

Bacillus subtilis: A Potential Plant Growth Promoting Rhizobacteria that Impacts Plant Disease

Deepak Kumari, N. K. Yadav, Rakesh Punia and Garima

Bacillus subtilis is a gram-positive, rod shaped, spore-forming bacterium which is extensively present in the environment. *B. subtilis* develops endospores that can withstand stress. Its sequenced genome have 4 214 630 base pairs which encoded about 4100 proteins. It is used as a model organism for research on sporulation and the behaviour of Gram-positive bacteria with low GC levels. It is not infectious. Proteases and amylases are two of the products it produces. Due to its commercial importance and simplicity for genetic manipulation *B. subtilis* has undergone extensive research. It has low GC content of 43.5%. It lacks an exterior membrane, as is typical of Gram-positive organisms, but it does have a cytoplasmic membrane and a strong cell wall.

Domain: Bacteria

Phylum: Firmicute

Class: Bacilli

Order: Bacillales

Family: Bacillaceae

Genus: *Bacillus*

Species: *Subtilis*

Mechanism of *B. subtilis* in plant disease control

Competition for nutrients and colonizing sites

Root exudates such as amino acids, fatty acids, polyamines, sterols, phenolics, organic acids,

Table 1: List of antibiotics produced by several strains of *B. subtilis*

<i>B. subtilis</i> strains	Plant pathogen	Disease in plant	Antibiotics produced
<i>B. subtilis</i> RB14	<i>Rhizoctonia solani</i>	Damping off of tomato	Iturin A and surfactin
<i>B. subtilis</i>	<i>F. oxysporum</i> f.sp. <i>ciceris</i>	Fusarium wilt of chickpea	Subtilin
<i>B. subtilis</i> AU195	<i>Aspergillus flavus</i>	Aflatoxin contamination	Bacillomycin D
<i>B. subtilis</i> QST713	<i>Botrytis cinerea</i> and <i>R. solani</i>	Damping off	Iturin A
<i>B. subtilis</i> BBG100	<i>Pythium aphanidermatum</i>	Damping off	Mycosubtili
<i>B. subtilis</i> UW85	<i>Phytophthora medicagins</i>	Damping off	Zwittermycin A, kanosamine
<i>B. subtilis</i> fmbj	<i>Aspergillus flavous</i>	Aflatoxin contamination	Bacillomycin D
<i>B. subtilis</i> B47	<i>Bipolaris maydis</i>	Southern corn leaf blight	Iturin A2
<i>B. subtilis</i> SQR9	<i>F. oxysporum</i> f.sp. <i>cucumerinum</i>	Wilt disease of cucurbits	Fengycin and Bacillomycin
<i>B. subtilis</i> CMB 32	<i>Colletotrichum gleosporioides</i>	Anthrachnose	Iturin A, Fengycin and surfactin A
<i>B. subtilis</i> PCL1608	<i>Fusarium oxysporum</i>	Wilt	Iturin A
<i>B. subtilis</i> PCL1612	<i>Rosellinia necatrix</i>	White root rot	Iturin A

Systemic position of the bacterium

The genus was first discovered as *Vibrio subtilis* by Christian Gottfried Ehrenberg in 1835. It was renamed by Ferdinand Cohn in 1872. The *Bacillus* genus was first reported by Cohn in 1872. Currently, the genus includes over 377 species.

nucleotides, organic acids and vitamins. Competition for these nutrients is fundamental key of *B. subtilis* for management of plant disease. Chemotaxis was suggested as the essential characteristic for colonisation. Colonization of plant roots by *B. subtilis* directly contributes to plant disease management. On

chir pine seedlings, *B. subtilis* BN1 showed excellent root colonization.

Plant growth promotion

B. subtilis produce several phytohormones including auxins, cytokinins, gibberellins and ethylene as well as enzymes like 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase that helps to regulate the growth and development of plants. Additionally, this encourages nitrogen fixation, enhanced root growth, nutrient mineralization (such as phosphate, potassium, and zinc solubilization) and enhance absorption ability of roots. Application of *B. subtilis* GB03 and *B. amyloliquefaciens* IN937 mixture helps to enhance the growth of *Arabidopsis* mutants deficient in IAA. Cucumber seedling growth was stimulated by *B. subtilis* B579 through phosphate solubilization, IAA and siderophore production.

