PSYCHIATRY AND PRECLINICAL PSYCHIATRIC STUDIES - REVIEW ARTICLE



Application of pharmacogenetics in clinical practice: problems and solutions

Andrius Baskys^{1,2,3}

Received: 1 March 2018 / Accepted: 12 June 2018 / Published online: 19 June 2018 © Springer-Verlag GmbH Austria, part of Springer Nature 2018

Abstract

This paper discusses difficulties of pharmacogenomic data integration into clinical practice. It emphasizes the need for developing simple and easy to use bioinformatics tools to help prescribers to rapidly access and use genetic data in clinical decision-making at the point of encounter.

Keywords Clinical informatics tools · Medication side effect estimation · Drug-drug interactions · Drug-gene interactions

Introduction

How would you individualize medication therapy? You have a new patient whom you have diagnosed with a major depressive disorder. There are over 20 antidepressants available on the market, and you would like medical evidencebased guidance to help you decide which one to prescribe. One approach is often referred to as personalized or precision medicine. Its early uses can be traced back to 1900, when Austrian Karl Landsteiner identified three blood groups, and to 1907 when Reuben Ottenberg performed the first successful blood transfusion at Mount Sinai Hospital in New York based on Landsteiner's findings (Landsteiner 1900). In 1956, the genetic basis for the selective toxicity of fava beans ("favism") and primaquine was discovered to be a deficiency in glucose-6-phospho dehydrogenase or G6PD (Alving et al. 1956). In 1977, cytochrome P450 (CYP) enzyme CYP2D6 was identified as responsible for increased duration and intensity of an anti-hypertensive drug debrisoquine effect, marking the beginning of the modern era of genetics-based treatments (Mahgoub et al. 1977).

Advances in pharmacogenomics have greatly increased our understanding of the relationship between genetic variations, and drug effectiveness or their side effects. Pharmacogenetic testing (PGx) to identify genetic mutations that predict patient responses to pharmacotherapy are emerging as a science-based method to select the optimal treatment regimen for individual patients. PGx-based testing services offering analysis of a patient's DNA have recently become commercially available. Since the adverse drug reactionrelated mortality ranks the 5th in the overall US mortality statistics (Lazarou et al. 1998), the attractiveness of PGx in informing medication prescribing should be difficult to contest. Nevertheless, PGx implementation into clinical practice has been slow, and acceptance by prescribers has been wavering (Patel et al. 2014; Hess et al. 2015; Peterson et al. 2016). The question that inevitably arises is why? To address this question let's review the basic concepts of PGx.

Overview

Recall from basic pharmacology that the relationship between a drug dose and its effectiveness can be separated into pharmacokinetic (dose \rightarrow concentration) and pharmacodynamic (concentration \rightarrow physiological function, pathological process, etc.) components (Katzung et al. 2009). Tissue proteins ultimately influence a patient's response to a drug; since proteins are encoded by genes and genes vary from person to person, drug responses will also vary. Genetic variants associated with the pharmacodynamic component are usually directly associated with a drug target. For example, carriers of a G

Andrius Baskys abaskys@gmail.com

Graduate College of Biomedical Sciences, Western University of Health Sciences, Pomona, CA, USA

Memory Disorders Clinic, Riverside Psychiatric Medical Group, Riverside, CA, USA

World Association of Genomic Medicine, A Coruña, Spain

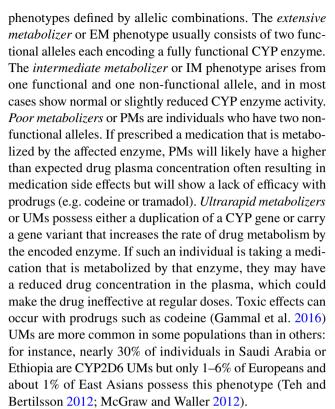
allele in the opiate mu-receptor 1 (*OPMR1*) in a single nucleotide polymorphism (referred to as SNP) known as 118A>G (rs1799971) have a greater sensitivity to pain and require two to four times more of an analgesic drug to achieve a comparable degree of analgesia as non-carriers (Ren et al. 2015).

