

Ohio EPA Drinking Water Laboratory Certification **Updates** and **Tips**

OTCO Water Laboratory Webinar

May 22, 2024



OVERVIEW

- Program Changes
- Lab Replacement/Remodeling Requirements
- Application Issues and Reminders
- Invoicing and Fee Payments
- 5-Year Rule Review
- 2025 Manual Edits
- Survey Tips
- HABs/qPCR Certification
- PFAS Certification



PROGRAM CHANGES

- Data request prior to survey
- 100% Electronic
- Remote surveys for IAs, HAB/qPCR
- Staffing



LAB REPLACEMENT/REMODELING REQUIREMENTS

- New labs, remodeled labs, and temporary labs
- Chapter 2 of the Micro and Chem manuals.
- Contact Lab Certification prior to construction
- Completed construction
- Does not have to be detail plans



All Applications

- Methods
- Analyst Names
- Send renewal applications on time.



Interim Authorization Applications

- Unacceptable parallel testing
 - Take samples at same time
 - Use acceptable trainer
- Acceptable Trainers
- Ensure performing only operational testing until certificate is issued
- Corrective Action Page
- Potential for reduced parallels



Name of Primary Contact					
for the Laboratory:		lirst		Middle Initial	Last
Email Address to Send Invoices:					
Date Laboratory Certification	/	/			

NOTICE

In order to be processed, the most current version of the application must be used, and it must be complete and legible. The most current version is located on our website at https://epa.ohio.gov/divisions-and-offices/drinking-and-ground-waters/public-water-systems/laboratory-certification. After acceptance of this application, an invoice will be generated. Additionally, the lab must have copies of all referenced methods and an acceptable SOP, or the most current version of the Ohio EPA lab certification manual.



OATH

I certify that all of the information included on this application is true, complete and correct to the best of my knowledge and belief and are made in good faith. I affirm the right of the Ohio Environmental Protection Agency to inspect the laboratory, its operations and pertinent records. I agree the personnel to be approved will analyze applicable unknown performance samples provided at the time of the survey and will report the values within a time period designated by the Laboratory Certification Officer.

Signature of Primary Contact for Laboratory:	Date:	/	/	
Title of Primary Contact for Laboratory:				

Send completed applications to:

DWLabCert@epa.ohio.gov



Interim Authorization Training Documentation

Laboratory Name:	Name of Operator-In-Training:
Date Training Started:	Date of Training Concluded:

Instructions: Analysts are required to analyze a minimum of seven samples per day, including the quality control (QC) samples. It is recommended that at least one potentially positive sample be included. Results must be generated in parallel with a trainer currently certified for SM 9223-B. Record the operator-in-training results in "OIT" boxes and trainer results in "T" boxes. To be considered acceptable, the OIT results must contain no false negatives and no more than one false positive in comparison to trainer results. Circle all results with a false negative or a false positive and describe any corrective action(s) on page 4.

				Date (Month/Day):						Date (Month/Day):				Date (Month/Day):									
	Test Method		QC		Samples			QC Samples			QC		Samples										
		+	-	1	2	3	4	5	+	1	1	2	3	4	5	+	-	1	2	3	4	5	
		OIT	+/+	-/-	-/-	-/+	-/-	-/-	-/-														
		Т																					
		OIT																					
		Т																					
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INVOICING AND FEE PAYMENTS

- Ensure payments are made by the deadline
- Reminder emails
- Electronic payments



FIVE-YEAR RULE REVIEW

- Updating references
- Updating reporting limit for Microcystin
- Adding reporting limits for regulated PFAS compounds
- Ensure you are signed up for Ohio EPA's electronic mailing list
- Support Login (custhelp.com)



2025 MICRO MANUAL EDITS

Operational Certification

Microbiological operational certification is defined in rule 3745-89-01 of the OAC as certification granted by the Director for an analyst to perform MMO-MUG (SM 9223 B) and Quanti-Tray (SM 9223 B), limited to set up and interpretation of samples, including positive and negative controls. Each operationally certified analyst must complete drinking water sample analysis at a minimum rate of one set of samples per month for each method [e.g., MMO-MUG (SM 9223 B); Quanti-Tray (SM 9223 B)] which the analyst is certified.



