

# **PRIMARY HUMAN MAST CELL ASSAY**

## RELIABLE EVALUATION OF DRUG EFFICACY USING HUMAN MAST CELL DEGRANULATION TEST

#### ABOUT MAST CELL ASSAY

Mast cells are granular immune cells that are characterized by c-kit and FccR1 expression and originate from bone marrow blood precursors. Mast cells are involved in:

- Allergy and anaphylaxis
- Wound healing and angiogenesis
- Immune tolerance
- Defense against pathogens
- Vascular permeability in brain tumors

Mast cell degranulation can be induced by: IgE-FccR1 complex aggregation, stem cell factor-induced c-kit dimerization, etc. During degranulation, mast cells release a number of immunomodulatory products that are implicated in both protective immune responses and clinically relevant disorders such as anaphylaxis.

### OUR ASSAYS WIDELY USED TO SYUDY BIOLOGY OF DISEASES

- ✓ Allergy
- ✓ Anaphylaxis
- ✓ Asthma
- ✓ IgE-mediated hypersensitivity reactions
- ✓ Mastocytosis
- Anti-helminth immunity
- ✓ Mast cell activation syndromes
- ✓ Cancer

At PicoImmune, we offer assays using human primary mature mast cells to determine the ability of a prospective therapeutic to modulate mast cell degranulation.

#### CHALLENGES OF MAST CELL RESEARCH

- Primary human mast cells are rare in peripheral blood and tissue isolation protocols yield low cell quantities, viability and purity.
- Differentiation of blood stem/precursor cells to mature mast cells is time-consuming and culture reagents are costly.
- Both human (HMC-1, LAD2, LUVA) and rodent (P815, RBL-1, FMA-3) mast cell lines exhibit aberrant phenotypes, including lack/loss of functional receptors such as FccR1, impaired cytokine production, long doubling time or insufficient cytosolic granules.







#### **OUR SERVICE FEATURES**

- CD34+ blood precursor-derived mast cells: Mimic mature human mast cells; express functional c-kit and FccR1; have a well condensed non-lobate nucleus and abundant cytosolic granules
- Variety of readouts: histamine/ tryptase/βhexosaminidase/cytokines/ PGD2/gene expression
- Degranulation Induction: IgE-dependent and -independent (Compound 48/80, cortistatin-14, substance P)
- High throughput: 96/384-well format, duplicate or triplicate; multiple donors can be tested concurrently; highly multiplex analysis at transcriptome/secretome/proteome levels
- Robust and highly reproducible: More predictive results than with cell lines or rodent cells
- Well-validated reagents and protocol: provide established differentiation reagents, mast cell agonists and antagonists

- Turnaround time: ~3 months
- State-of-art platforms: CytoFLEX Flow Cytometer; FlexMAP 3D (Luminex, 96 and 384-well format); Tecan Microplate Reader; NanoString nCounter
- 20+ years of experience of mast cell research: Expert data analysis and interpretation, scientific and technical support

#### HOW OUR ASSAY WORK?

- 1. CD34+ precursor cell isolation from peripheral blood.
- 2. Differentiation of CD34+ precursor to mature mast cells.
- 3. Evaluation of mast cell culture purity:
  - c-kit expression
  - FccR1 expression
- 4. Test compound treatment during stimulation
- 5. Readout quantification:
  - Cytokines production
  - Gene Expression
  - Degranulation products/markers







#### EXAMPLE DATA 1: MAST CELL PURITY

Primary CD34+ hematopoietic precursor cells isolated from 2 healthy donors were differentiated to mast cells *in vitro* by culturing cells in medium supplemented with recombinant stem cell factor (SCF), interleukin (IL)-6, and IL-3. Mast cell culture purity was evaluated after weeks of differentiation based on the expression of c-kit and FccR1 with flow cytometry.









#### EXAMPLE DATA 2: COMPOUND 48/80 INDUCED MAST CELL DEGRADNULATION

Mature CD34+ hematopoietic precursor-derived mast cells were incubated with varying concentrations of test compound or cromolyn (mast cell stabilizer) during stimulation with compound 48/80 (mast cell activator). After treatment,  $\beta$ -hexosaminidase, histamine, prostaglandin D<sub>2</sub> and tumor necrosis factor (TNF)- $\alpha$  in the cell-free supernatants were quantified. Data shown are mean ± SEM (3 donors). Statistical analysis was performed using one-way ANOVA with pairwise comparisons made to cells that received no compound treatment. \*, P < 0.05; \*\*, P < 0.01; \*\*\*, P < 0.001; \*\*\*\*, P < 0.001.