Pharmacokinetic processes govern absorption, distribution, and elimination of drugs; the genetic aspects of the latter process being most intensely studied. Drug metabolism by cytochrome P450 (CYP) mixed function oxidase system accounts for the elimination of 60-80% of all commercial drugs. Because of a high prevalence of SNPs, deletions, duplications, and splicing defects in CYP genes, effectiveness of CYP enzymes varies greatly between individuals and ethnic groups. A computational analysis of genetic and functional differences of the 57 CYP450 genes performed using the Human Genome Diversity Project and HapMap (http:// www.1000genomes.org/) data that included approximately 1694 individuals belonging to 62 human populations discovered a total of 449 SNPs distributed across the 57 CYP450 genes that could affect individual responses to drugs [Polimanti et al. 2012]. For example, CYP2C19 (Cytochrome P450 Family 2 Subfamily C Member 19) gene is responsible for the metabolism of antidepressants such as citalopram, clomipramine and amitriptyline, proton pump inhibitors and antiplatelet drug clopidogrel (Mrazec 2010). A SNP rs4244285 responsible for a nucleotide change from G to A creates an aberrant splice site resulting in reduced CYP2C19 enzyme function found in carriers of altered alleles (e.g. CYP2C19 *2, see de Morais et al. 1994). A loss of function by one or both alleles results in a partially or completely metabolically inactive enzyme and slows the elimination of drugs predominantly metabolized by CYP2C19 (e.g. sertraline). Alternatively, the presence of CYP2C19 *17 allele results in an accelerated drug metabolism and a potential loss of drug effectiveness.

Another example of genetic variation effects on drug pharmacokinetics is prodrugs such as codeine. Codeine is a week analgesic until it is metabolized by CYP2D6 enzyme yielding morphine, a potent analgesic. Mutations in *CYP2D6* gene impairing CYP2D6 enzyme-mediated metabolic activation of codeine result in low concentration of morphine and, consequently, poor analgesia, leaving the patient in pain even at high doses of codeine (Brousseau et al. 2007). On the other hand, duplication of *CYP2D6* gene can result in a rapid increase in morphine concentration and life-threatening toxicity due to accelerated metabolic activation of codeine.

Metabolizer types

Most PGx services report test results in terms of extensive, intermediate, poor or ultrarapid metabolizer phenotypes (EM, IM, PM and UM). These terms refer to metabolic



Taken together, four phenotypes have been identified in association with CYP genetic polymorphisms that can impact drug metabolic rates and, consequently, have an impact on adverse drug reactions (ADRs) as well as drug efficacy. The US Food and Drug Administration (FDA) has published a "Table of Pharmacogenomic Biomarkers in Drug Labeling" that includes *CYP2D6* and *CYP2C19* genotype-specific dosing recommendations for several popular psychotropic drugs when these drugs are being prescribed to PMs and UMs (Baskys 2017).

CYPs and drug-drug interactions

In addition to genetic polymorphisms, there are other factors influencing the expression and functions of CYPs. They include physiological states or processes such as age, sex, hormones, pregnancy, environment as well as pathological states such as cancer, inflammation and cholestasis. Epigenetic mechanisms such as DNA methylation and histone modifications also regulate CYP gene expression. These factors together with drug–drug interactions (DDIs) contribute to the clinical manifestations of ADRs that arise as a consequence of CYP-catalyzed reactions. CYP induction or inhibition is another major mechanism that underlies DDIs (Samer et al. 2013; Manikandan and Nagini 2018). For a detailed analysis of the molecular mechanisms underlying the induction or inhibition of specific CYPs, as well as epigenetic regulation of CYP expression, the reader is referred



to several excellent reviews on the topic (e.g. He et al. 2015; Manikandan and Nagini 2018). From a prescriber perspective, CYP enzyme induction and inhibition should be always considered in conjunction with the pharmacogenetic data. This can be illustrated by a phenomenon of *phenoconver*sion, which occurs when a person with an IM phenotype is prescribed a medication inhibiting the altered CYP enzyme and becomes a poor metabolizer as a result. Clinically, this may present as a sudden appearance of side effects associated with a drug that a patient has been taking for some time without problems. Significance of phenoconversion is reflected in regulatory agency (such as FDA) warnings and in manufacturers' package inserts. Thus, the FDA warns that fluoxetine (other examples are quinidine, paroxetine) inhibits the activity of CYP2D6 and may make individuals with a typical CYP2D6 metabolic activity resemble poor metabolizers. Prevalence of variant alleles responsible for the IM phenotype can be disproportionally high in some geographic areas and low in others. For example, 50–70% of East Asians are CYP2C19 IMs (Teh and Bertilsson 2012; McGraw and Waller 2012) and could be prone to developing ADRs to CYP2C19 substrate drugs (e.g. voriconazole) when a CYP2C19 inhibitor (e.g. fluvoxamine) is added to their regimen.

