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Review Article

Emerging Noninvasive Biomolecular Checkpoints for Healthy Microbiomes

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Dysbiosis commonly impacts otherwise healthy individuals over the age of 50 due to seemingly innocuous changes in diet, hydration, or physical activity. Noninvasive biomarkers are emerging as checkpoints for immune health, energy-yielding metabolization, and oxidative stress during the aging process. These biomarkers work individually and collectively to detect dysbiosis, injuries, and infections, providing opportunities for earlier interventions with greater certainty of a positive long-term outcome. Measurements of C-reactive protein (CRP), white blood cells (WBCs), and neutrophil elastase are reliable biomarkers of persistent immune response. The absence of urinary bikunin is an accurate measure of immune system recovery during injuries, surgeries, and infections. Oxidative stress by-products of 4-hydroxynonenal (HNE), such as HNE-albumin adduct, allow assessing immune exhaustion and poor cell oxygenation. Overwhelming the immune system reduces the ability of monocytes (CD14) to transform into macrophages and impairs the energy-yielding metabolism signaling of adipokines, lowering the ability to improve cardiorespiratory fitness (CRF) or achieve significant weight loss. Bacterial endotoxins in urine are reliable indicators of ongoing infections and dysbiosis of the gut. Efficient gut microbiome health is predicted by dietary metabolites spilled into urine such as β -hydroxybutyrate, 2-methylbutyrate, 1,5-anhydroglucitol, enterolactone, enterodiol, carboxy-4-methyl-5-propyl-2-furanpropanoic acid, p-cresol, hydroxytyrosol, ethyl glucuronide, and F2-isoprostane and with blood markers like ferritin, homocysteine, and total cysteine. Vitamins B12 and D and folate are also key biomarkers that can monitor nutrient absorption during the aging process.

Keywords: adiponectin receptor; health aging; infectious disease; innate immunity; macrocytic anemia; metabolic syndrome (metS); microbiome; nutrition; vitamin deficiencies

1. Introduction

Dysbiosis is a microbiome etiology whereby the commensalism and mutualistic bacteria needed for healthy digestion are overwhelmed by pathogenic bacteria causing onset of chronic inflammation and oxidative stress (Figure 1) [1–7]. Dysbiosis leads to dysregulation of energy-yielding metabolization and occurs with nutritional imbalances in carbohydrates, protein, and fats as well as by antibiotic usage [2, 8–12]. The gastrointestinal system digests complex biomolecules from food cells after commensal bacteria metabolize food cell wall membranes, which then releases carbohydrate, peptides, lipids and nutrients to the host's enterocytes lining the intestines and then into blood circulation (Figure 1) [13,

14]. Intestinal enterocytes have tight intracellular junctions preventing intact bacteria from crossing into the blood stream and allow intercellular digestion pathogenic bacteria into glycopeptides and endotoxins [13, 15]. During dysbiosis, enterocytes will release more endotoxins into blood circulation, causing a systemic activation of inflammasome stress responses [16, 17]. Prolonged systemic inflammasome stress reduces the immune system's ability to respond to localized injuries or infections and will eventually lead to immune exhaustion [18]. Nutritional interventions for dysbiosis reduce endotoxins and oxidative stress, thereby improving immune health [2, 8–11].

Chronic inflammatory stress during aging is often associated with metabolic syndrome (metS) and underlying

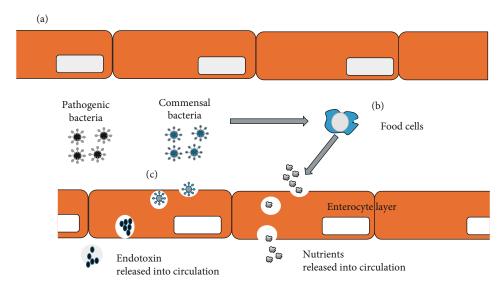


FIGURE 1: Gut metabolism of bacteria and nutrients. (a) The enterocyte layer of the gastrointestinal system prevents bacteria from crossing into the blood and invading the body. (b) Enterocytes digest complex biomolecules from food cells after commensal bacterial digestion and release carbohydrate, peptides, lipids and nutrients into blood circulation. (c) The enterocyte layer also digests excessive bacteria during dysbiosis and releases endotoxins and glycopeptides into blood circulation causing systemic oxidative stress and inflammation to tissues. Systemic endotoxins reduce the immune system's ability to response to specific injury and infection sites.

insulin resistance, which are well known to lead to higher morbidity and mortality risks associated with chronic diseases [19, 20]. Insulin resistance reduces the energy balance by impairing the body's ability to metabolize fatty acids and carbohydrates, which in turn increases obesity but also diminishes the immune response [21]. Diminished energy balance and immunity at age 60 and over are characterized by obesity, reduced mobility, poor fitness, loss of cognitive function, and poor gut microbiome [22, 23]. Persistent inflammatory responses damage tissues, leading to diabetes, autoimmune disorders, and epigenetic cellular changes in tissues [22, 24]. Healthy aging has been defined as increased life expectancy and quality of life, as measured by improvements in chronic inflammatory levels [25, 26]. Fitness is also an important healthy aging practice that improves chronic inflammatory levels [27, 28]. Poor nutrition is now well understood to alter gut health and impair energy-yielding metabolism, diabetes, and insulin resistance during aging [22, 23].

Diagnostics such as hemoglobin A1c (hbA1C) are examples of biomarkers that have been successfully used to promote healthy aging by monitoring the development of diabetes [29]. This biomarker measures the 30-day average of hyperglycemia, allowing a convenient quarterly check for improvements. The predictive value of biomarkers for health screening is greatly magnified when a noninvasive sample like saliva or urine allows repetitive measurement [30]. Calculation of biomarkers in saliva and urine requires accounting for specimen specific gravity [31]. Other common diagnostic biomarkers and risk factors such as lipidemia, hypertension, obesity, and the gut microbiome have failed to predict damaging impacts during aging early enough for interventions before such irreversible damage occurs [19, 20, 32]. Cardiorespiratory fitness (CRF) levels upon recovery after a defined exercise challenge are better measured by using digital tools and fitness equipment than

by a biomarker [33, 34]. Measuring respiratory function by CRF is important for managing recovery from respiratory infections and injuries [33, 35]. Step and heartbeat monitors are common digital tools used for motivating fitness. However, they offer poor diagnostic accuracy for determining overall fitness level (receiver operating characteristic area under the curve [ROC AUC score] of 0.50-0.60) in nonelderly populations (age < 60). The system biology of the interaction between the microbiome and human nutrition is an emerging topic [22, 23]. New biomarkers are emerging as a means for gauging a person's unique nutrigenomic profile as aging progresses due to ever-increasing sensitivity of new bioanalytical methods [36, 37]. Noninvasive biomarkers are evaluated herein for their ability to predict improved immune response, metabolic energy, and oxidative stress levels to provide a scientific foundation for developing an in vitro diagnostic (IVD) panel for healthy aging.

2. Healthy Aging Diagnostics

2.1. Immune Health

2.1.1. Innate Immunity Response. Healthy aging requires a fully functionating innate immune system to completely resolve managing injuries and infections [19, 38]. The impact of innate immune system response to injury, infection, and dysbiosis is shown in Figures 2a, 2b, and 2c. Pathogenic bacteria are commonly acquired during aging as upper respiratory tract infections, urinary tract infection, or in wounds and injuries, as well as entering the gastrointestinal system in food [37]. These microbes can progress to bacteremia or bacteriuria with $> 10^4$ colony-forming units (CFUs)/milliliter [37]. Pathogenic bacteria may overcome the commensal bacteria of the host by producing virulence factors which increase pathogenicity and survival against

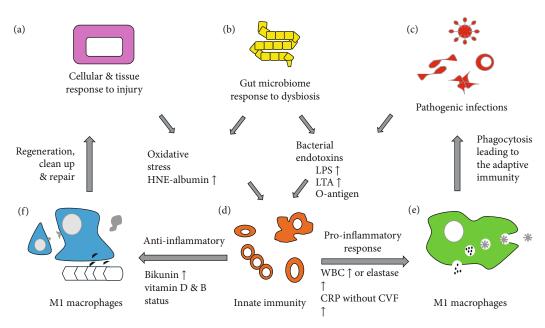


FIGURE 2: Innate immune response to injury, dysbiosis, and infection. (a) Epithelial cell, endothelial cell, smooth muscle, fibroblast, platelets, and other cells produce oxidative stress when tissues and organs are damaged during injuries, infections, and chronic disease. Oxidative stress triggers the innate immunity to release kallikrein, thrombin, plasmin, factors VII and X, and trypsin serine proteases to cause swelling, clotting, vascular dilation, and tissue remodeling. (b) The gut microbiome releases bacteria endotoxin into blood causing additional oxidative stress reflecting the balance of pathogenic and symbiotic microbiota during dysbiosis problems brought on by infectious illnesses, certain diets, or the prolonged use of antibiotics or other bacteria-destroying medications. (c) Microbes, parasites, and viruses which enter the body during infections through the respiratory, urinary tracts, gut, and wounds generate and release endotoxins into blood which causes additional oxidative stress and reflects cellular damage due to infection. (d) Neutrophilic innate immune cells migrate to inflamed tissues under oxidative stress and initiate a proinflammatory response to infections or tissue injury. Red blood cells respond by increasing oxygenation and initiating coagulation to repair hypoxic tissue under oxidative stress. Monocytes shed the C-terminal fragment of adiponectin receptor (AdipoR CTF) in response to oxidative stress to transform into M0 macrophages (CD14+/CD63+/TACE+) to initiate the adaptive immune response. (e) Neutrophilic innate immune cells release elastase to initiate pathogen destruction (phagocytosis). M1 macrophages (CD14+/CD63+/TACE+/CD206+ [mannose receptor]) initiate adaptive immunity for antigen presentation by responding to interferon gamma (IFN-y). (f) Increased elastase releases the serine protease inhibitor, bikunin, to initiate the anti-inflammatory cascade. M2 macrophages (CD14+/CD63+/TACE+/CD220+ [insulin receptor]) initiate extracellular membrane remodeling, cellular regeneration, and clean-up of apoptotic cells by responding to IL4 and IL-13.

the host's immune response and antibiotics [17]. Bacteria cell lysis releases lipopolysaccharide (LPS), lipoteichoic acid (LTA), and O-antigen cell wall components during infections and dysbiosis which act as endotoxins, causing oxidative stress. Trace levels of endotoxin are now measurable by new technologies allowing earlier assessments of nutritional interventions for dysbiosis, injuries, and infection (Table 1) [2, 7, 37].

