



# Molecular Diagnostics

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NACMID

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# Objectives

- Discuss background concepts related to molecular diagnostics
- Provide an overview of molecular diagnostic history
- Highlight 2 specific molecular diagnostic techniques
- Patient case application(s)

# INTRODUCTION



# Molecular diagnostics

- Using molecular biology to;
  - Expand the scientific knowledge of the natural history of diseases, identify people at risk for specific diseases, and diagnose diseases at the molecular level
  - Uses sequence specific information in macromolecules
    - Peptides/proteins
    - Polysaccharides
    - Polynucleotides/nucleic acids

# Molecular diagnostics

- Detection of pathogenic mutations in DNA and RNA samples
  - Aid in detection
  - Diagnosis and subclassification
  - Prognosis
  - Monitoring response to therapy
- > 80% of molecular tests performed address infectious disease detection and management

# Molecular diagnostics timeline

1865  
Gregor  
Mendel, Law  
of heredity

1949  
Sickle cell  
anemia  
mutation

1970  
Recombinant  
DNA  
technology

1985  
*In vitro*  
amplification  
of DNA (PCR)

2005-2011  
Sequencing  
technologies  
and genome  
sequencing



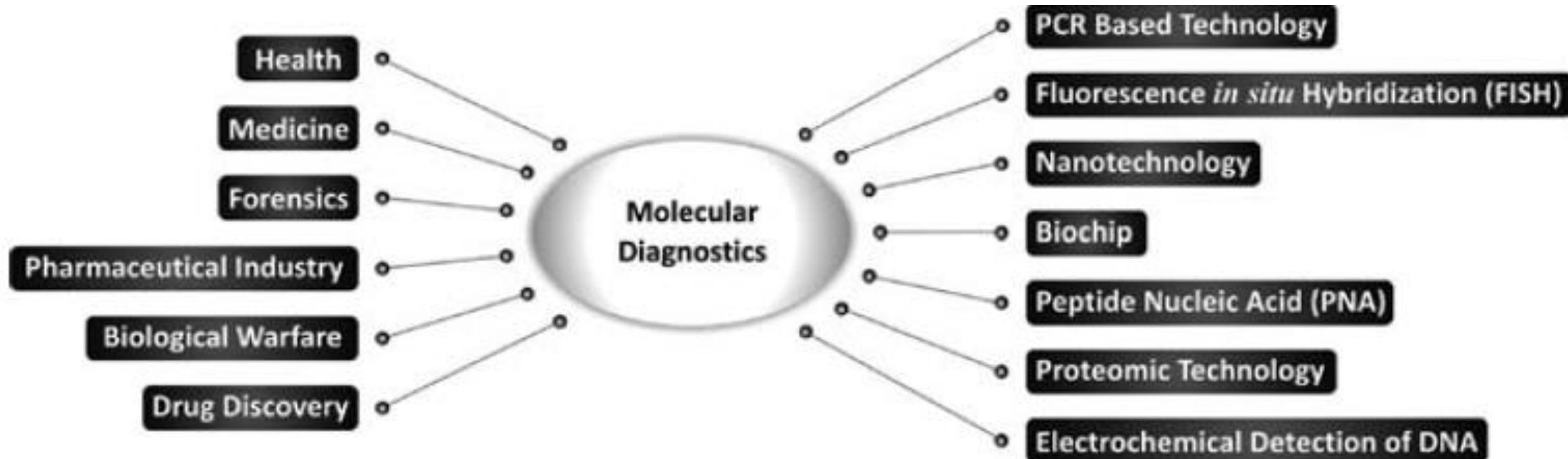
1866  
Purification of  
DNA-Johann  
Miescher  
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1953  
Watson and  
Crick,  
structure of  
DNA

