





Complete Genome Sequencing of the Novel *Pseudomonas* aeruginosa Phage UF_RH1

Abdolrazagh Hashemi Shahraki, Majid Vahed, Mehdi Mirsaeidi

^aDivision of Pulmonary, Critical Care and Sleep, College of Medicine-Jacksonville, University of Florida, Jacksonville, Florida, USA

ABSTRACT Here, we present the genome sequence of a novel *Pseudomonas aeruginosa* bacteriophage called UF_RH1. This lytic phage has a genome size of 42,567 bp and is classified as a member of the *Siphoviridae* family and the *Septimatrevirus* genus. UF_RH1 shares genetic similarities with *Stenotrophomonas* phage vB_SmaS-DLP_2.

seudomonas aeruginosa is an opportunistic pathogen that poses significant challenges for treatment (1). However, one effective approach to combat this pathogen is phage therapy, particularly with lytic phages (2). A P. aeruginosa phage named UF_RH1 was isolated by introducing 10 μ L of filtered wastewater collected from a sewage treatment plant (Alexander Orr Water Treatment Plant located at Miami, FL) to 400 μ L of *P. aeruginosa* (strain DJ06) in logarithmic phase and then performing single-plaque isolation using double-layer agar (3). The purity of the phage was confirmed after five rounds of single-plaque isolation. Phage DNA was extracted using a commercial kit (QIAamp DNA Mini kit; Qiagen, USA). The Nextera XT library preparation kit (Illumina, San Diego, CA) was used to create a phage DNA library, which was sequenced on the Illumina NovaSeq 6000 platform with 150-bp paired-end reads. The raw reads were cleaned up with the Cutadapt program v2.8 (4) to remove lowquality bases and sequencing adaptors and then searched against the genome of P. aeruginosa from the NCBI genome database using the read mapper of the STAR package to remove potential host DNA contamination (5). The unmapped pairedend reads were further assembled with the software MetaWRAP v1.2.0 (6), and the assembled consensus sequences with a length of more than 5,000 bp were evaluated by QUAST v5.0.2 (7) and analyzed with the metagenomic software Centrifuge v1.04b (8). The quality of the viral genome completeness and identification of closed genomes were assessed using the CheckV v1.01 package (9). The viral genome candidates were further analyzed with NCBI BLASTn to determine the taxonomic relationships of the phage (10), and Victor was used for phylogenetic analysis (11). Open reading frames (ORFs) were identified by GeneMarkS (12), and the genome was annotated based on PHASTER (13) and BLASTp results (10). PhageTerm was used to determine the phage termini (14). The tRNA sequences were determined by tRNAscan-SE v2.00 (15), while virulence factors and antibiotic resistance factors were detected using ResFinder v4.00 (16) and the Antibiotic Resistance Genes Database (17), respectively, with default parameters for all software tools.

The genome of *Pseudomonas* phage UF_RH1 is a liner double-stranded DNA (dsDNA) genome with a length of 42,567 bp, a GC content of 53.56%, 931,200 total reads, and $23,360\times$ average read coverage. PhageTerm predicted a circularly permuted genome for UF_RH1. It includes 57 open reading frames (ORFs), and it shares sequence similarity with members of the genus *Septimatrevirus* (Fig. 1). *Stenotrophomonas* phage vB_SmaS-DLP_2 (18) is a closer phage to UF_RH1 (Table 1). The annotated proteins of UF_RH1, such as dead box helicase, RecB exonuclease, central tail hub, and holin,

Editor Catherine Putonti, Loyola University

Copyright © 2023 Hashemi Shahraki et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Mehdi Mirsaeidi, m.mirsaeidi@ufl.edu.

The authors declare no conflict of interest.

Received 24 February 2023 **Accepted** 30 April 2023



VICTOR nt tree with suggested taxa (trimming, D0)

FIG 1 Whole-genome-based phylogenic tree (with average nucleotide identity grouping) and taxonomic position of UF_RH1 among closely related phages of genus *Septimatrevirus*. Three series of color boxes behind the tree indicate viral taxonomy position (family, genus, and species), and the last one shows the GC content (%) for each virus.

exhibit 99%, 98%, 98%, and 86.36% similarity, respectively, to the corresponding annotated proteins of *Stenotrophomonas* phage vB_SmaS-DLP_2. DNA polymerase and endolysin also show the highest similarity of 99.8% and 92%, respectively, to the proteins encoded by *Pseudomonas* phage TehO. The tail length tape-measure protein also displays the highest similarity of 99.58% to the tail protein of *Pseudomonas* phage vB_PaeS_C1. UF_RH1 does not contain tRNA, virulence, and antibiotic-resistant genes.

Data availability. The complete phage genome sequence was deposited in GenBank under the accession number OQ259603. The raw data are available in the NCBI Sequence Read Archive (SRA) under BioProject accession number PRJNA936202, SRA accession number SRS16838829, and BioSample accession number SAMN33343989.

