









Fortification of Staple Commodities and Microbial Safety Requirements for Human Food Production: Distant USAID F2F Program in Haiti

Trip Report (April 8, 2021)

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Fortification of Staple Commodities and Microbial Safety Requirements for Human Food Production

USAID F2F assignment: Haiti, Nashville, TN 12-7-2020

Tennessee State University, Nashville, TN



Summary:

I was privileged and honored to serve as a volunteer for this assignment in Haiti. Due to an ongoing global respiratory pandemic/endemic and consequently local and global travel restrictions, this assignment was completed solely via Zoom and distantly. Nevertheless, thanks to the support and cooperation of F2F colleagues from Washington and Haiti offices, and the enthusiastic participation of the host, this assignment was very impactful for capacity building and progress. The assignment was originally designed for only two weeks in July, however, due to requests from the host institution, the meeting was extended until December 2020.

During the first three months of the assignment, through various Zoom/Team calls and email conversations, the public health microbiology program was able to share and discuss resources and best practices about micronutrient malnutrition with the host institution including information from USAID Micronutrient Program and the World Health Organization.

Specifically, methods and best practices were discussed for fortification of flour with iron including Qualitative Test (Spot Test), Semi-Quantitative Test (Spot Test), Semi-Quantitative Test (Colorimetric method), Quantitative Test (Spectrophotometric method), and Quantitative Test (Atomic absorption spectrophotometric [AAS]) method. These assays could be incorporated as part of regulatory affairs in the field as well as during laboratory analyses to assure effectiveness and compliance of the intervention.

In the next phase of the assignment, a list of reagents and consumables was developed to assure the host institutions are capable of conducting these endeavors. The list was tailored to the need of the host institution by assuring assays and laboratory procedures could be completed domestically after acquiring the supplies. Cost-saving recommendations as well as suggestions to purchase service contract for parts and labor were made to assure sensitive instrumentation could remain functional for an extended time.

In November of 2020, some participants of the host program in Haiti joined a multiday food safety certification workshop hosted by the public health microbiology laboratory. While receiving the certifications, members of the host also shared valuable and insightful information about great progress in the food industry and fortification programs in Haiti that benefited the

participants of the workshop. Excerpts of the workshop presentations are provided in this trip report.

One of the most impactful parts of this assignment was a fortification workshop held in December of 2020 where detailed best laboratory practices, nutrition information, and food safety procedures were discussed with 13 senior-level participants of the workshop including members from governmental and non-governmental organizations and the food industry (specifically participants were from Ministry of Health; Oil Industry: Carribex & HUHSA; Salt Industry: Bon Sel D'Haiti; Wheat Flour Industry: LMH & Caribbean Milling; and RANFOSE).

Not only am very pleased about the great progress and deliverables of this extended assignment, but as well I would like to sincerely thank great colleagues from RANFOSE for their outstanding work and progress for food security and public health in their country. Specifically, am grateful for friendship and collaboration with Mr. Regis Yves-Laurent and Dr. Ruth Climat who inspired me by their outstanding progress, professionalism, and dedication to the fortification program in their home country.

Finally, I would like to thank all colleagues in the Washington and Haiti F2F office, specially Ms. Susanna Meyer, for the outstanding work they are doing and for harmonizing the events of this program. With their help and support and with assistance from the members of the public health microbiology laboratory, we were able to conduct this impactful and important training event during the unknown and stressful global respiratory pandemic/endemic times. Am pleased and excited about the capacity-building endeavors in this assignment and hope that the host company and the F2F office in Haiti continue to flourish in future years to come.

Sincere regards,

Aliyar Fouladkhah, PhD, MPH, CFS

Assistant Professor, Tennessee State University

Faculty Director, Public Health Microbiology Laboratory

Yale School of Public Health Alumnus

Consumables and Methods



Hello, Mr. Regis,

I put together the testing methods for iron in flour to seek your input. I just would want to make sure that I am on right track and if these are the tests that you have in mind.

If you would like to schedule a phone call I would complete some other tests by then and could report to you all more on these. For Iron testing I divided the tests in three categories (1) **Qualitative/spot test**. This would be helpful for inspection purposes and shows presence or absence of the micronutrient in the food vehicle, relatively inexpensive and easy to conduct. The second category (2), called them **semi-quantitative test** that are wet chemistry-based and do not need major instrumentations, 2.A. is similar to the spot test and 2.B is a spectrophotometric methods. Both 2.A. and 2.B. could be used in regional labs and also in the manufacturing facilities. Third categories are **quantitative methods**, those are obviously more labor intensive and more difficult to conduct as they require iron standards, 3.A. is spectrophometric tests and 3.B is Atomic absorption spectrophotometric methods. I just would want to make sure that I am on right track and if these are the tests that you have in mind. Once we schedule the next phone call, I will report on remaining of progress as well.

Testing Iron in Flour:

1. Qualitative Test (Spot Test):

Method: Iron qualitative method, AACC Method 40-40

Reagents:

Thiocyanate reagent (Dissolve 10 g KSCN in 100 ml water).

2N HCl,

Hydrogen peroxide, 3%.

Equipment: none

2.A. Semi-Quantitative Test: (Spot Test):

Method: Semi-quantitative spot test for iron INCAP method IV.

Reagents:

Hydrochloric acid, HCl

Hydrogen peroxide, H2O2

Potassium thiocyanate, KSCN

H2O2

2. B. Semi-Quantitative Test (Colorimetric method):

Method: Semi-quantitative colorimetric determination of iron in flour INCAP method V

Sodium acetate (CH3COONa.3H2O), 99%

Hydrochloric acid (HCl), 37%

a,a-dipyridyl (2,2' bipyridine)

Bathophenanthroline, 4,7-diphenyl-1,10-phenanthroline-disulfonic acid (C24H16N2O6S2)

Hydroxylamine hydrochloride (NH2OH.HCl)

Iron standards: Electrolytic iron, Merck 3810 or Baker 2234

3.A. Quantitative Test (Spectrophotometric method):

Method: Iron spectrophotometric method AACC method 40-41B

Reagents:

Orthophenanthroline solution (0.1 g o-phenanthroline in about 80 ml water).

Iron standard solution, (10 μg Fe/ml)

Hydroxylamine hydrochloride solution (10 g NH2OH.HCl in water and dilute to 100 ml)

Acetate buffer solution (8.3 g anhydrous sodium acetate in water)

3.B. Quantitative Test Atomic absorption spectrophotometric (AAS) method:

Method: Spectrophotometric analysis for quantitative, INCAP method VI

Reagents:

Sodium acetate (CH3COONa.3H2O), 99%

Hydrochloric acid (HCl), 37%, 1.19 g/mL

a,a-dipyridyl (2,2' bipyridine) (C10H8N2)

Bathophenanthroline, 4,7-diphenyl-1,10-phenanthroline-disulfonic acid (C24H16N2O6S2)

Hydroxylamine hydrochloride (NH2OH.HCl)

Iron standards: Electrolytic iron, Merck 3810

Spot Test for Iron in Flour

AACC Method 40-40: Iron Qualitative Method

References:

Schlesinger, H. I., and Van Valkenburgh, H. B. 1931. The structure of ferric thiocyanate and the thiocyanate test for iron. J. Am. Chem. Soc. 53:1212.

Johnson, Q., Mannar, V. and Ranum, P., 2004. Fortification handbook: Vitamin and mineral fortification of wheat flour and maize meal. Ottawa: Micronutrient Initiative.

	Suggested catalog number VWR	Suggested catalog number Fischer
Acetonitrile, HPLC grade	BDH83642.050	A998-1
Ascorbic acid, Analytical	BDH9242-250G	LC115309
reagent grade		
Deionized (DI), water	BDH1168-4LP	9180-32
Nanopure, 18.2 megaohm		
Flour Unenriched	n/a	n/a
Folic acid, 98% pure	AAJ60833-22	AC216630100
analytical reagent grade		
Glacial acetic acid, Analytical	BDH3093-2.2LP	02-002-123
reagent grade		
Hexane, HPLC grade	BDH1129-4LP	H303-1
Methanol, HPLC grade	BDH7941-1	A454-1
pH buffers, 4.00 and 7.00	BDH5046-500ML;	13-301-133
	BDH0196-55GL	
Phosphoric acid, Analytical	BDH7347-1	RABP0030500
reagent grade		
Potassium hydroxide,	BDH7548-4	P246-3
Analytical reagent grade		
Potassium phosphate, dibasic	BDH153184U	P285-500
Analytical reagent grade		
Reference flour	n/a	n/a
Sodium acetate, anhydrous	BDH9278-500G	ICN19549601
Analytical reagent grade		
Sodium chloride Analytical	BDH7466-1	S271-500
reagent grade		

Iron—Spectrophotometric Method

Inorganic Constituents AACC Method 40-41B

References:

AOAC International. 1995. Official Methods of Analysis of AOAC International, 16th ed. Method 965.09. The Association, Arlington, VA.

Gatehouse, B. M., and Willis, J. B. 1961. Performance of a simple atomic absorption spectrophotometer.

Spectrochim. Acta 17:710.

Johnson, Q., Mannar, V. and Ranum, P., 2004. Fortification handbook: Vitamin and mineral fortification of wheat flour and maize meal. Ottawa: Micronutrient Initiative.

	Suggested catalog number VWR	Suggested catalog number Fischer
Ortho phenanthroline solution	RC5522-32	NC0646077
Iron standard solution (from analytical grade Fe)	AA11381-GH	US-ICP-126
Fe (NH4)2(SO4)2·6H2O	700001-550	US-ICP-126
HCL	BT224145-4L	A508-P500
Hydroxylamine hydrochloride (from NH2OH·HCl)	EM1.04616.1000	NC1020874
Anhydrous sodium acetate	BDH9278-500G	S210-500
Mg (NO3)2·6H2O	AAH52101-MD	n/a
Redistilled HNO3	34126-320	A198C-212

B-Vitamins Tests by HPLC

Vitamins AACC Method 86-90

References:

Johnson, Q., Mannar, V. and Ranum, P., 2004. Fortification handbook: Vitamin and mineral fortification of wheat flour and maize meal. Ottawa: Micronutrient Initiative.

	Suggested catalog number VWR	Suggested catalog number Fischer	
Pentanesulfonic acid	BDH152813W	02-002-049	
Acetic acid	87003-239	SA36-1	
Calcium carbonate, HPLC	JT1301-5	LC126901	
grade			
Methanol	BDH7941-1	NC0446254	
Niacin	10791-832	18-604-780	
Niacinamide	10791-810	AAJ19480A1	
Pyridoxine hydrochloride	103510-526	18-605-992	
Riboflavin	97062-304	AC132350250	
Niacin	10791-832	18-604-780	
Niacinamide	10791-810	AAJ19480A1	
CaCO3	95032-222	18-609-528	

Elements by Atomic Absorption Spectrophotometry

Inorganic Constituents AACC Method 40-70

References:

Johnson, Q., Mannar, V. and Ranum, P., 2004. Fortification handbook: Vitamin and mineral fortification of wheat flour and maize meal. Ottawa: Micronutrient Initiative.

