Cyanobacteria Harvesting and Treatment Simulations for Removal of Microcystin from Natural Waters

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Agenda

- Harvesting methods
- Concentrating cyanotoxins
- Treatment simulations with permanganates
- Treatment simulations with powdered activated carbon
- Desktop simulations with GAC
- Treatment simulations with chlorine
- Overall HAB planning and treatment activities
- Questions

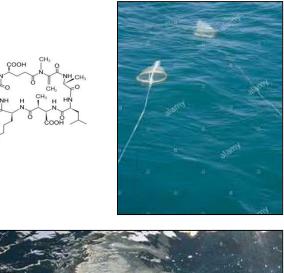
- Concentrated cyanotoxins can be prepared from natural surface water containing cyanobacteria
 - Apparent surface scums and growths
 - Direct microscopic analyses for presence in water
 - Cyanotoxin testing for presence in water
 - Generally water temperatures above 18°C
- Collect in phytoplankton net
 - Composite samples from source water into volume needed for treatment simulations
 - 1 liter or 2 liters
 - Preparation of cyanotoxin material
 - Analysis of toxin concentration



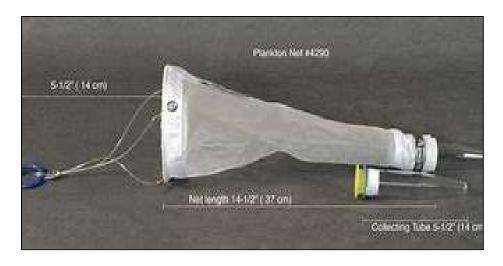


Relative abundance of cyanobacteria in some surface waters

- Drag net slowly by boat or offshore with pole
- Collect in phytoplankton net
 - Composite samples from source water into volume needed for treatment simulations
 - About 2 liters generally used for testing
 - Prepare concentrated cyanotoxin solution
 - Analyze toxin concentration









- Phytoplankton nets
 - Inexpensive device
 - Small mesh size (≈80 microns)
 - Three-point drag line
 - Collection bottle
- Net collects cyanobacteria
 - Rinse net with water to push cells downward into sample bottle
 - Pour cyanobacteria sample into composite container

Concentrating Cyanotoxins





- Mix composite sample thoroughly
- Freeze sample container
 - Thaw in warm water bath until liquified
 - Repeat at least 3 cycles
- Freeze/thaw cycles lyse cells and releases toxin material
- Mix final liquid and analyze toxin
 - Microcystin readily tested using customary methods

Concentrating Cyanotoxins

- Final concentrated solution appears dark green
 - Concentrated Microcystin level may range from 900 µg/L to 3,600 µg/L
- Concentrated solution applicable for jar testing dilutions
 - $C_1 V_1 = C_2 V_2$ then
 - $\bullet V_1 = C_2 V_2 / C_1$
 - Volume to add to jars



Concentrated Microcystin Solution

- Jar testing most common assessment method
 - Simulates specific treatment based on spiked water samples
 - Oxidant evaluations
 - $KMnO_4$, $NaMnO_4$, Cl_2 , ClO_2 , O_3 , H_2O_2 , etc.
 - Adsorption evaluations
 - Powdered activated carbon
 - Coagulation and softening impacts
- GAC adsorption evaluations
 GUD from W/DE (1009)
 - CUR from WRF (1998)
- Case Studies
 - 6 surface water plants



- Concentrated toxin solution used for jar tests
 - $\bullet C_1 V_1 = C_2 V_2$
- Dosages
 - 1 mg/L 5 mg/L
 - Residual half life in water is about 140 minutes
- Each of 5 jars spiked with ≈50 µg/L microcystin
 - 1 jar used as control (spike)
 - Remaining jars dosed with KMnO₄ or NaMnO₄ solution



Concentrated microcystin solution



KMnO₄ Oxidation for Microcystin Reduction - Attica



NaMnO₄ Oxidation for Microcystin Reduction - Lima



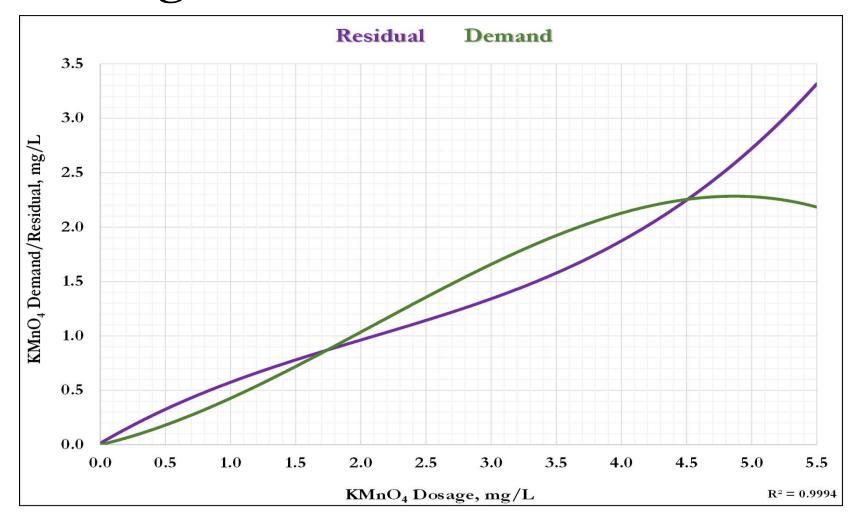
NaMnO₄ Oxidation for Microcystin Reduction - Defiance

