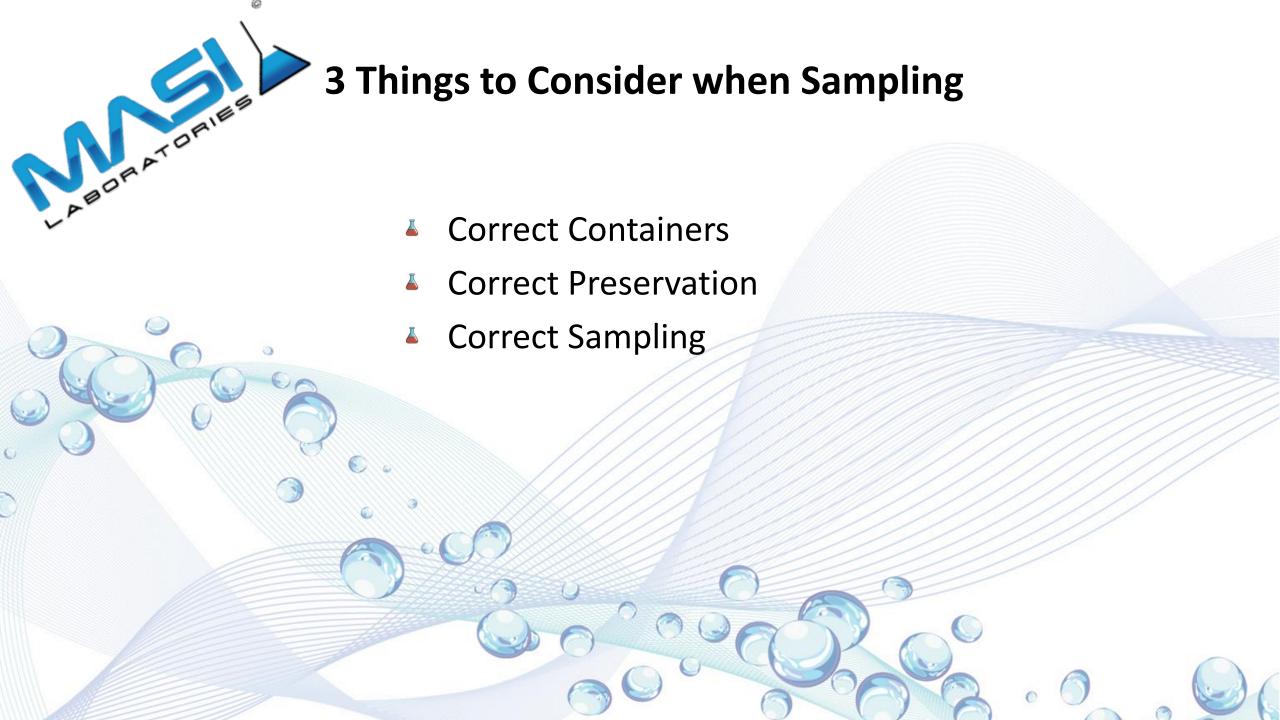
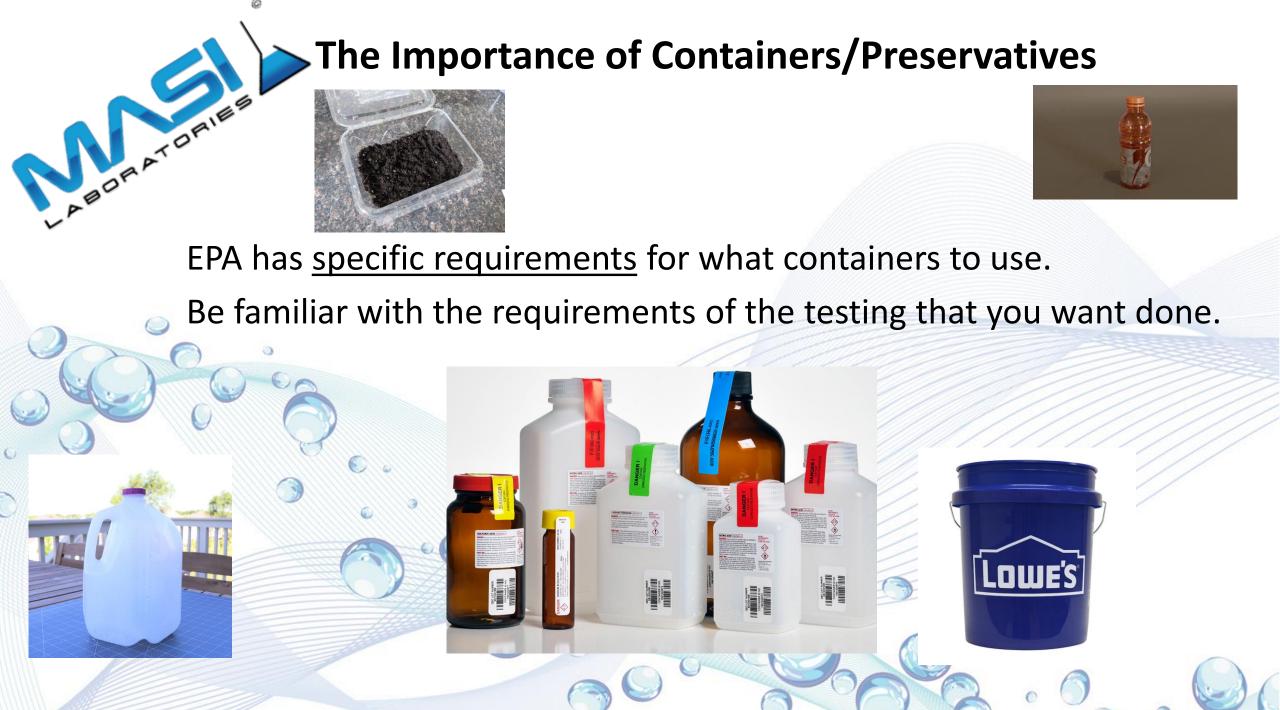