Chronic inflammation signifies a persistent innate immune cell response which is typical of chronic diseases of aging [20]. The innate immune system is driven by white blood cell (WBC), primarily composed of 60%–70% granulocytes (polymorphonuclear leukocytes), which include neutrophils, eosinophils, and basophils Figure 2d [38]. Granulocytes generate pro- and anti-inflammatory signals to fight off viruses, bacteria, fungi, or parasites and to repair injuries in tissues and organs. Neutrophils are the most common granulocytes, created in the bone marrow and typically circulating in the bloodstream for 6–10 h prior to self-destructing after one burst of activity. Injured cells and tissue, gut microbiome dysbiosis, and infectious pathogens all lead to endotoxins and oxidative stress in the circulatory sys-

tem, which are now measurable by new high-sensitivity bioanalytical methods [37, 39–42]. Elevation in the number of gram – and + bacteria beyond $> 10^4$ CFU/mL in a blood drop, urine, or saliva sample indicates an active pathogenic infection while $> 10^2$ CFU/mL a residual infection (Table 1) [37, 39–42]. Endotoxins (LPS, LTA, and O-antigens) are released from the gut and infections to the bloodstream for elimination into urine [2, 7, 37]. Notably, pathogen levels in the gut and during infections are being impacted by modern methods of food production and nutrition [43, 44].

The pro- and anti-inflammatory response of the innate immune system to endotoxins and oxidative stress is shown in Figures 2d, 2e, and 2f. Elevation of WBC and elastase in blood, urine, or saliva is a reliable indication of an active innate immune response generating a proinflammatory response [19, 40, 45–48]. Complete cell counts of leukocytes (CD45+), granulocytes (CD45+CD15+), and monocytes (CD45+CD14+) using microscopy, flow cytometry, or immunocytochemistry (ICC) demonstrate elevation in the number of leukocytes (CD45+) in a blood drop (>12,500/ μ L), urine (>10/ μ L), or saliva [30] sample (>230/ μ L) and therefore indicate an active innate immune response (see

TABLE 1: Noninvasive biomarkers for healthy aging.

Biomarkers	Biofluid	Process	Conditions detected
Immune health			
Bacterial endotoxins	Blood drop, saliva, urine	Phagocytosis	Infections, dysbiosis, and poor nutrition [2, 7, 37, 39-44]
WBC	Blood drop, saliva, urine	Active innate immunity	Proinflammatory response and CRF [19, 35, 40, 45-49]
Neutrophilic elastase	Blood, urine	Active innate immune response	Immune stress due to cytokines, INFy, and chemokines [19, 40, 45–48, 50, 51]
CRP	Blood drop, saliva	Proinflammatory	Proinflammatory response autoimmunity and infection [38, 52–60]
Bikunin	Urine	Anti-inflammatory	Anti-inflammatory response healing during infections and injury [19, 38]
Vitamin D	Blood drop, urine	Antioxidant capacity	Vitamin D deficiency and immune responsiveness [8, 61–69]
Active Vit B12 and folate Energy-yielding metabolism	Blood drop, saliva	Microbiome health	Immune response and good gut bacterial growth [66–87]
eta-Hydroxybutyrate	Urine	Metabolic efficiency	Good ketogenesis with fatty acid uptake, β -oxidation, and lipogenesis [88–90]
Adiponectin and leptin	Blood drop	Poor metabolism	Poor adipose health for signaling of oxidization of fatty acids (obesity) [19, 20, 51, 88, 91]
Autoantibodies to AdipoR CTF	Blood drop	Poor metabolism	Deactivation of the AMPK response for fatty acid uptake, β -oxidation, and lipogenesis [19, 20]
HbA1c	Blood drop	Poor metabolism	Prediabetic hyperglycemia and damaging advanced glycation end products [52, 90]
2-Methylbutyrate	Urine	Dysbiosis	Unhealthy nutrition with carbohydrate or protein overload causing fermentation stress of gut bacteria [10]
1,5-Anhydroglucitol	Urine	Dysbiosis	Hyperglycemia, overdoing high-carbohydrate diet (HCD) [92]
Enterolactone and enterodiol	Urine	Microbiome health	Sufficient fiber diet for good gut bacterial growth [93, 94]
Ferratin	Urine, saliva	Dysbiosis	High-protein diet causing oxidative stress and bacterial fermentation [10, 95, 96]
Carboxy-4-methyl-5-propyl-2-furanpropanoic acid	Urine	Microbiome health	Good fish protein and 3nFA diet for reduced oxidative stress [95, 97]
p-Cresol	Urine	Dysbiosis	Overdoing soy leading to cytotoxic microbiome imbalance [95]
Hydroxytyrosol and ethyl glucuronide	Urine	Dysbiosis	Excessive alcohol consumption leading to cytotoxic microbiome imbalance [98, 99]
F2-Isoprostane	Urine	Microbiome health	Good natural antioxidative diet levels including fruits, vegetables, cereals, and nuts [100–102]
Homocysteine/total cysT	Urine	Poor metabolism	Insufficient protein diet, unhealthy lean weight, and need of protein intake [10, 95, 96]
Oxidative stress			
Ferratin and transferrin hemoglobin	Blood, saliva	Cell senescence and poor oxygenation	Macrocytic age-related anemia [74, 75, 103-106]
Active Vit B12 and folate	Blood drop, saliva	RBC oxygenations	Macrocytic age-related anemia [74, 107, 108]
HNE-albumin ratio to albumin	Blood, saliva, urine	Oxidative stress, poor oxygenation	Overwhelmed immune system, tissue necrosis, vascular thrombus, and fibrosis damage to poor oxygenation [109–112]

Table 1) [19, 40, 45–48]. Human neutrophilic elastase is a well-established marker of neutrophilic exposure. A release above a threshold of 10 neutrophils/ μ L is significant in any biofluid (see Table 1) [39, 50, 51]. Neutrophils are motile, entering into the intestinal spaces of tissues upon vascular permeation due to proinflammatory signaling of swelling, clotting, vascular dilation, and tissue remodeling, allowing phagocytosis of pathogenic bacteria [7].

WBC counts are strongly correlated to CRF during aging [113, 114]. The National Health and Nutrition Examination Survey has demonstrated WBC counts as a measure of risk of chronic disease, infections, cancers, morbidity, and mortality [33, 49]. A person's physical fitness does improve personal WBC counts, cardiovascular risk, and respiratory function [33]. Other inflammatory biomarkers associated with elevated WBC such as cytokines, chemokines, and growth factors are also predictive of healthy aging in many cases [88, 115]. Fitness interventions complemented with restriction diets can lead to greater body mass index (BMI) loss and lower WBC counts within 1 year, with neutrophilic leukocytes and platelets most impacted. Other inflammatory markers showed no statistically significant changes upon intervention, such as Creactive protein (CRP), interleukins (IL-6, IL-8), tumor necrosis factor- α (TNF α), c-peptide (inactive insulin form), and T cell markers of adaptive immunity. Additionally, monocyte chemoattractant protein (MCP-1) and IL-18 biomarkers were statistically significantly activated neutrophilic leukocytes during autoimmune disease.

Continuous chronic inflammation increases oxidative stress and leads to excessive protease activity where receptors, like the adiponectin receptor (AdipoR), are deactivated and immune exhaustion is likely [19, 51]. Chronic inflammation can be reduced with a combination of nutrition and fitness [52, 53]. High-sensitivity CRP is a blood biomarker that has been used to monitor improvements in chronic inflammation (2–10 mg/L) in cardiovascular disease [54-56]. Values above 10 mg/mL reflect true clinical inflammation due to infection, injury, or autoimmune disease [38]. Saliva CRP measurements are only slightly reflective of blood value due to diurnal variation and elevate from 238.5 ± 94.78 to 1519.5 ± 660.4 pg/mL during inflammation [57]. Values of CRP only improve with significant weight loss and improved CRF [33, 58]. Notably, many common anti-inflammatory medications, such as aspirin and nutritional supplements, such as vitamins C and E, omega-3 fatty acids, and zinc, lower CRP levels without corresponding changes in the WBC counts [54, 59, 60].

2.1.2. Bikunin Response. Bikunin in urine is a reliable indicator of infection and/or recovery from injuries [19]. Bikunin is a serine protease inhibitor released by neutrophilic elastase to suppress proinflammatory serine proteases [19, 38]. As a drug named ulinastatin, bikunin is well established as protecting the body during acute circulatory failure, sepsis, ischemic injury, cardiac arrest, traumatic brain injury, pancreatitis, and many conditions from immune-mediated apoptosis [19, 116–119]. Bikunin is rapidly eliminated from the body during infection or injury so that the immune system can continue to repair tissue damage and eliminate patho-

gens (Table 1) [19]. Urinary bikunin is a well-studied biomarker for chronic inflammation and more sensitive than CRP, persisting until the immune system returns to normal status [19, 38]. Bikunin correlates strongly with WBC counts and is formed by neutrophilic elastase whereas CRP is produced by the liver in a delayed mechanism to activate C3 complement and cause opsonization. Bikunin reflects worsening inflammation in patients > age 50 with metS, diabetes, and chronic diseases in a range from 2.0 to 7.5 mg/L. These patients are generally overweight (32% with BMI 25-30) or obese (52% with BMI > 30). Bikunin correlates with lower risk of comorbidities when conventional diagnostics fail to predict death and complications. Bikunin values above 7.5 mg/mL are diagnostic of clinical injuries and systemic infections, upper respiratory infections, urinary tract infections, cardiovascular tissue injury, glomerulus nephritis, and pancreatitis [38].

