, Vachharajani V. Safety of phenylephrine infusion through peripheral intravenous catheter in the neurological intensive care unit. *J Intensive Care Med*. 2018;33(10):589–592. doi:10.1177/0885066617712214
291. Delgado T, Wolfe B, Davis G, Ansari S. Safety of peripheral administration of phenylephrine in a neurologic intensive care unit: a pilot study. *J Crit Care*. 2016;34:107–110. doi: 10.1016/j.jccr.2016.04.004
292. Tran DQH, Finlayson RJ. Use of stellate ganglion block to salvage an ischemic hand caused by the extravasation of vasopressors. *Reg Anesth Pain Med*. 2005;30(4):405–408.
293. Greenwald RA, Rheingold OJ, Chiprut RO, Rogers AI. Local gangrene: a complication of peripheral Pitressin therapy for bleeding esophageal varices. *Gastroenterology*. 1978;74(4):744–746.
294. Mogan GR, Wormser GP, Gottfried EB. Infected gangrene: a serious complication of peripheral vasopressin administration. *Am J Gastroenterol*. 1980;73(5):426–429.
295. Wormser GP, Kornblee LV, Gottfried EB. Cutaneous necrosis following peripheral intravenous vasopressin therapy. *Cutis*. 1982;29(3):249–252.
296. Anderson JR, Johnston GW. Development of cutaneous gangrene during continuous peripheral infusion of vasopressin. *Br Med J (Clin Res Ed)*. 1983;287(6406):1657–1658. doi:10.1136/bmj.287.6406.1657
297. Kahn JM, Kress JP, Hall JB. Skin necrosis after extravasation of low-dose vasopressin administered for septic shock. *Crit Care Med*. 2002;30(8):1899–1901. doi:10.1097/00003246-200208000-00038
298. Harris PA, Bradley S, Moss AL. Limiting the damage of iatrogenic extravasation injury in neonates. *Plast Reconstr Surg*. 2001;107(3):893–894
299. Andrés AM, Burgos L, López Gutiérrez JC, et al. Treatment protocol for extravasation lesions [in Spanish]. *Cir Pediatr*. 2006;19(3):136–139.
300. Brown AS, Hoelzer DJ, Piercy SA. Skin necrosis from extravasation of intravenous fluids in children. *Plast Reconstr Surg*. 1979;64(2):145–150.
301. Wilkins CE, Emmerson AJ. Extravasation injuries on regional neonatal units. *Arch Dis Child Fetal Neonatal Ed*. 2004;89(3):F274–275.
302. Crowther DM, Buck ML, McCarthy MW, Barton VW. Improving pediatric adverse drug event reporting through clinical pharmacy services. *J Pediatr Pharmacol Ther*. 2011;16(4):285–290. doi:10.5863/1551-6776-16.4.285
303. Chenoweth KB, Guo J-W, Chan B. The extended dwell peripheral intravenous catheter is an alternative method of NICU intravenous access. *Adv Neonatal Care*. 2018;18(4):295–301. doi: 10.1097/ANC.0000000000000515
304. Amaya R. Use of active Leptospermum honey (ALH) and dehydrated amniotic membrane allograft (DAMA) to manage extravasation wounds in neonates. *J Wound Ostomy Continence Nurs* 2016;43(3):S25.
305. Hyaluronidase. Package insert. Halozyme Therapeutics Inc; 2012.
306. Hallman N, Kulonen E, Forsander O. On the use of hyaluronidase on hypodermoclysis in infants. *Acta Paediatrica*. 1950;39(1):94–101.
307. Haire RD Jr. Use of alidase in prevention of painful arm in accidental perivascular injection of neosarsphenamine and mapharsen. *Rocky Mt Med J*. 1950;47(8):600–601.
308. Britton RC, Habib DV. Clinical uses of hyaluronidase: a current review. *Surgery*. 1953;33(6):917–942.
309. Zimmert SE. The prevention of cutaneous necrosis following extravasation of hypertonic saline and sodium tetradecyl sulfate. *J Dermatol Surg Oncol*. 1993;19(7):641–646.
310. Laurie SW, Wilson KL, Kernahan DA, Bauer BS, Vistnes LM. Intravenous extravasation injuries: the effectiveness of hyaluronidase in their treatment. *Ann Plast Surg*. 1984;13(3):191–194.
311. Compañía P, Míguez JM, de Toro Santos FJ, et al. The use of antidotes for calcium gluconate extravasation: an experimental study in mice. *Plast Reconstr Surg*. 2018;142(3):699–707. doi: 10.1097/PRS.0000000000004640

312. Terbutaline sulfate. Package insert. Akorn, Inc.; 2011.
313. Yucha CB, Hastings-Tolsma M, Szeverenyi NM. Effect of elevation on intravenous extravasations. *J Intraven Nurs*. 1994;17(5):231–234.
314. Hastings-Tolsma MT, Yucha CB, Tompkins J, et al. Effect of warm and cold applications on the resolution of the I.V. infiltrations. *Res Nurs Health*. 1993;16(3):171–178.
315. Davé AL. Third-degree burn following use of microwave-heated cryogel pack. *Clin Pediatr* 1993;32(3):191–192. doi:10.1177/000992289303200318
316. Goldminz D, Barnhill R, McGuire J, Stenn KS. Calcinosis cutis following extravasation of calcium chloride. *Arch Dermatol*. 1988;124(6):922–925.
317. Schumacher HR, Osterman AL, Choi SJ, Weisz PB. Calcinosis at the site of leakage from extravasation of calcium disodium edetate intravenous chelator therapy in a child with lead poisoning. *Clin Orthop Relat Res*. 1987;219:221–225.