Ordering and interpretation of PGx test results

Knowledge of a person's CYP polymorphisms is essential for the development of personalized drug therapy. The polymorphisms of CYP genes can be identified by examining a person's phenotype or genotype. Phenotyping involves administration of a probe drug (e.g. debrisoquine, dextrametorphan) followed by a subsequent measurement of drug/ metabolite concentration ratio in blood, urine or saliva. The advantage of CYP phenotyping is that it offers clinically relevant information reflected by genetic, environmental, and endogenous factors (Manikandan and Nagini 2018). Genotyping provides an accurate individual DNA sequence and information on loss-of-function or gain-of-function mutations coding for specific enzymes involved in metabolism of pharmacological agents. There are numerous methods of CYP genotyping ranging from PCR to high-throughput next generation sequencing (NGS). Comparison of these methods is beyond the scope of this review and interested readers are directed to several recently published excellent reviews that cover this topic (e.g. Samer et al. 2013; Manikandan and Nagini 2018). One important disadvantage of genotyping (which is the foundation of PGx) is the inability to measure the influence of the environmental factors on the CYP enzymes.

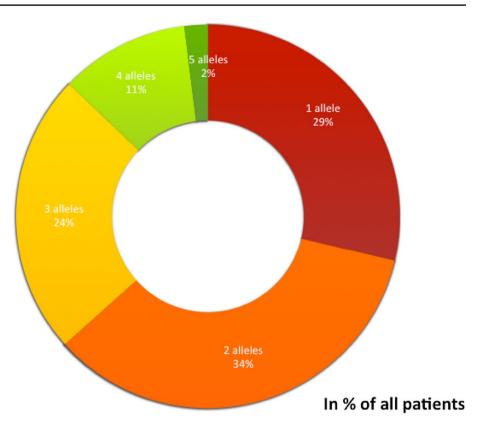
When should one order a pharmacogenetic test? The answer is rather simple: if it has not yet been done before, always order PGx when starting a new medication. There are numerous reasons for this. Thus, health, patient safety, financial and consumer satisfaction benefits of PGx have already been well documented in adult and pediatric populations (Snyder et al. 2014; Ferreri et al. 2014; Plumpton et al. 2016; Gammal et al. 2016; Aka et al. 2017; Moretti et al. 2018), and there are ongoing projects focusing on the implementation of this technology into clinical practice (Blagec et al. 2018). The Ubiquitous-PGx or U-PGx project is an effort of seven European countries to integrate PGx into routine clinical care, which is often hampered by insufficient or fragmented infrastructures. This project aims to set up and implement a unique multimodal, multilingual clinical decision support intervention system consisting of digital-, paper-, and mobile-based tools deployed across implementation sites in the participating countries (Blagec et al. 2018). Further, unlike traditional laboratory tests, PGx test results will remain valid for life and can be applied not only to drugs that the patient is currently taking but also to those that may be taken in the distant future. While it may be argued that preemptive genetic tests should be done only in certain circumstances, so far it has not been possible to apriori define such circumstances (Hinderer et al. 2017). Moreover, recent studies (Cheung et al. 2016; Ji et al. 2016) found that on average there are at least 1-2 actionable variant CYP genes per general psychiatry clinic patient (Fig. 1). These findings suggest that without the knowledge of genetic variants impacting drug metabolism there is a significant likelihood that undesirable drug-gene interactions will be encountered by most prescribers in most patients. Finally, compliance with drug labeling that clearly spell out dose adjustment requirements based on genotype along with fully informing the patient of all available options are undeniably prudent approaches.