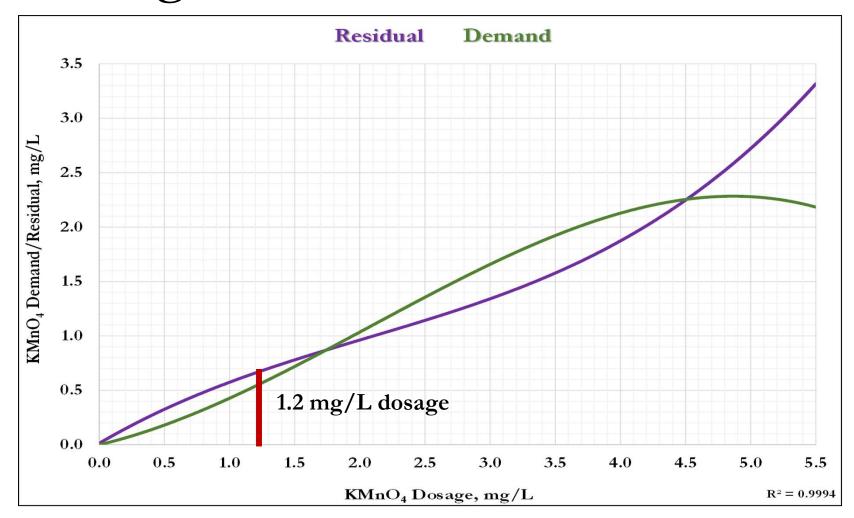


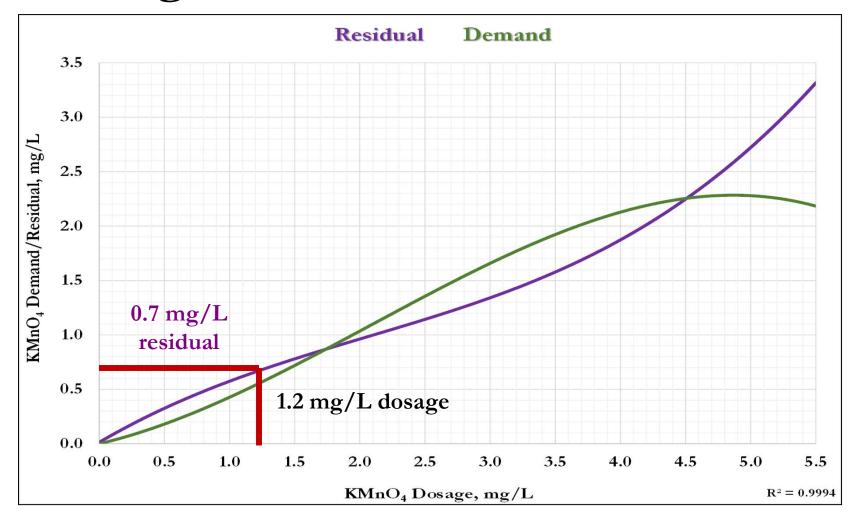
KMnO₄ Oxidation for Microcystin Reduction - Buffalo

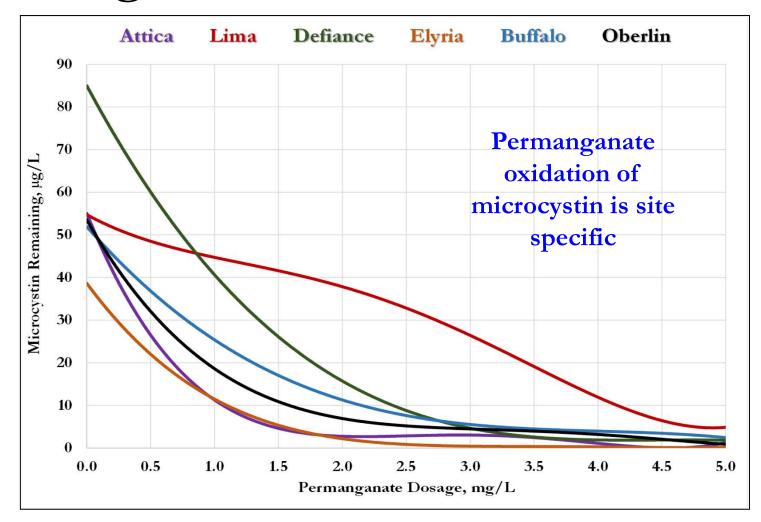
- Permanganate residual can be tested after about 10 minutes of reaction
 - DPD method provides oxidant residual in mg/L
 - Multiple by 0.89 to convert to permanganate residual, mg/L
 - Standard Methods technique and discussion
- Once residual and dosage are known, oxidant demand can be calculated