Bikunin as a serine protease inhibitor prevents prolonged proteolysis and is an anti-inflammatory response that protects cells and tissues from immune-mediated apoptosis leading to irreversible cellular changes and autoimmune disease (Figure 2e) [120]. Prolonged neutrophilic exposure by elevated WBC counts causes tissue damage by constant activation of immune-mediated apoptosis through proinflammatory signaling caused by constant release of inflammatory proteases (neutrophilic elastase, cathepsin G, proteinase 3, and granzyme B) leading to shedding of cytokines (IL-1, IL-6, IL-8, IL-10, and TNFα), interferon gamma (IFN-γ), chemokines (MCP-1, CXCL9, CCL11, and CXCL), and growth factors [19, 20, 38, 51]. Prolonged elevation of bikunin indicates a proinflammatory state where immune cells are causing damage to tissues and inducing poor wound healing [19, 20, 38]. Bikunin inhibits the release of cytokines, chemokines, and growth factors in the Janus kinase and signal transducer and activators of transcription (Jak-STAT) pathway, increasing protein kinase B (Akt) and phosphatidylinositol 3-kinase (PI3K) phosphorylation by blocking the potassium large conductance calcium-activated channel (KCNMA) [19, 116-119]. Bikunin also inhibits urokinase activation of mitogen-activated protein-extracellular signal-regulated kinase (MAPK/ERK) by plasmin, protease active receptor (PAR) activation of protein kinase C by trypsin, and granzyme B activation of caspase 8. It further blocks the extrinsic death factor receptor to help prevent immunemediated apoptosis and promote tissue generation [19]. Bikunin also inhibits epithelial cell, endothelial cell, smooth muscle, fibroblast, and platelet release of serine proteases (kallikrein, thrombin, plasmin, trypsin, and factors VII and X) from causing additional swelling, clotting, and vascular dilation [19, 38].

2.1.3. Macrophage Response. The macrophage response of the innate immune system is triggered by endotoxins and oxidative stress and an imbalance caused by injury, dysbiosis, and infection (Figures 2d, 2e, and 2f). Monocytes (CD45+CD14+) are the third most common type of innate immune cells and account for 2%–10% of leucocytes in the blood and are essential to immune health, with elevation of > $800/\mu L$ indicative of an active innate immune response

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(Table 1) [19, 20, 38, 51]. Monocytes transform into macrophages as they migrate into body tissues for phagocytosis by first differentiating into M0 macrophages (CD45+CD14 +CD63+) due to the release of neutrophil protease and the presence of oxidative stress in tissue [51]. M0 macrophages become proinflammatory M1 macrophages (CD45+CD14 +CD63+TACE+CD206+) in response to IFNy and bacterial endotoxins (LPS, LTA, and O-antigen) to activate phagocytosis [51]. Monocytes become insulin resistant during the aging process, becoming less responsive to endotoxins and reactive oxidative species (ROS) [51, 121-123]. M0 macrophages influence the classical and nonclassical processes of adaptive immunity and antigen presentation. Monocytes can perform phagocytosis using intermediary opsonizing proteins such as antibodies or complements that coat the pathogen, as well as by binding to the microbe directly via pattern-recognition receptors that recognize specific pathogens. Monocytes are also capable of killing infected host cells via antibody-dependent cell-mediated cytotoxicity. This process occurs with M0 macrophages differentiated into M2 anti-inflammatory macrophages (CD45+CD14+CD63 +CD220+) in response to the presence of IL-4 and IL-13 and cells killed by phagocytosis during repair of tissue.

Vitamin D has a direct endocrine function and regulatory impact on immune health, improving macrophage response, lowering WBC counts, and decreasing immune cell death [8, 61, 62]. Vitamin D improves the total antioxidant capacity and reduces microbiota-generated oxidative stress and production of ROS while increasing glutathione (GSH). Measuring the need for vitamin D supplementation to promote immune health is independent of the effects of common anti-inflammatory medication, and its deficiency is measured by a blood value of < 20 ng/mL (Table 1) [63]. Notably, concentrations of CRP are lowered to 2.02 mg/dL (1.95–2.08) from 2.60 mg/dL (2.41–2.82) after treatment for vitamin D deficiency [64]. Vitamin D deficiency is also measurable in saliva, with values of 17.4 ± 8.0 ng/dL improved to 20.8 ± 6.3 ng/dL upon supplementation [65].

2.2. Energy-Yielding Metabolism

2.2.1. Metabolic Response. Poor energy-yielding metabolism can be attributed to obesity and metS with a commensurate increased risk of morbidity and mortality [124, 125]. The underlying insulin resistance observed with metS and obesity impacts the energy metabolism of the body and impairs CRF by limiting the ability of the body to oxidize fatty acids and restore energy [19]. Progression to the onset of diabetes is diagnosed by an HbA1c level greater than > 6.5% or by a fasting blood glucose or oral glucose tolerance test. Continued hyperglycemia leads to advanced glycation end products attaching to proteins which in turn degrades cell function and destroys elastin and collagen in skin and tissue [19]. Treatments to reduce hyperglycemia help to reduce weight but do not necessarily address the underlying insulin resistance or progression of chronic diseases [19]. Insulin resistance can be measured by understanding the compartment model of pancreatic secretion of insulin, its utilization, and metabolization [19]. It is not sufficient to just measure pan-

creatic insulin secretion to determine insulin resistance. Fasting to determine glucose and insulin values over time must be considered [19]. Blood and urine levels of β -hydroxybutyrate provide a measure of metabolic efficiency for carbohydrate and fat burning under lower lipid peroxidation levels stress during caloric restriction diets without a fasting collection (Table 1) [89]. Insulin resistance impairs the ability of the body to respond to oxidized fatty acids and hyperglycemia, thereby limiting the ability to heal injuries and fight infections [19, 51, 126]. A reduced aerobic fat burning ability increases oxidative stress and occurs with impaired adipose tissue [19]. The health of the adipose can be measured by adipokine, adiponectin, and leptin levels as risk factors for an inability to tolerate a hypercaloric high-fat diet and a significant risk of developing metS and diabetes (Table 1) [19, 127]. Further, while modest elastin/collagen preservation with topical antioxidants can somewhat improve skin health, the underlying metabolic stress is not resolved and further degradation can be expected [128].

2.2.2. Adipokine Response. Adiponectin and leptin are adipokines which provide a measure of adipose health for optimal energy-yielding metabolism [19, 51, 88, 91]. Leptin predicts adipose health and appetite control through the Sh2b1 neuroreceptor directly linked to brown fat transformation which regulates body weight and insulin resistance and which could be impacted by artificial sweeteners [129]. Adiponectin is an insulin-sensitizing hormone released from adipose tissue that explains how cells are activated to oxidize fatty acids and store glycogen for future energy [19]. Adiponectin secretion from the adipose is suppressed during obesity and diabetes, offering a metabolic pathway explanation for insulin resistance [19, 51]. Adiponectin's mechanism of action is not through insulin's activation of Akt but rather through the adiponectin activation of AMP-activated protein kinase (AMPK), which stimulates glucose uptake and lipid oxidation to produce energy [19, 91]. Activation of AMPK reduces oxidative stress in the tissues and organs and increases oxidization of fatty acids in liver and muscle controlling whole-body glucose homeostasis [130].

Adiponectin signaling of AMPK is deactivated by proteolytic shedding of the C-terminal fragment from the adiponectin receptor (AdipoR CTF) (Figure 2e,f) [51]. AdipoR CTF shedding correlates with an increased inflammasome measured by bikunin [20]. In obesity/age models, proteolytic shedding of AdipoR CTF correlates with an impaired energy-yielding metabolism and increases with obesity, age, and poor diet leading to Type 1 and 2 diabetes [19, 51]. Pancreatic damage was observed with increased neutrophil infiltration, leading to AdipoR CTF shedding and the presence of helper T cells and antigen activation presentation [51]. Antibodies to AdipoR CTF can be measured in humans and increase with aging, obesity, and diabetes [19, 20]. A healthy immune system is characterized by continuous shedding of AdipoR CTF [20, 51]. A lack of AdipoR CTF shedding reflects immune system exhaustion and a generally impaired energy-yielding metabolism, increasing the risk of morbidity and mortality in chronic diseases [19, 20]. A prolonged inflammasome leads to loss of proteostasis

or excessive protease activity without inhibition, which in turn leads to complete shedding of AdipoR CTF and hyperglycemia with impaired monocyte immune response [51].

Deactivation of the AMPK response by AdipoR CTF shedding occurs naturally in monocytes transforming into macrophages (M0, M1, and M2) for fighting microbial pathogens and repairing injuries [51, 131]. Loss of AMPK signaling reduces the ability to oxidize fatty acids and increases immune cell sensitivity to oxidative stress [51, 131]. Shedding of AdipoR CTF occurs with the shedding of cytokines like TNF α , IL-6, and IL-12 by the TNF α convertase enzyme (TACE) during the proinflammatory response initiated by any neutrophil protease [51]. Immune cells lacking AdipoR CTF are also unable to bind to an insulin degradation enzyme, which correlates with high intracellular insulin levels characteristic of cellular insulin resistance [51]. Oxidative stress exceeding the active inflammasome response (loss of proteostasis) indicates exhaustion of AdipoR CTF shedding and limits the ability of the immune system to fight infection and make repairs [51]. Bikunin regulates proteostasis to maintain the energy-yielding metabolism of AMPK by inhibiting PAR activation and activating phosphorylation of Akt/PI3 by blocking KCNMA [19, 51].

2.2.3. Nutritional Response. Carbohydrate and protein fermentation by gut bacteria produces short-chain fatty acids like 2-methylbutyrate in blood and urine indicative of dysbiosis (Table 1) [10]. Hyperglycemia produces urinary 1,5anhydroglucitol indicative of an excessive carbohydrate diet (Table 1) [92]. The quantity and source of dietary proteins regulate the production of cytotoxic metabolites by gut microbiota, which alter the rectal mucosa of the host [10]. Amino acids, glycans, and other metabolites generated and digested by gut bacteria create environmental controls that are cytotoxic or promoting of bacterial species [8, 132–134]. Ferritin is indicative of excessive protein intake, while homocysteine and total cysteine predict unhealthy lean weight and need for protein intake (Table 1) [10, 95, 96]. Protein intake in a hypercaloric diet has been linked to metS, obesity, KD, LD, and DM [95, 135, 136].