One significant difficulty associated with PGx is a relatively large number of data points for a prescriber to consider. This brings us to the question posed at the beginning of this article: which antidepressant to prescribe? While PGx-guided selection of an antidepressant is a sciencebased answer to this question, it is not a simple answer. Given a large number of potential genotypes [449 SNPs distributed across the 57 CYP450 genes that could affect individual responses to drugs (Polimanti et al. 2012)], at least four phenotypes and drug actions on CYP enzymes (substrates, inducers and inhibitors), and their coding genes, the task of PGx-guided prescribing may be difficult if not impossible for a human mind. While most PGx companies do provide detailed genotyping reports, reading these reports could be unacceptably time consuming. For example, Ferreri et al. (2014) found that a clinical pharmacist on average spends 76.6 min with a



112 A. Baskys

Fig. 1 Frequencies of actionable CYP gene variants in a general psychiatry clinic patient population. More than 50% of the study samples were found to have at least one actionable CYP gene allele in six pharmacogenes (CYP2B6, CYP2C9, CYP2D6, CYP2C19, CYP3A4 and CYP3A5). On average, there were 2.22 variant alleles within the six pharmacogenes per subject (Adapted from Cheung et al. 2016 with permission)



patient communicating PGx test results. Further, static PGx reports typically do not account for the phenoconversion phenomenon. To address these problems and facilitate PGx acceptance by prescribers, user-friendly bioinformatics tools are urgently needed. There are several user-friendly static databases that allow to explore drug-drug or drug-genotype interactions. Examples include EuroPharmagenics drug-gene-disease interaction database by EuroEspes SA, (http://www.europharma genics.com), Stanford University managed PharmGKB database (Whirl-Carrillo et al. 2012) and SuperCYP, a comprehensive database on Cytochrome P450 enzymes that includes a tool for analysis of CYP-drug interactions (Preissner et al. 2012). An innovative interactive tool that allows drug-gene and drug-drug interaction visualization and the Dynamic Medication SelectionTM algorithm is available for download from http://www.medpicker.com.

In summary, strengths of PGx and other molecular techniques are in their accuracy and a potential to improve outcomes, patient safety and experiences. Their drawbacks are large quantities of data that cannot be swiftly integrated into the clinical decision process without resorting to bioinformatics tools. The successful implementation of PGx in clinical practice depends on the development of user-friendly bioinformatics tools that can process and present genetic and drug interaction data to clinicians at the point of service.

References

Aka I, Bernal CJ, Carroll R, Maxwell-Horn A, Oshikoya KA, Van Driest SL (2017) Clinical pharmacogenetics of cytochrome P450-associated drugs in children. J Personalized Med 7:E14

Alving AS, Carson PE, Flanagan CL, Ickes CE (1956) Enzymatic deficiency in primaquine-sensitive erythrocytes. Science 124:484–485

Baskys A (2017) Pharmacogenomic information for common psychotropic drugs. In: Procyshyn RM, Bezchlibnyk-Butler KZ, Jeffries JJ (eds) Clinical handbook of psychotropic drugs, 22nd edn. Hogrefe Publishing, Boston, pp 399–409

Blagec K, Koopmann R, Crommentuijn-van Rhenen M, Holsappel I, van der Wouden CH, Konta L, Xu H, Steinberger D, Just E, Swen JJ, Guchelaar HJ, Samwald M (2018) Implementing pharmacogenomics decision support across seven European countries: the Ubiquitous Pharmacogenomics (U-PGx) project. J Am Med Inform Assoc. https://doi.org/10.1093/jamia/ocy005 (Epub ahead of print)

Brousseau DC, McCarver DG, Drendel AL, Divakaran K, Panepinto JA (2007) The effect of CYP2D6 polymorphisms on the response to pain treatment for pediatric sickle cell pain crisis. J Pediatr 150:623–626

Cheung Y, Summerour RB, Cui X, Baskys A (2016) Testing for CYP polymorphisms is associated with a reduction in the frequency of changes in psychotropic prescriptions made by community psychiatrists. J Genom Med Pharmacogenomics 1:1–5

de Morais SM, Wilkinson GR, Blaisdell J, Nakamura K, Meyer UA, Goldstein JA (1994) The major genetic defect responsible for the polymorphism of S-mephenytoin metabolism in humans. J Biol Chem 269:15419–15422

Ferreri SP, Greco AJ, Michaels NM, O'Connor SK, Chater RW, Viera AJ, Faruki H, McLeod HL, Roederer MW (2014) Implementation