1977  
DNA  
sequencing

2001  
Human  
Genome  
Project

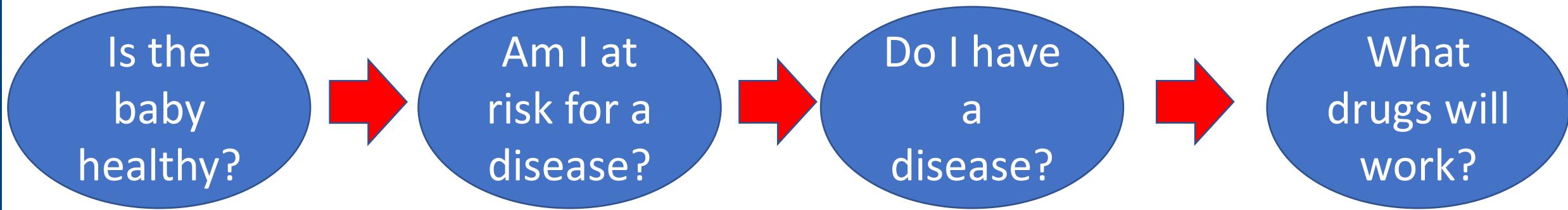
# Common applications



**Fig. 1.1** Applications of molecular diagnostics in the field of clinical diagnostics

# Common applications

- Molecular diagnostics is > \$3 billion worldwide
- Growing at > 20% annually



# TECHNIQUES



# Old vs new molecular diagnostics

- Old
  - Grow cells/pathogen then test
  - Problematic, slow, and costly
- New
  - Direct test
  - Immunological
  - DNA/RNA based

# Non-amplified probe technology

- Detection done via
  - Chromogenic enzyme
  - Fluorophore
  - Radioisotope
- Adjuvant to culture based methods
- Applied to cultures with 1<sup>st</sup> growth of an organism