Measuring nutritional status can improve the microbiota to promote an optimized metabolism [2, 6, 7]. A restricted Mediterranean-inspired diet in the PREDIMED-Plus Trial clearly demonstrated improved glucose metabolism-related parameters (fasting glucose, HbA1c, and insulin resistance) in a diet high in n-3 fatty acids (n-3 FAs) [88, 90]. Weight loss in metS occurred after 12 months and correlated with β -hydroxybutyrate and phospholipids shifting toward n-3 FAs from n-6 fatty acids with improved lipogenesis [88, 90]. The best sources of n-3 FAs are fish followed by red meat, linseed oil, and canola oil [95, 97]. Fish diet and n-3 FAs are measured by urinary carboxy-4-methyl-5-propyl-2-furanpropanoic acid and have shown beneficial effects in glycemic control and insulin sensitivity [95, 97]. The intake of fish increases plasma and erythrocyte n-3 FAs, allowing eicosapentaenoic acid and docosahexaenoic acid concentrations to reduce oxidative stress [137]. Dietary intake of palmitate and oleate also has a broad impact on systemic and tissue lipid profiles in humans [138].

Tolerance to hypercaloric western diets which are heavy in simple carbohydrates, protein, fat, and preservative varies greatly between person and race [139]. Metabolic adaptation in lipogenic transcription factors for fatty acid uptake, β -oxidation, and lipogenesis alters the α -ketoglutarate pathway to remove excess acetyl-CoA carboxylase from the cell mitochondria in the mammalian rapamycin (mTOR) pathway, reducing a person's ability for weight loss and ketogenesis [90]. Mitochondria are the oxidative energy reaction centers in cells that have their own DNA and can produce an excess of ROS, which can then induce programed cell death (apoptosis) and reduce mitochondrial efficiency further, which already diminishes during aging. Intermittent fasting by spacing caloric loads of carbohydrates and fat food reduces oxidative stress [89]. Reducing intake of foods with toxic metabolites reduces mitochondrial stress. For example, urinary excretion of cytotoxic p-cresol is increased in an excessive soy protein diet [95]. Excessive alcohol consumption of 0.2-0.1 g (2-1 drinks)/day increases oxidative stress, which can be measured by urinary hydroxytyrosol and ethyl glucuronide levels [98, 99].

Dietary interventions with indigestible carbohydrates such as whole grain and rye wheat fibers have not yielded improved energy metabolism but did increase commensal gut microbes and reduce lipidemia [128]. A sufficient fiber diet can be measured by urinary enterolactone and enterodiol lignan metabolites [93, 94]. Antioxidant foods including fruits, vegetables, cereals, and nuts include natural antioxidants such as vitamin C, vitamin E, flavanols, anthocyanins, quercetins, and polyphenols at subcytotoxic levels that induce robust cellular in vivo signaling [100-102, 140]. Antioxidant vitamins such as C, D, and E and omega-3 fatty acids have been associated with reduced oxidation, stress, and inflammation [59, 141, 142]. Vitamin C supplementation causes a reduction in inflammation (CRP and IL-6) improved energy-yielding metabolism by blood glucose, triglycerides, and hbA1C after 8 weeks of treatment [143–145]. Concentrations of n-3 and n-6 fatty acids in saliva are reflective of food intake and not chronic oxidative stress [146]. Vegetarian diets with natural vitamin E (< or 100 IU/day resveratrol) improve WBC counts and CRP and may have antithrombotic effects. However, supplemented vitamin E requires levels in the cytotoxic range (400 IU/day α -tocopherol) to decrease lipid oxidative stress [147, 148]. Metabolites of F2-isoprostanes measure the effects of an antioxidative diet [100-102]. The effects of ingesting tomatoes, plantains, grapes, legumes, nuts, and citrus are detected by urinary glycerates, stachydrine, hypaphorine, tryptophan, and proline derivatives [92, 136, 149]. A lack of plant-based diet and overdoing protein intake can be detected by urinary gamma glutamyl peptides due to overwhelming of GSH homeostasis [95].

Precision nutrition (nutrigenomics) is gaining interest as a means to address nutrient and vitamin deficiency on a personalized basis by measuring specific biomarkers [150]. Vitamin B deficiency (B1, 2, 3, 5, 6, and 12, folate, and biotin) is well studied and known to increase with age, metS, and an inadequate diet [70–72]. Gut absorption of vitamin B does not decline with age but rather the cell metabolism

and signaling diminish as well [73–80]. Vitamin Bs are known to regulate bacterial growth and are strongly associated with improved immune cell response and an optimized gut microbiome [151–153]. Markers of gut microbiome health are important to judge the idiosyncratic and highly individualized response to vitamin and nutrient supplements [6, 154]. Treatments with vitamins B and D boast immune health claims against various pathogens, reducing inflammation and oxidative stress and improving metabolic processes [61, 66–69]. Supplementation for vitamin B and D deficiency is known to reduce the risk or severity of cardiovascular disease, stroke, sepsis, dementia, and other conditions [81, 82, 152].

Testing for vitamin B12 and folate deficiency is a wellaccepted component of the optimization of an energyyielding metabolism (Table 1) [72]. Vitamin B deficiency can be measured by immunoassay for the holotranscobalamin complex (active B12) that promotes the uptake of cobalamin by all cells via specific receptors, including leukocytes, neutrophils, and monocytes [83-85]. Active B12 is normally at 200-900 pg/mL in serum, while values of 30-200 and <30g/mL indicate borderline and clinical deficiency [65, 66, 83-87]. Folic acid deficiency leads to vitamin B deficiency at <2 ng/mL in serum, with >4 ng/mL considered normal. Endogenous folate is abundant in saliva due to diet intake of 0.32 pmol/ng saliva, but systemic deficiency can be measured by immunoreactive folate-binding proteins [155]. Active B12 and folic acid-binding protein are found in saliva, but normal and deficiency ranges are yet to be established (Table 1) [155, 156].

2.3. Oxidative Stress

2.3.1. Cell Senescence Response. Cell senescence and autophagy during aging can be predicted by impaired immune health and increased oxidative stress biomarker levels (Table 1 and Figure 2d,f) [25, 157, 158]. Tissue survival and regeneration rely on oxygenation for thriving cellular health [159]. Oxygenation is a key factor in tissue health, and management of oxygenation (blood oxygen) declines in pulmonary and vascular diseases associated with aging (Figure 2d) [160]. Vascular thrombus and fibrosis damage impair oxygenation, a self-perpetuating cycle leading to cardiac stress and tissue necrosis, which can be detected by standard coagulation and cardiac panels [55, 103]. Pathological lack of oxygenation (hypoxia) or low partial pressure of oxygen (hypoxemia) leads to tissue necrosis and loss of autophagy in all tissues, including the brain and heart. Necrosis and fibrosis markers specific to vascular and neurological systems are important indicators of ongoing damage [118, 161-164].

Oxidative carbonyl species impact the cellular function of the mTOR pathway implicated in many age-related disorders [51, 90, 165]. Poor injury recovery is characterized by poor monocyte polarization into M1/M2 macrophages and lack of clean-up of cellular materials damaged by reactive oxygen species and resulting oxidative stress [51]. Oxidative adducts of lipids induce monocytes to produce proinflammatory M1 macrophages through the liver X receptor and

to activate clearing microbial pathogens and wounds [109, 148]. Oxidative stress is therefore indicative of poor immune health and persistent biomolecular and cell damage [19, 146]. Increased oxidation of lipids and proteins has been implicated in cardiovascular comorbidities, hypertension, and autoimmune disease. Thus, poor aging is correlated to cellular signaling in the NF-kappa B and Jak-STAT pathways which regulate the death Fas receptor (APO-1) for the natural programmed cell death (apoptosis) need for recycling of cellular materials (autophagy) in order to eliminate by-products of oxidative stress [165, 166]. Further, neutrophilic infiltration causes activation of the Jak-STAT-2,4 pathways through TACE proteolysis and leads to this immune-mediated apoptosis through cytokine and growth factor release [51]. However, continuous proinflammatory stress prevents the signaling switch to the Jak-STAT-1,3 pathway and JNK stress-activated protein kinases needed for tissue regeneration and regulation of senescence through repression of p38 MAP kinase and expression of tumor protein p53, the "guardian of the genome" [167]. The Jak-STAT-1,3 pathway also activates PIWI-dependent nuclear receptor complex formation in this process. The PIWI nuclear receptor binds to piRNAs with a length of ~26-31 nucleotides for directing DNA methylation in epigenetic gene control of cellular differentiation during tissue remodeling [168]. Tissue self-renewal and regeneration are initiated by tissue pluripotent stem cells which rely heavily on growth factor activation of specific stem cell factors, hormone receptors, and hormones. Impaired tissue self-renewal occurs with diminished hormonal response cell responses in aging leading to incorrect cellular differentiation and loss of tissue function [169].

2.3.2. 4-Hydroxynonenal (HNE) Response. Persistent chronic inflammation without repair is characterized and measurable by oxidative stress biomarkers such as HNE [7, 19, 70, 170]. Oxidative stress pathways including superoxide anion, singlet oxygen, hydroxyl radical, hydrogen peroxide, peroxynitrite anion, and nitric oxide all lead to protein, enzyme, lipid, DNA, and cellular damage, inactivation, and programmed death (apoptosis) [70, 171]. Oxidative stress, including hypoxia, results in nitrosative stress, endoplasmic reticulum stress, mitochondrial dysfunction, and carbonylic stress. Nitrosative and oxidative stress are measured by and associated with ROS. Every individual has a highly variable and often dynamic ability to tolerate unhealthy oxidative stress levels. Measuring unhealthy oxidative stress can be inferred through ROS levels but more accurately assessed through fragments, adducts, or metabolites of tissue, cellular, or nucleic damage. Malondialdehyde (MDA) and HNE are aldehydes that result from lipid peroxidation of polyunsaturated fatty acids [70]. Many biomarkers of oxidative stress as MDA, HNE, 8-hydroxy-2'-deoxyguanosine (OHdG), and nitrotyrosine are not stable in biological fluids and are poor markers [7, 19, 70, 170]. The formation of HNE is the end of the chain of reaction of all ROS and nitrosative species. Reactive HNE reacts with proteins, lipids, and glycoconjugate to form stable measurable adducts, allowing the quantitation of a 30-day average level of oxidative stress in an organism [110, 172].

