

I am not robot!

Life cycle of maize weevil pdf

View PDFVolume 8, Issue 12, December 2022, e11859Author links open overlay panel, rights and contentUnder a Creative Commons licenseopen accessBotanicalMaizePest managementStorageWeevils[©] 2022 The Author(s). Published by Elsevier Ltd. Advances in Agriculture/2016/Article/Research Article | Open AccessVolume 2016 | Article ID 7836379 | Adebayo Ojo1 and Adebayo Amos Omoloye2Academic Editor: Kassim Al-KhatibThe maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae), is one of the most destructive pests of stored cereals. Knowledge of the life history and biology is important to the development of an integrated pest management (arva and pupa) and adult stages varied significantly among the cereal grains, mean fecundity was highest on maize () and lowest on millet (). Number of immature (larva and pupa) and adult stages varied significantly among the cereal grains. There exist four larval instar development and head capsule width, with a mean total instar larval development of grains, cereals, and other processed and unprocessed stored products in sub-Saharan Africa [-4]. S. zeamais causes qualitative and quantitative damage to stored products, with uncited and products of rais. Causes qualitative and quantitative damage to stored products, which include storage structures and physical and chemical properties of the produce. Heavy infestation of adults and larva of maize weevil, which cause postharvest losses have become increasingly important constraints to storage entomology [8] and food security in the tropics. The common control methods for this pest of maize or method stored cereals in other to elucidate here eleopmental biology of predeomethoral biology of predot stored cereals in other to elucidate here weevil on maize of adults and larvae of maize everity of maize everity of maize everity of maize everity in the tropics. The common control methods for this pest are the use of chemical insecticides, biological control, and betanical properties of these biology of predot substrate [1, 16].



The mean developmental period from egg to adult varied, being highest on maize (34.7 d) and lowest on sorghum (33.5 d).1. IntroductionThe maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae), is one of the most destructive stored products in sub-Saharan Africa [1-4]. S. zeamais causes qualitative and quantitative damage to stored products, with grain weight loss ranging between 20 to 90% for untreated stored maize [5-7], and the severity of damage depends on factors which include storage structures and physical and chemical properties of the produce. Heavy infestation of adults and larvae of maize weevil which cause postharvest losses have become increasingly important constraints to storage entomology [8] and food security in the tropics. The common control methods for this period ecology and pest monitoring and management without a better understanding of the phenology on preferred food substrate [1, 16]. Therefore, this study seeks to investigate the developmental biology of S. zeamais under laboratory conditions on four main stored cereals in other to elucidate some important aspects of its life history.2. Material and MethodsA culture was established of maize week-lod S. zeamais, using a modified method as described by Ojo and Omoloye [17] that were first collected from cultures in the Entomology Research Laboratory, Department of Crop Protection and Environmental Biology, University of Ibadan, Twenty five pairs of one-week-lod S. zeamais were introduced into 100 g grains of maize in 4.5 kg capacity Kilher jars covered with mesh lids, replicated five times (2013), each of maize (var. TIA 306), sorghum (var. samsorg 17), and mille (local witer was using morphological control was observed adult was excluded from the period the study was conducted in Kilner jars with adult S. zeamais were kept under ambient temperature of 25-28°C, 60-70% relative humidity, and 12-hour photophase. A daily examination and dissection of the infested grains started on eighth day following weevil removal.

Grain was carefully removed to allow for study of the grains for eggs and larval development of S. zeamais. Acid fuchsin and 250 mL glacial acetic acid to 750 mL of distilled water. The staining of egg plugs was determined following procedures of Pedersen [19].



There was linear relationship and significant correlation between the stages of larval development and head capsule width. The mean development and head capsule with with was capsule and opended by for untreated stored products in sub-Saharan Africa [1-4]. S. zeamais causes qualitative and quantitative and quantitative