AOAC International. 1995. Official Methods of Analysis of AOAC International, 16th ed. Method 965.09. The Association, Arlington, VA.

Gatehouse, B. M., and Willis, J. B. 1961. Performance of a simple atomic absorption spectrophotometer. Spectrochim. Acta 17:710.

Zook, E. G., Greene, F. E., and Morris, E. R. 1970. Nutrient composition of selected wheats and wheat products. VI. Distribution of manganese, copper, nickel, zinc, magnesium, lead, tin, cadmium, chromium, and selenium as determined by atomic absorption spectroscopy and colorimetry. Cereal Chem. 47:720.

	Suggested catalog number VWR	Suggested catalog number Fischer
Distilled-deionized water	MKH45309	9180-32
CaCO3	103514-732	LC127007
3N HCl	BDH7375-1	R3720000120
Copper (pure Cu)	JT1720-1	AC206330050
HNO3	BT127005-1LB	60-010-99
Iron (pure Fe)	JT4348-22	AC169430050
6N HCl	BDH7204-4	18-610-924
Magnesium (pure Mg)	AAL08120-0E	AA1023322
Manganese (pure MnO2)	AA10236-A7	AA3631514
Zinc (pure Zn metal)	103524-180	Z5-500
La2O3	AA11264-30	AC193291000
1% HCl	470301-230	n/a

Excerpts of Food Safety Workshop Presentations and Certificants





Foodborne Diseases of Public Health Importance



Bacteria

•	If conditions are ideal, bacteria can
	multiply once every 20 minutes

- It is unlikely you'll ever start with just ONE bacterium
- Some pathogens can make people sick with a dose of **10 cells or less**
- What conditions are optimal?
 - Food source
 - Moisture
 - Right temperature

Time	# of Bacteria
20 min	2
40 min	4
1 hour	8
80 min	16
100 min	32
2 hours	64
4 hours	4096
6 hours	262,144
8 hours	16,777,216
	/

Emerging pathogens

Diversity, moving towards "fitness" and Emerging Pathogens

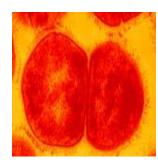
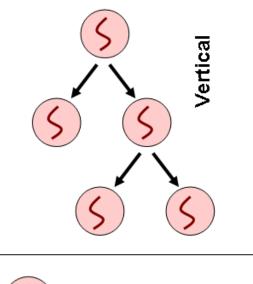
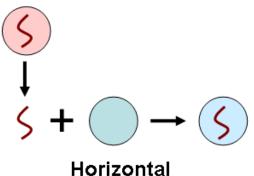
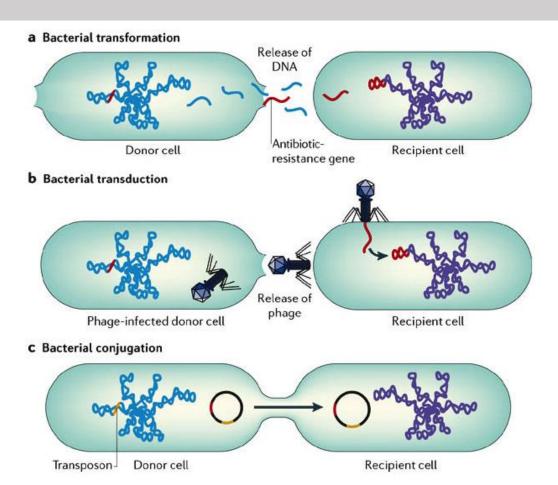


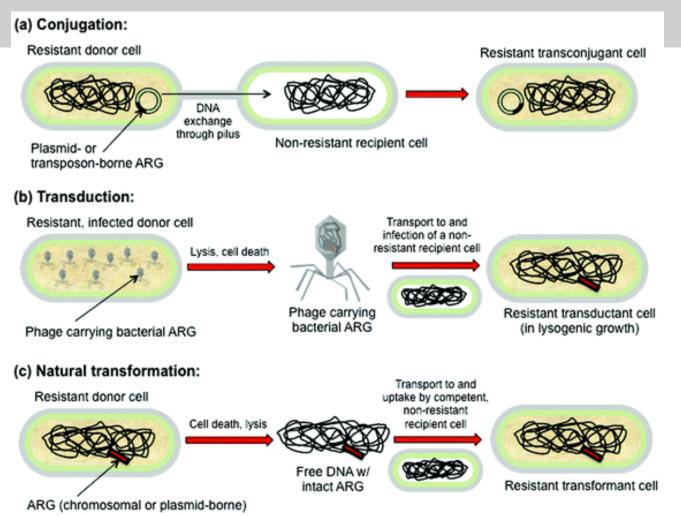
Photo Courtesy: http://www.daviddarling.info/encyclopedia /B/binary_fission.html



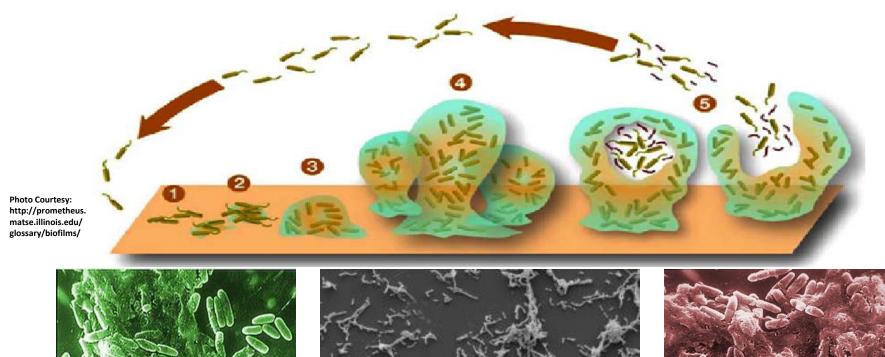




Horizontal Gene Transfer



Planktonic cells and Biofilm Communities



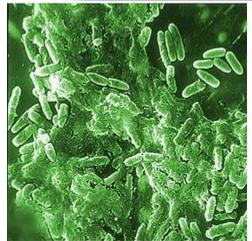


Photo Courtesy: http://microwriters.egybio.net/blog/?tag=antibiotic-resistance

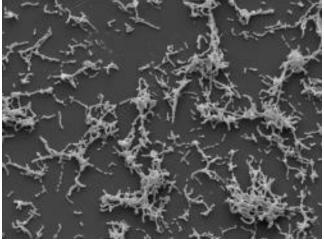


Photo Courtesy: http://www.ifenergy.com/50226711/boosting_microbial_fuel_cells_with_biofilm.php

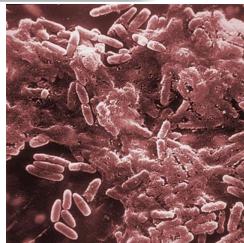
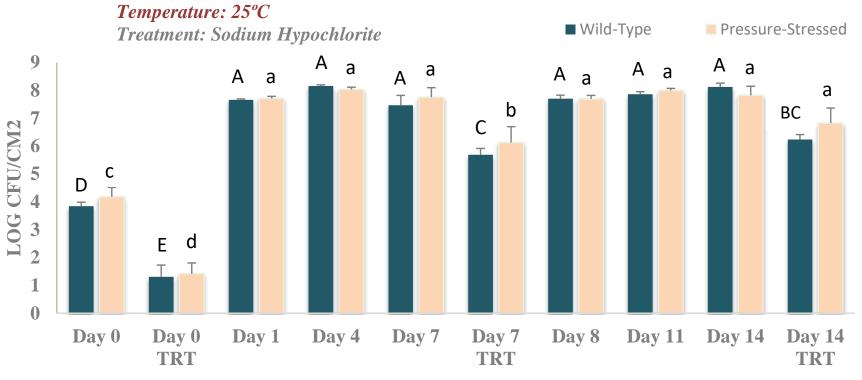


Photo Courtesy: http://www.microbiologybytes.com/blog/category/biofilms/

Cronobacter sakazakii Two outbreaks in Tennessee (1998, Memphis; 2001 Knoxville)

Biofilm Formation and Decontamination of Wild-Type and Pressure-Stressed *Cronobacter Sakazakii*





Allison et al., 2020



DAYS AND TREATMENT (MEAN±SE)

Quorum Sensing and Biofilm formation Shiga toxin-Producing E. coli and antibiotics treatment

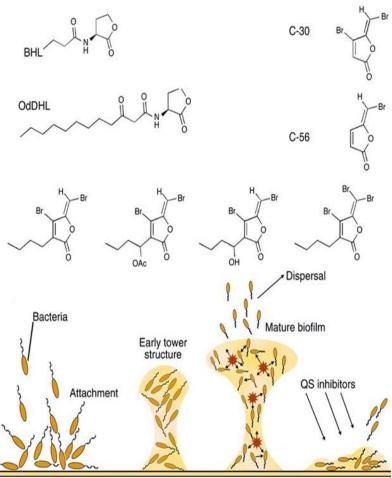


Photo Courtesy: http://www.jci.org/articles/view/20074/figure/2

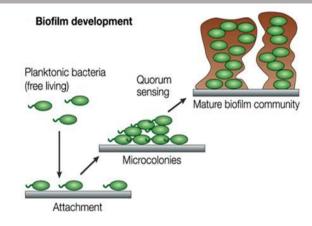
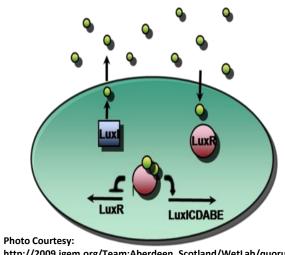


Photo Courtesy: http://labrat.fieldofscience.com/2010/07/quorum-sensing-and-biofilms.html



http://2009.igem.org/Team:Aberdeen_Scotland/WetLab/quorumsensing

Infectious Diseases in Animals and Human is a Moving Target...

- It is estimated only 1% of microbial community has been identified.
- Currently etiological agent of 80.3% of foodborne illnesses, **56.2% of hospitalization**, and 55.5% of deaths remain unknown.