2025 MICRO MANUAL EDITS

- a. Reagent Grade Water: Only satisfactory reagent water from deionization units may be used to prepare media, reagents and dilution/rinse water for performing microbial analyses.
 - 1. If a resistivity indicator light is used, sensitivity of the light must be set at 0.5m. > 0.5 megohms resistance or < 2 micromhos/cm.

Prior to use, the quality of the reagent water should be tested and meet the criteria as listed in Table 1.

Table 1: Required Reagent Grade Water Criteria

Parameter	Limits	Frequency ¹
Conductivity	> 0.5 megohms resistance or < 2 micromhos/cm (microsiemens/cm) at 25°C	Monthly ^{2,3}
Total Chlorine Residual	< 0.1 mg/L	Monthly ^{2,3}
Pb, Cd, Cr, Cu, Ni, Zn	Per Contaminant < 0.05 mg/L Collectively < 0.1 mg/L	Annually⁴

¹ If the laboratory purchases bottled reagent grade water, Table 1 does not apply.

Microbio	ological Laboratory Reagent Grade Water Record	
	See Table 1 for criteria.	
boratory	Date of Annual Trace Metals Analysis	_

Analyst	Date	Resistivity Indicator Light (Green or Red)	Conductivity (microsiemens/cm)	Total Chlorine Residual (mg/L)	Comments/Corrective Actions Taken



² If the meter has a resistivity indicator light (i.e., green/red light), record color of light on Microbiological Laboratory Reagent Grade Water Record. The water should only be used if the light is green at the time of use. No conductivity or chlorine residuals are necessary.

³ If no resistivity light, must analyze with each new batch of reagent water.

⁴ Must be analyzed by an Ohio EPA Drinking Water certified or accepted laboratory.

2025 MICRO MANUAL EDITS

MMO-MUG Reagent Quality Control Record To be recorded for each new lot or annually. If performed by a different laboratory, results must be retained. Laboratory Test Results Klebsiella/Variicola **Incubation Start** Incubation End Reagent **Brand/Lot Number** Date/Time Date/Time Color Change Change Change Sample Bottle Sterility/Fluorescence Record To be recorded for each lot received or batch sterilized

	Date	Brand/Lot	Number of Bottles	Growth Growth		Growth Results Interpretation		UV Re	esults	
Analyst	Received/ Sterilized	Number	Received/ Sterilized	Start Date/Time	Incubation End Date/Time	Number Positive	Number Negative	Number Positive	Number Negative	Comments

	Media Quality Control Record	
	To be checked and recorded for each new prepared batch and annually	
Laboratory		

Analyst	Media	Growth Incubation Growth Incubation Growth Incubation Interpretation			Comments		
					Positive	Negative	
				<u> </u>			



2025 CHEMISTRY MANUAL EDITS

Monthly Fluoride QC Sample Record

Analyst	Date	Results (mg/L)	Certified Value Range per PT Provider (mg/L)	Results Within Range (Y/N)	QC Sample Provider Name	Sample Lot #



2025 CHEMISTRY MANUAL EDITS

Monthly Alkali	nity Titrant Standardization Record
aboratory	
Standard Concentration	

Analyst	Date	Reagent Water Volume (mL)	Blank Verification Result * (mL/drops)	Standard Volume (mL)	Titration #1	Titration #2	Titrant Lot Number/Date Prepared	Corrective Action Taken If Out of Range

^{*}Blank verification must be <0.2 mL or 4 drops.



2025 CHEMISTRY EDITS

Inorganic Analytical Methods

Analysis of inorganic constituents in drinking water must be performed following Ohio EPA accepted analytical methods referenced in rule 3745-81-27(A) of the OAC. Unless otherwise specified below, quality control (QC) acceptance limits listed in the individual method must be followed. In addition to individual method's QC requirements, the Laboratory Certification Section requires that, **at minimum**, the following program specific inorganic analysis QC be met. See Chapter 2, Section B of this manual for Quality Assurance Plan requirements.