- of a pharmacogenomics service in a community pharmacy. J Am Pharm Assoc 54:172–180
- Gammal RS, Crews KR, Haidar CE, Hoffman JM, Baker DK, Barker PJ, Estepp JH, Pei D, Broeckel U, Wang W, Weiss MJ, Relling MV, Hankins J. (2016) Pharmacogenetics for safe codeine use in sickle cell disease. Pediatrics 138:e20153479
- He ZX, Chen XW, Zhou ZW, Zhou SF (2015) Impact of physiological, pathological and environmental factors on the expression and activity of human cytochrome P450 2D6 and implications in precision medicine. Drug Metab Rev 47:470–519
- Hess GP, Fonseca E, Scott R, Fagerness J (2015) Pharmacogenomic and pharmacogenetic-guided therapy as a tool in precision medicine: current state and factors impacting acceptance by stakeholders. Genet Res (Camb) 97:e13
- Hinderer M, Boeker M, Wagner SA, Lablans M, Newe S, Hülsemann JL, Neumaier M, Binder H, Renz H, Acker T, Prokosch HU, Sedlmayr M (2017) Integrating clinical decision support systems for pharmacogenomic testing into clinical routine—a scoping review of designs of user-system interactions in recent system development. BMC Med Inform Decis Mak 17:81
- Ji Y, Skierka JM, Blommel JH, Moore BE, VanCuyk DL, Bruflat JK, Peterson LM, Veldhuizen TL, Fadra N, Peterson SE, Lagerstedt SA, Train LJ, Baudhuin LM, Klee EW, Ferber MJ5, Bielinski SJ, Caraballo PJ, Weinshilboum RM, Black JL (2016) Preemptive pharmacogenomic testing for precision medicine. A comprehensive analysis of five actionable pharmacogenomic genes using next-generation DNA sequencing and a customized CYP2D6 genotyping cascade. J Mol Diagn 18:438–445
- Katzung BG, Masters SB, Trevor AJ (2009) Basic and clinical pharmacology, 11th edn. McGraw-Hill Medical, New York
- Landsteiner K (1900) Zur Kenntnis der antifermentativen, lytischen und agglutinierenden Wirkungen des Blutserums und der LympheZentralbl. Bakteriol 27:357–362
- Lazarou J, Pomeranz BH, Corey PN (1998) Incidence of adverse drug reactions in hospitalized patients: a meta-analysis of prospective studies. JAMA 279:1200–1205
- Mahgoub A, Idle JR, Dring LG, Lancaster R, Smith RL (1977) Polymorphic hydroxylation of Debrisoquine in man. Lancet 2:584–586
- Manikandan and Nagini (2018) Cytochrome P450 structure, function and clinical significance: a review. Curr Drug Targets 19:38–54
- McGraw J, Waller D (2012) Cytochrome P450 variations in different ethnic populations. Expert Opin Drug Metab Toxicol 8:371–382
- Moretti ME, Lato DF, Berger H, Koren G, Ito S, Ungar WJ (2018) A cost-effectiveness analysis of maternal CYP2D6 genetic testing

- to guide treatment for postpartum pain and avert infant adverse events. Pharmacogenomics J 18:391–397
- Mrazec DA (2010) Psychiatric Pharmacogenomics. Oxford University Press, Oxford
- Patel HN, Ursan ID, Zueger PM, Cavallari LH, Pickard AS (2014) Stakeholder views on pharmacogenomic testing. Pharmacotherapy 34:151–165
- Peterson JF, Field JR, Shi Y, Schildcrout JS, Denny JC1, McGregor TL, Van Driest SL, Pulley JM, Lubin IM, Laposata M, Roden DM, Clayton EW (2016) Attitudes of clinicians following large-scale pharmacogenomics implementation. Pharmacogenomics J 16:393–398
- Plumpton CO, Roberts D, Pirmohamed M, Hughes DA (2016) A systematic review of economic evaluations of pharmacogenetic testing for prevention of adverse drug reactions. Pharmacoeconomics 34:771–793
- Polimanti R, Piacentini S, Manfellotto D, Fuciarelli M (2012) Human genetic variation of CYP450 superfamily: analysis of functional diversity in worldwide populations. Pharmacogenomics 16:1951–1960
- Preissner S, Kroll K, Dunkel M et al. (2012) SuperCYP: a comprehensive database on cytochrome P450 enzymes including a tool for analysis of CYP-drug interactions. Nucleic Acids Res 2010:D237–D243
- Ren ZY, Xu XQ, Bao YP, He J, Shi L, Deng JH, Gao XJ, Tang HL, Wang YM, Lu L (2015) The impact of genetic variation on sensitivity to opioid analgesics in patients with postoperative pain: a systematic review and meta-analysis. Pain Phys 18:131–152
- Samer CF, Lorenzini KI, Rollason V, Daali Y, Desmeules JA (2013) Applications of CYP450 testing in the clinical setting. Mol Diagn Ther 17:165–184
- Snyder SR, Mitropoulou C, Patrinos GP, Williams MS (2014) Economic evaluation of pharmacogenomics: a value based approach to pragmatic decision making in the face of complexity. Public Health Genom 17:256–264
- Teh LK, Bertilsson L (2012) Pharmacogenomics of CYP2D6: molecular genetics, interethnic differences and clinical importance. Drug Metab Pharmacokinet 27:55–67
- Whirl-Carrillo M, McDonagh EM, Hebert JM et al (2012) Pharmacogenomics knowledge for personalized medicine. Clin Pharmacol Ther 92:414–417