Protein adducts of HNE for serum proteins such as albumin allow measurement of systemic oxidative stress, whereas cell-specific proteins measure specific stress to tissue (Table 1) [109-112]. The attachment of HNE to proteins occurs immediately to form an adduct which can be detected by immunoassays. Adducts to proteins are a stable method for assessment of overall oxidative stress [31, 172, 173]. Adducts of HNE to human serum albumin (HNE-albumin) have been shown highly reproducible within a patient, reflecting a 30-day average oxidative stress correlating with nitrotyrosine, OHdG, and MDA [174]. Albumin has a fairly constant protein presence in blood at 3.4-5.4 g/L, saliva at 0.1-2 mg/L, and urine at 0.5-80 mg/L [19, 31]. HNE-albumin in urine is 0.59 ± 0.09 and 2.0 ± 0.5 mg/dL for low and high inflammation, respectively (Table 1) [31]. Systemic oxidative stress (>1.5 mg/mL HNE-albumin) and chronic inflammation (2.0-7.5 mg/L bikunin) are observed in 69% of adults over 50 without diabetes or cardiovascular disease and 82% of adults with diabetes in the absence of known infections or recent surgeries [31]. The ratio of HNE to albumin is typically s 1:1-1:5 or ~1.5-2.2 nmol of adduct [31]. Oxidative stress is measured relative to the total amount of albumin in the sample (HNE-albumin/albumin) [31]. Measuring oxidative stress adds to the assessment of chronic inflammation by determining if the immune system is addressing oxidative stress factors or is in fact overwhelmed [19, 31, 51]. Antioxidant supplementation like vitamins D and C decreases carbonylic reactive species like MDA and HNE [104, 175, 176]. Vitamin C also decreases oxidative stress by interaction with GSH and paraoxonase-1 [104, 105, 175, 176].

The kidney is a highly sensitive oxygen sensor and mediates red blood cell (RBC) production in response to hypoxia due to release of erythropoietin and activation of the hypoxia inducible factor 1 to increase blood oxygen-carrying capacity production [177, 178]. Slight hypoxia, such as caused by higher altitudes (up to 8000 ft, 2500 m), increases the blood's oxygen-carrying capacity [178, 179]. Slight hypoxia, when combined with good metabolic health, results in increased blood hemoglobin (Hgb) and delays the onset of replicative senescence in neurological cultured cells and extends lifespan in animal models [180, 181]. However, unhealthy blood oxygen saturation (<90%) caused by low-perfusion conditions or high altitudes above 14,000 ft/4300 M closely simulates poor aging [182, 183]. The brain requires an ongoing and stable oxygen supply to support its underlying functions. The role of oxygen in the functional activity of the brain is important in the relationship of healthy aging and its potential pathologies. Clinical signs and symptoms of poor oxygenation are fatigue, dyspnea on exertion, vertigo, palpitation, low blood pressure, pallor, and headaches resulting from diminished delivery of oxygen to tissues. Age-related reduction of mental quality of life is caused by fatigue, headaches, weight loss, impaired balance, mood changes, muscle loss, neuropathy, dementia, weakness, leukopenia, neuropsychiatric behavior, and gastrointestinal issues [72].

2.3.3. Hgb Response. Markers of anemia seek to address the inability of RBCs to transport oxygen [74, 75, 105]. Agerelated anemia is estimated to impact 900 million people

globally over the age of 65 [85, 184]. Improving detection and treatment of age-related anemia improves mental quality of life and plays an important role in the effectiveness of aging care [71, 72]. The primary diagnostic method of general anemia is low Hgb concentration or hematocrit value in blood [103]. Anemia pathophysiology is categorized as being due to RBC efficiency, accelerated RBC destruction, and impaired RBC production and judged by the size and shape of the RBC as distinguished by cytology and hematology. Age-related anemia due to accelerated RBC destruction causing Hgb deficiency can be due to autoimmune disease, toxins, infections, cancers, or medicines and can be detected by Hgb in blood. Age-related pernicious macrocytic anemia is typically associated with impaired vitamin and nutrient absorption [74, 107, 108]. Abnormal RBC response to oxidative stress damage (ferroptosis) has been used to explain impairment [86]. Macrocytic anemias are commonly due to related vitamin B deficiency, hypothyroidism, testosterone deficiency (40-50-year men), or medication such as metformin, aspirin, and nitrous oxide. Monitoring of these hormones is typically performed in an endocrinology assessment of macrocytic anemia [185]. Active B12 and folate measurements are useful. Additionally, biomarkers identified for chronic alcohol consumption, malnutrition, vegetarian diets, and genetic factors further contribute to agerelated macrocytic anemia (Table 1).

Vitamin and nutrient supplements with Vit B, iron, zinc, and iodine are a safe, well-accepted treatment for resolving anemia due to Hgb deficiency and improving nutritional status [104, 106]. Treatments can be monitored by blood immunoassays and iron panels [103]. Hgb deficiency is likely when the serum ferritin level is less than 50 ng per milliliter (112.35 pmol per liter). Serum ferritin values greater than or equal to 100 ng per milliliter (224.70 pmol per liter) generally exclude iron deficiency anemia (Table 1). Transferrin is also measured to estimate the total iron-binding capacity. Lower transferrin with higher ferritin indicates a lower total ironbinding capacity and could indicate increased oxidative stress to RBCs. The normal range for transferrin in serum is 215-380 mg/dL. A predictable amount of transferrin is present per Hgb found. Anemia has also been detected in saliva, where the mean level of salivary ferritin in subjects with iron deficiency was significantly higher at $139.37 \pm 47.90 \,\mu\text{g/dL}$ when compared to the levels in nonanemic subjects at 94.18 ± $62.90 \,\mu\text{g/dL}$ [186–188]. Transferrin is typically < $4.0 \,\text{mg/L}$ in saliva and correlates with the amount of Hgb in saliva, which normally ranges from 0.045 to 2 mg/dL Hgb [45, 189]. The measurement of transferrin and ferritin in saliva for anemia requires accounting for bleeding of the gums by additionally measuring and ratioing to Hgb [31].

3. Challenges and Prospects

Progressing noninvasive biomarker panels to widespread clinical practice will require new technologies enabling one-touch and rapid measurements utilizing novel point of care systems [37]. Analytical validation of autocalibration, standardization, high reliability, and error-free use of these systems must be proven capable when administered by

untrained users. Noninvasive urine or saliva analysis must correct for sample variation caused by changes in sample concentration. Blood finger-prick samples of $\sim\!\!1\,\mu\mathrm{L}$ must correct for collection variations in hematocrit values. Additional clinical validation studies will be needed with larger populations to set normal reference ranges, define quantitation requirements, and establish relationships needed for promoting healthy gut microbiomes [30]. Each biomarker will have to undergo independent validation before the integration of multiplexed panel results can be applied to machine learning analysis of multimodal data and further validated. Outcome studies using precise nutritional interventions lasting only 3–6 months will have to demonstrate an improvement of dysbiosis to support widespread clinical adoption.

4. Conclusion

Healthy aging biomarkers are showing value for establishing trends in normal immune health, normal energy-yielding metabolism, and oxidative stress. Chronic inflammation and oxidative stress leading to dysbiosis, tissue damage, and infections can be monitored before signs of clinical pathology become evident. Biomarkers of diet, vitamins, and nutrition related to improved digestive health and energy conversion can potentially keep oxidative stress from exceeding the inflammasome response and allow continued immune system health. Biomarkers of tissue oxygenation health relate directly to mobility and mental quality of life, with mobility itself preventing an increase in the byproducts of dysfunction, cellular senescence, and impaired regeneration. Maintaining healthy gut microbiomes can be aided by early monitoring of biomarkers that enable adherence to a personalized nutritional plan and when used in combination with broad coverage of key aging factors.

Nomenclature

HNE

Active B12	holotranscobalamin complex
AdinoR	adinonectin recentor

AdipoR adiponectin receptor

AdipoR CTF C-terminal fragment from the AdipoR AGE advanced glycation end products

Akt protein kinase B

AMPK AMP-activated protein kinase

BMI body mass index cluster of differentiation CD CFU colony-forming units cardiorespiratory fitness CRF CRP C-reactive protein CXC cysteine-X-cysteine CCL CXC ligands n-3 FAs n-3 fatty acids **GSH** glutathione hemoglobin A1c hbA1C Hgb hemoglobin

HNE-albumin adducts of HNE to human serum albumin

4-hydroxynonenal

IFN-γ interferon gamma
IL interleukin

IVD in vitro diagnostics

Jak-STAT Janus kinase and signal transducer and

activators of transcription

KCNMA potassium large conductance calcium-

activated channel lipopolysaccharide lipoteichoic acid

MAP mitogen-activated protein

MAPK/ERK kinase-extracellular signal-regulated

kinase

MCP monocyte chemoattractant protein

MDA malondialdehyde metS metabolic syndrome

mTOR mammalian target of rapamycin
OHdG hydroxy-2'-deoxyguanosine
PAR protease active receptor
PI3K phosphatidylinositol 3-kinase

RBC red blood cell

ROC AUC score receiver operating characteristic area

under the curve

ROS reactive oxidative stress TACE TNF α convertase enzyme tumor necrosis factor- α WBC white blood cell

Data Availability Statement

Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

Disclosure

LPS

LTA

The authors have conducted this biomarker research independently in accordance with a policy on objectivity in research.

Conflicts of Interest

The authors declare no conflicts of interest.

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References

- H. T. John, T. C. Thomas, E. C. Chukwuebuka, et al., "The Microbiota-Human Health Axis," *Microorganisms* 13, no. 4 (2025): 948, https://doi.org/10.3390/microorganisms13040948.
- [2] C. P. Lozano, L. R. Wilkens, Y. B. Shvetsov, et al., "Associations of the Dietary Inflammatory Index With Total Adiposity and Ectopic Fat Through the Gut Microbiota, LPS, and C-Reactive Protein in the Multiethnic Cohort-Adiposity Phenotype Study," *American Journal of Clinical Nutrition* 115, no. 5 (2022): 1344–1356, https://doi.org/10.1093/ajcn/nqab398.
- [3] M. D'Accolti, I. Soffritti, E. Mazziga, et al., "A Sustainable Combined Approach to Control the Microbial Bioburden in the School Environment," *Microorganisms* 13, no. 4 (2025): 791, https://doi.org/10.3390/microorganisms13040791.

