

Non-Enzymatic Colorimetric Assay for Nitric Oxide

Catalog Number: A22070 (AOXRE Catalog Number 22070)
For Research Use Only

INTRODUCTION

This kit allows you to measure total nitric oxide (NO) produced for *in vitro* experimental systems

following conversion of nitrate to nitrite by metallic cadmium.

Nitric oxide can be spectrophotometrically assayed by measuring the accumulation of its stable

degradation products, nitrate and nitrite. The ratio of these two products in biological fluids, tissue culture media, etc. may vary substantially. Hence, for accurate assessment of the total nitric oxide generated, one must monitor both nitrate and nitrite. An excellent solution to this problem involves the conversion of nitrate to nitrite catalyzed by cadmium. This is then followed by the quantitation of nitrite using the Griess reaction.

This kit is suitable for the quantitative determination of total nitric oxide in samples that have high protein concentrations. The cadmium catalyst proves far more robust when in the presence of harsh deproteinating reagents than the alternative, nitrate reductase.

PRINCIPLES OF PROCEDURE

In aqueous solution, nitric oxide rapidly degrades to nitrate and nitrite. Spectrophotometric quantitation of nitrite using Griess Reagent is straightforward, but does not measure nitrate. This kit employs metallic cadmium for quantitative conversion of nitrate to nitrite (Figure 1) prior to quantitation of nitrate using Griess reagent — thus providing for accurate determination of total NO production.

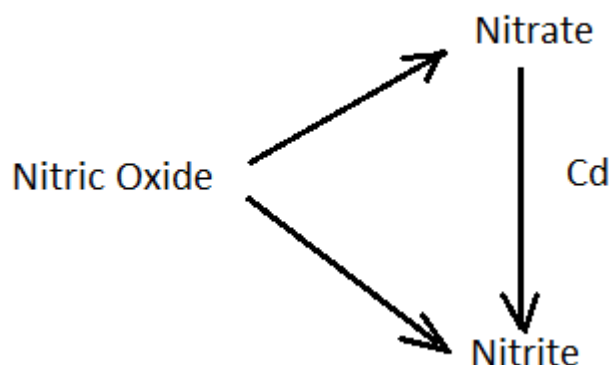


Figure 1: The reduction of nitrate to nitrite

This kit can be used to accurately measure as little as 1 pmol/mL (~1µM) NO produced in aqueous

solutions. Very little sample is required (1 to 100µL depending on the [NO] in the sample). The

completed reaction is read at 540 nm.

MATERIALS PROVIDED

Component	Contents	Quantity	Storage
Cadmium Beads	Granulated Cadmium (Cd ⁺⁺) for Nitrate reduction	25 g	4°C
ZnSO ₄ Solution	30% (w/v) solution of Zinc Sulfate for deproteinization	2 mL	4°C
Microcentrifuge Tubes	1.5 mL microcentrifuge tubes for sample incubation	50 tubes	4°C
Color Reagent #1	Sulfanilamide (<i>p</i> -Aminobenzenesulfonamide) in 3N HCl	7 mL	4°C
Color Reagent #2	N-(1-Naphthyl) ethylenediamine dihydrochloride in H ₂ O	7 mL	4°C
Nitrite Standard	500 pmol/μL NaNO ₂ (equivalent to 500 μM NO)	1.5 mL	4°C
Microplate	96-well low-binding flat-bottom microplate	1 plate	4°C
0.1 M HCl	Acidic solution used to wash Cadmium Beads	125 mL	4°C
0.1 M NH ₄ OH	Basic solution used to wash Cadmium Beads	125 mL	4°C

MATERIALS NEEDED BUT NOT PROVIDED

1. Adjustable pipettes (10-1,000 μL) and disposable tips
2. Microcentrifuge tubes
3. Microplate reader with 540 nm filter
4. Vortex mixer
5. Deionized water

STORAGE CONDITIONS

1. The Cadmium Beads should be stored in a dedicated dry environment.
2. Store this kit and its components at 4°C until use.
3. Do not freeze.

WARNINGS AND PRECAUTIONS

1. Use aseptic technique when opening and dispensing reagents.
2. This kit is designed to work properly as provided and instructed. Additions, deletions, or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.
3. Exercise universal precautions during the performance or handling of this kit or any component contained therein.