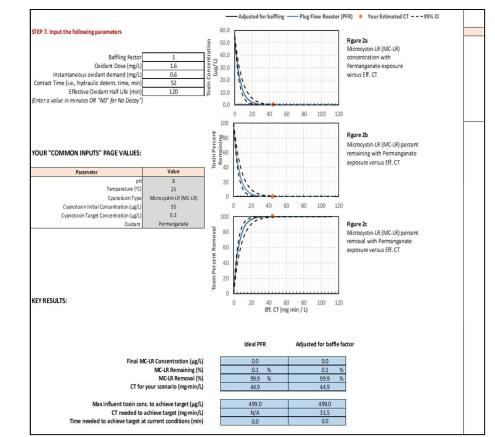




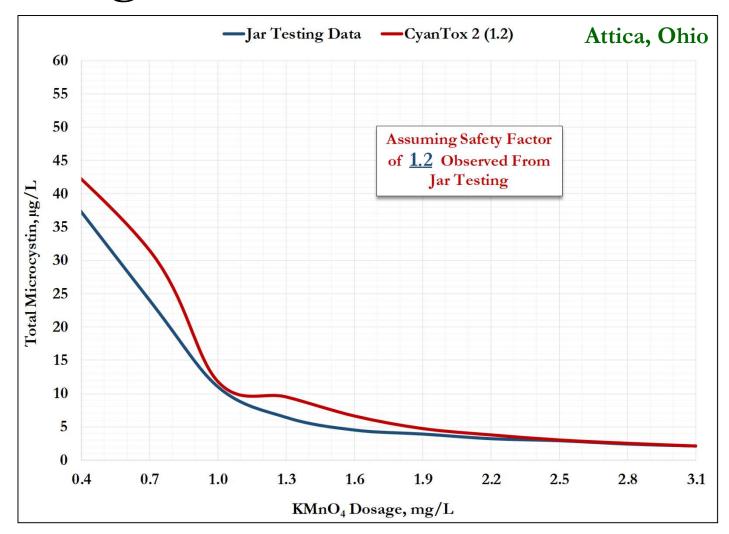


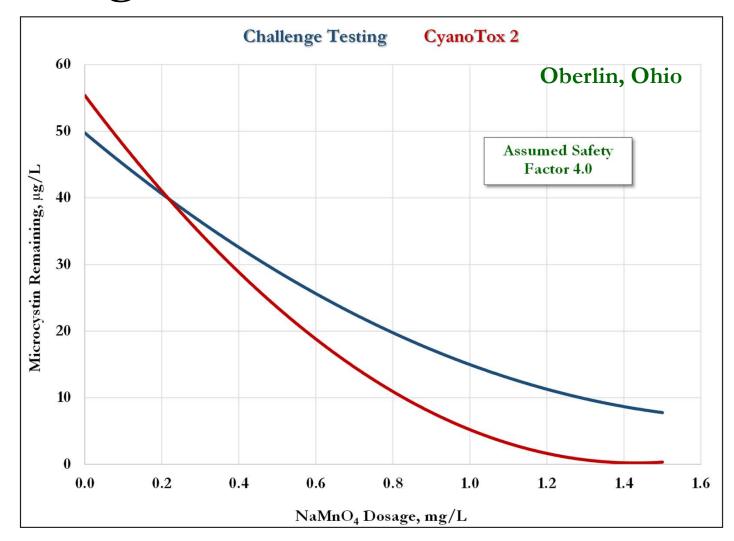


- Comparison of jar tests with AWWA CyanoTox2 Model for Cyanotoxins
 - 1st Assumed no safety factor
 - Input actual permanganate demand and dosage ranges
 - Compared model output with experimental data
 - 2^{cd} Identified approximate Safety Factor (site specific)



AWWA CyanoTox2 Model





- CyanoTox2 model appears to be useful for some KMnO₄ or some NaMnO₄ reactions for Microcystin removals
 - Does not correlate well for all water plants
 - Assume reasonable safety factor
 - Actual safety factor determined from correlations to bench-scale tests
- Model reaction kinetics based on KMnO₄ addition (Rodriguez, et al)
 - NaMnO₄ reaction kinetics likely differ from model equations
 - Assume no reasonable safety factor could be determined if computer modeling shows correlation factor greater than 2.5

- Permanganate oxidation effective for Microcystin removals
 - Dosage dependent
 - Reaction time dependent
 - KMnO₄ appears to be better than NaMnO₄
 - Typical residual range 0.15 mg/L to 0.5 mg/L
- Correlations to AWWA's CyanoTox 2 model possible for some treatment plants



- Powdered activated carbons (PAC) evaluated
 - Lignite-based (Hydrodarco W)
 - <u>500</u> iodine number, high mesopore volume
 - Wood-bituminous blend (WaterCarb 800)
 - <u>800</u> iodine number, high mesopore volume
 - WPH bituminous
 - <u>800</u> iodine number, high mesopore volume
 - WPH 1000 bituminous
 - <u>**1,000</u>** iodine number, high mesopore volume</u>
 - AquaSorb CB-1-W
 - <u>**1,000</u>** iodine number, high mesopore volume</u>

- Carbon solutions
 - Prepared from dry samples or obtained from slurry tank
 - Concentration must be known
 - Mix for at least 30 minutes to displace air from carbon pores
 - Generally dosed up to 60 mg/L
 - Simulate carbon contact time in full-scale treatment
 - Long contact times can be adjusted to full-scale if jar tests are shorter contact time



- Carbon slurry used for testing
- Carbon dosing
 - $\bullet C_1 V_1 = C_2 V_2$
- Each of 5 jars spiked with $\approx 50 \ \mu g/L$ microcystin
 - 1 jar used as control (spike)
 - Remaining jars dosed with carbon solution