- [4] F. Reis, L. M. R. Ferreira, E. Ortega, and S. Viana, "Nutrition and Gut Microbiota-Immune System Interplay in Chronic Diseases," *Nutrients* 17, no. 8 (2025): 1330, https://doi.org/ 10.3390/nu17081330.
- [5] N. Kumar, S. Sain, and B. Solanki, "Deciphering Microbial Carbohydrate Metabolism: Unveiling Mechanistic Insights Into Type 2 Diabetes," Advanced Gut & Microbiome Research 2025 (2025): 17, 4395850, https://doi.org/10.1155/agm3/ 4395850.
- [6] M. J. Barratt, C. Lebrilla, H. Y. Shapiro, and J. I. Gordon, "The Gut Microbiota, Food Science, and Human Nutrition: A Timely Marriage," *Cell Host & Microbe* 22, no. 2 (2017): 134–141, https://doi.org/10.1016/j.chom.2017.07.006.
- [7] H. J. Zheng, J. Guo, Q. Jia, et al., "The Effect of Probiotic and Synbiotic Supplementation on Biomarkers of Inflammation and Oxidative Stress in Diabetic Patients: A Systematic Review and Meta-Analysis of Randomized Controlled Trials," *Pharmacological Research* 142 (2019): 303–313, https://doi.org/10.1016/j.phrs.2019.02.016.
- [8] R. Y. Chen, I. Mostafa, M. C. Hibberd, et al., "A Microbiota-Directed Food Intervention for Undernourished Children," New England Journal of Medicine 384, no. 16 (2021): 1517– 1528, https://doi.org/10.1056/NEJMoa2023294.
- [9] P. J. Turnbaugh, F. Bäckhed, L. Fulton, and J. I. Gordon, "Diet-Induced Obesity Is Linked to Marked but Reversible Alterations in the Mouse Distal Gut Microbiome," *Cell Host & Microbe* 3, no. 4 (2008): 213–223, https://doi.org/10.1016/j.chom.2008.02.015.
- [10] M. Beaumont, K. J. Portune, N. Steuer, et al., "Quantity and Source of Dietary Protein Influence Metabolite Production by Gut Microbiota and Rectal Mucosa Gene Expression: A Randomized, Parallel, Double-Blind Trial in Overweight Humans," American Journal of Clinical Nutrition 106, no. 4 (2017): 1005–1019, https://doi.org/10.3945/ajcn.117.158816.
- [11] A. S. Raman, J. L. Gehrig, S. Venkatesh, et al., "A Sparse Covarying Unit That Describes Healthy and Impaired Human Gut Microbiota Development," *Science* 365, no. 6449 (2019): https://doi.org/10.1126/science.aau4735.
- [12] H. Holland, P. N. Bezan, B. F. Vercesi, P. P. Ovídio, L. N. Z. Ramalho, and A. A. Jordão, "Effects of Resistant Starch Supplementation on Metabolic Parameters and Oxidative Stress in C57BL/6 Mice Fed With a High-Fat Diet," Advanced Gut & Microbiome Research 2024 (2024): 14, 5534697, https://doi.org/10.1155/2024/5534697.
- [13] N. Miron and V. Cristea, "Enterocytes: Active Cells in Tolerance to Food and Microbial Antigens in the Gut," *Clinical & Experimental Immunology* 167, no. 3 (2012): 405–412, https://doi.org/10.1111/j.1365-2249.2011.04523.x.
- [14] X. S. Zhang, Y. Wang, H. Sun, et al., Gut Microbiota Phospholipids Regulate Intestinal Gene Expression and Can Counteract the Effects of Antibiotic Treatment (bioRxiv, 2025).
- [15] P. Dean, S. Quitard, D. M. Bulmer, A. J. Roe, and B. Kenny, "Cultured Enterocytes Internalise Bacteria Across Their Basolateral Surface for, Pathogen-Inhibitable, Trafficking to the Apical Compartment," *Scientific Reports* 5, no. 1 (2015): 17359, https://doi.org/10.1038/srep17359.
- [16] N. B. Molina, B. León, S. Grenóvero, A. Bosch, M. López, and M. D. Sparo, "Diarrheagenic and Uropathogenic Escherichia coli: Biofilm Production and Virulence Factors in Clinical Isolates From Argentina," Advanced Gut & Microbiome Research 2025 (2025): 8, 3243778, https://doi.org/10.1155/ agm3/3243778.

- [17] B. Dabuo, A. Abubakari, F. E. Sankah, and H. A. Aryee, "Antibiotics and Antimicrobial Resistance Genes in a Gut Microbiota as a Reservoir—A Review," *Advanced Gut & Microbiome Research* 2025 (2025): 12, 6574751, https://doi.org/10.1155/agm3/6574751.
- [18] S. Cahill and F. Humphries, "Inflammasomopathies: Mechanisms and Disease Signatures," *Trends in Immunology* 46, no. 5 (2025): 372–385, https://doi.org/10.1016/j.it.2025.03.008.
- [19] M. J. Pugia, "Inflammatory Pathways in Diabetes," in *Progress in Inflammation Research* XVISpringer International Publishing, 2015), 219.
- [20] M. J. Pugia, M. Pradhan, R. Qi, et al., "Utilization of Electronic Health Records for the Assessment of Adiponectin Receptor Autoantibodies During the Progression of Cardio-Metabolic Comorbidities," *Archives of Autoimmune Diseases* 1, no. 1 (2020): 17–27, https://doi.org/10.46439/autoimmune.1.004.
- [21] J. M. Olefsky and G. M. Reaven, "Insulin Binding to Monocytes and Total Mononuclear Leukocytes From Normal and Diabetic Patients," *Journal of Clinical Endocrinology & Metabolism* 43, no. 1 (1976): 226–231.
- [22] C. E. Stewart and A. P. Sharples, "Aging, Skeletal Muscle, and Epigenetics," *Plastic and Reconstructive Surgery* 150 (2022): 27S-33S, https://doi.org/10.1097/PRS.0000000000009670.
- [23] P. J. Turnbaugh and J. I. Gordon, "The Core Gut Microbiome, Energy Balance and Obesity," *Journal of Physiology* 587, pt 17 (2009): 4153–4158, https://doi.org/10.1113/jphysiol.2009.174136.
- [24] J. Zhang, S. Wang, and B. Liu, "New Insights Into the Genetics and Epigenetics of Aging Plasticity," *Genes (Basel)* 14, no. 2 (2023): 329, https://doi.org/10.3390/genes14020329.
- [25] N. Sayed, Y. Huang, K. Nguyen, et al., "An Inflammatory Aging Clock (iAge) Based on Deep Learning Tracks Multimorbidity, Immunosenescence, Frailty and Cardiovascular Aging," *Nature Aging* 1 (2021): 598–615, https://doi.org/ 10.1038/s43587-021-00082-y.
- [26] L. Ferrucci and E. Fabbri, "Inflammageing: Chronic Inflammation in Ageing, Cardiovascular Disease, and Frailty," *Nature Reviews Cardiology* 15, no. 9 (2018): 505–522, https://doi.org/10.1038/s41569-018-0064-2.
- [27] D. Brigger, C. Riether, R. van Brummelen, et al., "Eosinophils Regulate Adipose Tissue Inflammation and Sustain Physical and Immunological Fitness in Old Age," *Nature Metabolism* 2, no. 8 (2020): 688–702, https://doi.org/10.1038/s42255-020-0228-3.
- [28] P. P. Coll, *Healthy Aging: A Complete Guide to Clinical Management* (Springer International Publishing, 2019), Imprint: Springer,: Cham. p. 1 online resource (XV, 399 pages 84 illustrations, 73 illustrations in color.
- [29] J. A. Ahern, D. F. Kruger, P. M. Gatcomb, W. A. Petit Jr., and W. V. Tamborlane, "The Diabetes Control and Complications Trial (DCCT): The Trial Coordinator Perspective. Report by the DCCT Research Group," *Diabetes Educator* 15, no. 3 (1989): 236–241.
- [30] M. J. Pugia, M. Murakami, J. A. Lott, et al., "Screening for Proteinuria in Japanese Schoolchildren: A New Approach," *Clinical Chemistry and Laboratory Medicine* 38, no. 10 (2000): 975–982.
- [31] M. J. Pugia and J. M. Kulick, "Method for Correction for Urine Volume" (Patent Application, 2022), WO 2022/082013 A1.
- [32] P. J. Turnbaugh, R. E. Ley, M. A. Mahowald, V. Magrini, E. R. Mardis, and J. I. Gordon, "An Obesity-Associated Gut