- Laboratory analyte reporting limits must meet reporting limit concentrations referenced in the appendix to rule 3745-89-03 of the OAC.
- An Initial Demonstration of Capability (IDC) study, (i.e., a blank and 4 LFBs), must be completed and documented for each analyst certified for drinking water method analysis.
- For methods not included in this manual, certified analysts must generate a curve at least once annually
 for all analytical methods which they are certified.
- Curve generation is limited to 1st or 2nd order. Calibration curves must result in a Correlation Coefficient
 (R) greater than 0.995 or a Coefficient of Determination (R²) greater than 0.990 to be acceptable for
 drinking water analysis. It is recommended that curves not be forced through zero. (Calibration curves
 must be at least 3 standards and a blank, unless otherwise specified in the method.)
- Any concentrations above the highest standard in the calibration curve must be diluted to fall within the calibration range.
- At least once every three months, a drinking water sample must be analyzed using the inorganic analytical methods for which the laboratory is certified.
- An annual Method Detection Limit (MDL) study must be performed using the most recent version of USEPA's "Definition and Procedure for the Determination of the Method Detection Limit" in accordance with the 40 Code of Federal Regulations (C.F.R.).
- A Reporting Limit Verification (RLV) sample must be analyzed with each analytical run. The RLV
 concentration is equal to the reporting limit concentration for each analyte of interest. If there is no
 regulatory reporting limit, use the lowest calibration concentration point as the RLV. The acceptance

- Initial Demonstration of Capability study must be documented for each analyst certified for drinking water methods (for methods not included in the manual).
- Method Detection Limit study must be performed annually for each lab.



INITIAL DEMONSTRATION OF CAPABILITY (IDC)

- Used to determine the analyst's ability to perform the method with acceptable precision and recovery
- Method specific
- Often is a variation of a blank and 4 LFBs within a recovery of ±10%
- Ensure all lab SOPs are updated to reflect this requirement



METHOD DETECTION LIMITS (MDLS)

EPA 821-R-16-006 – Definition and Procedure for the Determination of the Method Detection Limit, Revision 2, December 2016

- Applies to all drinking water MDLs except HABs and Hach TNT methods
- Ensure all lab standard operating procedures are updated to reflect this revision.

Annual MDLs should be submitted to dwlabcert@epa.ohio.gov for review.



METHOD DETECTION LIMITS (MDLS)

- Initial MDL: at least seven spikes/blank prepared and analyzed on three different days
- Once an initial MDL is established, the MDL is re-calculated annually including all spike/blank values over the last 24 months
- If seven points are not achieved over a 24-month period, a new initial MDL must be established
- The correct Student-t value must be used corresponding to the number of spikes in calculation (n-1)



METHOD DETECTION LIMITS (MDLS)



MDL Procedure: epa.gov/sites/default/files/201 6-12/documents/mdlprocedure_rev2_12-13-2016.pdf



MDL Frequently Asked
Questions: epa.gov/cwamethods/method-detectionlimit-frequent-questions



Expanded Student t Value Table: itl.nist.gov/div898/handbook/ eda/section3/eda3672.htm



2025 CHEMISTRY MANUAL EDITS

- Fluoride by SPADNS 2 (Arsenic-Free)
 Method 10225
- Nitrate by Hach TNT plus 835/836
 Method 10206
- Orthophosphate by Hach Method 8048 (EPA 365.1)

Fluoride Analysis by SPADNS 2 (Arsenic-Free) Method 10225

Quick Reference	Standard/Reagent/Equipment	Requirements	
	SPADNS 2 Reagent	Manufacturer's Recommendations	
Standard/Reagent Storage	0.5/1.0/1.5 mg/L Standards	Manufacturer's Recommendations	
	100 mg/L Stock Standard	Manufacturer's Recommendations	
	Standard/Reagent	Expiration	
Standard/Reagent	SPADNS 2 Reagent	1 Year After Opening/ Manufacturer's Expiration Date	
Expiration	0.5/1.0/1.5 mg/L Standards	1 Year After Opening/ Manufacturer's Expiration Date	
	100 mg/L Stock Standard	1 Year After Opening/ Manufacturer's Expiration Date	
	QC Procedure	Frequency	
Barriard Quality Cantual	Meter Calibration Verification	Once Every Three Months	
Required Quality Control	Blank, QCS	Once Per Batch	
	QC Sample Analysis	Once Per Month	
	Preservation	Maximum Hold Time	
Sample Collection	None	48 Hours [See OAC rule 3745-83-01(F) (4)(b)] or 1 Month [See OAC rule 3745-81-23(J)]	