TABLE 1 Genome sequence coverage and nucleotide identity of UF_RH1 with their closest relatives^a

Phage	Sequence coverage (nucleotide identity) of:							
(GenBank accession no.)	UF_RH1	vB Pae-Kakheti25	vB SmaS-DLP_2	Epa41	Epa40	phipa4	vB PaeS_SCUT-S4	PA73
UF_RH1 (OQ259603.1)	100 (100)	95 (96.2)	97 (97.8)	92 (94.8)	91 (94.8)	95 (94.1)	94 (97.2)	94 (94)
vB Pae-Kakheti25 (JQ307387.1)	95 (96.2)	100 (100)	95 (97.08)	91 (96.8)	90 (96.8)	97 (97.9)	98 (96.8)	93 (97.6)
vB SmaS-DLP_2 (KR537871.1)	97 (97.8)	95 (97.08)	100 (100)	89 (97.36)	89 (98.37)	95 (97)	95 (97.3)	93 (98.33)
Epa41 (MT118305.1)	92 (94.8)	91 (96.8)	89 (97.36)	100 (100)	99 (100)	89 (97.5)	89 (98.34)	92 (97.67)
Epa40 (MT118304.1)	91 (94.8)	90 (96.8)	89 (98.37)	99 (100)	100 (100)	89 (97.5)	89 (98.34)	92 (97.75)
phipa4 (OK539825.1)	95 (94.1)	97 (97.9)	95 (97)	89 (97.5)	89 (97.5)	100 (100)	98 (96.88)	94 (98)
vB PaeS_SCUT-S4 (MK165658.1)	94 (97.2)	98 (96.8)	95 (97.3)	89 (98.34)	89 (98.34)	98 (96.88)	100 (100)	91 (97.64)
PA73 (DQ163913.1)	94 (94)	93 (97.6)	93 (98.33)	92 (97.67)	92 (97.75)	94 (98)	91 (97.64)	100 (100)

^a All values are %. All phages are classified as *Pseudomonas* phage except vB_SmaS-DLP_2, which is classified as *Stenotrophomonas* phage.

Month YYYY Volume XX Issue XX 10.1128/mra.00139-23

ACKNOWLEDGMENTS

This work was supported by funding from the dean's office of University of Florida, College of Medicine Jacksonville.

We thank Naresh Kumar of the University of Miami for providing the raw sewage. Additional support came from the University of Florida ICBR Bioinformatics Core Facility, especially from Fahong Yu.

REFERENCES

- Shariati A, Azimi T, Ardebili A, Chirani AS, Bahramian A, Pormohammad A, Sadredinamin M, Erfanimanesh S, Bostanghadiri N, Shams S, Hashemi A. 2018. Insertional inactivation of oprD in carbapenem-resistant Pseudomonas aeruginosa strains isolated from burn patients in Tehran, Iran. New Microbes New Infect 21:75–80. https://doi.org/10.1016/j.nmni.2017.10.013.
- Chegini Z, Khoshbayan A, Taati Moghadam M, Farahani I, Jazireian P, Shariati A. 2020. Bacteriophage therapy against Pseudomonas aeruginosa biofilms: a review. Ann Clin Microbiol Antimicrob 19:45. https://doi.org/10 .1186/s12941-020-00389-5.
- Jiang Y-H, Liu J-Q, Zhao C-Y, Yu S, Sun Y-B, Shi H-Y, Huang H-L. 2020. Isolation and genome sequencing of a novel Pseudomonas aeruginosa Phage PA-YS35. Curr Microbiol 77:123–128. https://doi.org/10.1007/s00284-019-01792-8.
- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. EMBnet j 17:10–12. https://doi.org/10.14806/ej.17.1.200.
- Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR. 2013. STAR: ultrafast universal RNA-seq aligner. Bioinformatics 29:15–21. https://doi.org/10.1093/bioinformatics/bts635.
- Uritskiy GV, DiRuggiero J, Taylor J. 2018. MetaWRAP—a flexible pipeline for genome-resolved metagenomic data analysis. Microbiome 6:158. https://doi.org/10.1186/s40168-018-0541-1.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Kim D, Song L, Breitwieser FP, Salzberg SL. 2016. Centrifuge: rapid and sensitive classification of metagenomic sequences. Genome Res 26:1721–1729. https://doi.org/10.1101/gr.210641.116.
- Nayfach S, Camargo AP, Schulz F, Eloe-Fadrosh E, Roux S, Kyrpides NC. 2021. CheckV assesses the quality and completeness of metagenomeassembled viral genomes. Nat Biotechnol 39:578–585. https://doi.org/10 .1038/s41587-020-00774-7.
- Donkor ES, Dayie NT, Adiku TK. 2014. Bioinformatics with basic local alignment search tool (BLAST) and fast alignment (FASTA). J Bioinform Seq Anal 6:1–6.

- Meier-Kolthoff JP, Göker M. 2017. VICTOR: genome-based phylogeny and classification of prokaryotic viruses. Bioinformatics 33:3396–3404. https://doi.org/10.1093/bioinformatics/btx440.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res 29:2607–2618. https://doi.org/10.1093/nar/29.12.2607.
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better faster version of the PHAST phage search tool. Nucleic Acids Res 44:W16–W21. https://doi.org/10.1093/nar/gkw387.
- Garneau JR, Depardieu F, Fortier L-C, Bikard D, Monot M. 2017. PhageTerm: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. Sci Rep 7:8292. https://doi .org/10.1038/s41598-017-07910-5.
- Lowe TM, Chan PP. 2016. tRNAscan-SE on-line: integrating search and context for analysis of transfer RNA genes. Nucleic Acids Res 44:W54–W57. https://doi .org/10.1093/nar/gkw413.
- Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippon A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender JK, Stingl K, Nguyen M, Coppens J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf I, Nykäsenoja S, Olkkola S, Wieczorek K, Amaro A, Clemente L, Mossong J, Losch S, Ragimbeau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. J Antimicrob Chemother 75:3491–3500. https://doi.org/10 .1093/jac/dkaa345.
- 17. Liu B, Pop M. 2009. ARDB—antibiotic resistance genes database. Nucleic Acids Res 37:D443–D447. https://doi.org/10.1093/nar/qkn656.
- Peters DL, Lynch KH, Stothard P, Dennis JJ. 2015. The isolation and characterization of two Stenotrophomonas maltophilia bacteriophages capable of cross-taxonomic order infectivity. BMC Genom 16:664. https://doi.org/10.1186/s12864-015-1848-y.