"Emerging" Pathogens:

- Vertical and horizontal gene transfer spores and biofilm formation
- Quorum sensing and cell to cell communication

"It is the microbes who will have the last word."
-Louis Pasteur

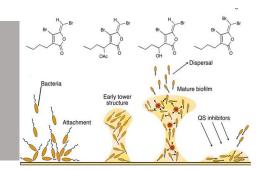
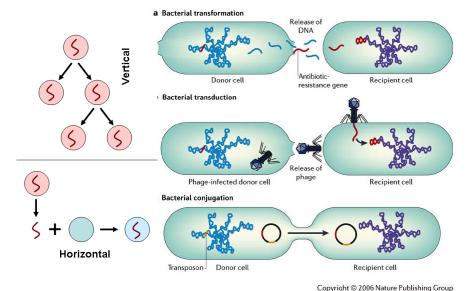
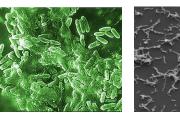
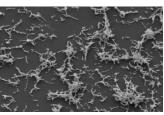


Photo Courtesy: http://www.jci.org/articles/view/20074/figure/2









Nature Reviews | Microbiology

Photo Courtesy: http://www.microbiologybytes.com/blog/category/biofilms/ http://www.ifenergy.com/50226711/boosting_microbial_fuel_cells_with_biofilm.php http://micro-writers.egybio.net/blog/?tag=antibiotic-resistance

A superbug resistant to every available antibiotic in the U.S. kills Nevada woman

79424









BY HELEN BRANSWELL, STAT January 13, 2017 at 10:01 AM EST



(HealthDay)—A recent multidrug-resistant (MDR) Salmonella enterica serotype Newport outbreak, affecting patients in 32 states, was associated with soft cheese and beef consumption, according to a report published in the Aug. 23 issue of the U.S. Centers for Disease Control and Prevention Morbidity and Mortality Weekly Report.

Rare strain of E. coli strikes across Canada: source unknown

BY NEWS DESK | JANUARY 14, 2017

A dozen cases of E. coli O121 have been confirmed in three Canadian provinces, according to matching genetic fingerprint data, but the source of the outbreak has not yet been identified.

The Public Health Agency of Canada reports four of the rare O121 cases were confirmed in British Columbia, four in Saskatchewan and four in Newfoundland and Labrador. The illness onset dates were in November and December of 2016.

Four of the v Those indivi the process o determine th coli O121 cor

The Public H improper has eating raw gr common sou common sou





Photo illustration

CDC says outbreak traced to raw clover sprouts has come to an end

By News Desk on April 23, 2020

The Food and Drug Administration's investigation of an outbreak of E. coli 0103 in clover sprouts has been completed, and the Centers for Disease Control and Prevention has declared that the outbreak is over.

The FDA, along with CDC and state and local partners, investigated 51 illnesses

Epidemiology of Foodborne Diseases

• Based on data from 1990s: (Mead et al., 1999)

76 million illnesses, 323,000 hospitalizations, **5,200 deaths** in the United States.

• More recent estimates show: (Scallan et al., 2011)

47.8 million illnesses, 127,839 hospitalizations, and more than **3,037** deaths in the United States.

- 9.4 million illnesses, 55,961 hospitalizations, and 1,351 deaths are cause by 31 known foodborne agents.
- In addition to consumer insecurity, foodborne diseases cause around \$77.7 billion for losses in productivity and economical losses.
- Approximately 30% of population are especially "at risk" for foodborne diseases (The **YOPI**'s: The young, the old, Pregnant, and Immunocompromised)

Significant foodborne pathogens... based on Mead et al., 1999 and Scallan et al., 2011 studies

- Leading etiological agents for illnesses: *Norovirus* (58%), Nontyphoidal *Salmonella* serovars (11%), *Clostridium perfringens* (10%), and *Campylobacter* spp (9%).
- Leading etiological agents for hospitalization: Nontyphoidal Salmonella serovars (35%), Norovirus (26%), Campylobacter spp (15%), and Toxoplasma gondii (8%).
- Leading etiological agents for death: Nontyphoidal *Salmonella* serovars (28%), *T. gondii* (24%), *Listeria monocytogenes* (19%), and *Norovirus* (11%).

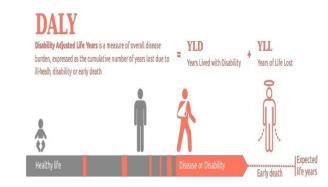
Signs and Symptoms of Foodborne Diseases

- Mild illness (no medical care sought)
- Guillain–Barré syndrome (Campylobacter and Salmonella)
- Post-infectious irritable bowel syndrome (Campylobacter and Salmonella)
- Reactive arthritis (Campylobacter and Salmonella)
- Haemolytic uraemic syndrome (E. coli O157)
- End-stage renal disease (E. coli O157)
- Death

Significant Foodborne Pathogens of Public Health Concern: Considering DALY and QALY (Scallan et al., 2015)

- Disability Adjusted Life Year (DALY). Loss of life and health due to illness compared with 'perfect' health
- Non-typhoidal Salmonella (329000)
- Toxoplasma (32700)
- Campylobacter (22500)
- Norovirus (9900)
- Listeria monocytogenes (8800)
- Clostridium perfringens (4000)
- Escherichia coli O157 (1200)

62% bacterial agents; 29% parasitic agents; 9% viral agents



- Mild illness (no medical care sought)
- Guillain–Barré syndrome (Campylobacter and Salmonella)
- Post-infectious irritable bowel syndrome (Campylobacter and Salmonella)
- **Reactive arthritis** (*Campylobacter* and *Salmonella*)
- Haemolytic uraemic syndrome (*E. coli* O157)
- End-stage renal disease (E. coli O157)
- Death

CDC Estimates of Food Safety Burden

http://www.cdc.gov/foodborneburden/attribution-image.html#foodborne-illnesses

Toxoplasma and Vibrio vulnificus.



Are these outbreaks associated with corporates and lager manufactures?

Prevalence of Pathogens in Medium-sized Poultry Operations

• 200–300 ft houses, 3000 to 5000 birds, conventional operation

(Alali et al., 2010)

	Salmonella serovars
Fecal samples (n=420)	38.8%
Feed (n=140)	27.5%

• Total of 135 sample from commercial free-range chicken producers (Bailey et al., 2005)

	Salmonella serovars		
Chicken Carcasses in Operation 1	64%		
Chicken Carcasses in Operation 2	31%		

Prevalence of Pathogens in Small Poultry Farms

 Study of 60 Small poultry slaughterhouses (fewer than 200 birds slaughtered per day)

Sampling sites	Salmonella serovars (Albany, Hadar, Indiana, and Enteritidis sub-species)
Carcasses after slaughter	42%
Utensils	23.1%
Storage freezers and refrigerators	71.4%

 The Study concluded "The widespread occurrence of Salmonella in small slaughterhouses reinforces the need for implementation of effective control measures..."

Climate Change and Public Health Microbiology

Non-typhoidal Salmonella enterica serovars

- o **Global death:** 50,000 global death in 2010 (WHO, 2020)
- Public Health Burden in the U.S.: >1 million annual cases in 2011 (CDC, 2011)

Climate Change:

- 1 °C increase : 5 to 10% increases in Salmonellosis (WHO, 2010)
- o 2500 to 5000 additional global death
- 50,000 to 100,000 U.S. morbidity

At our current rate:

- \circ >1.5 °C by 2040
- o >4.8 °C by 2100







Editoria

Safety of Food and Water Supplies in the Landscape of Changing Climate

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In response to evolving environmental, production, and processing conditions, microbial communities have tremendous abilities to move toward increased diversity and fitness by various pathways such as vertical and horizontal gene transfer mechanisms, biofilm formation, and quorum sensing [1,2]. As such, assuring the safety of water and food supplies from various natural and anthropogenic microbial pathogens is a daunting task and a moving target. Recent outbreaks of Listeria monocytogenes in South Africa associated with a ready-to-eat product (affecting close to 1000 individuals) and the 2018 outbreak of Shiga toxin-producing Escherichia coli O26 associated with ground meat in the United States (leading to the recall of more than 132,000 pounds of products) are bitter reminders of the devastating influences of foodborne diseases on the public health and food manufacturing [3,4].

Recent epidemiological studies of world populations indicate that 420,000 people lose their lives every year due to foodborne diseases, with around one-third of those being 5 years of age or younger. It is further estimated that every year, 1 in 10 individuals experience foodborne diseases around the globe, leading to an annual loss of 33 million healthy life years [5]. These episodes of food and water

Main Bacterial Pathogens Associated with Animal and Human Health Diseases

Foodborne Pathogens of Public Health Concerns >200 foodborne diseases

- Salmonella serovars
- Staphylococcus aureus
- Campylobacter spp.
- Bacillus cereus
- Shiga Toxin-Producing Escherichia coli (STEC)
- Vibrio spp.
- Yersinia enterocolitica
- *Streptococcus* spp.
- Shigella spp.
- Listeria monocytogenes
- Mycobacterium bovis
- Cronobacter sakazakii





Salmonella serovars

- Annual illness (death): 1,027,561 (378) in American adults and children
- Infection causes nausea, vomiting, diarrhea, fever, headache
- **Primary sources**: Intestinal tract of people and animals
- **Transmitted by** meat, poultry, eggs, raw milk, unpasteurized juice, many other foods (nuts, spices, produce, chocolate, flour) [Low-moisture environment]
- Contributing factors: cross-contamination, undercooked food, poor agricultural practices

Growth parameters	Minimum	Optimum	Maximum	
Temperature	41°F (<mark>5.2°C</mark>)	95-109°F (35-43°C)	115°F (46.2°C)	
рН	3.7	7-7.5	9.5	
a _w	0.94	0.99	>0.99	
Other	Non-spore former			
Atmosphere	Facultative - grows with or without oxygen			

Sources: ICMSF 1995 and Bad Bug Book 2nd edition, Scallan et al., 2011, and FSPCA

Salmonella serovars

- Carriers: Reptiles (turtles, lizards, and snakes); Amphibians (frogs and toads);
 Poultry (chicks, chickens, ducklings, ducks, geese, and turkeys); Other birds (parakeets, parrots, and wild birds); Rodents (mice, rats, hamsters, and guinea pigs); Other small mammals (hedgehogs); Farm animals (goats, calves, cows, sheep, and pigs); Dogs; Cats; Horses. [Pretty much ubiquitous!]
- Dogs and cats that become ill from Salmonella infection generally will have diarrhea that may contain blood or mucus
- Some cats do not have diarrhea, but will have a decreased appetite, fever, and excess salivation.

Prevention:

• Minimizing direct contact, washing hands, and cleaning up after the pets could minimize the risk of transmission from infected animals to human.