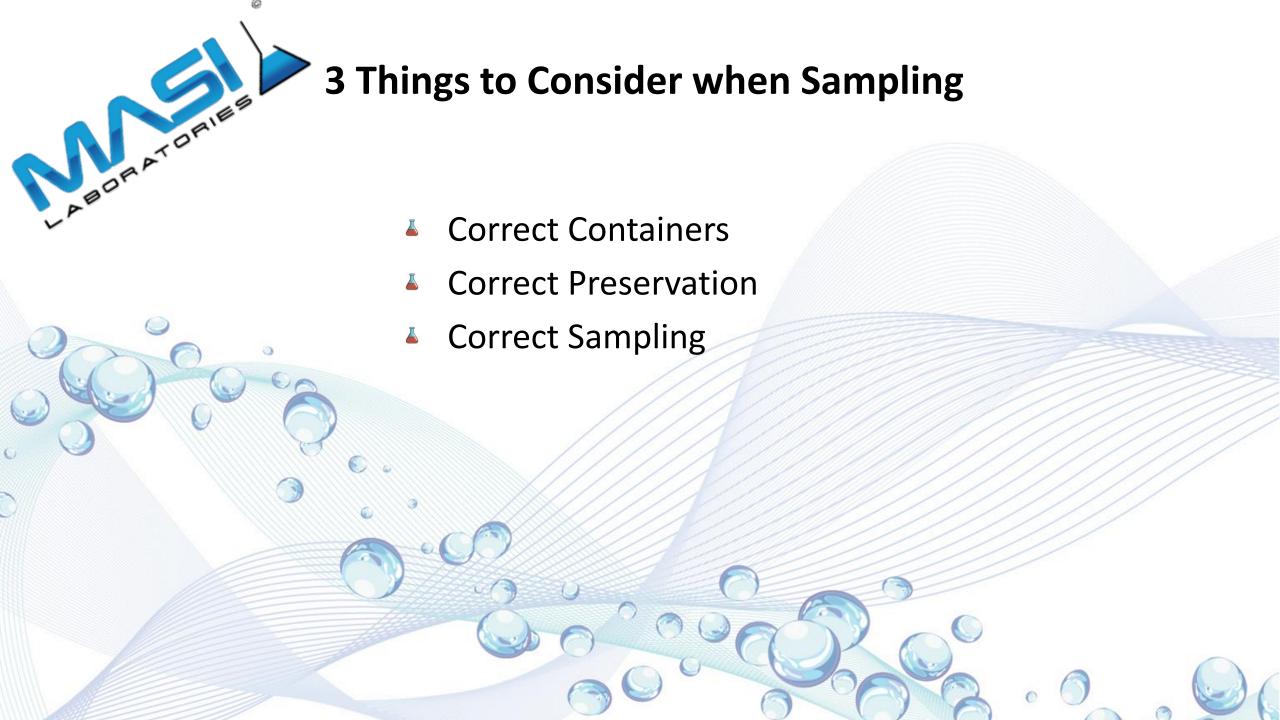


