Unveiling the Role of Melanin in Fungal Pathogenicity and Environmental Stress Resistance

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Abstract

Melanin, which is produced by various microorganisms including fungi and bacteria, plays essential roles with a broad range of functions. Melanisation fortifies fungal microorganisms against various environmental stressors including radiation, heavy metals, desiccation, hydrolytic enzymes, oxidative agents, heat, and cold amplifying their virulence as protective armour. This study illuminates the roles of melanin in plant-pathogenic fungi, including its function as a shield against host defence mechanisms, a facilitator of penetration and colonization, and a promoter of pathogen establishment and dissemination.

Introduction

Melanin is formed through the oxidative polymerization of phenolic compounds into high molecular weight pigments. A substance qualifies as amelanin if it is dark in color, insoluble in both aqueous and organic solvents, resilient concentrated acids, and vulnerable to oxidizing agents bleaching action. Fungi, bacteria, and helminths host produce melanin pigments. dihydroxyphenols are the general class of monomers for conjugated polymers. Each polymeric melanin residue incorporates two ortho oxygens. Melanins, extracted mostly from fungi, are derived from the 1,8-dihydroxynaphthalene precursor using the pathway polyketide in ascomycetes deuteromycetes. Basidiomycetes utilize g-glutaminyl-4-hydroxybenzene and catechol as additional monomeric precursors for melanin production. It is a stable free radical, as determined by Electro Spin Resonance (ESR) characteristics, defines melanin as a pigment. These microbes' virulence and pathogenicity for their host plants are associated with their melanin production capability. The prominent discussion concerns the role of melanin in enhancing fungal pathogens' virulence, which has been the most extensively investigated mechanism.

Melanin biosynthesis: Many soil fungi produce melanin, which shields them from various

environmental injuries. The production of large quantities of melanin by melanotic fungi indicates its significance as a fungal metabolic product. 30% of Agaricus bisporus dry weight comprises melanin. A. alternata and Cladosporium sphaerospermum among them have been identified in the Chernobyl nuclear reactor site, an environment where radiation levels surpass 10 thousand times the normal limit is mainly due to the presence of melanin in their cell wall. 1,8di-hydroxy naphthalene and L-3,4dihydroxyphenylalanine as the precursors of DHNmelanin and DOPA-melanin which are the two types mainly involved in tyrosine and polyketide pathway. (Table 1)

Table 1. Types of melanin biosynthetic pathways in fungus

Synthesis	Enzyme	Example
DHN- melanin	Polyketide Synthase (PKS)	Colletotrichum lagenarium Magnaporthe oryzae, Verticillium dahlia Wangiella dermatitidis Alternaria alternata Aspergillus fumigatus
DOPA- melanin	Tyrosinase or Laccase	Cryptococcus neoformans

Relation of fungal melanin to virulence

Many potentially invasive fungi, not all of which are pathogenic, produce melanin. These are called dematiaceous or phaeohyphomycetous fungi (Table 2). During appressorial differentiation and late stationary mycelial growth, fungal plant pathogens synthesize DHN melanin. An asexual spore's germ tube gives rise to the appressorium as a solitary cell. The appressorium is mostly melanized, with the exception of its pore, through which the fungus interacts with the host plant cell. The lack of melanin at the appressorium pore enables the discharge of cell wall degrading enzymes and intake of nutrients. Melanin synthesis during appressorial growth is controlled by developmental factors. These fungi viz., Magnaporthe grisea, Colletotrichum lindemuthianum,



Colletotrichum lagenarium require melanin formation in their appressorium for successful invasion of rice, bean, and cucumber, respectively. The penetration peg, rooted in the appressorium pore, pierces the host cell. The immense turgor pressures, over 80 bar, aid in penetrating the host cells epidermal cuticle. Melanin in melanized appressoria acts as a permeability barrier to glycerol. The penetration peg of Colletotrichum graminicola exerts about 17 micronewtons of pressure on the host cell.

Table 2. Mechanisms developed by fungus against various host produced chemicals

various nost produced chemicals					
Mechanism	Target	Activity			
	pathogen				
Protection	Cryptococcus	Reduction of Fe ³⁺			
against	neoformans	toFe ²⁺			
oxidants	Aspergillus	Redox buffer			
	niger	Against			
	Alternaria	permanganate			
	alternata	andhypochlorite			
	Sporothrixsc	Against ROS through			
	henckii	redox buffer action			
	Aspergillus	ROS quenching			
	fumigatus	mechanism			
	Magnaporthe				
	grisea	Hydrogen peroxide			
	Proteus	scavengers			
T(C)	mirabilis	A1. 1			
Effect on	Cryptococcus	Altered surface			
Phagocytosi	neoformans	hydrophobicity and			
S	Exophiala	charge			
	dermatitis	Produce defensins,			
	Sporothrixsc henckii	microbicidal peptides			
D 1.					
Resistance	Yeast	Capsofungin activity			
toantimicro	Moniliniafru	Against			
bial	iticola	dicarboximide			
compounds	Botrytis	Conversion of			
	cinerea	resveratrol by laccase-			
		Fungal toxin			

Role of melanin in environmental protection

The melanin production of free-living microbes is linked to an environmental survival advantage. Melanin protects against UV damage by absorbing a wide spectrum of electromagnetic radiation. Compared to non-melanized fungi, melanized fungi exhibited greater resistance. The Chernobyl Reactor no 4 is most notably colonized by black fungi, demonstrating exceptional resilience to

radiation. It is involved in absorption and degradation of environmental pollutants, protection of microorganisms from radiation and oxidation, neutralises the ROS released into the environment. Various mechanisms underlying resistance was listed below in Table 3.