INITIAL DEMONSTRATION OF CAPABILITY FOR ADDED METHODS

Initial Method Detection Limit (MDL) study Initial Precision and Recovery (IPR) study

Method Detection Limit (MDL) Study

		*To be performed i	n a single run		
Laboratory:			Analyst:		
Instrument:					
				(number)	(unit)
Date:			True Value:		
	Replicate	Value (mg/L)			
	1				
	2				
	3				
	4				
	5				
	6				
	7				
	Average	#DIV/0!			
	Std Deviation	#DIV/0!			
	MDL Result	#DIV/0!			
	MDL Acceptable	#DIV/0!			

Initial Precision and Recovery Study

*To be performed in a single run						
Laboratory:			Analyst:			
Insrument:						
				(number)	(unit)	
Date:			True Value:			
	Replicate	Value (mg/L)	% Recovery			
	1		#DIV/0!			
	2		#DIV/0!			
	3		#DIV/0!			
	4		#DIV/0!			
	Average	#DIV/0!	#DIV/0!			
	Std Deviation	#DIV/0!				
	%RSD	#DIV/0!				
Accuracy (% Re	ecovery) Passing?	#DIV/0!				
Precision	(%RSD) Passing?	#DIV/0!				



- Reagent water quality (indicator light) verify prior to use
- Incubator temperatures must be recorded on weekends if samples are being incubated
- Autoclave timer must be checked only at times used (e.g., 15, 30, 45); use proper procedure
- Balance verification must be done prior to use
- Sampling instructions for micro samples requires analyzing for chlorine residual after disinfection of sample tap



MMO-MUG Analysis for Total Coliform and *E. coli* by Colilert and Colisure

Quick Reference	Standard/Reagent/Equipment	Requirements	
	MMO-MUG Reagent	Colilert – Dark Environment and Manufacturer's Recommendations Colisure – Refrigerated and Manufacturer's Recommendations	
	Chemical Reagents	Manufacturer's Recommendations	
Standard/Reagent/Equipment	Dehydrated Media	Manufacturer's Recommendations	
Storage	Media Performance Check Cultures	Manufacturer's Storage Requirements	
	Prepared Media	Refrigerated/Room Temperature	
	pH Electrodes	pH 7 Buffer/Manufacturer's Storage Solution	
	pH Buffers	Room Temperature	
	Standard/Reagent	Maximum Storage Time	
		Manufacturer's Expiration Date	
	Chemical Reagents	Manufacturer's Expiration Date	
	Dehydrated Media	6 Months After Opening or 1 Year After Opening if Stored in Desiccator	
Standard/Reagent Expiration	10% Sodium Thiosulfate	1 Year After Preparation/ Manufacturer's Expiration Date	
σ	Media Performance Check Cultures	Manufacturer's Expiration Date	
	Prepared Media	3 Months Refrigerated (screw-capped tubes/flasks/vessels) or 1 Week Room Temperature (sealed/covered)	
	pH Buffers	6 Months After Opening/ Manufacturer's Expiration Date	
	QC Procedure	Frequency	
	Total Coliform/E. coli positive	Once Per Month Per Analyst	
Standard/Reagent/Equipment Storage Chemical Reagents Dehydrated Media Media Performance Check Cultures Prepared Media PH Electrodes PH Buffers Standard/Reagent Chemical Reagent Dehydrated Media 10% Sodium Thiosulfate Media Performance Check Cultures Prepared Media PH Buffers QC Procedure Total Coliform/E. coli positive Sample/Test Bottle Sterility Check Sample/Test Bottle Sterility Check Sample/Test Bottle Fluorescence Check Media Performance Check Media Performance Check Glass/Electronic Thermometer Data Logger Calibration Dial Thermometer Calibration Equipment Timers PH Meter Calibration Equipment Timers PH Meter Calibration Part		One Per Batch Prepared or 1% Per	
	Lot Received (maximum of 4 per lot)		
		Every Sample/Test Bottle Prepared or 1% Per Lot Received (maximum of 4	
Standard/Reagent MMO-MUG Reagent Colliert – Dark Environme Manufacturer's Recomme Colisure – Refrigerated a Manufacturer's Recomme Colisure – Refrigerated a Manufacturer's Recomme Manufacturer's Recomme Manufacturer's Recomme Media Performance Check Cultures Prepared Media Manufacturer's Storage Refrigerated/Room Temp PH Jeietrodes PH Buffers Standard/Reagent Manufacturer's Solution PH Buffers Room Temperature Standard/Reagent Manufacturer's Expiration Manufacturer's Expiration Standard/Reagent Manufacturer's Expiration Dehydrated Media Manufacturer's Expiration Manufact	F =		
Required Quality Control	Glass/Electronic Thermometer/	<u> </u>	
		Once Every Three Months	
		Once Every Three Months	
	pH Meter Calibration	Prior to Use	
		Prior to Use	
		Prior to Use	
	Refrigerator Record	Daily	
	Incubator Record	Twice Daily	
	Preservation	Maximum Holding Time	
Sample Collection	10% Sodium Thiosulfate	30 Hours	