Proper Sampling Protocols for Emerging Contaminates













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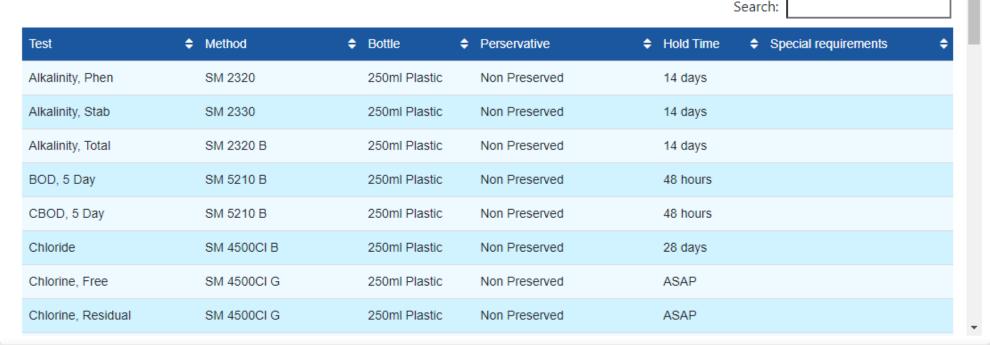


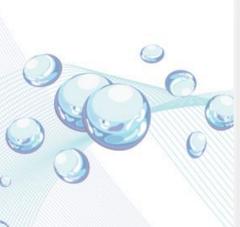
Most Labs have reference material on their websites for containers, **Drinking Water Capabilities**

sampling instructions, and hold times

Waste Water Capabilities





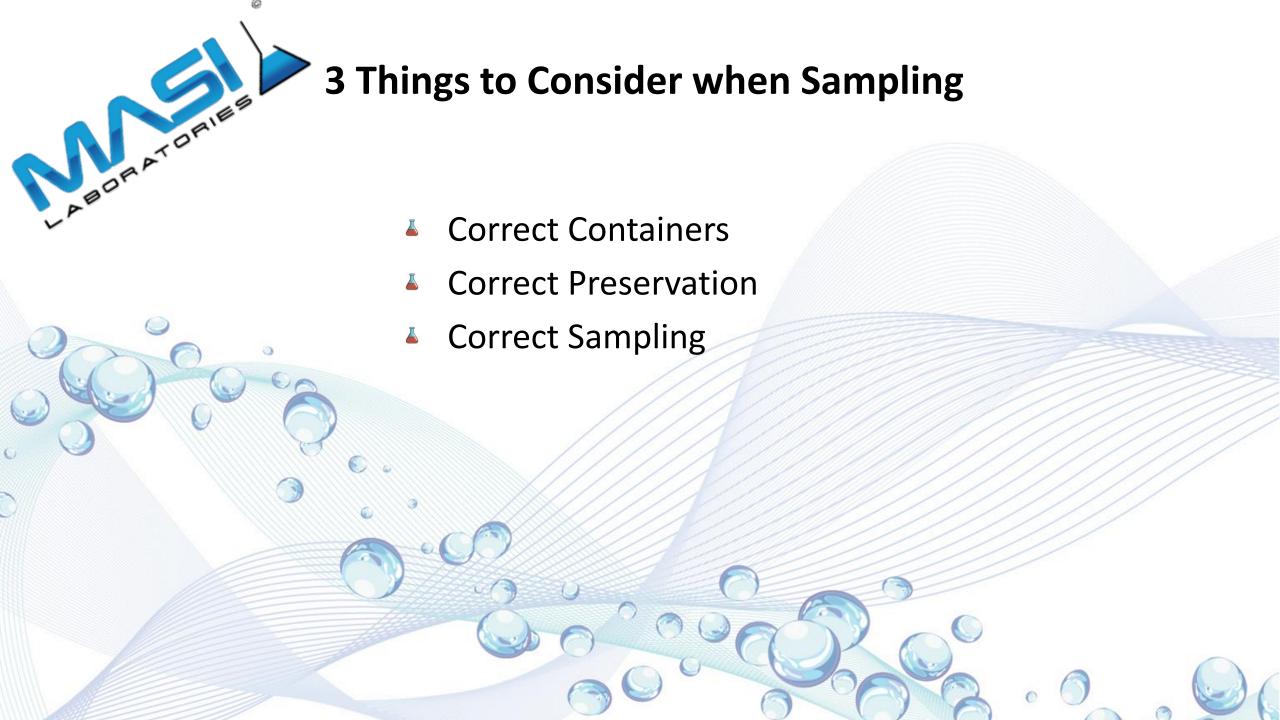




Dechlorinator vs Preservative

- A dechlorinator will remove any residual chlorine from the sample.
 - Sodium Thiosulfate
 - Ascorbic Acid
 - Sodium Sulfite
 - Ammonium Chloride
- A preservative will maintain the sample as it is at sample collection
 - Acids (Nitric, Hydrochloric, Phosphoric, Sulfuric)