Staphylococcus aureus

Foodborne Diseases

- Infection
- Intoxication
- Toxico-infection

- Annual illness (death): 241,148 (6) Americans every year
- Both causes infection and toxico-infection
- Produces heat stable toxins after extensive growth
- Primary sources: Boils, nasal passages and skin (around 20% positive on nasal passage, >10% hands)
- Transmitted by recontaminated cooked foods, and foods with high salt or high sugar (Gram-positive, poor competitor)
- Contributing factors: Recontamination and time/temperature abuse

Growth Minimum C		Opt	timum	Maximum		
parameters	Growth	Toxin	Growth	Toxin	Growth	Toxin
Temperature	45°F (7°C)	50°F (10°C)	99°F (37°C)	104-113°F (40-45°C)	122°F (50°C)	118°F (48°C)
рН	4	4	6-7	7-8	10	9.8
a _W	0.83	0.85	C).98	>0.	99
Other	Poor competitor, non-sporeformer					
Atmosphere	Facultative –	grows with	n or without	oxygen, but s	slower with	out

Sources: ICMSF 1995 and Bad Bug Book 2nd edition, Scallan et al. 2011, and FSPCA

Staphylococcus aureus

- Methicillin-resistant Staphylococcus aureus (MRSA) a major concern for animal and human health.
- Humans working closely in animal feeding operations are in elevated risk of exposure to this pathogen particularly the antibiotic resistant phenotypes
- A contagious bacterium responsible for vast majority of environmental Mastitis in dairy operations.
- Cause of "bumblefoot" in chickens
- A major pathogen of farm rabbits



Campylobacter spp.

- Annual illness (death): 845,024(76)
- Infection causes diarrhea, and potential nerve damage
- **Primary sources**: Intestinal tract of animals
- Transmitted by raw poultry, raw milk products, contaminated water, poultry (dump tank, nearly 80%). Relatively high infective dose
- Contributing factor: cross contamination and undercooking

Growth parameters	Minimum	Optimum	Maximum
Temperature	86°F (30°C)	108-109°F (42-43°C)	113°F (45°C)
рН	4.9	6.5-7.5	9.5
a _w	>0.987	0.997	-
Other	Non-spore former		
Atmosphere	3-5% oxygen optimum		

Sources: ICMSF 1995 and Bad Bug Book 2nd edition and FSPCA

Campylobacter spp.

- Bacterium exist in GI track of many healthy farm and companion animals.
- Dogs six week and younger are prone to Campylobacteriosis with symptoms:
 - -Fever
 - -Vomiting
 - -Loss of appetite
 - -Enlarged lymph nodes
- Staying in kennels that increases the exposure to fecal matter and contact with contaminated food and water are main sources of disease in dogs.

Bacillus cereus

- Annual illness (death): 63,400 (0)
- Produces toxins and extensive growth is required for illness
- **Primary source**: soil and GI track
- Transmitted by: rice and starchy foods, meats, vegetables, milk products, sauces
- Contributing factors: temperature abuse

Growth parameters	Minimum	Optimum	Maximum	
Temperature	39°F (4°C)	82-95° F (28-35°C)	131°F (55°C)	
рН	4.3	6.0-7.0	9.3	
a _W	0.92	-	-	
Other	Spore former; one toxin is heat stable			
Atmosphere	Facultative – grows with or without oxygen			

Sources: Seafood Hazards Guide, ICMSF 1995, Bad Bug Book, Scallan et al. 2011, and FSOCA

Bacillus cereus

- Some studies indicate the bacterium could behave as an agent of mammary gland infection in cows and goats thus causing mastitis.
- Cases of food poisoning in dogs and cats had also been reported, although not very frequent in nature.
- Many agricultural animals carry the bacterium in their intestinal area without symptoms.



Shiga Toxin-Producing Escherichia coli (STEC)

- Annual illness (death): 176,152 (20)
- Notable outbreak: 1992-1993 outbreak in pacific northwest- Very important regulatory status (adulterant)
- Infection causes bloody diarrhea, and sometimes kidney failure and death [HUS in kids]
- **Primary sources**: Intestinal tract of ruminant animals (e.g., cows, sheep)
- Transmitted by raw and undercooked beef, poultry, leafy greens, and unpasteurized milk and juices
- Contributing factors: poor GAP, inadequate heating, and person-to-person

Growth parameters	Minimum	Optimum	Maximum
Temperature	44°F (6.5°C)	95-104°F (35-40°C)	121°F (49.4°C)
рН	4	6-7	10
a _w	0.95	0.995	-
Other	Non-spore forming		
Atmosphere	Facultative - grows with or without oxygen		

Sources: ICMSF 1995 and Bad Bug Book 2nd edition, Scallan et al. 2011, and FSPCA

Shiga Toxin-Producing Escherichia coli (STEC)

- Animals that can spread E. coli O157 to humans include:
 - -cows, especially calves
 - -goats
 - -sheep
 - -deer
- E. coli infection very common in cats and puppies younger than one week.
- **Colostrum**, plays a pivotal role in protecting a newborn the animal's undeveloped immune system against *E. coli* infection.
- As high as 80% of agricultural animals could carry various serogroups of shiga-toxigenic E. coli
 without having symptoms

Vibrio spp.

- Causing about 80,000 illness and 100 death annually in the United States.
- Infection symptoms vary depending on strain, ranging from diarrhea to high fever
- Vibrio is a halophilic bacterium and is a major concern in aquaculture industry
- Primary sources: Salt water environments and seafood
- Requires salt to reproduce (halophile)

Growth parameters	Minimum	Optimum	Maximum	
Temperature	41°F (5°C)	99°F (37°C)	114°F (45.3°C)	
рН	4.8	7.8-8.6	11	
a_W	0.94	0.98	0.996 (10% NaCl)	
Other	Non-sporeformer, requires salt			
Atmosphere	Facultative - grows with or without oxygen			

Sources: Seafood Hazards Guide 2011, ICMSF 1995 and Bad Bug Book 2nd edition

Yersinia enterocolitica

- Not a reportable disease, no statistics available
- Infection causes abdominal pain, fever and diarrhea. May mimic appendicitis.
- **Primary sources**: Raw pork, raw milk
- Contributing factors: Cross-contamination between raw pork products and RTE foods

Growth parameters	Minimum	Optimum	Maximum
Temperature	30°F (-1.3°C)	77-99°F (25-37°C)	108°F (42°C)
рН	4.2	7.2	10
a _W	0.945	-	-
Other	Non-spore former, raw milk in fridge?		
Atmosphere	Facultative - grows with or without oxygen		

Sources: Seafood Hazards Guide, ICMSF 1995, and Bad Bug Book

Foodborne Streptococcus spp.

- Not a reportable disease, no statistics available (not part of active surveillance data of CDC)
- Infection causes sore throat, tonsillitis and fever
- Primary sources: Infected sites of humans and animals, raw milk
- Contributing factors: Infected workers handling food and consumption of raw milk or meat products.
- **Symptoms**: meningitis, sepsis, and pneumonia (>200,000 sepsis cases per year, not foodborne)
- Found in: cattle, horses, dogs, rabbits, guinea pigs and mice
- Important cause of mastitis in cows.

Growth parameters	Minimum	Optimum	Maximum
Temperature	50°F (<mark>10°C)</mark>	99°F (37°C)	<113°F (<45°C)
рН	4.8-5.3	7	>9.3
%NaCl (salt)	-	-	<6.5
Other	Non-sporeformer		
Atmosphere	Facultative - grows with or without oxygen		

Sources: Seafood Hazards Guide, ICMSF 1995, and Bad Bug Book

Shigella spp.

- Infection in humans causes diarrhea, which may be watery to bloody. The infection is also known as dysentery [Taxonomy similar to Salmonella serovars]
- **Primary sources**: Human and Animal intestinal tract
- Transmitted by fecal contamination from contaminated water or infected food.
- Clinical signs are rare in dogs and cats, mostly mild diarrhea.
- Many species such dogs, cats, rodents and nonhuman primates could carry the pathogen asymptomatically.

Growth parameters	Minimum	Optimum	Maximum
Temperature	43°F (6.1°C)	-	117°F (47.1°C)
рН	4.8	-	9.3
a _W	0.96	-	-
Other	Non-spore former		
Atmosphere	Facultative - grows with or without oxygen		

Sources: Seafood Hazards Guide 2011, ICMSF 1995 and Bad Bug Book 2nd edition

Mycobacterium bovis

- Infection causing respiratory symptoms and tuberculosis
- Primary sources: Cattle and raw milk
- Other source: bison, elk, and deer.
- Contributing factors: Lack of milk pasteurization and exposure to aerosols from infected animals
- Grows very slowly and under reduced oxygen (microaerophilic)
- The US has nearly eliminated M. bovis infection from cattle, over one million animal is tested for the bacterium by inspectors. [USDA FSIS]
- M. bovis can be found in wild animals such as bison, elk, and deer; uninfected cattle that come into contact with these wild animals can become infected.

Listeria monocytogenes

- Infection causes severe illness in susceptible people mortality 15-30%
- **Primary sources**: Occurs widely in agriculture (soil, plants and water) –(Important during pregnancy)
- Transmitted by: Refrigerated RTE foods that support growth (South Africa, Largest in History in 2018)
- Contributing factors: Environmental pathogen spread by environmental contamination, equipment, people, incoming raw ingredients (ubiquitous in nature)
- Common in domesticated ruminates particularly sheep, poultry, and birds.
- Could cause sporadic and farm outbreaks in ruminants
- Could cause: Encephalitis, late abortion, and GI problems in ruminants.

Growth parameters	Minimum	Optimum	Maximum
Temperature	31°F (-0.4°C)	99°F (37°C)	113°F (45°C)
рН	4.4	7.0	9.4
a _W	0.92	-	-
Other	Non-sporeformer		
Atmosphere	Facultative - grows with or without oxygen		

Sources: ICMSF 1995 and Bad Bug Book 2nd edition



Cronobacter Sakazakii

- Recently reclassified bacteria (2006-07), formerly known as Enterobacter sakazakii
- The Genus Cronobacter was derived from the Greek term "Cronos," a Titans of ancient mythology who swallowed each of his infants as soon as they were born (he was afraid to be replaced by his infants).
- The **species name**, **sakazakii**, is named in honor of the Japanese microbiologist, Riichi Sakazaki, when the bacterium was first explained in 1980.
- Gram-negative, rod-shaped bacteria.
- Facultative anaerobic
- The growing temperature range is 6°C-45°C
- Primarily associated with Powered Infant Formula
- There has been several outbreaks associated with the bacterium and neonatal meningitis and death including two outbreaks in Tennessee (1998 and 2001).