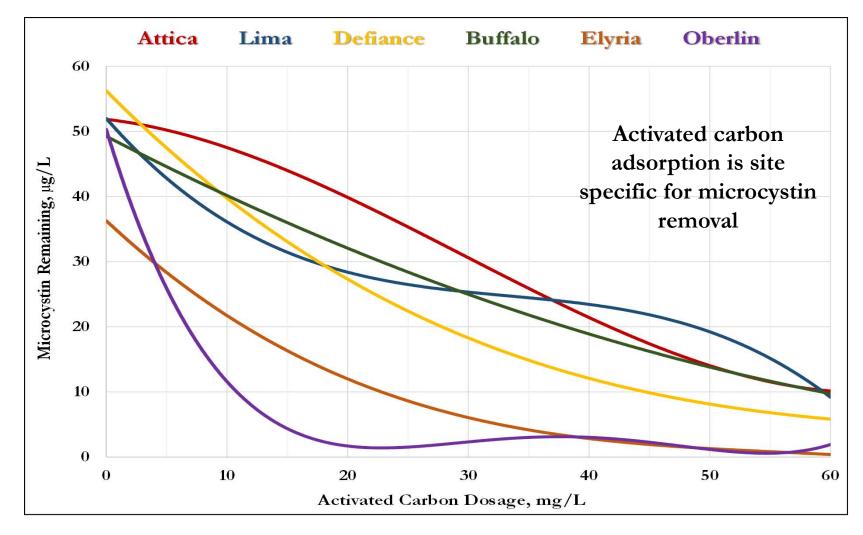
Concentrated Microcystin Solution

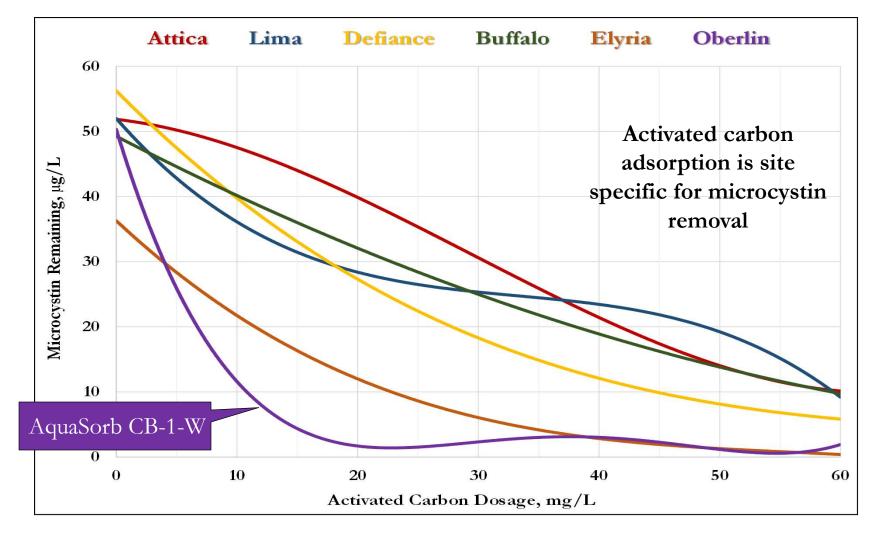


Activated carbon evaluation for Microcystin reduction - Attica

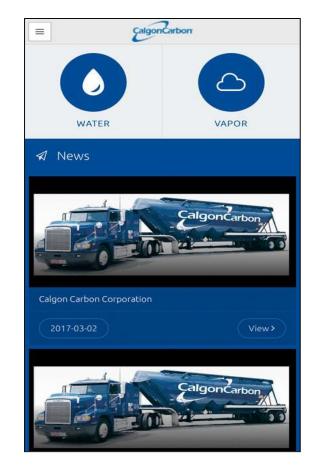


Activated carbon evaluation for Microcystin reduction - Buffalo





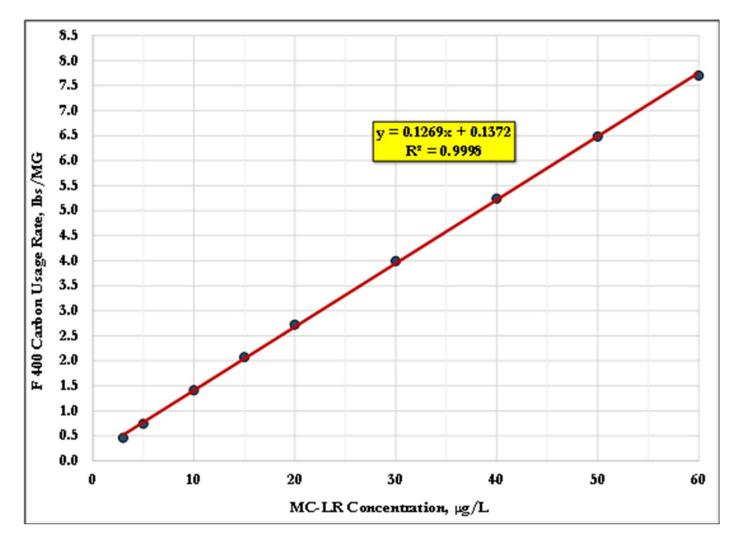
- GAC contactor desktop evaluations
 - Specific GAC designs
 - 10-min/20-min EBCT
 - Run to TOC breakthrough
 - Up to 2-year life cycle historically
- Carbon usage rate (CUR) for Microcystin-LR
 - CCURE app (assumed safety factor of 2)
 - 0.00648 pounds per 1,000 gallons treated @ 50 µg/L
 - 0.00141pounds per 1,000 gallons treated @ 10 µg/L



$$CUR, lbs / 1,000 gallons = \frac{EBCT * \rho GAC * 10^{3}}{T * 7.48 * 1,440}$$

Where CUR = Carbon Usage Rate, pounds per 1,000 gallons EBCT= empty bed contact time, minutes QGAC = carbon density, pounds/cubic foot 7.48 = 7.48 gallons per cubic foot 1,440 = 1,440 minutes per day

1998 WRF report - "Removal of DBP Precursors by GAC Adsorption"



34



- Microcystin that would likely enter <u>Lima's</u> GAC contactors
 - Pretreatment reduction with NaMnO₄ and activated carbon
- HAB event may reduce GAC life about 20 days every 2-year replacement cycle
 - Negligible impact to GAC treatment for TOC reduction
 - Effluent 0.3 µg/L Microcystin or less