PROCEDURAL NOTES

1. Reagents can be used immediately upon removal from refrigeration.
2. Performance of the entire kit at once is not required. When performing this kit in part, please adhere to the following:
 - All unused components should be returned to their respective storage areas.
 - Create a standard curve for each performance of the assay. The prepared Standards may be stored at 4°C for up to 24 hours. Sufficient stock solution of standard is provided for two standard curves.
3. To minimize error due to handling, wipe the exterior bottom of the microplate wells with a lint-free paper towel.
4. Take note of the volume of each sample at the various stages of preparation and assay procedure to ensure proper dilution factor and NO concentration calculation.

REAGENT PREPARATION

1. **Cadmium Beads:** Cadmium Beads should be washed following the procedure below prior to use. Cadmium Beads may be prepared days prior to its intended use and should be stored in a sealed container under inert gas such as nitrogen or argon.
 - a. Place the desired amount of Cd⁺⁺ in an appropriately sized, sealable container. Please consider

that for every 0.5 g of Cd⁺⁺ there will be 1.0 mL of wash solution applied.

b. Apply 1.0 mL of Deionized Water for every 0.5 g of Cd⁺⁺ to be used. Seal container and mix by inversion.

c. Decant the remaining solution and repeat Step b one time. Remove residual amounts of solution

with a probing pipette or by dabbing with a lint-free paper towel.

d. Apply 1.0 mL of 0.1 M HCl Solution for every 0.5 g of Cd⁺⁺ to be used. Seal container and mix by inversion.

e. Decant the remaining solution and repeat Step d one time. Remove residual amounts of solution

with a probing pipette or by dabbing with a lint-free paper towel.

f. Apply 1.0 mL of 0.1 M NH₄OH Solution for every 0.5 g of Cd⁺⁺ to be used. Seal container and mix by inversion.

g. Decant the remaining solution and repeat Step f one time. Remove residual amounts of solution

with a probing pipette or by dabbing with a lint-free paper towel.

h. The washed and dried Cd⁺⁺ is now ready for use. If you are not using the washed Cd⁺⁺ immediately, it should be stored under inert gas such as nitrogen or argon in a sealed container.

SAMPLE PREPARATION

This kit is designed for use with samples possessing high concentrations of protein such as serum, culture medium or tissue homogenates. Each sample should be diluted according to its presence of NO metabolites (nitrate and nitrite). Pilot studies or other research should serve as a precedent for the optimal dilution.

1. Adjust 10-50 µL of sample volume to 190 µL with Deionized Water.

2. Add 10 µL of ZnSO₄ Solution to the 190 µL of diluted sample, vigorously mix and incubate at room temperature (RT) for 15 minutes.

3. Centrifuge sample at 3,000 x g for 5 minutes.

4. Transfer the resulting supernatant to a collection tube for sample storage at -20°C or proceed to the Sample Incubation Procedure located below.

SAMPLE INCUBATION PROCEDURE

1. Place approximately 0.5 g of washed and dried Cadmium Beads in a dedicated centrifuge tube for each representative sample.

2. Add the deproteinated and clarified sample directly to dedicated centrifuge tube with the Cd⁺⁺ present. Incubate at RT overnight with agitation.

3. After incubation, transfer the sample to a clean microcentrifuge tube. Centrifuge for 5 minutes at

3,000 x g. Begin the assay within 1 hour for best results. The used Cadmium Beads should be

collected and washed as indicated in the Reagent Preparation section above.

STANDARD PREPARATION

The stock nitrite standard is provided as a 500 µM NO equivalents (500 pmol/µL of NaNO₂). Prepare the standards according to the following Table 1. The Standards do not need to be incubated with the Cadmium Beads.