Cronobacter Sakazakii

Prevention (CDC & WHO guidelines)

- Breastfeed
- Practice careful hygiene
- Clean and sanitize properly
- Prepare Powered Infant Formula as recommended

Symptoms:

- poor feeding response,
- irritability,
- jaundice,
- grunting respirations,
- instability of body temperature,
- Could lead to: seizures, brain abscess, hydrocephalus, and developmental delay, or death



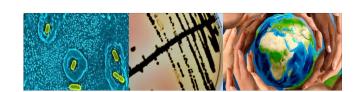
https://www.mdpi.com/2076-2607/7/3/77



Thank you



Food Safety Modernization Act Certification



Aliyar Cyrus Fouladkhah
Public Health Microbiology Laboratory
Cooperative Extension Program
Tennessee State University



Food Safety Modernization Act (FSMA)

- Signed to law in January of 2011, FSMA is the largest expansion of U.S. food safety authorities since the 1930s.
- Many sectors of agriculture and manufacturing will undergo strict regulations for the first time in the history of the country.
- Shifting responses from food safety problems to proactively prevent the episodes
- FSMA, a large and comprehensive legislation broaden FDA's ability to:
 - · Mandatory recall of contaminated food products
 - Enhanced surveillance to investigate foodborne illness outbreaks
 - Established **new preventive controls** and food safety plans at some food processing facilities and farms
 - Enhanced FDA's traceability capacity
 - Increased inspection frequencies of high-risk food facilities (both domestic and foreign facilities)
 - Expanded authority and oversight capabilities with regard to foreign companies

Regulatory Landscape of Food Industry Before FSMA

Very small companies:

Exemption from federal requirements, need to follow state policies

Restaurant operations:

Exemption from federal requirements, need to follow state policies (food code)

Food Safety Inspection Service (FSIS) of USDA:

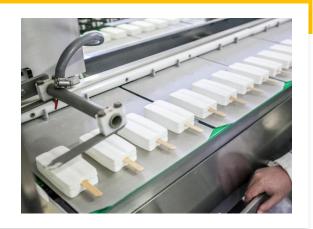
 ${\it Meat, Poultry and Egg products, HACCP requirements}$

Food and Drug Administration of DHHS:

High Risk Foods: Juices, seafood, and shell egg, HACCP requirements

Farmers and other food products:

No federal regulation



Mandated by FSMA

- Food manufacturing (processors)
- Farmers and growers (producers)
- Transportation, retailers
- Imported foods
- Third party laboratories
- Local, state, and federal agencies
- Foreign governments



Not mandated by FSMA

- FSMA does not directly address sectors under pre-existing jurisdictions.
 HACCP will remain the dominant regulation for:
- Meat, poultry, and egg products (USDA-FSIS)
- Juices, seafood, and shell eggs (DHHA-FDA)
- Very small producers and processors could receive exception from FSMA requirements (cottage industry).
- FSMA does not mandate GM products, antibiotic resistant organisms, organic production, and pesticide and fertilizer use.

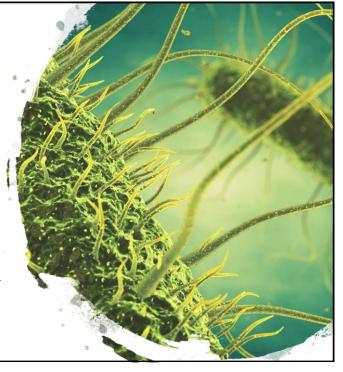
FSMA Implementation Schedule

FSMA was signed into law on January, 2011

Regulations were supposed to be finalized within one to two years of enactment (roughly **January 2012 and January 2013**)

Revised implementation dates: (all drafts are currently publically available)

- Preventative controls: FSMA §103(a) and(c): August 30, 2015
- Foreign supplier verification program: FSMA §301(a): October 31, 2015
- Accreditation of third party auditors: FSMA §307): October 31, 2015
- Produce safety Rule: FSMA §105(a): October 31, 2015 [Week 11+ Survey]
- Sanitary transportation practices for food and feed: FSMA §111: March 31, 2016
- Intentional adulteration of food: FSMA §106(b): May 31, 2016.



Produce and Preventive Rules and Land-grant Institutions

- Standards for the Growing, Harvesting, Packing, and Holding of Produce for Human Consumption (Produce Rule): Producers
- Current Good Manufacturing Practice and Hazard Analysis and Risk-Based Preventive Controls for Human Food (Preventive Rule): Processors
- Large producers and processors
- Small and medium size producers and processors
- Very small (hobbyists) producers and processors (local and cottage industry)
- Many of small and medium size entrepreneur will require assistance from the nations 75 land-grant institution for safe and economical access to market.



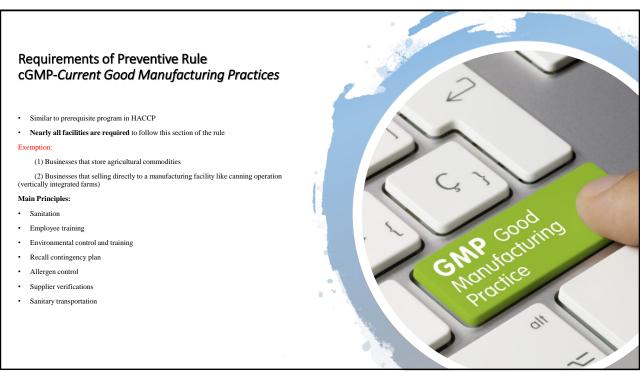
Preventive Control for Human Food Rule: Overview (PC QI)

- Regulate "processors"
- Under the regulation all "facilities" have to be registered with FDA
- The rule has two sections: Hazard Analysis (HARPc) and GMP, facilities obligated to have one or both.
- Exemptions: Juice, seafood, and shell egg sectors and businesses that store agricultural comities. (differs with preventive rule)

Modified Requirements:

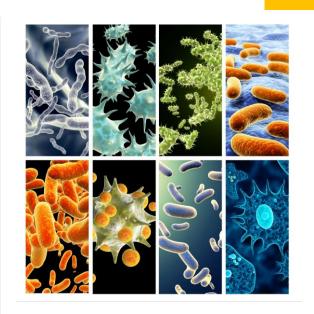
- Three-year average sales less than \$500K, AND
 - Direct sales to restaurants and consumers within 275 mile radius, or
 - Within states sales in 275 mile radius.



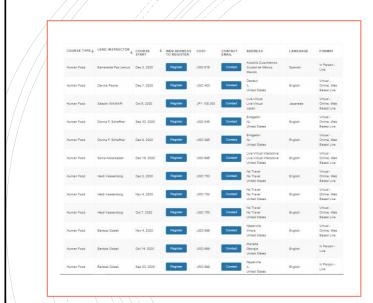


Requirements of Preventive Rule Hazard Analysis and Risk-Based Preventative Controls (HARPC)

- Previous a 7-step plan for FSIS HACCP, 12-step plan for Codex HACCP, and currently 5-step plan for HARPC:
- · Hazard analysis
- Identification and implementation preventive controls.
- Monitoring the performance of controls.
- Developing corrective actions for preventative deviation.
- **Verification and recordkeeping** of preventative controls effectiveness
- 2.5 day workshop Preventive Control Qualified Individuals (PC QI)







Preventive Control for Human Food: PC QI

- Our course 10-08-20202 to 10-29-2020
- Funded by USDA
- Animal Food PC QI:

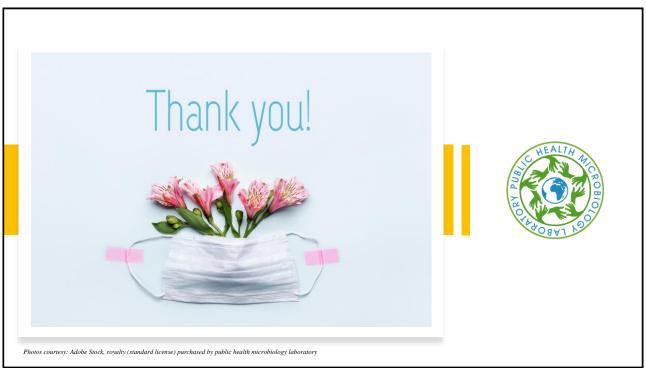
https://fspca.force.com/FSPCA/s/courselist?language=en_US



United States Department of Agriculture National Institute of Food and Agriculture









CERTIFICATE OF TRAINING

is awarded to

Dr. Ruth Climat

in recognition for having successfully completed the Food Safety Preventive Controls Alliance course:

FSPCA Preventive Controls for Human Food

delivered by Lead Instructor

Dr. Aliyar Cyrus Fouladkhah

completed on 11/27/2020

Robert Brackett. VP and Director Institute for Food Safety and Health



ILLINOIS INSTITUTE OF TECHNOLOGY



Certificate # c6db1b28

Mandernach, Executive Director ciation of Food and Drug Officials





CERTIFICATE OF TRAINING

is awarded to

Yves-Laurent Regis

in recognition for having successfully completed the Food Safety Preventive Controls Alliance course:

FSPCA Preventive Controls for Human Food

delivered by Lead Instructor

Dr. Aliyar Cyrus Fouladkhah

completed on 11/27/2020

Robert Brackett. VP and Director Institute for Food Safety and Health



ILLINOIS INSTITUTE OF TECHNOLOGY



Certificate #ff3d55c5

Mandernach, Executive Director ciation of Food and Drug Officials



Food Fortification Workshop





Fortification of Staple Commodities and Microbial Safety Requirements for Human Food Production

USAID F2F assignment: Haiti, Nashville, TN 12-7-2020

Tennessee State University, Nashville, TN

A. Fouladkhah: Faculty Director, Public Health Microbiology Laboratory





1

Presentation Content

Part I: Brief Introduction to my Program

Part II: Recent Food Safety Requirements

Part III: Dietary Guidelines and Fortifications









United States Department of Agriculture National Institute of Food and Agriculture





Part I: Brief Introduction to my Program

3

- Microbiology and Food Safety, PhD
- Applied Statistics and Data Analysis, Graduate Certificate
- Food Science & Human Nutrition, MS

Yale school of public health

- Biostatistics and Epidemiology, Advanced Professional MPH
- Food and Drug Regulatory Affairs, Graduate Certificate
- Climate Change and Health, Graduate Certificate







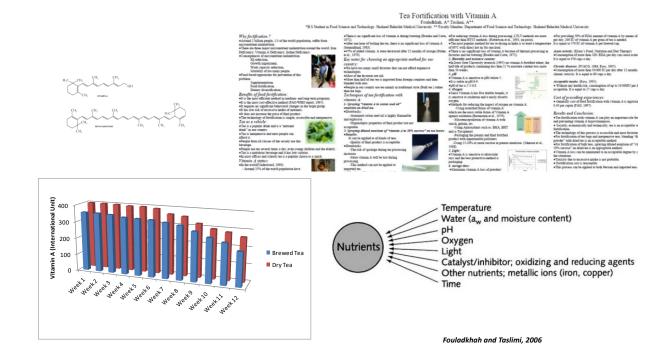






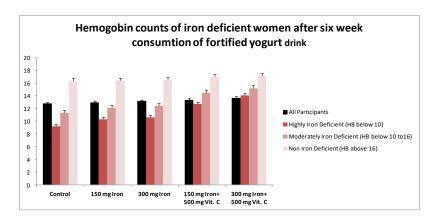


Website: https://publichealth.yale.edu/advanced/ Video: https://www.youtube.com/watch?v=IGVN9Jfolt