- Microcystin that would likely enter <u>Defiance's</u> planned GAC contactors
 - Pretreatment reduction with NaMnO₄ and activated carbon
- HAB event may reduce GAC life about 5 days every 5month replacement cycle
 - Negligible impact to GAC treatment for TOC reduction
 - Effluent 0.3 µg/L Microcystin or less

GAC Treatment Simulations



- Microcystin that would likely enter <u>Elyria's</u> proposed GAC contactors
 - Pretreatment reduction with KMnO₄ and activated carbon
- HAB event may reduce GAC life about 3 days every 9month replacement cycle
 - Negligible impact to GAC treatment for TOC reduction
 - Effluent 0.3 µg/L Microcystin or less

GAC Treatment Simulations



- Microcystin that would likely enter <u>Oberlin's</u> potential GAC filter caps
 - Pretreatment reduction with NaMnO₄ and activated carbon
- HAB event may reduce GAC life about 1 day every 3-month replacement cycle
 - Negligible impact to GAC treatment for TOC reduction
 - Effluent 0.3 µg/L Microcystin or less

Carbon Treatment Simulations

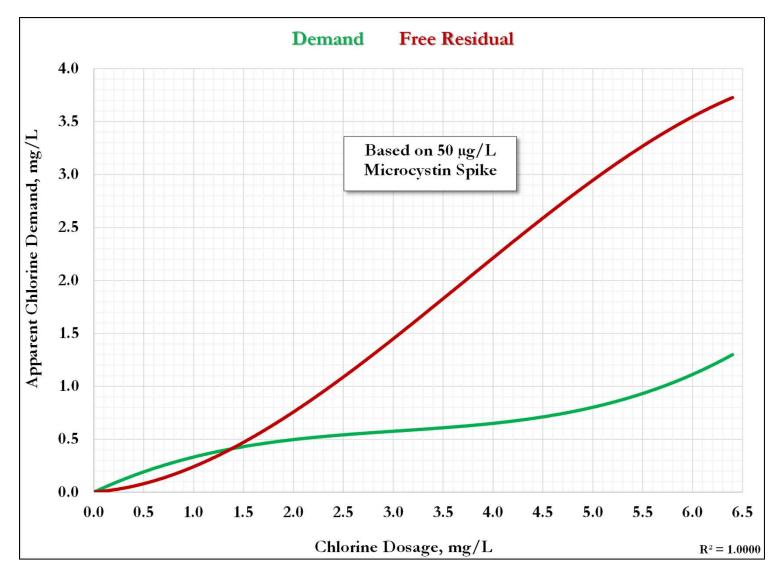
- Activated carbon adsorption effective for Microcystin reduction
 - Pore distribution dependent (carbon selection important)
 - Dosage and EBCT dependent
 - TOC dependent (organic character impacts)
 - PAC contact time dependent (60 minutes minimum)
 - High iodine number tends to reduce carbon dosages and produce greater removals
- Need to evaluate PAC impacts to filtration processes
 - Carbon carryover may limit actual dosages due to impacts identified
 - May limit carbon dosing to about 10 mg/L in conventional treatment
 - Could have minimal impacts if using <u>tube settlers</u> or <u>plate settlers</u>

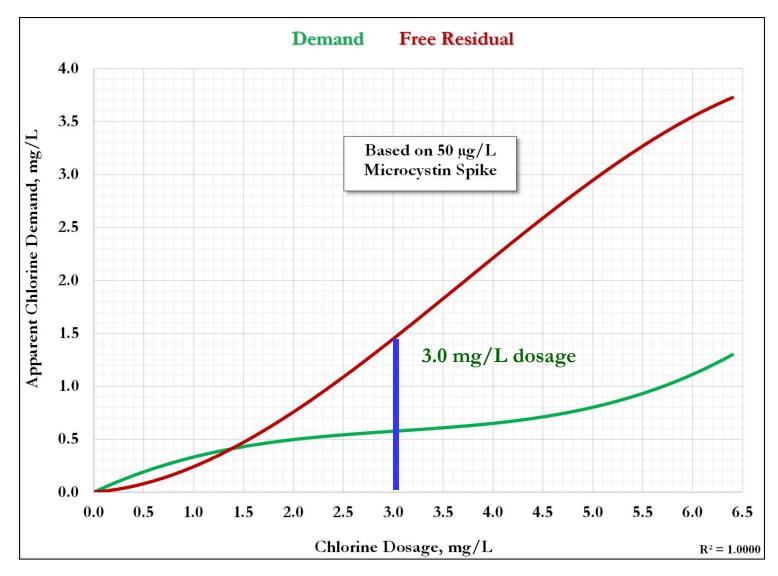
- Concentrated toxin solution used for jar tests
 - $\bullet C_1 V_1 = C_2 V_2$
- Dosages
 - 2 mg/L to 5 mg/L
 - Bracket current chlorine dosing
 - Filtered water used for simulations
- Each of 5 jars spiked with ≈50 µg/L microcystin
 - 1 jar used as control (spike)
 - Remaining jars dosed with chlorine solution