NOTE: Non-preserved samples usually have shorter hold times





✓ Collect samples in an area free of excessive dust, rain, snow, or other sources of contamination.

✓ Select a cold-water faucet for sampling which is free of contaminating devices such as screens, aerations devices, hoses, purification devices or swiveled faucets. Check the faucet to be sure it is clean. If the faucet is in a state of disrepair, select another sampling location.

✓ Collect samples from faucets which are high enough to put a bottle underneath, generally the bathtub or kitchen sink, without contacting the mouth of the container with the faucet.



✓ Open the faucet and thoroughly flush. Generally, 5-10 minutes will suffice. Once the lines are flushed, adjust the flow so it does not splash against the walls of the bathtub, sink or other surfaces.

Follow the collection instructions provided for the analytes of interest. Remember PPE when handling containers with acid preservatives.

General Sampling Procedures

✓ Fill out the chain of custody form with the sample collection information.

✓ Deliver or ship samples to the laboratory to ensure that the holding times are met. Holding time starts at the sample collection and ends at the preparation and/or analysis. Be sure to allow time for the laboratory to process the samples.



Specific Sampling Procedures and Considerations

Total Coliform

- Bottles are sterile so care must be taken not to contaminate the bottle or cap.
- Coliforms are common in the environment, and some are not harmful to humans. Contamination of your sample from soil, dust or other outside factors must me minimized to prevent false positive samples.
- Sanitize the tap before taking the sample.

Specific Sampling Procedures and Considerations

VOC/THM/BTEX/Halomethanes

Care must be taken to prevent headspace in the sample vial.

Potential contar

Provide multiple

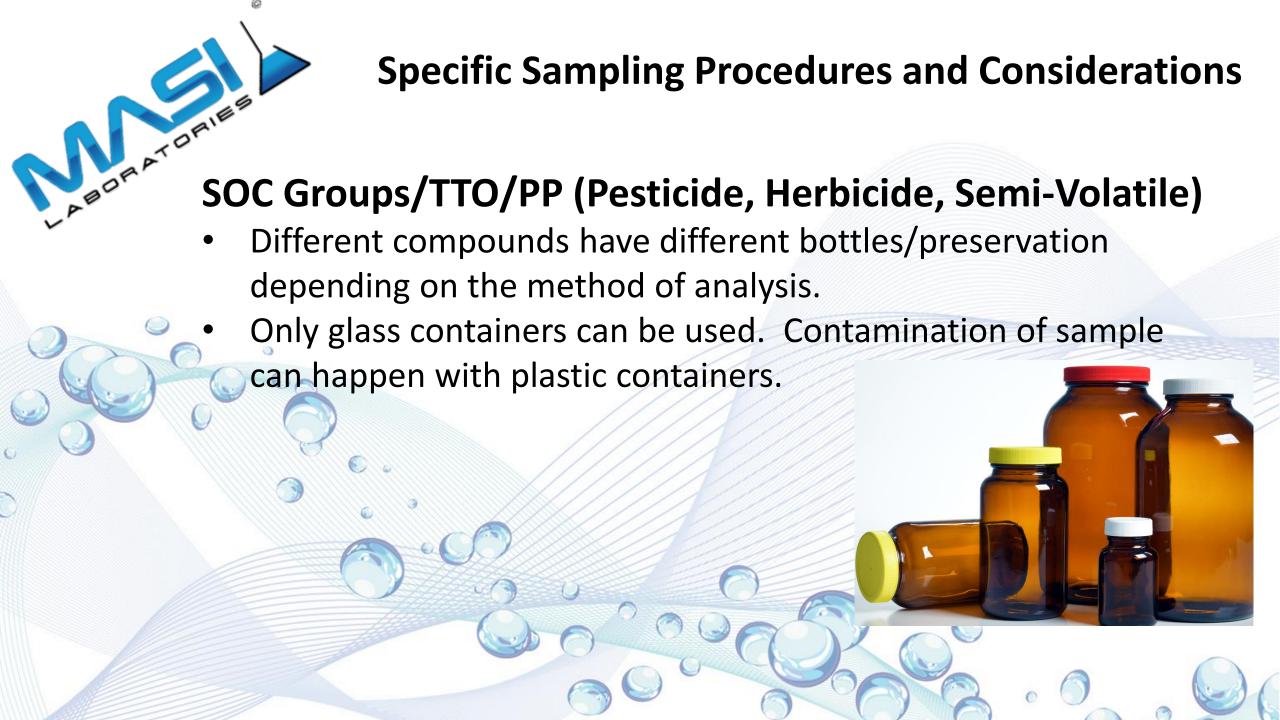
 Be careful not to bottle. This cou chlorine which we the lab for analy