- Microbiome With Increased Capacity for Energy Harvest," *Nature* 444, no. 7122 (2006): 1027–1031.
- [33] M. K. Edwards and P. D. Loprinzi, "All-Cause Mortality Risk as a Function of Sedentary Behavior, Moderate-to-Vigorous Physical Activity and Cardiorespiratory Fitness," *Physician* and Sportsmedicine 44, no. 3 (2016): 223–230, https:// doi.org/10.1080/00913847.2016.1221751.
- [34] C. L. Coolbaugh, I. B. Anderson, M. D. Wilson, D. A. Hawkins, and E. A. Amsterdam, "Evaluation of an Exercise Field Test Using Heart Rate Monitors to Assess Cardiorespiratory Fitness and Heart Rate Recovery in an Asymptomatic Population," *PLoS One* 9, no. 5 (2014): e97704, https://doi.org/10.1371/journal.pone.0097704.
- [35] S. Rooney, A. Webster, and L. Paul, "Systematic Review of Changes and Recovery in Physical Function and Fitness After Severe Acute Respiratory Syndrome-Related Coronavirus Infection: Implications for COVID-19 Rehabilitation," *Physical Therapy* 100, no. 10 (2020): 1717–1729, https://doi.org/ 10.1093/ptj/pzaa129.
- [36] M. Fenech, "Nutrigenomics and Nutrigenetics: The New Paradigm for Optimising Health and Preventing Disease," supplement, *Journal of Nutritional Science and Vitaminology* 61, p. S209, https://doi.org/10.3177/jnsv.61.S209.
- [37] M. Pugia, T. Bose, M. Tjioe, et al., "Multiplexed Signal Ion Emission Reactive Release Amplification (SIERRA) Assay for the Culture-Free Detection of Gram-Negative and Gram-Positive Bacteria and Antimicrobial Resistance Genes," *Analytical Chemistry* 93, no. 17 (2021): 6604–6612, https://doi.org/10.1021/acs.analchem.0c00453.
- [38] M. J. Pugia, R. Valdes Jr., and S. A. Jortani, "Bikunin (Urinary Trypsin Inhibitor): Structure, Biological Relevance, and Measurement," *Advances in Clinical Chemistry* 44 (2007): 223–245, https://doi.org/10.1016/S0065-2423(07)44007-0.
- [39] M. J. Pugia, R. Sommer, P. Corey, et al., "The Uristatin Dipstick Is Useful in Distinguishing Upper Respiratory From Urinary Tract Infections," *Clinica Chimica Acta* 341, no. 1-2 (2004): 73–81, https://doi.org/10.1016/j.cccn.2003.10.019.
- [40] S. A. Jortani, M. J. Pugia, R. J. Elin, et al., "Sensitive Noninvasive Marker for the Diagnosis of Probable Bacterial or Viral Infection," *Journal of Clinical Laboratory Analysis* 18, no. 6 (2004): 289–295.
- [41] J. H. Gordon, M. LaMonte, R. J. Genco, et al., "Is the Oral Microbiome Associated With Blood Pressure in Older Women?," *High Blood Pressure & Cardiovascular Prevention* 26, no. 3 (2019): 217–225, https://doi.org/10.1007/s40292-019-00322-8.
- [42] O. Opota, A. Croxatto, G. Prod'hom, and G. Greub, "Blood Culture-Based Diagnosis of Bacteraemia: State of the Art," *Clinical Microbiology and Infection* 21, no. 4 (2015): 313–322, https://doi.org/10.1016/j.cmi.2015.01.003.
- [43] J. Rohr, C. B. Barrett, D. J. Civitello, et al., "Emerging Human Infectious Diseases and the Links to Global Food Production," *Nature Sustainability* 2, no. 6 (2019): 445–456, https://doi.org/10.1038/s41893-019-0293-3.
- [44] P. Katona and J. Katona-Apte, "The Interaction Between Nutrition and Infection," *Clinical Infectious Diseases* 46, no. 10 (2008): 1582–1588, https://doi.org/10.1086/587658.
- [45] C. Theda, S. H. Hwang, A. Czajko, Y. J. Loke, P. Leong, and J. M. Craig, "Quantitation of the Cellular Content of Saliva and Buccal Swab Samples," *Scientific Reports* 8, no. 1 (2018): 6944, https://doi.org/10.1038/s41598-018-25311-0.

- [46] T. Gillum, M. Kuennen, Z. McKenna, M. Castillo, A. Jordan-Patterson, and C. Bohnert, "Exercise Does Not Increase Salivary Lymphocytes, Monocytes, or Granulocytes, but Does Increase Salivary Lysozyme," *Journal of Sports Sciences* 35, no. 13 (2017): 1294–1299, https://doi.org/10.1080/02640414.2016.1221522.
- [47] M. J. Magbanua, M. Pugia, J. S. Lee, et al., "A Novel Strategy for Detection and Enumeration of Circulating Rare Cell Populations in Metastatic Cancer Patients Using Automated Microfluidic Filtration and Multiplex Immunoassay," *PLoS One* 10, no. 10 (2015): e0141166, https://doi.org/10.1371/ journal.pone.0141166.
- [48] P. E. B. Calouius, "The Leukocyte Count in Saliva," Oral Surgery, Oral Medicine, Oral Pathology 11, no. 1 (1958): 43–46, https://doi.org/10.1016/0030-4220(58)90219-6.
- [49] G. A. Mensah, D. W. Brown, J. B. Croft, and K. J. Greenlund, "Major Coronary Risk Factors and Death From Coronary Heart Disease: Baseline and Follow-up Mortality Data From the Second National Health and Nutrition Examination Survey (NHANES II)," 5 supplement 1, American Journal of Preventive Medicine 29, 68–74.
- [50] M. J. Pugia, "Technology Behind Diagnostic Reagent Strips," Laboratory Medicine 31, no. 2 (2000): 92–96, https://doi.org/ 10.1309/15R1-46YG-CV95-C36K.
- [51] D. Frabutt, N. Stull, A. R. Pineros, et al., "Adiponectin Receptor Fragmentation in Mouse Models of Type 1 and Type 2 Diabetes," *Archives of Autoimmune Diseases* 1, no. 1 (2020): 3–13, https://doi.org/10.46439/autoimmune.1.002.
- [52] Y. Gu, J. J. Manly, R. P. Mayeux, and A. M. Brickman, "An Inflammation-Related Nutrient Pattern Is Associated With Both Brain and Cognitive Measures in a Multiethnic Elderly Population," *Current Alzheimer Research* 15, no. 5 (2018): 493–501, https://doi.org/10.2174/1567205015666180101145619.
- [53] N. A. Duggal, G. Niemiro, S. D. R. Harridge, R. J. Simpson, and J. M. Lord, "Can Physical Activity Ameliorate Immunosenescence and Thereby Reduce Age-Related Multi-Morbidity?," *Nature Reviews Immunology* 19, no. 9 (2019): 563–572, https://doi.org/10.1038/s41577-019-0177-9.
- [54] H. Yu and N. Rifai, "High-Sensitivity C-Reactive Protein and Atherosclerosis: From Theory to Therapy," *Clinical Biochemistry* 33, no. 8 (2000): 601–610.
- [55] N. Rifai and P. M. Ridker, "High-Sensitivity C-Reactive Protein: A Novel and Promising Marker of Coronary Heart Disease," *Clinical Chemistry* 47, no. 3 (2001): 403–411.
- [56] B. M. Scirica, C. P. Cannon, M. S. Sabatine, et al., "Concentrations of C-Reactive Protein and B-Type Natriuretic Peptide 30 Days After Acute Coronary Syndromes Independently Predict Hospitalization for Heart Failure and Cardiovascular Death," *Clinical Chemistry* 55, no. 2 (2009): 265–273, https://doi.org/10.1373/clinchem.2008.117192.
- [57] J. Wetterö, S. von Löhneysen, F. Cobar, M. Kristenson, P. Garvin, and C. Sjöwall, "Pronounced Diurnal Pattern of Salivary C-Reactive Protein (CRP) With Modest Associations to Circulating CRP Levels," Frontiers in Immunology 11 (2021): 607166, https://doi.org/10.3389/fimmu.2020.607166.
- [58] C. Rheaume, B. J. Arsenault, M. P. Dumas, et al., "Contributions of Cardiorespiratory Fitness and Visceral Adiposity to Six-Year Changes in Cardiometabolic Risk Markers in Apparently Healthy Men and Women," *Journal of Clinical Endocrinology & Metabolism* 96, no. 5 (2011): 1462–1468, https://doi.org/10.1210/jc.2010-2432.

- [59] K. H. Costenbader, L. MacFarlane, I. M. Lee, et al., "Effects of One Year of Vitamin D and Marine Omega-3 Fatty Acid Supplementation on Biomarkers of Systemic Inflammation in Older US Adults," *Clinical Chemistry* 65, no. 12 (2019): 1508–1521, https://doi.org/10.1373/clinchem.2019.306902.
- [60] A. S. Prasad, "Zinc in Human Health: Effect of Zinc on Immune Cells," Molecular Medicine 14, no. 5-6 (2008): 353–357, https://doi.org/10.2119/2008-00033.Prasad.
- [61] S. J. Wimalawansa, "Rapidly Increasing Serum 25(OH)D Boosts the Immune System, Against Infections-Sepsis and COVID-19," *Nutrients* 14, no. 14 (2022): 2997, https://doi.org/10.3390/nu14142997.
- [62] W. Saengsiwaritt, J. Jittikoon, U. Chaikledkaew, T. Tawonsawatruk, S. Honsawek, and W. Udomsinprasert, "Effect of Vitamin D Supplementation on Circulating Level of Autophagosome Protein LC3A, Inflammation, and Physical Performance in Knee Osteoarthritis," Clinical and Translational Science 16, no. 12 (2023): 2543–2556, https://doi.org/ 10.1111/cts.13646.
- [63] K. S. Willis, D. T. Smith, K. S. Broughton, and D. E. Larson-Meyer, "Vitamin D Status and Biomarkers of Inflammation in Runners," *Open Access Journal of Sports Medicine* 3 (2012): 35–42, https://doi.org/10.2147/OAJSM.S31022.
- [64] S. M. L. Namaste, J. Ou, A. M. Williams, M. F. Young, E. X. Yu, and P. S. Suchdev, "Adjusting Iron and Vitamin A Status in Settings of Inflammation: A Sensitivity Analysis of the Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) Approach," *American Journal of Clinical Nutrition* 112 (2020): 458S–467S, https://doi.org/10.1093/ajcn/nqaa141.
- [65] A. Bahramian, P. Falsafi, T. Abbasi, et al., "Comparing Serum and Salivary Levels of Vitamin D in Patients With Recurrent Aphthous Stomatitis and Healthy Individuals," *Journal of Dentistry* 19, no. 4 (2018): 295–300.
- [66] T. Quasim, D. McMillan, D. Talwar, A. Vasilaki, D. St J O'Reilly, and J. Kinsella, "The Relationship Between Plasma and Red Cell B-Vitamin Concentrations in Critically-Ill Patients," Clinical Nutrition 24, no. 6 (2005): 956–960.
- [67] R. W. Beal and W. M. Read, "Studies on the Plasma Protein Biing of [⁵⁸Co]-Vitamin B₁₂. V. Relationship of Granulocytes to Binding Capacity," *Australian Journal of Experimental Biology and Medical Science* 47, no. 3 (1969): 387–391.
- [68] F. Haidari, B. Abiri, M. Iravani, et al., "Effects of Vitamin D and Omega-3 Fatty Acids Co-Supplementation on Inflammatory Biomarkers, Tumor Marker CEA, and Nutritional Status in Patients With Colorectal Cancer: A Study Protocol for a Double Blind Randomized Controlled Trial," *Trials* 20, no. 1 (2019): 682, https://doi.org/10.1186/s13063-019-3719-3.
- [69] A. Yarparvar, I. Elmadfa, A. Djazayery, Z. Abdollahi, and F. Salehi, "The Association of Vitamin D Status With Lipid Profile and Inflammation Biomarkers in Healthy Adolescents," *Nutrients* 12, no. 2 (2020): 590, https://doi.org/ 10.3390/nu12020590.
- [70] I. Milisav, S. Ribarič, and B. Poljsak, "Antioxidant Vitamins and Ageing," *Biochemistry and Cell Biology of Ageing: Part I Biomedical Science* 90 (2018): https://doi.org/10.1007/978-981-13-2835-0_1.
- [71] K. Mikkelsen and V. Apostolopoulos, "B Vitamins and Ageing," in *Biochemistry and Cell Biology of Ageing: Part I Biomedical Science. Subcellular Biochemistry* 90, Springer, 2018), 451–470.