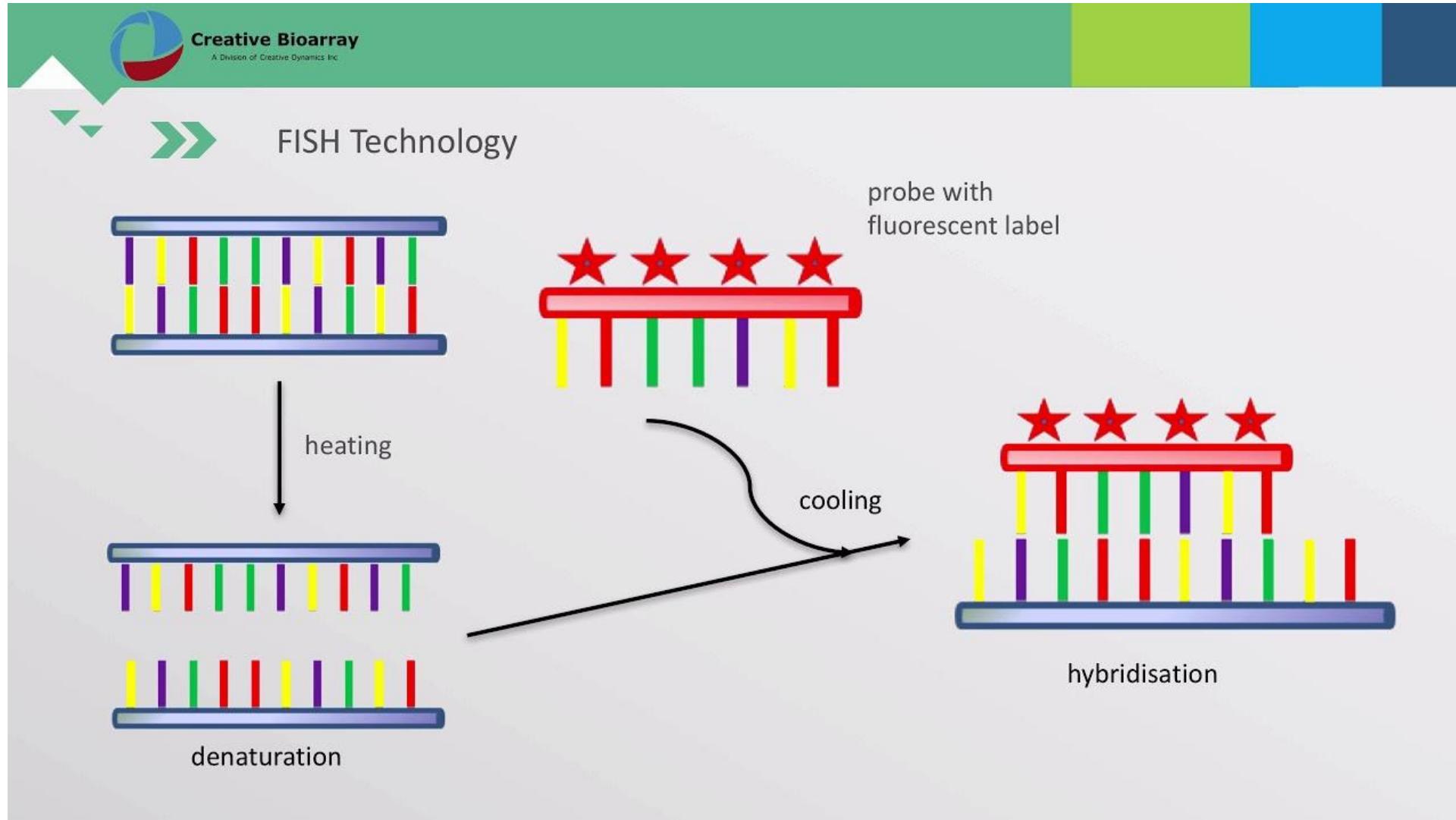
# Non-amplified probe technology

- Among the 1<sup>st</sup> molecular *in vitro* diagnostics (IVDs) to be FDA approved
- Does not include nucleic acid amplification
- Usually require *in vitro* microbial growth
- Molecular probes hybridize with microbial nucleic acids
- Molecular probes identify a targeted sequence of the organism

# Non-amplified probe technology

- Fluorescent *in situ* hybridization (FISH)
  - Popular in early 2000s
  - Individual probes directed against common pathogens causing bacteremia

# FISH



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[Creative Bioarray - YouTube](#)

# Nucleic acid amplification

- Advent in 1980s with development of polymerase chain reaction (PCR) by Mullis
- Synthesize double stranded DNA from a single stranded DNA template with a primer
- The primer allows for amplification of a targeted DNA sequence

# Nucleic acid amplification

- High sensitivity (e.g., single-digit copies of starting template)
- Amplification occurs through multiple heating/cooling temperature cycles
- Each cycle theoretically doubles the amplicons
- Current innovations allow for ~ 40 cycle reactions in ~ 30 minutes.