Autoclave Sterility Check

- Required once every three months, per autoclave
- May use biological indicator ampules following manufacturer's instructions
- May use TSB or BHI, inoculated with a known coliform culture
- Ensure recorded on Autoclave Sterilization Record

Thermometer Calibration Record

- Must first include the NIST thermometer's temperature at ice point
- Recommend including each thermometers serial number
- MRTs are not calibrated with NIST
- Autoclave Dial (Display) Thermometers are not required to be calibrated unless fast exhaust is used.



Media Preparation(e.g., TSB, BHI)

- Balance Calibration Record
- pH Meter Slope/Linearity Verification
- Media Quality Control Record
- Autoclave Sterilization Record
 - TSB or BHI at temperature 12-15 min
 - Autoclave door must be opened no later than 45 min after closing

Pre-Made Purchased TSB

- Use manufacturer's expiration date prior to opening.
- Keep all paperwork.

Microbiological Test Data Sheets

 All data from our bench sheets must be recorded to avoid invalidation of sample results.



Maximum Registering Thermometers (MRTs)

- Calibrated by Lab Certification staff at the renewal survey
- Ohio Revised Code 3734.63, Sale of mercury-containing thermometer for promotional purposes.
 - If required to comply with federal law, these can be sold and distributed.
- Dial autoclave thermometers are not permitted.







SURVEY TIPS - CHEMISTRY

- QC requirements on first page of each method in the manual
- Never pipette directly out of a standard bottle
- Stability by saturation is to be filtered using a fine porosity fast-flow glass fiber filter paper
- Dry secondary chlorine standards with lint-free wipes
- Verification of alkalinity endpoint by pH 4.5
- Hach TU 5200 has a different Method Number

You may NOT perform analysis unless IA is granted, or a survey is successfully completed.



Alkalinity Analysis by Sulfuric Acid Titration Method

Quick Reference	Standard/Reagent	Requirements	
	0.020 N Sulfuric Acid (H ₂ SO ₄)	Manufacturer's Recommendations	
Standard/Reagent	Indicator (Bromcresol Green/ Methyl Red)	Manufacturer's Recommendations	
Storage	Sodium Thiosulfate	Manufacturer's Recommendations	
	0.020 N Sodium Carbonate (Na ₂ CO ₃) Standard	Manufacturer's Recommendations	
	Standard/Reagent	Expiration	
	0.020 N Sulfuric Acid (H ₂ SO ₄)	1 Year After Opening/ Manufacturer's Expiration Date	
Standard/Reagent Expiration	Indicator (Bromcresol Green/ Methyl Red)	1 Year After Opening/ Manufacturer's Expiration Date	
	Sodium Thiosulfate	1 Year After Opening/ Manufacturer's Expiration Date	
	0.020 N Sodium Carbonate (Na ₂ CO ₃) Standard	1 Year After Opening/ Manufacturer's Expiration Date	
	QC Procedure	Frequency	
Required Quality Control	Standardize Titrant	Once Per Month	
	pH 4.5 Endpoint Verification	Once Per Month	
Sample Collection	Preservation	Maximum Hold Time	
	4°C	14 Days	