Table 1: Preparation of Standard Curve

Standard	NO Equivalents (µM)	Deionized Water (mL)	Transfer Volume	Transfer Source	Final Volume (mL)
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			(mL)		
S ₇	100	2.0	0.5	Standard Stock	2.0
S ₆	50	0.5	0.5	S ₇	0.5
S ₅	25	0.5	0.5	S ₆	0.5
S ₄	10	0.75	0.5	S ₅	0.75
S ₃	5	0.5	0.5	S ₄	0.5
S ₂	1	2.0	0.5	S ₃	2.0
S ₁	0.5	0.5	0.5	S ₂	1.0
B ₀	0	0.5	---	---	0.5

ASSAY PROCEDURE

1. Add 100 μ L of Standards or Samples to the microplate in duplicate. **Note:** Samples may require further dilution with deionized water if the NO concentration exceeds the standard curve parameters.
2. Add 50 μ L Color Reagent #1 to each well and shake briefly.
3. Add 50 μ L Color Reagent #2 to each well. Shake for 5 minutes at room temperature.
4. Read the plate at 540 nm.

Scheme I:

	1	2	3	4	5	6	7	8	9	10	11	12
A	S7	S7	U1	U1	U9	U9	U17	U17	U25	U25	U33	U33
B	S6	S6	U2	U2	U10	U10	U18	U18	U26	U26	U34	U34
C	S5	S5	U3	U3	U11	U11	U19	U19	U27	U27	U35	U35
D	S4	S4	U4	U4	U12	U12	U20	U20	U28	U28	U36	U36
E	S3	S3	U5	U5	U13	U13	U21	U21	U29	U29	U37	U37
F	S2	S2	U6	U6	U14	U14	U22	U22	U30	U30	U38	U38
G	S1	S1	U7	U7	U15	U15	U23	U23	U31	U31	U39	U39
H	B0	B0	U8	U8	U16	U16	U24	U24	U32	U32	U40	U40

CALCULATIONS

1. Average the O.D. values for each replicate of sample and standard.
2. Subtract the average O.D. value of the blank wells (B0) from all other pairs of wells.
3. Plot the standard curve using the standard concentration (X-axis) vs. the corresponding O.D. (Y-axis). A typical standard curve is shown on the next page.

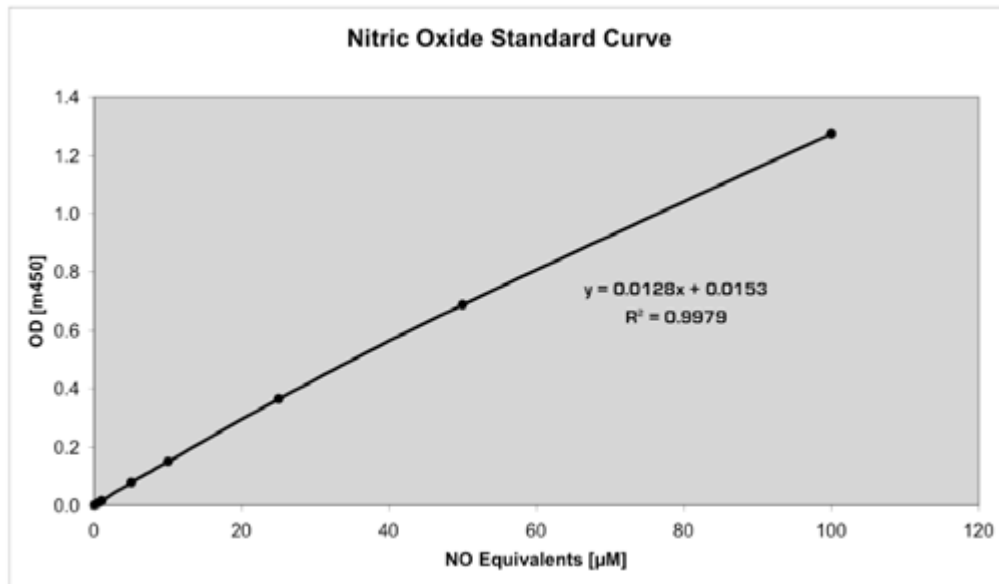
4. Determine the concentration of each sample by interpolation from the standard curve using the Yintercept equation. Remember to multiply by any dilution factors from the preparation steps.

Note: The standard curve is demonstrated in μM Nitric Oxide equivalents but can alternatively be

demonstrated in pmol/mL of NaNO_2 .

1 nM NO = 1 pmol/mL NaNO_2

Figure 2: Typical Standard Curve



REFERENCES

1. Schmidt, H.H., *et. al.*, (1995) *Biochemica* 2:22-23

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