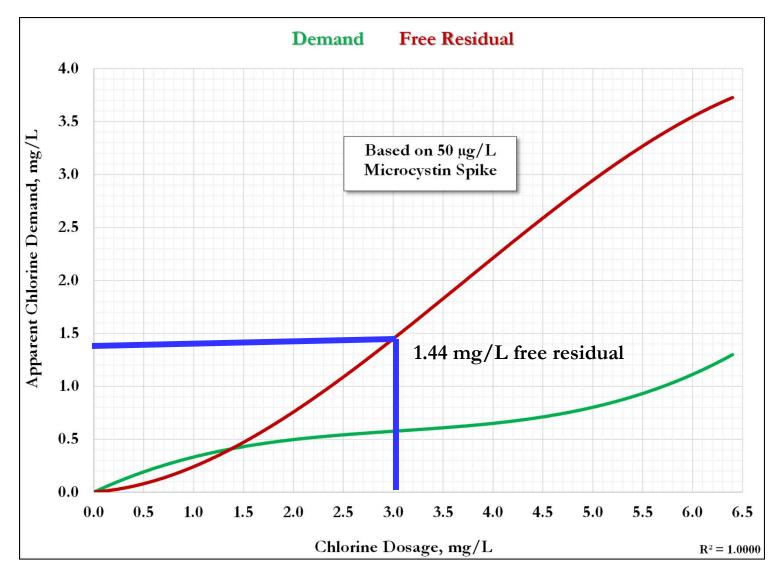
Concentrated microcystin solution

- Chlorine residual can be tested after about 10 minutes of reaction
 - DPD method provides chlorine residuals in mg/L
 - Free chlorine and total chlorine needed for evaluations
 - Free chlorine allows calculation of CT values for microcystin removal
- Once residual and dosage are known, chlorine demand can be calculated
- CT simulations demonstrate potential impacts of pH and contact time on microcystin reductions

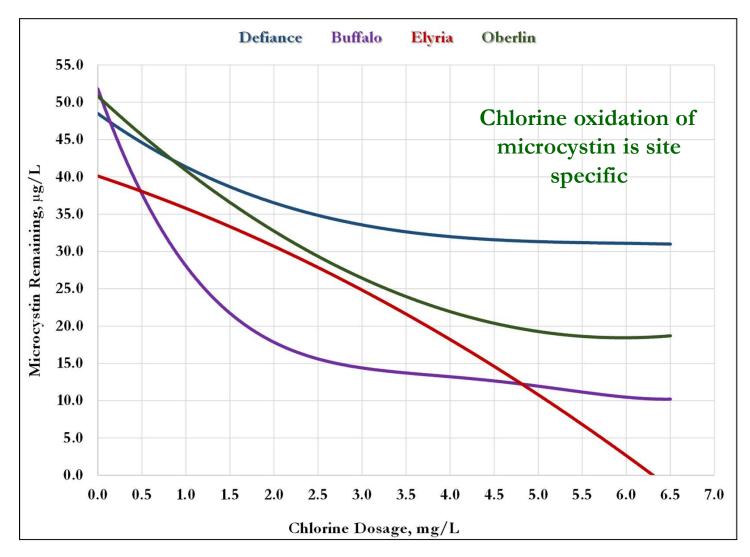


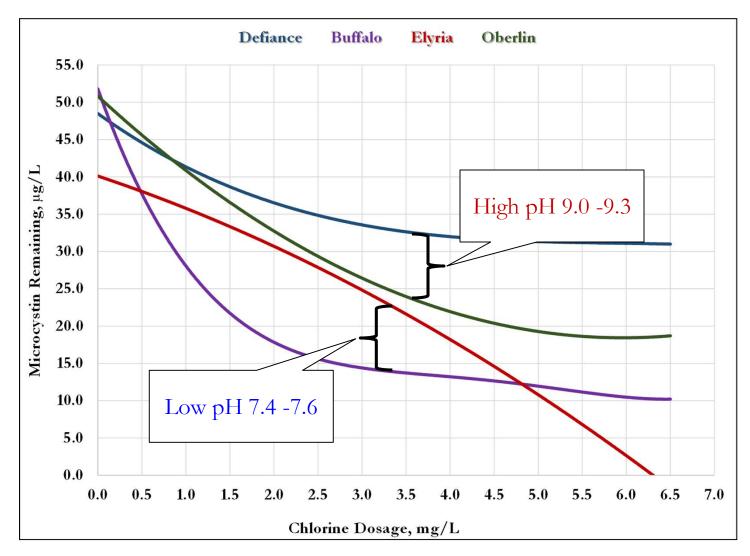


43

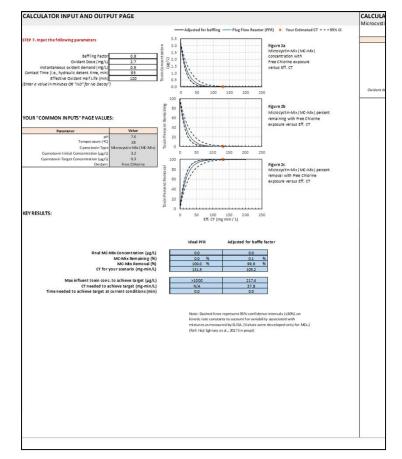


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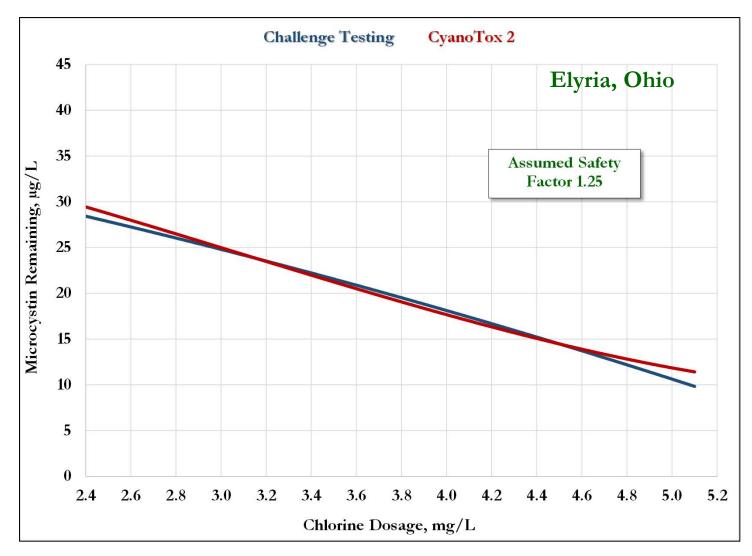