Knowledge of the life history and biology is important to the development program. Investigation was carried out on developmental biology of S. zeamais on four main cereal crops, maize, orghum, and millet, under laboratory conditions. Egg incubation, oviposition periods, and larval instar development were not different significantly among the cereal grains; mean fecundity was highest on maize () and lowest on millet, under laboratory conditions. There exist four larval developmental period of 23.1, 22.2, 22.2, and 21.6 d on maize, rice, sorghum, and millet, respectively. There was linear relationship and significant correlation between the stages of larval development and head capsule width. The mean developmental period from large to 400% for large depends on maize (34.7 d) and lowest on sorghum (33.5 d).1. IntroductionThe maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae), is one of the most destructive stored products in sub-Saharan Africa [1–4]. S. zeamais causes equalitative and quantitative and quantitative and quantitative and purporties to to 90% for untreated stored maize [5–7], and the severity of damage depends on factors which include storage entructures and physical and chemical properties of the products in sub-Saharan Africa [1–4]. S. zeamais cause postharvative and quantitative and quantitative and quantitative and quantitative and quantitative and maize (see of the products, sub-scalar and file curcles (see of the products, biological control, and botanical insecticides [9–11] among others. There cannot be a realistic success in applied ecology and pest monitoring and management without a better understanding of the phenology and dynamics of insects' life cycle [12]. Several studies have been conducted on the reproductive biology of maize weevil on maize or modified maize developmental biology of S. zeamais under laboratory conditions on downey the search laboratory. Department of Crop Protection and Environmental Biology, University of Ibadan, Ibadan. Twenty five pairs of



Number of eggs laid varied significantly among the cereal grains; mean fecundity was highest on maize () and lowest on millet (). Number of immature (larva and pupa) and adult stages varied significantly among the cereal grains. There exist four larval instars with a varied mean head capsule width, with a mean total instar larval developmental period of 23.1, 22.2, 22.2, and 21.6 d on maize, rice, sorghum, and millet, respectively. There was linear relationship and significant correlation between the stages of larval development and head capsule width. The mean developmental period from egg to adult varied, being highest on maize (34.7 d) and lowest on sorghum (33.5 d).1. IntroductionThe maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae), is one of the most destructive stored product pests of grains, cereals, and other processed and unprocessed stored products in sub-Saharan Africa [1-4]. S. zeamais causes qualitative and quantitative damage to stored products, with grain weight loss ranging between 20 to 90% for untreated stored maize [5-7], and the severity of damage depends on factors which include storage structures and physical and chemical properties of the produce. Heavy infestation of adults and larvae of maize weevil which cause postharvest losses have become increasingly important constraints to storage entomology [8] and food security in the tropics. The common control methods for this pest are the use of chemical insecticides, biological control, and botanical insecticides [9-11] among others.

There cannot be a realistic success in applied ecology and pest monitoring and management without a better understanding of the phenology and dynamics of insects' life cycle [12]. Several studies have been conducted on the reproductive biology of maize weevil on maize or modified maize diet [13-15]. However, there is a paucity of information on developmental biology of S. zeamais under laboratory conditions on four main stored cereals in other to elucidate some important aspects of its life history.2. Material and MethodsA culture was established of maize weevils, S. zeamais, using a modified method as described by Ojo and Omoloye [17] that were first collected from cultures in the Entomology Research Laboratory, Department of Crop Protection and Environmental Biology, University of Ibadan, Ibadan. Twenty five pairs of one-week-old S. zeamais were introduced into 100 g grains of maize in 4.5 kg capacity Kilner jars covered with mesh lids, replicated five times (). Weevils were allowed to feed, mate, and oviposit for 7 days and then removed. Culture arenas were observed daily until new progenies emerged; they were removed and sexed using morphological characters described by Halstead [18]. This stock culture was used as source of Sitophilus zeamais throughout the period this study was conducted in 2013.Presterilized samples (200 g), each of maize (var. TZPB-SW-R), rice (var. ITA 306), sorghum (var. samsorg 17), and millet (local variety), were weighed and placed in Kilner jars with mesh lids. To each jar containing grain, 200 unsexed adult S. zeamais were added from the laboratory culture for a total of five replicates per grain type (). Grain jars with adult S. zeamais were kept under ambient temperature of 25-28°C, 60-70% relative humidity, and 12-hour photophase.