Production of cell wall hydrolytic enzymes

Various strains of *B. subtilis* synthesize a wide range of hydrolytic enzymes i.e. cellulases, proteases, and β -glucanases. The diminished growth of the *Rhizopus stolonifer* fungus on *B. cereus* AR156 and *B. subtilis* SM21 treated peach fruit was due to over expression of the genes for 1,3-glucanase, chitinase, and phenylalanine-ammonium-protein lyases.

Antibiotics production

B. subtilis produces several antibiotics that are very helpful in suppressing the growth and multiplication of the pathogens.

Induction of plant disease resistance

The use of PGPR such as *B. subtilis* may activate host defense mechanisms which results changes in ultrastructure and cytochemical alteration (ISR) against the pathogen. One of the main mechanisms of *B. subtilis* was the elicitation of ISR for controlling plant diseases. It is well accepted that signal transduction pathways elicited by *B. subtilis* are dependent on production of Jasmonic acid (JA), ethylene, and the regulatory gene NPR1, but

independent of salicylic acid (SA), a signal of systemic acquired resistance elicited by pathogens.

Table 2: Commercially available formulations of *B. subtilis*

Commercial product	Active biological agent
Kodiak	<i>B. subtilis</i> GB03
Gallipro	<i>B. subtilis</i> DSM 17299
Calsporin	<i>B. subtilis</i> C-3102
Clostat	<i>B. subtilis</i> PB6
Biotop	<i>B. subtilis</i> CCT 7611
Natto powder- 710	<i>B. subtilis</i> Natto
Fertitacto	<i>B. subtilis</i> PBR5-1
Fertitacto	<i>B. subtilis</i> PBR5-2
Serenade	<i>B. subtilis</i> QST 713
Integral	<i>B. subtilis</i> MBI 600
Companion	<i>B. subtilis</i> GB03
Green Dual	<i>B. subtilis</i>
Quantum-400	<i>B. subtilis</i> GB03
YIB	<i>B. subtilis</i> QST 713

Production of Bacteriocins

Ribosomal synthesized peptides which are produced by numerous bacteria and might be useful against pathogenic bacteria are known as Bacteriocins (Zou et al., 2018). Bacteriocins interfere with the cell wall synthesis or by forming pores in the cell membrane acts against target pathogen. *Bacillus* spp. exhibits a broad-spectrum of antibacterial activity due to the production of bacteriocins. Some bacteriocins and bacteriocin-like substances (BLSs) such as amylolysin, amysin, subtilin, subtilisin A, subtilisin B and thuricin were isolated from various *Bacillus* spp.

Production of siderophores

Siderophores are Fe-chelating, low molecular weight, non-ribosomal peptides which are produced by some microorganisms and plants under iron starvation conditions. Siderophores chelate with Fe, allowing its solubilization and extraction from

minerals and organic compounds by making Fe unavailable for pathogens., *Bacillus* spp. produce a wide variety of siderophores such as bacillibactin, pyoverdine, pyochelin, schizokinen, petrobactin etc. For example, *B. subtilis* acts as promising biological control agent against *Bipolaris sorokiniana* due to siderophores, chitinase and cellulase production of (Villa-Rodríguez et al., 2019). Some of commercially available products are listed along with their Trade name and biologically active strain of *B. subtilis*.

Conclusion

B. subtilis represent an eco-friendly approach to improving crop productivity through various mechanisms of biological control, biofertilization and biostimulation processes. Although it helps to boost crop output and reduce disease incidence. The interaction of bacteria with plants, potential pathogens, and the environment determines whether

B. subtilis can display advantageous characteristics. *B. subtilis* play a significant role in both economic and ecological processes, so more practically significant species must be discovered together with advanced approaches for rapid and comprehensive study and effective application.

References

- Villa-Rodriguez, E., Parra-Cota, F., Castro-Longoria, E., López-Cervantes, J. and de los Santos-Villalobos, S. (2019). *B. subtilis* TE3: a promising biological control agent against *Bipolaris sorokiniana*, the causal agent of spot blotch in wheat (*Triticum turgidum* L. subsp. *durum*). *Biological control*, 132, 135-143.
- Zou, J., Jiang, H., Cheng, H., Fang, J. and Huang, G. (2018). Strategies for screening, purification and characterization of bacteriocins. *International Journal of Biological Macromolecules*, 117, 781–789.

* * * * *