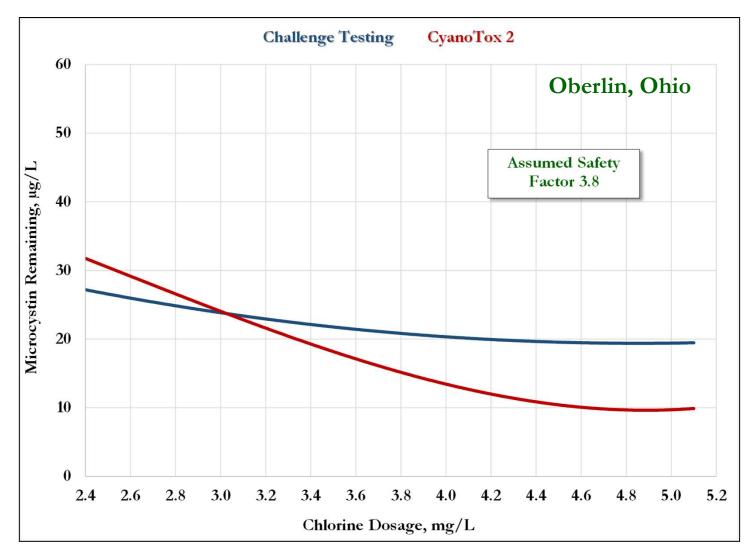


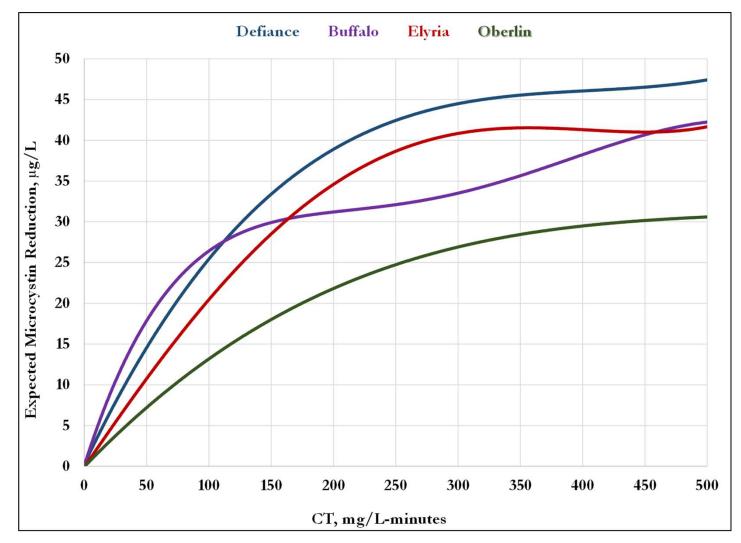
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 - Input actual chlorine demand and dosage ranges
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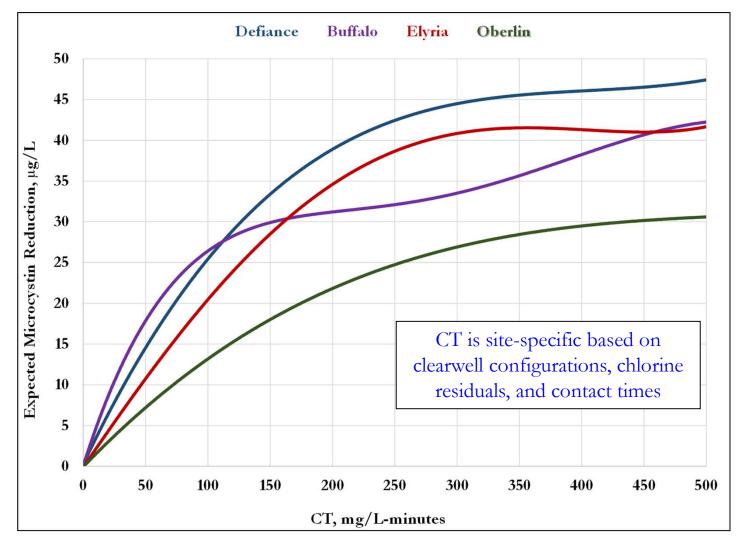


AWWA CyanoTox2 Model





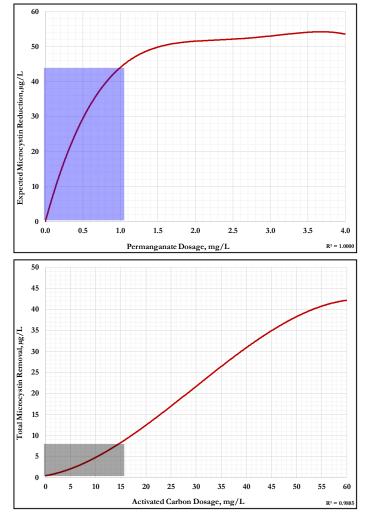


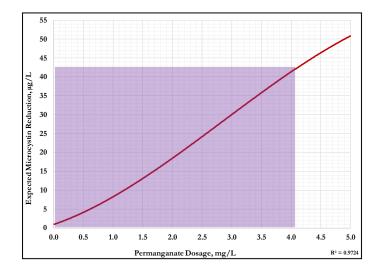


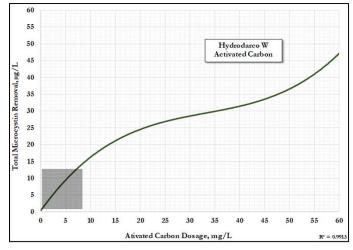


- Chlorine oxidation treatment effective for Microcystin reduction
 - Dosage dependent
 - Contact time dependent
 - pH dependent
 - Lower pH generally provides better removals
- Correlations to AWWA's CyanoTox 2 model possible for some treatment plants