Fortification of Yogurt Drinks with Iron and vitamin C

• >53 increase in HB counts in 6 weeks



Fouladkhah et al., 2007

Public Health Microbiology Laboratory Tennessee State University

MPH Curriculum Food Safety and Applied Epidemiology

- Stablished in 2015:
- Currently \$7,000/year from Cooperative Extension Program
- * Extramural Funding
- ❖ National Institute of Health: \$33,680 (PD of Sub-award, 2020-21)*
- Pressure BioScience Inc.: \$35,000 (Role: PD, 2019-2024)
- USDA-NIFA CBG: \$350,000 (Role: PD, 2018-2022)
- USDA-NIFA HEC: \$50,000 (Role: PD, 2018-2021)
- USDA-NIFA FSOP: \$165,000 (Role: PD, 2018-2021)
- Pressure BioScience Inc.: \$23,500 (Role: PD, 2017-2019)
- USDA-NIFA FSOP: \$59,750 (Role: PD, 2016-2019)
- Pressure BioScience Inc.: \$9,400 (Role: PD, 2017-2019)
- ❖ USDA-NIFA FSOP.: \$880,000 (Role: CO-PD, 2019-2023)**
- ❖ USDA-NIFA FSOP.: \$1,197,751 (Role: CO-PD, 2015-2020)**
- USDA-NIFA CBG.: \$300,000 (Role: CO-PD, 2018-2022)





Website: https://publichealthmicrobiology.education/



PROSPECTIVE STUDENTS, EDUCATORS, AND STAKEHOLDERS



Website performance: 4/22/2020

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Public Health Microbiology Laboratory Current Members

- Current Graduate Students (Primary Advisor/Degree Chair: A. Fouladkhah):
- <u>Sadiye Aras (2018-),</u> Graduate Research Assistant, (PhD candidate, Biological Sciences c. Food Microbiology)
- <u>Jyothi George (2019-)</u>, Graduate Research Assistant (PhD student, Biological Sciences c. Food Microbiology)
- Sabrina Wadood (2020-), Graduate Research Assistant (MS student, c. Food Microbiology)
- Current Research Technician, Associates, and Interns (Primary Supervisor: A. Fouladkhah):
- <u>Shahid Chowdhury</u>, BS. Research Technician (2016-present)
- <u>Niamul Kabir</u>, PhD. Post-doctoral Research Associate (2018-present).
- <u>Amir Kashipazha, MS</u>. Data Visualization Intern and Web Editor (2018-present).
- Dr. Bagheri, PhD, MSPH, Visiting Scholar (2020-present).
- Current Graduate Student Committee:
- -Shreya Singh Hamal, PhD candidate, Biological Sciences con. Food Microbiology (Committee member).
- Yun Tian, PhD student, Biological Sciences con. Genomics & Immunology (Committee member).
- Zedonia Williams, MS student. M.S. degree in Food and Animal Sciences (Committee member).



Fall 2019 Graduates
Abimbola Allison, PhD
Tyler Keene, MS
Monica Henry-Smith, MS



^{*}Pending account setting and internal administrative approval.

^{**} Sub-awardee of Southern Center Main Awards



- Fall 2019 Graduates
- · Abimbola Allison, PhD
- Tyler Keene, MS
- Monica Henry-Smith, MS







Department of **Human Resources**

Tennessee Leaders of Tomorrow Internship

Monica Smith 1282 Vantage Pointe Rd – Apt. 304 Ashland City, TN 37015

Congratulations! You have been selected for an internship with Tennessee State Government Being selected for this internship is very competitive and is a great honor. So, be proud of your accomplishment! We are excited to have you as a member of our customer focused team. We hope that this opportunity provides you with the professional development experience that

Below are the details of your internship offer

Job Title: Tennessee Department of Agriculture - Intern

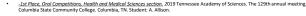
9

Students Awards

Students Success Available at:

https://publichealthmicrobiology.education/students-awards

Adviser: A. Fouladkhah



- -2nd Place, Oral Competitions, Health and Medical Sciences section, 2019 Tennessee Academy of Sciences. The 129th annual meeting, Columbia State Community College. Columbia, TN. Student: S. Aras.
- -1st Place (tied), Poster Competitions, Health and Medical Sciences section, 2019 Tennessee Academy of Sciences. The 129th annual meeting, Columbia State Community College. Columbia, TN. 2019 Summer Intern: A. Chowdhury
- -2nd Place, Poster Competitions, Health and Medical Sciences section. 2019 Tennessee Academy of Sciences. The 129th annual meeting, Columbia State Community College. Columbia, TN. Student: S. Aras.
- -2nd Place, Oral Graduate Competitions, 2019 Annual State-wide Competition for Food Safety Modernization Act, Food Safety, and Food Science Students. Tennessee (Volunteer) section Institute of Food Technologists. Student: A. Allison
- <u>-1st Place, Oral Graduate Competitions</u>, 2019 Annual State-wide Competition for Food Safety Modernization Act, Food Safety, and Food Science Students. Tennessee (Volunteer) section Institute of Food Technologists. Student: M. Henry
- -1st Place, Oral Competitions, Health and Medical Sciences section. 2018 Tennessee Academy of Sciences. The 128th annual meeting, Austin Peay State University, Clarksville, TN. Student: M. Henry
- -2nd Place, Oral Competitions, Health and Medical Sciences section. 2018 Tennessee Academy of Sciences. The 128th annual meeting
- -1st Place (tied), Graduate Poster Competitions, 2018 Annual State-wide Poster Competition for Food Safety Modernization Act, Food Safety, and Food Science Students. Tennessee (Volunteer) section Institute of Food Technologists. Student: M. Henry
- -1st Place, Undergraduate Poster Competitions, 2018 Annual State-wide Poster Competition for Food Safety Modernization Act, Food Safety, and Food Science Students. Tennessee (Volunteer) section Institute of Food Technologists. Student: B. Simpson.
- -2nd Place, Undergraduate Poster Competitions, 2018 Annual State-wide Poster Competition for Food Safety Modernization Act, Food Safety, and Food Science Students. Tennessee (Volunteer) section Institute of Food Technologists. Student: A. Sumlin.
- -2nd Place, Oral Competitions, Graduate Life and Physical Sciences Oral Division, 40th Annual University-Wide Research Symposium, Tennessee State University, Nashville, TN, 2018. Student: M. Henry
- -2nd Place, Oral Competitions, Graduate Life and Physical Sciences Division, 40th Annual University-Wide Research Symposium, Tennessee State University, Nashville, TN, 2018. Student: A. Allison.
- -1st Place, 2017 Tennessee Academy of Science, Health and Medical Sciences Student Oral Competition (Student: K. Sampson), Presentation: Sampson, K. Day, K., Allison, A., Chowdhury, S., Fouladkhah, A. 2017. Fate and Biofilm formation of Wild-type and rifampicin-resistant Coronobacter skazakii in biotic and abutice environments. Health and Medical Sciences section, 127th Meeting of Tennessee Academy of Science, November 17, 2017, University of Tennessee at Martin, Martin, Tennessee.
- -2nd Place, Oral Competitions, Graduate Biological Sciences Division, 39th Annual University-Wide Research Symposium, Tennessee State University, Nashville, TN, 2017. Student: A. Allison, PhD Student



2019 Tennessee Academy of Science Health and Medical Sciences Division

1st (A. Allison); 2nd (S. Aras); 3rd (M. Henry) Adviser: A. Fouladkhah



2018 Tennessee Academy of Science Health and Medical Sciences Division 1st (M. Henry); 2nd (A. Allison); 3rd (J. Adhikari) Adviser: A. Fouladkhah





A. Allison, Outstanding PhD Student in College of Agriculture, Receiving an Award from Dean Reddy.

Teaching in Tennessee and Internationally



2019 Evaluation:

- "Dr. Fouladkhah is an excellent professor. He does the absolute best job of making students feel comfortable making discussion in class and is exceptionally knowledgeable in the area of food sciences. The in class exercises are definitely helpful to make sure the lectures are being retained and assists in requiring little to no studying outside of the class meetings."
- "This course is top notch, one of the best courses I have ever taken, Much gratitude to the lead instructor Dr. Fouladkhah. I learned so much in the class and my knowledge on food policies and regulation has increased a thousandfold."

2018 Evaluation:

- "This man is so amazing. Learned so much in his class thank you Dr. Fouladkhah."
- "He is very helpful and always very encouraging. He helped me planned my studies and even future goals."

International Travel Reports Available at:

https://publichealthmicrobiology.education/international-programs





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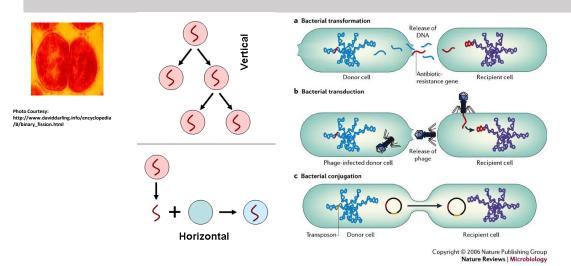


Part II: Recent Food Safety Requirements



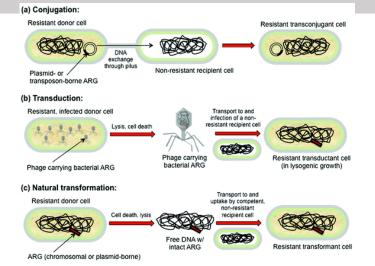
Emerging pathogens

Vertical and Horizontal Gene Transfer and Emerging Pathogens



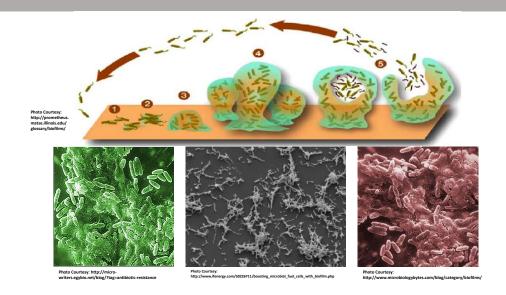
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Horizontal Gene Transfer