A daily examination and dissection of the infested grains started on eighth day following weevil removal. Grain was carefully removed to allow for study of the grains for eggs and larval development of S. zeamais. Acid fuchs in stain was used by adding 3.5 g acid fuchs in and 250 mL grains were first stained with acid fuchs in solution to locate and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were first stained with acid fuchs in solution to locate and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected dissected and study the egg plug on individual cereal grain were dissected and study the egg plug on individual cereal grain and to track egg maturation. A total of 20-30 infested grains were dissected and study the egg plug on individual cereal grains were dissected and study and longevity, five pairs (1) of Scamal second track egg maturation and egg slaid were determined following standard procedures as described earlier and adult longevity was determined the atomotidity. M



The mean developmental period from egg to adult varied, being highest on maize (34.7 d) and lowest on sorghum (33.5 d).1. IntroductionThe maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae), is one of the most destructive stored product pests of grains, cereals, and other processed and unprocessed stored products in sub-Saharan Africa [1-4].

S. zeamais causes qualitative and quantitative damage to stored products, with grain weight loss ranging between 20 to 90% for untreated stored maize [5-7], and the severity of damage depends on factors which include storage structures and physical and chemical properties of the produce. Heavy infestation of adults and larvae of maize weevil which cause postharvest losses have become increasingly important constraints to storage entomology [8] and food security in the tropics. The common control methods for this pest are the use of chemical insecticides, biological control, and botanical insecticides [9-11] among others. There cannot be a realistic success in applied ecology and pest monitoring and management without a better understanding of the phenology and dynamics of insects' life cycle [12]. Several studies have been conducted on the reproductive biology of maize weevil on maize or modified maize diet [13-15]. However, there is a paucity of information on developmental biology on preferred food substrate [1, 16]. Therefore, this study seeks to investigate the developmental biology of S. zeamais under laboratory conditions on four main stored cereals in other to elucidate some important aspects of its life history.2. Material and MethodsA culture was established of maize weevils, S. zeamais, using a modified method as described by Ojo and Omoloye [17] that were first collected from cultures in the Entomology Research Laboratory, Department of Crop Protection and Environmental Biology, University of Ibadan, Ibadan. Twenty five pairs of one-week-old S. zeamais were introduced into 100 g grains of maize in 4.5 kg capacity Kilner jars covered with mesh lids, replicated five times (). Weevils were allowed to feed, mate, and oviposit for 7 days and then removed. Culture arenas were observed daily until new progenies emerged; they were removed and sexed using morphological characters described by Halstead [18].

This stock culture was used as source of Sitophilus zeamais throughout the period this study was conducted in 2013.Presterilized samples (200 g), each of maize (var. TZPB-SW-R), rice (var. TZPB-SW-R), rice

S. zeamais has seven life stages comprising egg (), four larval instars (), prepupa/pupa (), and adult (). No significant difference (> 0.05, F = 0.56, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; > 0.05, F = 0.17, and DF = 3; and 7 d; the lowest mean oviposition period ranged between 3 and 7 d; the lowest mean was recorded on rice with 5.1 d and the highest was observed on millet with 5.4 d. Total average number of eggs laid was not significantly varied (> 0.05, F = 1.22, and DF = 3) among the cereal grains, with the highest and lowest mean fecundity being found on maize () and millet (), respectively. Also, there was significant different (< 0.05, F = 3.99, and DF = 3) adult longevity found among the cereals used; adult maize weevil significantly lived longest on maize and millet (and d) than on rice and sorghum (and .07 days), respectively (Table 1). Adult S, zeamais burrows into cereal grains with the aid of its strong rostrum, creating a cavity into which it lays a single egg. The eggs are oval in shape, whitish in colour, and rounded at the bottom. Female S, zeamais will then cover the egg with a gelatinous egg plug which she deposits. Egg plugs were, on average, mm wide and mm long.

Eggs hatch into a creamy white apodous larva with a sclerotized light brown head. The immature stages of maize weevil varied in relation to the grains on which it is cultured. The developmental period and body measurements of first instar larvae were not significantly different (> 0.05, F = 1.45, and DF = 3; > 0.05, F = 1.73) from one another regardless of the cereal grains tested. A significant difference (< 0.05, F = 1.45, and DF = 3) was observed from third instar stage, there were significant (< 0.05, F = 1.99, and DF = 3) shortest developmental days on sorghum, rice, and millet (5.5 and 5.7 days), the longest developmental period recorded being on maize (6.5 days). The newly hatched 1st instar larvae (0.54 mm long) remained inside the grain, feeding voraciously until the end of the 4th instar stage and shortest on milet (5.3 days). There was significantly (< 0.05, F = 4.4) longest on maize at 4th instar stage and shortest on rice and sorghum (4.7 days). There was significantly (< 0.05, F = 10.7; < 0.05, F = 9.17] longer between immature weevils on maize, rice, and sorghum (mm, mm, and mm), respectively, but significantly longer than weevil body length on millet, long the food host used; second instar larva levelopmental period of 5 days on maize, rice, millet, and sorghum, respectively. The mean dev