Method Reference

Standard Methods 22nd Edition (2320)

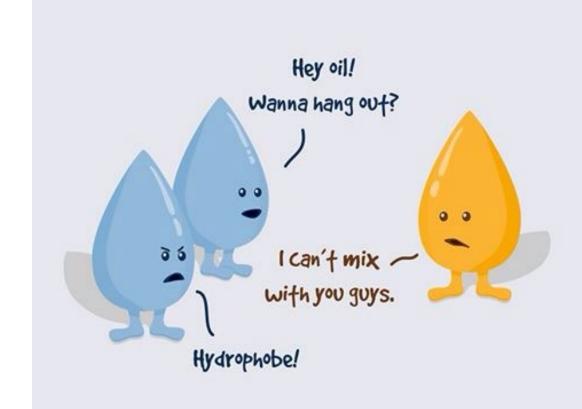
On-Site Survey Requirements

- Each certified analyst must be able to perform the alkalinity titrant standardization described in Section 7.0 of this method.
- Operationally certified analysts will be required to analyze a plant tap sample and may be required to analyze a performance sample.
- Procedural technique will be observed.
- All reagents, standards and solutions used for this method will be audited for correct labeling and dating.
- All records will be audited

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SURVEY TIPS - GENERAL

- Update bench sheets to most recent version available.
- Ensure all laboratory records are recorded using ink and are printed legibly.
- Scribbling/ writing over is unacceptable.
- Errors? Cross out with 1 line, initial, add correct information. **No White Out!!**
- Include results to the 10th (e.g., 121 is 121.0)
- Avoid eating or drinking in the lab.
- Annual review of manual(s) REQUIRED





SURVEY TIPS- GENERAL

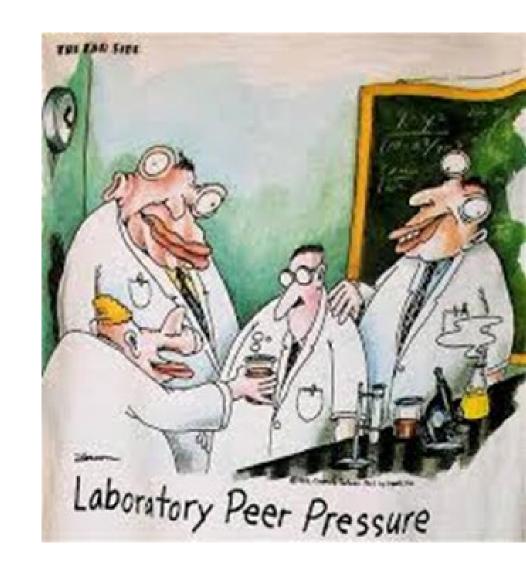
Annual Laboratory Manual Review Record						
Laboratory	Methods Reviewed					

Analyst No.	Analyst Signature	Date of Review	Analyst No.	Analyst Signature	Date of Review



SURVEY TIPS - GENERAL

- If it's not written down, it didn't happen.
- Sorry, "But we've always done it that way..." doesn't supersede current requirements.





CYANOTOXIN & CYANOBACTERIA CERTIFICATION

- Annual MDLs and curves as well as associated test data are to be sent to the dwlabcert@epa.ohio.gov email.
- Adding new analysts for Cyanotoxin and/or Cyanobacteria certification between renewal periods.
- SOPs for microcystin and qPCR are available on our Lab Certification website.
- Please review MDLs and curves prior to submitting and don't send if they have clearly failed.





PFAS CERTIFICATION

- Once Ohio EPA's PFAS rules are promulgated, we will begin certifying laboratories for EPA Methods 533 and/or 537.1.
- The State's lab is currently working on determining the reporting limits for the six regulated compounds.
- Lab Certification has provided acceptance for 16 laboratories to analyze for PFAS.
- The six regulated compounds will be PFOA, PFOS, GenX, PFBS, PFHxS, and PFNA.



Thank You

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