318. Hironaga M, Fujigaki T, Tanaka S. Cutaneous calcinosis in a neonate following extravasation of calcium gluconate. *J Am Acad Dermatol*. 1982;6(3):392–395.
319. Chiang MC, Chou YH, Wang CR, Huang CC. Extravasation of calcium gluconate concomitant with osteomyelitis in a neonate. *Acta Paediatr Taiwan*. 2004;45(1):35–37.
320. Chen TK, Yang CY, Chen SJ. Calcinosis cutis complicated by compartment syndrome following extravasation of calcium gluconate in a neonate: a case report. *Pediatr Neonatol*. 2010;51(4):238–241. doi:10.1016/S1875-9572(10)60045-9
321. Chinn M, Colella MR. Prehospital dextrose extravasation causing forearm compartment syndrome: a case report. *Prehosp Emerg Care*. 2017;21(1):79–82. doi:10.1080/10903127.2016.1209263
322. DeLorenzo RA, Vista JP. Another hazard of hypertonic dextrose. *Am J Emerg Med*. 1994;12(2):262–263.
323. Kasdan ML, June LA. Extravasation of phenytoin and diazepam requiring surgical debridement and skin grafting. *Orthopedics*. 1993;16(12):1355–1357.
324. Kumar MM, Sprung J. The use of hyaluronidase to treat mannitol extravasation. *Anesth Analg*. 2003;97(4):1199–1200.
325. Sacks GS, Mir TL, Lee M. Skin necrosis induced by extravasation of glycerol-containing peripheral parenteral nutrition formulation. *J Miss State Med Assoc*. 1999;40(9):307–311.
326. Handler EG. Superficial compartment syndrome of the foot after infiltration of an intravenous fluid. *Arch Phys Med Rehabil*. 1990;71(1):58–59.
327. Lau B, Lee NH. Acyclovir extravasation: case report and review of the literature. *Austin J Orthopade Rheumatol*. 2016;3(1):1026.
328. Simoni P, Scatciolla L, Maréchal C, Mustapha SB, Zobel BB. Cordarone extravasation inducing volkmann's like syndrome. *Cardio Res*. 2011;2(6):307–309.
329. Baker GL, Franklin JD. Management of arginine monohydrochloride extravasation in the forearm. *South Med J*. 1991;84(3):381–384. doi:10.1097/00007611-199103000-00018
330. Abraham MB, van der Westhuyzen J, Khanna V. Arginine extravasation leading to skin necrosis. *J Paediatr Child Health*. 2012;48(3):E96–E97. doi:10.1111/j.1440-1754.2011.02074
331. Hoff JV, Beatty PA, Wade JL. Dermal necrosis from dobutamine. *N Engl J Med*. 1979;300(22):1280.
332. Hardie GH, Hunter DC. Skin necrosis with intravenous use of norepinephrine. *Med Bull (Ann Arbor)*. 1955;21(7):213–219.
333. Davidson DC, Gilbert J. Severe extravasation injury. *Br Med J (Clin Res Ed)*. 1985;291(6489):217.
334. Hogue RJ Jr. Skin necrosis from extravasation of intravenous fluids in children. *Plast Reconstr Surg*. 1980;65(2):244.
335. Ahn SK, Kim KT, Lee SH, et al. The efficacy of treatment with triamcinolone acetonide in calcinosis cutis following extravasation of calcium gluconate: a preliminary study. *Pediatr Dermatol*. 1997;14(2):103–109.
336. Dougherty L, Oakley C. Advanced practice in management of extravasation. *Cancer Nurs Pract*. 2011;10(5):16–22.
337. Wissing FR, Ryu SM, Menke H. Scar contracture with hyperextension in the wrist due to extravasation in the childhood: two-stage scar correction with the dermal regeneration template (Integra®) and skin graft [article in German]. *Handchir Mikrochir Plast Chir*. 2017;49(5):337–340. doi:10.1055/s-0043-119690
338. Cisler-Cahill L. A protocol for the use of amorphous hydrogel to support wound healing in neonatal patients: an adjunct to nursing skin care. *Neonatal Netw*. 2006;25(4):267–273.
339. Thomas S, Rowe H, Keats J, Morgan R. The management of extravasation injury in neonates. *World Wide Wounds*. 1997:7p.
340. Mohr LD, Reyna R, Amaya R. Neonatal case studies using active Leptospermum honey. *J Wound Ostomy Continence Nurs*. 2014;41(3):213–218. doi:10.1097/WON.0000000000000028
341. Gibboney W, Lemons JA. Necrotizing fasciitis of the scalp in neonates. *Am J Perinatol*. 1986;3(1):58–60. doi:10.1055/s-2007-999829
342. Sindal MD, Nakhwa CP. Metastatic serratia endophthalmitis associated with extravasation injury in a preterm neonate. *Oman J Ophthalmol*. 2015;8(2):114–116. doi:10.4103/0974-620X.159261
343. Santoshi JA, Pallapati SC, Thomas BP. Hand contracture: an unusual sequel of intravenous fluid extravasation in the neonatal period. *J Postgrad Med*. 2008;54(4):244–245.
344. Nissim L, Gilbertson-Dahdal D. An unusual complication of an infiltrated intravenous catheter: heterotopic ossification in a newborn. *J Radiol Case Rep*. 2008;2(2):13–15. doi:10.3941/jrcr.v2i2.30
345. Koeppel R. Wound care after peripheral intravenous extravasation: what is the evidence? *Newborn Infant Nurs Rev*. 2006;6(4):202–211.