Trip blanks are a contamination.

ne motors.

orinator in the esult for residual e to be rejected by

dentify sampling





Lead/Copper Rule

- Select a cold-water faucet for sampling which is free from water softeners or point of use filters.
- Do NOT remove any screens or aeration devices.
- Sample tap must sit idle for at least 6 hours.
- Place a 1L bottle under the tap and collect the first water out of the tap.
- Bottles are unpreserved so watch holding times.



Specific Sampling Procedures and Considerations Low-Level Mercury (Clean Hands/Dirty Hands)

"Clean Hands"

- does not touch "non-clean" materials
- does not touch secondary container bags
- does not touch shipping containers
- handles all "clean" materials
- directly contacts sample source
- opens/closes primary container bags
- directly holds sample container

"Dirty Hands"

- does not disturb sample source
- does not touch primary container bags
- does not touch sample containers
- does not touch "clean equipment"
- handles all "non-clean" materials
- opens/closes shipping containers
- opens/closes secondary container bags
- operates pump/metallic equipment

Specific Sampling Procedures and Considerations

PFAS

t**ive Equipment** ontaining Gore-Tex®.

de of polyurethane and poly olyurethane and PVC.

- Do not use materials containing Tyvek® or polytetrafluoroet...... as Teflon®
- Do not use fabric softener on clothing to be worn in field.
- Do not use cosme

Fie

Do not use prohik

Food Considerations

No food or drink a

Field Equipment

contain must be

react field books can be used

proof field books can be used.

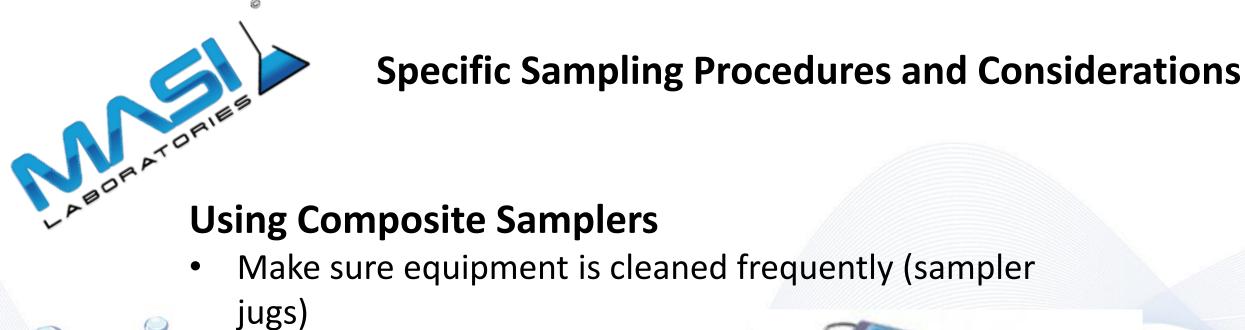
clipboards, binders, or spiral hard cover notebooks can be used and permanent markers not allowed; regular ball point pens are samples in separate cooler, away from sampling containers that

Coolers filled with regular ice only - Do not use chemical (blue) ice packs.

, or other related products the morning of sampling. lant.

of bottled water.

sity polyethylene (LDPE) m gh-density polyethylene (H



Make sure equipment is cleaned frequently (sampler

- Replace tubing
- Sampling sticks are clean
- Use gloves all the time



Important Reminders for CoC

- Containers should be CLEARLY labeled with your lab's ID / sample identifier.
- Sample locations are noted correctly (influent / effluent).
- All pertinent information for reporting potable parameters is provided and correct. For example, if you do not list the correct SMP ID, you will not get credit for that sampling.
- Container and chain of custody should match, for parameters needed.