Food host also influenced the body measurement of maize weevil pupa, with longest body length and width being recorded on maize (mm and mm). The food hosts significantly influenced emerging body measurements of adult maize weevil was significantly influenced emerging body measurements of adult maize weevil was significantly influenced emerging body measurements of adult maize weevil was significantly influenced emerging body measurements of adult maize weevil was significantly influenced emerging body measurements of adult maize weevil was significantly influenced emerging body measurements of adult maize weevil was significantly (< 0.05, F = 1.73) bigger on maize (mm long and mm wide) although not bigger than its male counterpart (Table 2). The measurement of head capsule width of larval instars daily showed four frequency peaks as confirmed by Dyar's rule representing four larval instars (Table 3 and Figure 2). The head capsule width was regular, and perfect geometric larval instar stages ranging from 1.2 to 1.5, with the mean growth ratio of 1.3. The relationship between the larval developmental period and mean head capsule width was regular, and perfect geometric larval geometric larval instar across the food hosts (Figure 1). Linear regression analysis depicted significant relationship between larval instar stage (Table 3 and Figure 1). Linear regression analysis depicted significant relationship between larval instar stage (Table 3 and Figure 1). Linear regression analysis depicted significant relationship between larval instar stage (Table 3) (b) (c) (d) (a) (b)

The developmental biology of S. zeamais could be influenced by this moderate fecundity and oviposition, shorter larval period, and ability to breed easily on any cereal crop.Regardless of the food hosts, this study showed that there were four larval instars of S. zeamais when the larval head capsule width was measured with successive instars and frequency distribution of head capsule multimodal curves which show four modal peaks. There was a distinct difference between the values of head capsule width for the successive larval instars. Body measurement is usually assumed to be normally distributed from insect of the same morphogenetic stage [24, 25]; this supports four peaks observed in the study.

The ratio varied from 1.2 to 1.5 across the four food hosts with a mean growth ratio of 1.3 on all the cereal crops used; these values showed that growth progresses at a constant rate in each molt which is relatively close to the constant Dyar's ratio of 1.4 for lepidopterous insects although it has been adopted for other insect orders [26, 27]; the slightest difference observed could be because S. zeamais is a coleopteran. Nevertheless, mean growth ratios obtained in this study were similar to what was obtained in other curculionids: 1.4 for Conotrachelus psidii [23]; 1.3 for Dendroctonus valens [28]; and 1.3 for Sitophilus linearis [17]. A linear relationship obtained between head capsule width and a high regression coefficient of 0.92 (on maize), 0.94 (on rice), 0.94 (on sorghum), and 0.98 (on millet) attested to the fact that no stadium was overlooked. The total larval developmental period was 23.1 days (on maize), 22.2 days (on rice), 22.2 days (on sorghum), and 21.6 days (on millet), and a prepupa/pupa developmental period was six to seven days which was different from what was obtained in other curculionids: 16 days for Conotrachelus psidii [23], 9.5 days for Hypothenemus hampei [29], and 14 to 16 days for Sitophilus linearis [17]. The prepupa/pupa period of 6.3 to 6.7 days observed across the food hosts used is relatively similar to what was obtained in other curculionids: 10 days for D. valens [28], 8 days for S. linearis [17], and 7.95 days for H. hampei [29]. Factors like food host conditions, type of insect species, geographical locations, and experimental conditions could play significant role in the developmental study of maize weevil. This study also showed that the comparative biological cycle of S. zeamais from egg to adult was 34.7 days (on rice), 34.1 days (on sorghum), and 33.5 days (on millet); this was similar to the S. zeamais mean developmental period obtained by other researchers using this insect species: 31-37 days [2], and 34.8 days [2], and 34.8 days [4].