- Attica treatment scenarios
 - Continue to treat reservoir with algaecide to limit Microcystin production
 - 1.1 mg/L KMnO₄ pretreatment removes up to 41 μg/L MC
 - 15 mg/L WaterCarb 800 needed for MC adsorption to 0.3 µg/L or less
 - Post filtration chlorine used as extra barrier treatment

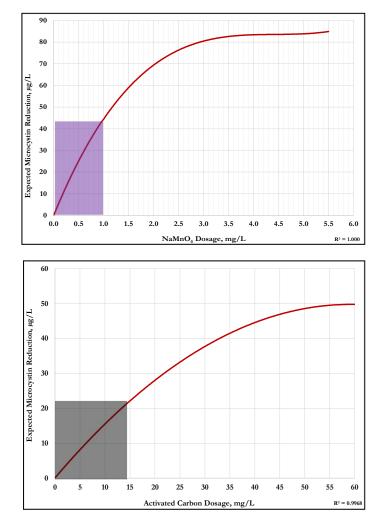


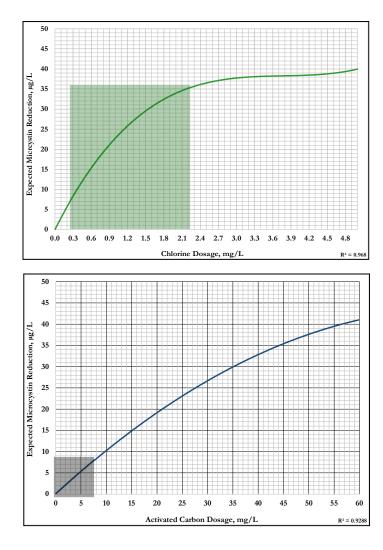




- Lima treatment scenarios
 - 1 mg/L NaMnO₄ dosing until MC extracellular
 - Up to 4 mg/L NaMnO₄ pretreatment removes up to 41 µg/L MC extracellular
 - 5 mg/L to 8 mg/L Hydrodarco W needed for MC adsorption to 0.3 µg/L or less
 - GAC used as extra barrier
 - Post GAC chlorine used as extra barrier

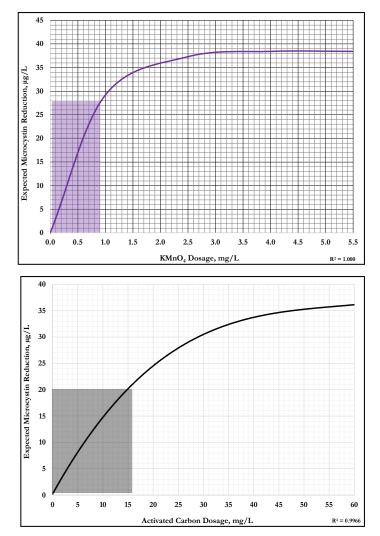
- Defiance treatment scenarios
 - Continue to treat reservoir with algaecide to limit Microcystin production
 - 1 mg/L NaMnO₄ pretreatment removes up to 44.5 μg/L MC
 - 8 mg/L Hydrodarco B needed for MC adsorption to 0.3 µg/L or less
 - GAC contactors being installed as extra barrier
 - Post GAC chlorine used as extra barrier
 - Can oxidize up to 15 µg/L at normal dosages

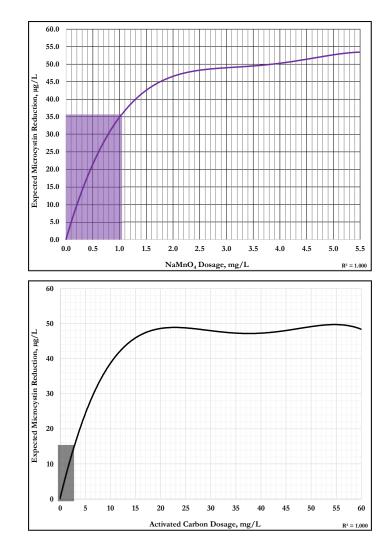




- Buffalo treatment scenarios
 - Routine 2 mg/L pre-chlorination treatment removes up to 35 µg/L MC
 - Low TOC source, no THM issues
 - 15 mg/L WPH needed for MC adsorption to 0.3 µg/L or less
 - Post filtration chlorine used as extra barrier
 - Can oxidize up to 26 µg/L at normal dosages

- Elyria treatment scenarios
 - 0.85 mg/L KMnO₄ pretreatment removes up to 26.5 µg/L MC
 - 15 mg/L WPH needed to remove up to 21 µg/L MC (<u>48 µg/L</u> combined)
 - Post filtration chlorine used oxidize remaining MC
 - Can remove up to 15 µg/L at normal dosages





- Oberlin treatment scenarios
 - 1.0 mg/L NaMnO₄ dosing removes up to 35 µg/L MC
 - 3 mg/L to 4 mg/L AquaSorb CB-1-W needed for MC adsorption to 0.3 µg/L or less
 - Post filtration chlorine used as extra barrier
 - Can oxidize up to 27.5 µg/L at normal dosages

Cyanobacteria Harvesting and Treatment Simulations for Removal of Microcystin from Natural Waters

Questions

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