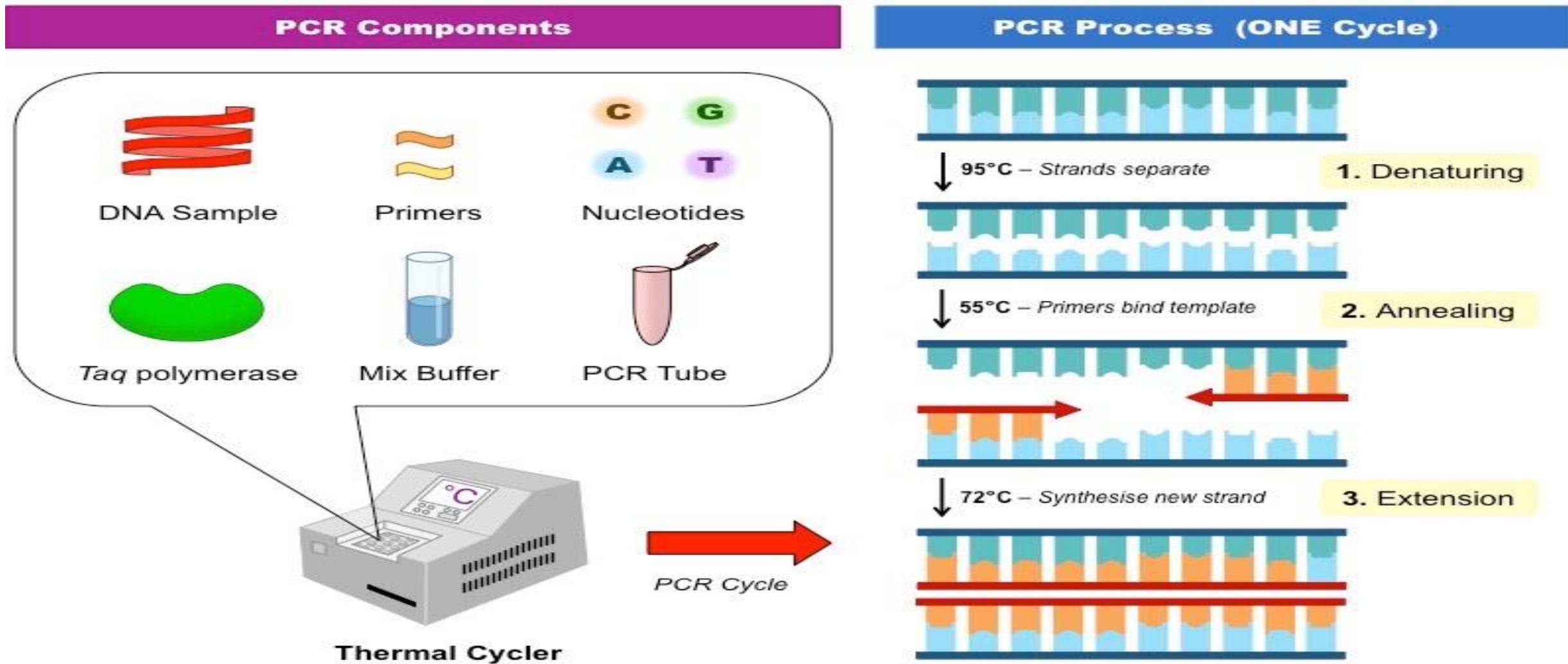
# Nucleic acid amplification

- 3 component steps
  - Specimen processing and nucleic acid extraction
  - Target amplification
  - Detection/characterization of amplified product
- Allows for qualitative and quantitative determination

# Nucleic acid amplification

- Real time PCR is most commonly application in use
- Amplification occurs through multiple heating/cooling temperature cycles
- Each cycle theoretically doubles the amplicons
- Current innovations allow for ~ 40 cycle reactions in ~ 30 minutes
- Single-plex: single pathogen detection
- Multi-plex: multiple pathogen detection