Donn, 2012

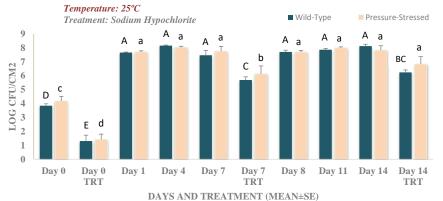
Planktonic cells and Biofilm Communities



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Cronobacter sakazakii Two outbreaks in Tennessee (1998, Memphis; 2001 Knoxville)

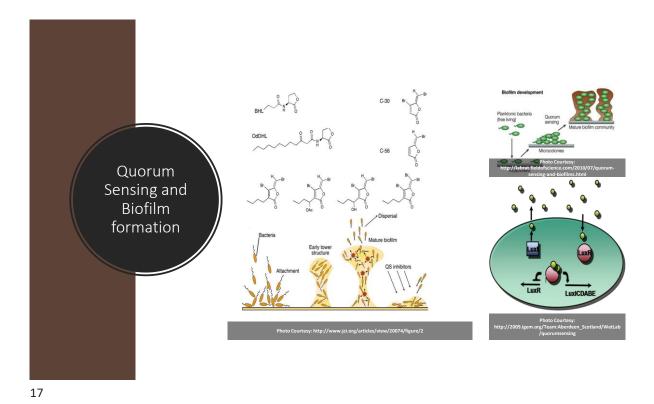
Biofilm Formation and Decontamination of Wild-Type and Pressure-Stressed *Cronobacter Sakazakii*





Allison et al., 2020





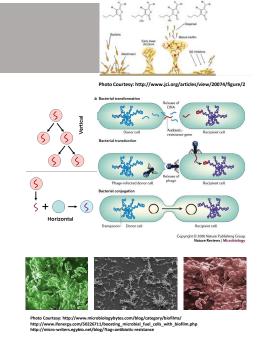
Infectious Diseases in Animals and Human is a Moving Target...

- It is estimated only 1% of microbial community has been identified.
- Currently etiological agent of 80.3% of foodborne illnesses, 56.2% of hospitalization, and 55.5% of deaths remain unknown.

"Emerging" Pathogens:

- Vertical and horizontal gene transfer spores and biofilm formation
- Quorum sensing and cell to cell communication

"It is the microbes who will have the last word."
-Louis Pasteur



Burden of Food Safety in the United States

• Based on data from 1990s: (Mead et al., 1999)

76 million illnesses, 323,000 hospitalizations, 5,200 deaths in the United States.

• More recent estimates show: (Scallan et al., 2011)

47.8 million illnesses, 127,839 hospitalizations, and more than **3,037** deaths in the United States.

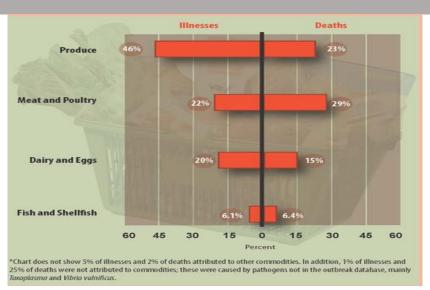
- 9.4 million illnesses, 55,961 hospitalizations, and 1,351 deaths are cause by 31 known foodborne agents.
- In addition to consumer insecurity, foodborne diseases cause around \$77.7 billion for losses in productivity and economical losses.
- Approximately 30% of population are especially "at risk" for foodborne diseases (The YOPI's: The young, the old, Pregnant, and Immunocompromised)



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CDC Estimates of Food Safety Burden

http://www.cdc.gov/foodborneburden/attribution-image.html#foodborne-illnesse:





Food Safety Modernization Act (FSMA)

- Signed to law in January of 2011, FSMA is the largest expansion of U.S. food safety authorities since the 1930s.
- Many sectors of agriculture and manufacturing will undergo strict regulations for the first time in the history of the country.
- Shifting responses from food safety problems to proactively prevent the episodes
- FSMA, a large and comprehensive legislation broaden FDA's ability to:
 - · Mandatory recall of contaminated food products
 - · Enhanced surveillance to investigate foodborne illness outbreaks
 - Established new preventive controls and food safety plans at some food processing facilities and farms
 - · Enhanced FDA's traceability capacity
 - Increased inspection frequencies of high-risk food facilities (both domestic and foreign facilities)
 - Expanded authority and oversight capabilities with regard to foreign companies

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Regulatory Landscape of Food Industry Before FSMA

Very small companies:

 ${\it Exemption from federal requirements, need to follow state} \\ policies$

Restaurant operations:

Exemption from federal requirements, need to follow state policies (food code)

Food Safety Inspection Service (FSIS) of USDA:

Meat, Poultry and Egg products, HACCP requirements

Food and Drug Administration of DHHS:

High Risk Foods: Juices, seafood, and shell egg, HACCP requirements

Farmers and other food products:

No federal regulation



Mandated by FSMA

- Food manufacturing (processors)
- Farmers and growers (producers)
- Transportation, retailers
- Imported foods
- Third party laboratories
- Local, state, and federal agencies
- Foreign governments



Not mandated by FSMA

- FSMA does not directly address sectors under pre-existing jurisdictions.
 HACCP will remain the dominant regulation for:
- Meat, poultry, and egg products (USDA-FSIS)
- Juices, seafood, and shell eggs (DHHA-FDA)
- Very small producers and processors could receive exception from FSMA requirements (cottage industry).
- FSMA does not mandate GM products, antibiotic resistant organisms, organic production, and pesticide and fertilizer use.

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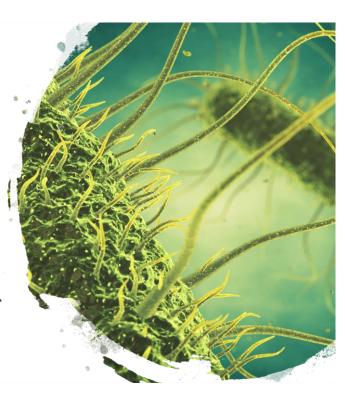
FSMA Implementation Schedule

FSMA was signed into law on January, 2011

Regulations were supposed to be finalized within one to two years of enactment (roughly January 2012 and January 2013)

Revised implementation dates: (all drafts are currently publically available)

- Preventative controls: FSMA §103(a) and(c): August 30, 2015
- Foreign supplier verification program: FSMA §301(a): October 31, 2015
- Accreditation of third party auditors: FSMA §307): October 31, 2015
- Produce safety Rule: FSMA §105(a): October 31, 2015 [Week 11+ Survey]
- Sanitary transportation practices for food and feed: FSMA §111: March 31, 2016
- Intentional adulteration of food: FSMA §106(b): May 31, 2016.





Produce and Preventive Rules and Land-grant Institutions

- Standards for the Growing, Harvesting, Packing, and Holding of Produce for Human Consumption (Produce Rule):
- Current Good Manufacturing Practice and Hazard Analysis and Risk-Based Preventive Controls for Human Food (Preventive Rule): Processors
- · Large producers and processors
- Small and medium size producers and processors
- Very small (hobbyists) producers and processors (local and cottage industry)
- Many of small and medium size entrepreneur will require assistance from the nations 75 land-grant institution for safe and economical access to market.



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Preventive Control for Human Food Rule: Overview (PC QI)

- Regulate "processors"
- Under the regulation all "facilities" have to be registered with FDA
- The rule has two sections: Hazard Analysis (HARPc) and GMP, facilities obligated to have one or both.
- Exemptions: Juice, seafood, and shell egg sectors and businesses that store agricultural comities. (differs with preventive rule)

Modified Requirements:

- · Three-year average sales less than \$500K, AND
 - Direct sales to restaurants and consumers within 275 mile radius, or
 - Within states sales in 275 mile radius.



Requirements of Preventive Rule cGMP-Current Good Manufacturing Practices

- · Similar to prerequisite program in HACCP
- · Nearly all facilities are required to follow this section of the rule

Exemption

- (1) Businesses that store agricultural commodities
- (2) Businesses that selling directly to a manufacturing facility like canning operation (vertically integrated farms)

Main Principles:

- Sanitation
- · Employee training
- · Environmental control and training
- Recall contingency plan
- Allergen control
- · Supplier verifications
- · Sanitary transportation



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Requirements of Preventive Rule Hazard Analysis and Risk-Based Preventative Controls (HARPC)

- Previous a 7-step plan for FSIS HACCP, 12-step plan for Codex HACCP, and currently 5-step plan for HARPC:
- · Hazard analysis
- Identification and implementation preventive controls.
- · Monitoring the performance of controls.
- Developing corrective actions for preventative deviation.
- Verification and recordkeeping of preventative controls effectiveness
- 2.5 day workshop Preventive Control Qualified Individuals (PC QI)





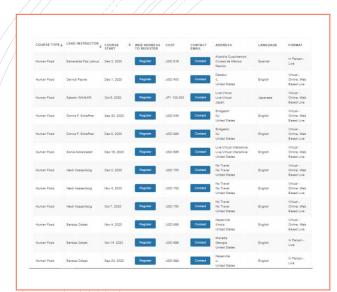
- Implementation date: August 30, 2015
- Compliance date:
 - Very small facility (\$2.5*m and below): 3 year
 - Small facility (less than 500 <u>employee</u> and does not qualified for exception): <u>2 years</u>
 - "Other" facilities: 1 years

Modified Requirements:

- Three-year average sales less than \$500K, AND
 - Direct sales to restaurants and consumers within 275 mile radius, or
 - · Within states sales in 275 mile radius.

*Total annual sale; the categories differ in preventive and produce rules.

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Preventive Control for Human Food: PC QI

• 2020 Workshops:

• Workshop 1: 10-08-20202 to 10-29-2020

Workshop 2: 11-23- to 25-2020

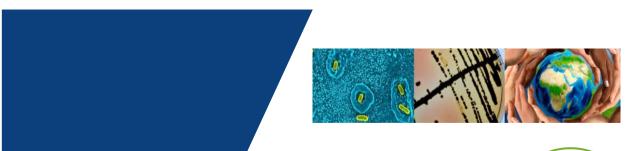
Funded by USDA



United States Department of Agriculture National Institute of Food and Agriculture



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<u>Part III:</u> Dietary Guidelines and Fortifications



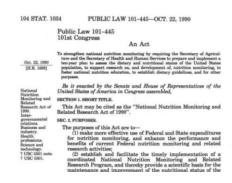


National Nutrition Monitoring and Related Research Act of 1990

- Poor diet and physical inactivity
- Epidemic of overweigh and obesity
 - · Men, women and children
 - · All segments of the society
- Poor diet and physical inactivity: a leading cause of premature morbidity and mortality
- Absence of harmonized national policy and guidelines for food, nutrition and health
- Lead to enactment of:

National Nutrition Monitoring and Related Research Act of 1990

(Public Law 101-445, Title III, 7 U.S.C. 5301 et seq.)



https://www.gpo.gov/fdsys/pkg/STATUT E-104/pdf/STATUTE-104-Pg1034.pdf

Oral Comment Meeting

On August 11, 2020, USDA and HHS hosted a virtual meeting to hear comments from the public on the Scientific Report of the 2020 Dietary Guidelines Advisory Committee. USDA and HHS will consider the Committee's Scientific Report, along with public and agency comments, as the Departments develop the next edition of the Dietary Guidelines.