The significant variation in sizes between adult S. zeamais could be a result of the type of food crops used; also female maize weevil was observed to be bigger than their male counterpart regardless of the food host. S. zeamais bred on maize was bigger (4.11 mm long for male and 4.18 mm long for female) than those bred on other cereals, with a relative smallest body size recorded on both rice (3.34 mm long for female) and millet (3.45 mm long for female). Therefore, kind of food host coupled with the prevailing environmental condition played a significant role in maize weevil body size, as basic nutrients influenced the metabolic activities in insect.

The results obtained in this study on S. zeamais comparative phenology and dynamics of its life cycle on different main cereals provide a base for researcher for proper understanding of maize weevil ontogeny and bioecology which are needed in formulation of sustainable pest management practices and approach, as maize has been field to store pest of economic importance attacking different kinds of processed and unprocessed crop products worldwide.Competing Interests regarding the publication of this article.AcknowledgmentsThe authors wish to acknowledge Adeniyi Aderinokun and Emmanuel Nnodim for their technical assistance in the laboratory. J. E. Throne, "Life history of immature maize weevils (Coleoptera: Curculionidae) on corn stored at constant temperatures and relative humidities in the laboratory," Environmental Entomology, vol. 23, no. 6, pp. 1459-1471, 1994.View at: Publisher Site | Google ScholarD. P. Rees, Insects of Stored Products, CSIRO Publishing, 2004.R. Plarre, "An attempt to reconstruct the natural and cultural history of the granary weevil, Sitophilus granarius (Coleoptera: Curculionidae)," European Journal of Entomology, vol. 107, no. 1, pp. 1-11, 2010.View at: Publisher Site | Google ScholarD. P. Giga, S. Mutemerewa, G. Moyo, and D. Neeley, "Assessment and control of losses caused by insect pests in small farmers' stores in Zimbabwe," Crop Protection, vol. 10, no. 4, pp. 287-292, 1991.View at: Publisher Site | Google ScholarS. Muzemu, J. Chitamba, and B. Mutetwa, "Evaluation of Eucalopapa as stored maize grain protectants against Sitophilus zeamais (Motsch.) (Coleoptera: Curculionidae)," Agriculture, Forestry and Fiscov, vol. 2, no. 5, pp. 196-201, 2013.View at: Publisher Site | Google ScholarE. N. Nukenine, B. Monglo, L. Awason, L. S. T. Ngamo, F. F. N. Tchuengue, and M. B. Ngasoum, "Farmer's percepted on some aspects of maize production, and infestation levels of stored maize without," Campital and carica papaya as in the Ngaoundere region of Cameroon," Cam, J. Biol.

Biochem. Sci, vol. 12, no. 1, pp. 18–30, 2002.View at: Google ScholarR. H. Markham, N. A. Bosque-Perez, C. Borgemeister, and W. Meikle, "Developing pest management strategies for Sitophilus zeamais and Prostephanus truncatus in the tropics," FAO Plant Protection Bulletin, vol. 42, no. 3, pp. 97–116, 1994.View at: Google ScholarC. A. Akob and F. K. Ewete, "The efficacy of ashes of four locally used plant materials against Sitophilus zeamais (Coleoptera: Curculionidae) in Cameroon," International Journal of Tropical Insect Science, vol. 27, no. 1, pp. 21–26, 2007.View at: Publisher Site | Google ScholarT. W. Phillips and J. E. Throne, "Biorational approaches to managing stored-product insects," Annual Review of Entomology, vol. 55, pp. 375–397, 2010.View at: Publisher Site | Google ScholarA. Holzmann, "Latest developments in the registration of SPP chemicals in Germany and European Symposium. Stored Product Protection. 'Stress on chemical products'," Julius-Kühn-Archiv, vol. 429, pp. 89–93, 2010.View at: Google ScholarA. Merville, A. Vallier, S. Venner et al., "Determining the instar of a weevil larva (Coleoptera: Curculionidae) using a parsimonious method," European Journal of Entomology, vol. 111, no.