Terms and Conditions

Springer Nature journal content, brought to you courtesy of Springer Nature Customer Service Center GmbH ("Springer Nature").

Springer Nature supports a reasonable amount of sharing of research papers by authors, subscribers and authorised users ("Users"), for small-scale personal, non-commercial use provided that all copyright, trade and service marks and other proprietary notices are maintained. By accessing, sharing, receiving or otherwise using the Springer Nature journal content you agree to these terms of use ("Terms"). For these purposes, Springer Nature considers academic use (by researchers and students) to be non-commercial.

These Terms are supplementary and will apply in addition to any applicable website terms and conditions, a relevant site licence or a personal subscription. These Terms will prevail over any conflict or ambiguity with regards to the relevant terms, a site licence or a personal subscription (to the extent of the conflict or ambiguity only). For Creative Commons-licensed articles, the terms of the Creative Commons license used will apply.

We collect and use personal data to provide access to the Springer Nature journal content. We may also use these personal data internally within ResearchGate and Springer Nature and as agreed share it, in an anonymised way, for purposes of tracking, analysis and reporting. We will not otherwise disclose your personal data outside the ResearchGate or the Springer Nature group of companies unless we have your permission as detailed in the Privacy Policy.

While Users may use the Springer Nature journal content for small scale, personal non-commercial use, it is important to note that Users may not:

- 1. use such content for the purpose of providing other users with access on a regular or large scale basis or as a means to circumvent access control;
- 2. use such content where to do so would be considered a criminal or statutory offence in any jurisdiction, or gives rise to civil liability, or is otherwise unlawful;
- 3. falsely or misleadingly imply or suggest endorsement, approval, sponsorship, or association unless explicitly agreed to by Springer Nature in writing:
- 4. use bots or other automated methods to access the content or redirect messages
- 5. override any security feature or exclusionary protocol; or
- 6. share the content in order to create substitute for Springer Nature products or services or a systematic database of Springer Nature journal content

In line with the restriction against commercial use, Springer Nature does not permit the creation of a product or service that creates revenue, royalties, rent or income from our content or its inclusion as part of a paid for service or for other commercial gain. Springer Nature journal content cannot be used for inter-library loans and librarians may not upload Springer Nature journal content on a large scale into their, or any other, institutional repository.

These terms of use are reviewed regularly and may be amended at any time. Springer Nature is not obligated to publish any information or content on this website and may remove it or features or functionality at our sole discretion, at any time with or without notice. Springer Nature may revoke this licence to you at any time and remove access to any copies of the Springer Nature journal content which have been saved.

To the fullest extent permitted by law, Springer Nature makes no warranties, representations or guarantees to Users, either express or implied with respect to the Springer nature journal content and all parties disclaim and waive any implied warranties or warranties imposed by law, including merchantability or fitness for any particular purpose.

Please note that these rights do not automatically extend to content, data or other material published by Springer Nature that may be licensed from third parties.

If you would like to use or distribute our Springer Nature journal content to a wider audience or on a regular basis or in any other manner not expressly permitted by these Terms, please contact Springer Nature at

onlineservice@springernature.com