# NAAT-PCR



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PCR | BioNinja

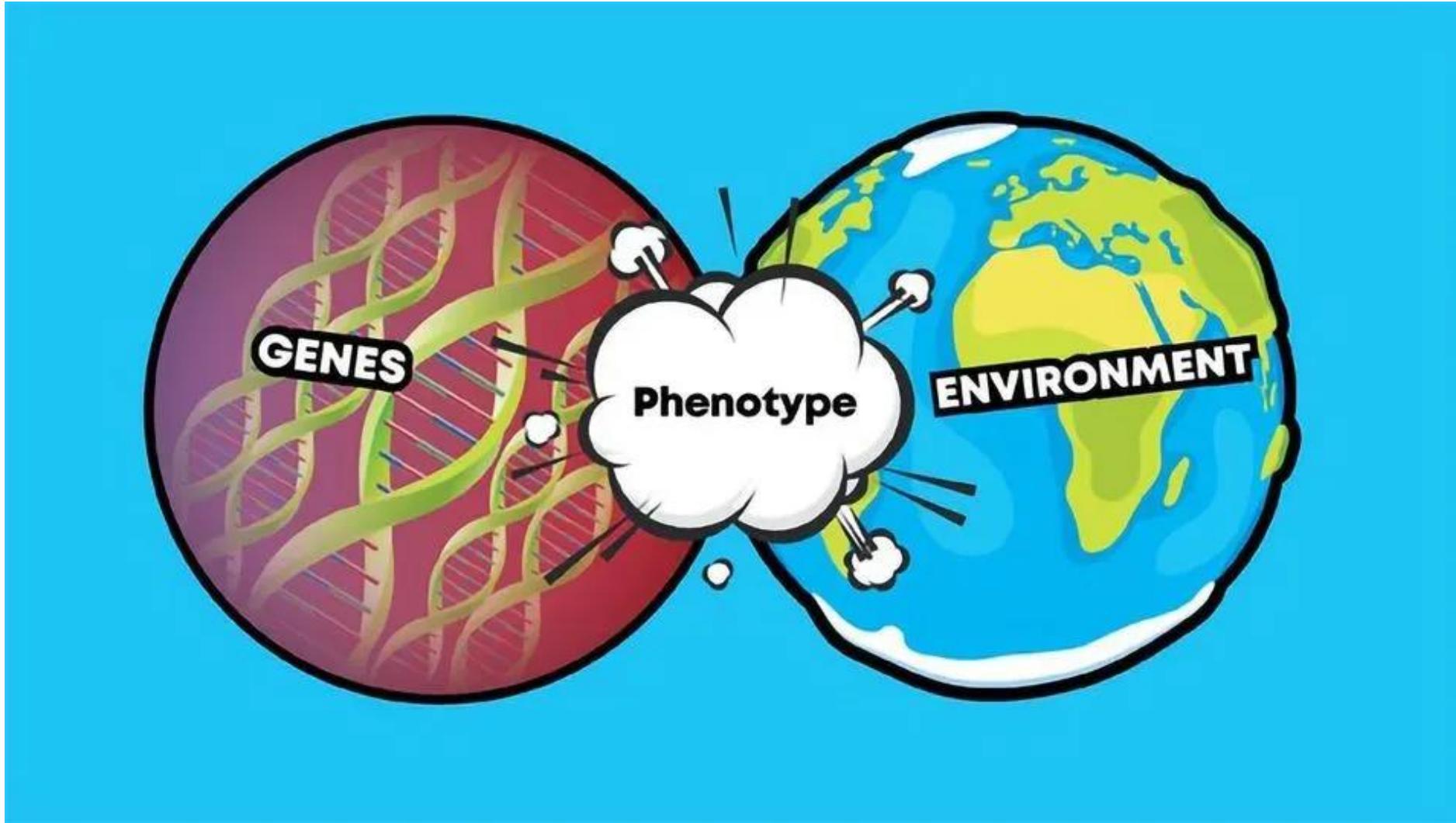
# Advantages

- Wide range of detection
- Time to detection
- Sensitivity
- Specificity

# Disadvantages

- False positives - specimen contamination
- False positive/negative - genotype vs. phenotype
- False negative - off panel target
- Cost
- Rigor of technique and QC

# Genotype vs phenotype



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[Genotype vs Phenotype: Examples and Definitions | Technology Networks](#)

# Genotype

- The genetic constitution of an individual organism
- Unique sequence of DNA
- Molecular diagnostics test for genes
- Presence of a particular gene does not equate to presence of a disease
- Presence of a particular gene does not equate to pres

# Phenotype

- The observable traits/characteristics of an organism
- Based on genotype and the genotypes interaction with the environment
- Detectable expression of a genotype (e.g., clinical presentation)

# Resistance gene detection

Gene	Gram-positive	Gram-negative
<i>mecA/C</i>	+	
<i>mecA/C</i> and MREJ	+	
<i>vanA/B</i>	+	
CTX-M		+
KPC		+
NDM		+

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*J Clin Micro* 2019;57(2):e01597; *BMC Infectious Diseases* 2022;22:794

# CASE 1



# Case 1

- 4 year old patient short bowel syndrome, colostomy, GJ tube, BPD
- Central line for chronic TPN and hydration management
- Presentation
  - Temperature 102 °F
  - Blood pressure 68/40 mmHg
  - Respiratory rate 25 breathes/minute
  - Heart rate 110 beats/minute
  - Capillary refill time 4 seconds

# Case 1

- Day 1 management
  - Isotonic fluids 60-100 mL/kg
  - Vasoactives
  - Blood cultures
  - Vancomycin 15 mg/kg/dose IV q6h
  - Cefepime 50 mg/kg/dose IV q12h then q6h

# Case 1

- Day 2 management
  - Isotonic fluids
  - Vasoactives
  - Repeat blood cultures
  - Meropenem 40 mg/kg/dose IV q8h
  - Blood culture from day 1 → gram-negative rods
  - Multi-plex nucleic acid blood culture test → KPC producing *Klebsiella pneumoniae*

# Case 1

- Day 2 management
  - Regimen changed to meropenem/vaborbactam 40 mg/kg/dose of the meropenem component IV q6h infused over 3 hours

# **Case 1**

## **Pharmacokinetics of the Meropenem Component of Meropenem-Vaborbactam in the Treatment of KPC-Producing *Klebsiella pneumoniae* Bloodstream Infection in a Pediatric Patient**