Meeting Materials

View the <u>agenda</u> and <u>list of oral commenters</u>. The archived webcast is available below. The meeting transcript will be posted to this page once available.



View Archived Webcast

Advisory Committee Meetings

The 2020 Dietary Guidelines Advisory Committee's work is now complete. The Advisory Committee held 6 public meetings. Information from these meetings, including meeting materials and archived webcasts, can be found here.





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DGA main concepts

- (1) Maintain calorie balance over time to achieve and sustain a healthy weight (quantity of diet)
 - · Decrease the calories consumption
 - · Increase the calories expenditure through physical activity
- (2) Consuming nutrient-dense foods and beverages (quality of diet)
 - Reduction in consumption of:
 - Sodium,
 - Calories from solid fats, (Trans fatty acids, another great topic for the paper)
 - Added sugars,
 - · Refined grains.
 - Increase in consumption of:
 - Vegetables, fruits, and whole grains,
 - Fat-free or low-fat milk and milk products,
 - Seafood, lean meats and poultry, eggs,
 - · Beans and peas, and nuts and seeds.



Foods and Nutrients to Increase Recommendations Specific Age Groups

Women capable of becoming pregnant

- Choose foods that supply heme iron, which is more readily absorbed by the body, additional iron sources, and enhancers of iron absorption such as vitamin C-rich foods.
- Consume 400 micrograms (mcg) per day of synthetic folic acid (from fortified foods and/or supplements) in addition to food forms of folate from a varied diet.
- [Low childhood mortality in the United States]
- [Since 1998, folic acid (B9) required breads, cereals, pasta, flour, rice, cornmeal and other processed grain products] Whole Grain Cereals Vs. Processed Cereals ... Pizza Crust?

Women who are pregnant or breastfeeding

- Consume 8 to 12 ounces of seafood per week from a variety of seafood types.
- Due to their methyl mercury content, limit white (albacore) tuna to 6 ounces per week and do not
 eat the following four types of fish: tilefish, shark, swordfish, and king mackerel.
- If pregnant, take an iron supplement as recommended by an obstetrician or other health care
 provider.

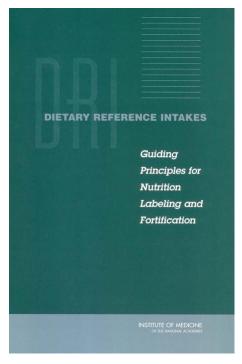
Individuals ages 50 years and older

· Consume foods fortified with vitamin B12, such as fortified cereals, or dietary supplements.









Iron Fortifications

- Substantial numbers of women who are capable of becoming pregnant, including adolescent girls, are **deficient in iron**.
- They can improve their iron status by choosing foods that supply heme iron, which is more readily absorbed by the body, as well as additional iron sources and enhancers of iron absorption such as vitamin C-rich foods.
- Sources of heme iron include lean meat and poultry and seafood.
- Sources of non-heme iron include: beans, lentils, and spinach, as well as foods enriched with iron, such as most breads and cereals.
- Non-heme iron is **not as readily absorbed** by the body.
- Women who are pregnant are advised to take an iron supplement as recommended by an obstetrician or other health care provider.







Testing Iron in Flour:

- 1. Qualitative Test (Spot Test):
- Method: Iron qualitative method, AACC Method 40-40
- Reagents: Thiocyanate reagent (Dissolve 10 g KSCN in 100 ml water); 2N HCl, Hydrogen peroxide, 3%.
- Equipment: none
- 2.A. Semi-Quantitative Test: (Spot Test):
- Method: Semi-quantitative spot test for iron INCAP method IV.
- Reagents: Hydrochloric acid, HCl; Hydrogen peroxide, H2O2; Potassium thiocyanate, KSCN; H2O2
- 2. B. Semi-Quantitative Test (Colorimetric method):
- Method: Semi-quantitative colorimetric determination of iron in flour INCAP method V
- Sodium acetate (CH3COONa.3H2O), 99%
- Hydrochloric acid (HCI), 37%
- a,a-dipyridyl (2,2' bipyridine)
- Bathophenanthroline, 4,7-diphenyl-1,10-phenanthroline-disulfonic acid (C24H16N2O6S
- Hydroxylamine hydrochloride (NH2OH.HCl)
- Iron standards: Electrolytic iron. Merck 3810 or Baker 2234







Testing Iron in Flour:

3.A. Quantitative Test (Spectrophotometric method):

- Method: Iron spectrophotometric method AACC method 40-41B
- Reagents:
- Orthophenanthroline solution (0. I g o-phenanthroline in about 80 ml water).
- Iron standard solution, (10 μg Fe/ml)
- Hydroxylamine hydrochloride solution (10 g NH2OH.HCl in water and dilute to 100 ml)
- Acetate buffer solution (8.3 g anhydrous sodium acetate in water)

${\bf 3.B.\ Quantitative\ Test\ Atomic\ absorption\ spectrophotometric\ (AAS)\ method:}$

- Method: Spectrophotometric analysis for quantitative, INCAP method VI
- Reagents:
- Sodium acetate (CH3COONa.3H2O), 99%
- Hydrochloric acid (HCI), 37%, 1.19 g/mL
- a,a-dipyridyl (2,2' bipyridine) (C10H8N2)
- Bathophenanthroline, 4,7-diphenyl-1,10-phenanthroline-disulfonic acid (C24H16N2O6S2)
- · Hydroxylamine hydrochloride (NH2OH.HCl)
- Iron standards: Electrolytic iron, Merck 3810





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General Requirements for Records

Form

Could be true copies or electronic

Content

- Actual values or observations
- Accurate, permanent (e.g., in ink) and legible
- Real time recording
- Adequate detail





Computerized Records

- Must be equivalent to paper records and hand written signatures
- An electronic record-keep system must:
 - Be authentic, accurate and protected
 - Provide accurate and complete copies of records
 - Protect records for later retrieval
 - Limit access to authorized individuals
 - Provide a secure record for audit trail
 - Be reviewed by a trained individual or PC QI





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Basic Information on Records





5 Implementation Records

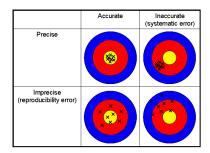
- (1) Monitoring records for preventive controls
- (2) Corrective action records
- (3) Verification records, when required
 - Validation
 - Verification of monitoring and corrective action
 - Calibration of monitoring and verification instruments
 - Product testing
 - Environmental monitoring
 - Records reviews
 - Reanalysis
- (4) Supply-chain program and supporting documentation
- (5) Training records, as appropriate



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Testing Considerations

- Considerations for calibration:
 - Design of the monitoring device
 - Reliability and sensitivity of the device (precision and accuracy)
 - Environment or conditions in which it is used



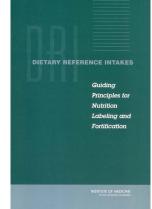


Product Testing Procedures Must...

Must be scientifically valid (AOAC, AOCS, BAM, USDA microbiology and

chemistry guidebooks)

- Must identify:
 - The microorganism or analyte
 - Relationship to lots (e.g. from geometrical center)
 - The number of samples, frequency, and analytical unit
 - Analytical test method
 - Laboratory
 - Corrective action procedures









Photos courtesy: Adobe Stock, royalty (standard license) purchased by public health microbiology laboratory

Recommendations





John Ogonowski and Doug Bereuter Farmer-to-Farmer Program

Volunteer Recommendations Form

Name of Volunteer:	Aliyar Cyrus Fouladkhah, PhD, MPH					
Country of Service:			14 to December 7, 2020			
	# of Persons Formally Trained – male: # of Persons Formally Trained – female:		6			
			7			
	# of Persons Formally Trained – Non-Binary:					
	# of Persons Formally Trained who are Youth:					
	# of Persons Formall	v Trained – total:	13			

Recommendations Made by the Volunteer:²

Please summarize the recommendations you made to the people/groups/organizations you assisted. Details of the recommendations should be included in the trip report – this is a summary table only.

Recommendation	Category*	Host	Time frame to implement change
Development, validation, and implementation of spot test for measuring iron content of the fortified product based on provided information.	1,3	RANFOSE and F2F Office in Haiti	Within two years
Incorporating food safety best practices based on information discussed in the workshops to assure regulatory compliance and safety of fortified products.	2	RANFOSE and F2F Office in Haiti	Within two years
Dissemination of the information discussed in the fortification workshop to relevant stakeholders.	2, 3	RANFOSE and F2F Office in Haiti	Within one year
Development of semi-quantitative and quantitative testing methodologies for measuring iron in flour-based commodities.	1,4	RANFOSE and F2F Office in Haiti	Within two years
Determining the stability of the fortificant during the shelf-life of flour-based products.	2, 4	RANFOSE and F2F Office in Haiti	Within two years

¹ **Persons Formally Trained:** number of persons who received technical/instructional training *in a "formal" setting:* classroom, workshop, institute/university or on-the-job setting with specific learning objectives and outcomes

^{**}Please review footnotes for definitions of "persons trained" and "persons directly assisted"**

² **Recommendations Made by the Volunteer:** The definition of "recommendation" is quite subjective, but might include an improved procedure, a technological or management innovation, a useful product or marketing tool, etc. Volunteers might make numerous detailed recommendations to a variety of hosts. Recommendations should be written in a way that is clear and measurable. *Please try to limit recommendations to no more than six per host.*

Development, optimization, and validation of laboratory best practices for ensuring effectiveness of testing based on provided information.	1,3	RANFOSE and F2F Office in Haiti	Within two years
Utilization of developed methods and consumables list for implementing new testing programs at field and laboratory levels.	3,4	RANFOSE and F2F Office in Haiti	Within two years
Dissemination of information about food quality control and food safety plan to relevant stakeholders to assure success of fortification programs.	2,3	RANFOSE and F2F Office in Haiti	Within one year
Expanding the currently in place fortification program to other staple commodities such as rice, table salt, and pasta.	1,2	RANFOSE and F2F Office in Haiti	Within five years

- * All recommendations should fall under one of four categories:
 - 1. Economic: improvement of profitability of the farm, business, or enterprise
 - 2. Organizational: improvement to organizational effectiveness, management, and sustainability
 - 3. Environmental: improvement of environmental management and natural resource conservation
 - 4. Financial: improvement in the provision of financial services