Table 3. Melanin induced resistance against various environmental stresses

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to UV light Phaeococcomyces Alternaria alternata Suspensions generated from fungacell walls	Mechanism	Target pathogen	Activity			
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Cladosporium sp. Oidiodendron cerealis Bind to Cryptococcus heavy neoformans metals Protection from neoformans predators Protection from solution predators Protection from Sclerotinia spp. from Sclerotinia spp. enzyme degradation Cladosporium sp. generated from fungation silver nitrate susceptible for cell wat degrading enzymes Enzyme sequestration Thielaviopsiabasicola		Alternaria alternata	suspensions			
Oidiodendron cerealis Bind to Cryptococcus heavy neoformans metals Protection Cryptococcus from neoformans predators Protection Rhizoctonia spp. from Sclerotinia spp. enzyme degradation Oidiodendron cerealis from fungacell walls Binds to silver nitrate susceptible for cell wa degrading enzymes Enzyme sequestratio		Cladosporium sp.	-			
Bind to Cryptococcus Binds to silver nitrate metals Protection Cryptococcus from neoformans susceptible for cell wa degrading enzymes Protection Rhizoctonia spp. Enzyme sequestration of the degradation Thielaviopsiabasicola		Oidiodendron cerealis	from fungal			
heavy neoformans silver nitrate metals Protection Cryptococcus Less from neoformans susceptible predators for cell wa degrading enzymes Protection Rhizoctonia spp. from Sclerotinia spp. enzyme Verticillium sp degradation Thielaviopsiabasicola			_			
metals Protection from neoformans predators Protection from predators Protection from from from Sclerotinia spp. Enzyme sequestratio enzyme degradation Thielaviopsiabasicola	Bind to	Cryptococcus	Binds to			
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from Sclerotinia spp. sequestratio enzyme Verticillium sp degradation Thielaviopsiabasicola	Protection	Rhizoctonia spp.	Enzyme			
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Phomosis sn	_	Thielaviopsiabasicola				
Thomosio sp.	0	Phomosis sp.				
Alternaria kikuchiana		Alternaria kikuchiana				
Phaeococcomycessp.		Phaeococcomycessp.				
Magnaporthe grisea		Magnaporthe grisea				
Moniliniafruiticola		Moniliniafruiticola				
Aspergillus sp.		Aspergillus sp.				

Conclusion: Melanin synthesis in *C. neoformans* satisfies the stringent conditions for being recognized as a virulence factor and serves as a prime model for grasping the role of melanogenesis in microbial pathogenesis. This treatment can trigger severe tumour reactions, potentially harming the host. The significance of melanization in microbial pathogenesis is ensured by its link to virulence and the widespread synthesis of melanin pigments by various pathogens. Melanotic fungi synthesize and modify melanin within their cell walls during growth. It enhances fungal pathogenicity by providing protection against environmental and host defenses, strengthening the cell wall, and increasing resistance to antifungal agents. Effective cell division and morphogenesis



require synchronization with melanin synthesis and degradation processes. Understanding melanin's role could lead to novel approaches in managing fungal infections. We are hopeful that this will significantly boost research in this mysterious field.

References

- Piattelli, M., Fattorusso, E., Nicolaus, R.A., & Magno, S. (1965). The structure of melanins melanogenesis: and Ustilagomelanin. Tetrahedron, 21(11), 3229-3236.
- Aghajanyan, A., Mikaelyan, A., Martirosyan, H., &Melyan, G. (2024). The role of plantderived melanin in enhancing in vitro growth and nutrient accumulation in potato varieties. Bioactive Compounds in Health and Disease, 7(10), 511-524.
- Ramachandra, S.K., Kaushik, S., Austin, H., Kumar, R., Parthiban, M. (2023). The Role of melanin in plant pathogenic fungi: insights into structure, biosynthesis, and function. Journal of Plant Biology and Crop Research, 6, 1090.
- Glagoleva, A.Y., Shoeva, O.Y. & Khlestkina, E.K. (2020). Melanin pigment in plants: Current and future knowledge perspectives. Frontiers in Plant Science, 11, 770.

- Wolkow, P.M., Sisler, H.D., & Vigil, E.L. (1983). Effect of inhibitors of melanin biosynthesis on structure and function of appressoria of lindemuthianum. Colletotrichum Physiological plant pathology, 23(1), 55-71.
- Butler, M.J., Gardiner, R.B., & Day, A.W. (2005). Degradation of melanin or inhibition of its synthesis: are these a significant approach as a biological control of phytopathogenic fungi? Biological Control, 32(2), 326-336.
- Nosanchuk, J.D., & Casadevall, A. (2003). The contribution of melanin to microbial pathogenesis. Cellular microbiology, 5(4), 203-223.
- Howard, R.J., & Ferrari, M.A. (1989). Role of melanin in appressorium function. Experimental Mycology, 13(4), 403-418.
- Toledo, A.V., Franco, M.E.E., Lopez, S.M.Y., Troncozo, M.I., Saparrat, M.C.N., & Balatti, P.A., 2017. Melanins in fungi: Types, localization and putative biological roles. Physiological and molecular plant pathology, 99, 2-6.