Alexandra M. Hanretty,<sup>1</sup> Ishminder Kaur,<sup>1,2</sup> Alan T. Evangelista,<sup>1</sup> Wayne S. Moore II,<sup>3</sup> Adela Enache,<sup>4</sup> Arun Chopra,<sup>2,5,6</sup> and Jeffrey J. Cies<sup>1,2,3\*</sup> 

<sup>1</sup>St. Christopher's Hospital for Children, Philadelphia, Pennsylvania; <sup>2</sup>Drexel University College of Medicine, Philadelphia, Pennsylvania; <sup>3</sup>The Center for Pediatric Pharmacotherapy, Pottstown, Pennsylvania; <sup>4</sup>Atlantic Diagnostic Laboratories, Bensalem, Pennsylvania; <sup>5</sup>NYU Langone Medical Center, New York, New York; <sup>6</sup>NYU School of Medicine, New York, New York

# **Case 1**

<b>Drug</b>	<b>MIC</b>
Amikacin	S
Gentamicin	S
Tetracycline	S
Tigecycline	S
Tobramycin	S
Ceftazidime/avibactam	S
Colistin	S
Meropenem/vaborbactam	S

# CASE 2



## Case 2

- Methicillin resistance in *staphylococci* is mediated by penicillin binding protein 2a (PBP 2a)
- Encoded by *mecA* on mobile staphylococcal cassette chromosome *mec* (SCC*mec*) element
- MRSA is a *S. aureus* isolate that displays phenotypic resistance to beta-lactams

## **Case 2**

ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, Aug. 2011, p. 3765–3773  
0066-4804/11/\$12.00 doi:10.1128/AAC.00187-11

Vol. 55, No. 8

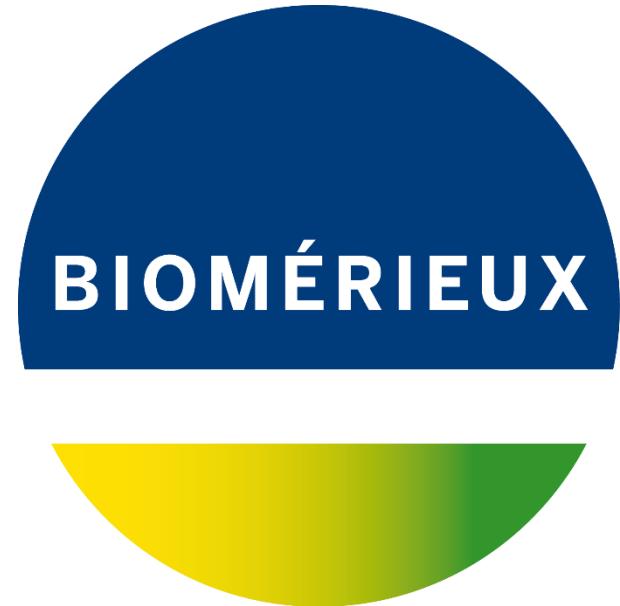
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# Detection of Staphylococcal Cassette Chromosome *mec* Type XI Carrying Highly Divergent *mecA*, *mecI*, *mecR1*, *blaZ*, and *ccr* Genes in Human Clinical Isolates of Clonal Complex 130 Methicillin-Resistant *Staphylococcus aureus*<sup>▽†</sup>

Anna C. Shore,<sup>1</sup> Emily C. Deasy,<sup>1</sup> Peter Slickers,<sup>2</sup> Grainne Brennan,<sup>3</sup> Brian O'Connell,<sup>3,4</sup> Stefan Monecke,<sup>5</sup> Ralf Ehricht,<sup>2</sup> and David C. Coleman<sup>1\*</sup>

## Case 2

- Two *S. aureus* isolates from an Irish hospital
- GeneXpert real time PCR assay
  - Identified as methicillin susceptible *S. aureus*
- Phenotypically PBP 2a (+) → methicillin resistant *S. aureus*
- Lacked *mecA* by conventional PCR and DNA microarray screening
- Whole genome sequencing
  - SCC*mec* type XI
  - Bovine source
  - Not identified with commercially available PCR technologies



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PIONEERING DIAGNOSTICS