4, pp. 567-573, 2014.View at: Publisher Site | Google ScholarR. T. Arbogast, "Beetles: coleopteran," in Ecology and Management of Food Industry Pests, J. R. B. Gorham, Ed., pp. 133-150, Association of Official Analytical Chemists, Arlinghton, Va, USA, 1991.View at: Google ScholarC. P. Haines, Insects and Arachnids of Tropical Stored-Products: Their Biology and Identification, Natural Resources Institute, Gillingham, UK, 2nd edition, 1991.M. Danho, C. Gaspar, and E. Haubruge, "The impact of grain quantity on the biology of Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae): Oviposition, distribution of eggs, adult emergence, body weight and sex ratio," Journal of Stored Products Research, vol. 38, no. 3, pp. 259-266, 2002.View at: Publisher Site | Google ScholarR. W. Howe, "The biology of the rice weevil, Calandra oryzae (L.)," Annals of Applied Biology, vol. 39, no. 2, pp. 168-180, 1952.View at: Publisher Site | Google ScholarJ. A. Ojo and A.

A. Omoloye, "Life history of the tamarind weevil, Sitophilus linearis (Herbst) (Coleoptera: Curculionidae), on tamarind seed," Journal of Insects, vol. 2015, Article ID 429579, 5 pages, 2015. View at: Publisher Site | Google ScholarD. G. Halstead, "External sex differences in stored-products Coleoptera," Bulletin of Entomological Research, vol. 54, no. 1, pp. 119-134, 1963. View at: Publisher Site | Google ScholarJ.

R. Pedersen, Selection of oviposition sites on wheat kernels by Sitophilus spp.: effect of moisture, temperature and kernel size [Ph.D. thesis], Kansas State University, Manhattan, Kan, USA, 1979.S. Sharifi and R. B. Mills, "Radiographic studies of Sitophilus zeamais (Motsch.) in wheat grains," Journal of Stored Products Research, vol. 7, pp. 195-206, 1979.View at: Google ScholarV. Choubey, R. Bhandari, and N.

Kulkarni, "Life history and morphology of seed weevil, Sitophilus rugicollis Casey (Coleoptera: Curculionidae) infesting sal seeds in Madhya Pradesh," Journal of Entomological Research, vol. 37, no. 3, pp. 259-267, 2013. View at: Google ScholarK. C. Narayana, G. P. Swamy, E. Mutthuraju, E. Jagadeesh, and G.

T. Thirumalaraju, "Biology of Sitophilus oryzae (L.) (Coleoptera: Curculionidae) on stored maize grains," Current Biotica, vol. 8, no. 1, pp. 76-81, 2014. View at: Google ScholarO. E.

Bailez, A. M. Viana-Bailez, J. O. G. de Lima, and D. D.

O. Moreira, "Life-history of the guava weevil, Conotrachelus psidii Marshall (Coleoptera: Curculionidae), under laboratory conditions," Neotropical Entomology, vol. 32, no.

2, pp.

203-207, 2003.View at: Publisher Site | Google ScholarJ. A. Logan, B. J.

Bentz, J.

C. Vandygriff, and D. L. Turner, "General program for determining instar distributions from headcapsule widths: example analysis of mountain pine beetle (Coleoptera: Scolytide) data," Environmental Entomology, vol. 27, no. 3, pp. 555–563, 1998. View at: Publisher Site | Google ScholarG. Hunt and R. E. Chapman, "Evaluating hypotheses of instargrouping in arthropods: a maximum likelihood approach," Paleobiology, vol. 27, no. 3, pp. 466–484, 2001. View at: Publisher Site | Google ScholarH. G. Dyar, "The number of moults of lepidopterous larvae," Psyche, vol. 5, no. 175-176, pp. 420–422, 1890. View at: Publisher Site | Google ScholarR.

L. Taylor, "On 'Dyar's rule' and its application to sawfly larvae," Annals of the Entomological Society of America, vol. 24, no. 3, pp. 451-466, 1931. View at: Publisher Site | Google ScholarZ. Liu, B. Xu, and J. Sun, "Instar numbers, development, flight period, and fecundity of Dendroctonus valens (Coleoptera: Curculionidae: Scolytinae) in China," Annals of the Entomological Society of America, vol. 107, no.

1, pp. 152-157, 2014.View at: Publisher Site | Google ScholarJ. Gómez, B.

Y. Chávez, A. Castillo, F. J. Valle, and F. E. Vega, "The coffee berry borer (Coleoptera: Curculionidae): how many instars are there?" Annals of the Entomological Society of America, vol. 108, no. 3, pp. 311-315, 2015. View at: Publisher Site | Google ScholarCopyright © 2016 James Adebayo Ojo and Adebayo Amos Omoloye. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.