- [72] E. J. de Koning, N. van der Zwaluw, J. van Wijngaarden, et al., "Effects of Two-Year Vitamin B₁₂ and Folic Acid Supplementation on Depressive Symptoms and Quality of Life in Older Adults With Elevated Homocysteine Concentrations: Additional Results From the B-PROOF Study, an RCT," Nutrients 8, no. 11 (2016).
- [73] I. Iglesia, I. Huybrechts, M. González-Gross, et al., "Folate and Vitamin B12 Concentrations Are Associated With Plasma DHA and EPA Fatty Acids in European Adolescents: The Healthy Lifestyle in Europe by Nutrition in Adolescence (HELENA) Study," *British Journal of Nutrition* 117, no. 1 (2017): 124–133, https://doi.org/10.1017/S0007114516004414.
- [74] D. L. Zhang, M. C. Ghosh, H. Ollivierre, Y. Li, and T. A. Rouault, "Ferroportin Deficiency in Erythroid Cells Causes Serum Iron Deficiency and Promotes Hemolysis Due to Oxidative Stress," *Blood* 132, no. 19 (2018): 2078–2087, https:// doi.org/10.1182/blood-2018-04-842997.
- [75] J. B. Zhang, J. Tong, D. Y. Sun, J. T. Fu, D. J. Li, and P. Wang, "Targeting Ferroptosis in Cardio-Metabolic-Diseases: Mechanisms and Therapeutic Prospects," *Medicinal Research Reviews* 43, no. 3 (2023): 683–712, https://doi.org/10.1002/med.21934.
- [76] A. L. Kau, P. P. Ahern, N. W. Griffin, A. L. Goodman, and J. I. Gordon, "Human Nutrition, the Gut Microbiome and the Immune System," *Nature* 474, no. 7351 (2011): 327–336, https://doi.org/10.1038/nature10213.
- [77] A. Bordoni, S. Hrelia, A. Lorenzini, et al., "Dual Influence of Aging and Vitamin B6 Deficiency on Delta-6-Desaturation of Essential Fatty Acids in Rat Liver Microsomes," *Prosta*glandins, Leukotrienes and Essential Fatty Acids 58, no. 6 (1998): 417–420.
- [78] J. Lazarov, "Resorption of Vitamin B1–XII. Changes in the Resorption and the Phosphorylation of Thiamine in Rats in Relation to Age," *Experimental Gerontology* 12, no. 1-2 (1977): 75–79.
- [79] A. W. McEvoy, J. D. Fenwick, K. Boddy, and O. F. James, "Vitamin B12 Absorption From the Gut Does Not Decline With Age in Normal Elderly Humans," *Age and Ageing* 11, no. 3 (1982): 180–183.
- [80] A. Hamfelt, "Age Variation of Vitamin B6 Metabolism in Man," Clinica Chimica Acta 10, no. 1 (1964): 48–54, https:// doi.org/10.1016/0009-8981(64)90214-1.
- [81] W. Roth and M. Mohamadzadeh, "Vitamin B12 and Gut-Brain Homeostasis in the Pathophysiology of Ischemic Stroke," *EBioMedicine* 73 (2021): 103676, https://doi.org/10.1016/j.ebiom.2021.103676.
- [82] L. Qi, W. Ma, Y. Heianza, et al., "Independent and Synergistic Associations of Biomarkers of Vitamin D Status With Risk of Coronary Heart Disease," *Arteriosclerosis, Thrombosis, and Vascular Biology* 37, no. 11 (2017): 2204–2212, https://doi.org/10.1161/ATVBAHA.117.309548.
- [83] L. E. Chapman, A. L. Darling, and J. E. Brown, "Association Between Metformin and Vitamin B₁₂ Deficiency in Patients With Type 2 Diabetes: A Systematic Review and Meta-Analysis," *Diabetes & Metabolism* 42, no. 5 (2016): 316–327, https://doi.org/10.1016/j.diabet.2016.03.008.
- [84] A. C. Ham, A. W. Enneman, S. van Dijk, et al., "Associations Between Medication Use and Homocysteine Levels in an Older Population, and Potential Mediation by Vitamin B₁₂ and Folate: Data From the B-PROOF Study," *Drugs & Aging* 31, no. 8 (2014): 611–621, https://doi.org/10.1007/s40266-014-0192-2.

- [85] K. Porter, L. Hoey, C. F. Hughes, M. Ward, and H. McNulty, "Causes, Consequences and Public Health Implications of Low B-Vitamin Status in Ageing," *Nutrients* 8, no. 11 (2016).
- [86] T. Nagao and M. Hirokawa, "Diagnosis and Treatment of Macrocytic Anemias in Adults," *Journal of General and Family Medicine* 18, no. 5 (2017): 200–204, https://doi.org/10.1002/jgf2.31.
- [87] R. A. Ghashut, D. McMillan, J. Kinsella, and D. Talwar, "Erythrocyte Concentrations of B1, B2, B6 but Not Plasma C and E Are Reliable Indicators of Nutrition Status in the Presence of Systemic Inflammation," *Clinical Nutrition* ESPEN 17 (2017): 54–62, https://doi.org/10.1016/ j.clnesp.2016.10.007.
- [88] J. Salas-Salvadó, A. Díaz-López, M. Ruiz-Canela, et al., "Effect of a Lifestyle Intervention Program With Energy-Restricted Mediterranean Diet and Exercise on Weight Loss and Cardiovascular Risk Factors: One-Year Results of the PREDIMED-Plus Trial," *Diabetes Care* 42, no. 5 (2019): 777–788, https://doi.org/10.2337/dc18-0836.
- [89] K. Kessler, S. Hornemann, K. J. Petzke, et al., "Diurnal Distribution of Carbohydrates and Fat Affects Substrate Oxidation and Adipokine Secretion in Humans," *American Journal of Clinical Nutrition* 108, no. 6 (2018): 1209–1219, https://doi.org/10.1093/ajcn/nqy224.
- [90] K. L. Whytock, K. D. Corbin, S. A. Parsons, et al., "Metabolic Adaptation Characterizes Short-Term Resistance to Weight Loss Induced by a Low-Calorie Diet in Overweight/Obese Individuals," *American Journal of Clinical Nutrition* 114, no. 1 (2021): 267–280, https://doi.org/10.1093/ajcn/nqab027.
- [91] M. Yamaguchi, "Adiponectin: Production, Regulation, and Roles in Disease," in *Endocrinology Research and Clinical Developments* x, Nova Science Publishers, 2012), 170.
- [92] H. Kim, B. Yu, X. Li, et al., "Serum Metabolomic Signatures of Plant-Based Diets and Incident Chronic Kidney Disease," *American Journal of Clinical Nutrition* 116, no. 1 (2022): 151–164, https://doi.org/10.1093/ajcn/nqac054.
- [93] P. Vitaglione, I. Mennella, R. Ferracane, et al., "Whole-Grain Wheat Consumption Reduces Inflammation in a Randomized Controlled Trial on Overweight and Obese Subjects With Unhealthy Dietary and Lifestyle Behaviors: Role of Polyphenols Bound to Cereal Dietary Fiber," *American Jour*nal of Clinical Nutrition 101, no. 2 (2015): 251–261, https:// doi.org/10.3945/ajcn.114.088120.
- [94] A. K. Eriksen, C. Brunius, M. Mazidi, et al., "Effects of Whole-Grain Wheat, Rye, and Lignan Supplementation on Cardiometabolic Risk Factors in Men With Metabolic Syndrome: A Randomized Crossover Trial," *American Journal of Clinical Nutrition* 111, no. 4 (2020): 864–876, https://doi.org/10.1093/ajcn/nqaa026.
- [95] Z. Miao, F. F. Zeng, Y. Tian, et al., "Furan Fatty Acid Metabolite CMPF Is Associated With Lower Risk of Type 2 Diabetes, but Not Chronic Kidney Disease: A Longitudinal Population-Based Cohort Study," American Journal of Clinical Nutrition 118, no. 3 (2023): 637–645, https://doi.org/10.1016/j.ajcnut.2023.07.016.
- [96] A. K. Elshorbagy, E. Nurk, C. G. Gjesdal, et al., "Homocysteine, Cysteine, and Body Composition in the Hordaland Homocysteine Study: Does Cysteine Link Amino Acid and Lipid Metabolism?," American Journal of Clinical Nutrition 88, no. 3 (2008): 738–746.
- [97] R. Blanco-Rojo, J. Delgado-Lista, Y. C. Lee, et al., "Interaction of an S100A9 Gene Variant With Saturated Fat and Carbohy-

- drates to Modulate Insulin Resistance in 3 Populations of Different Ancestries," *American Journal of Clinical Nutrition* 104, no. 2 (2016): 508–517, https://doi.org/10.3945/ajcn.116.130898.
- [98] Y. Zheng, B. Yu, D. Alexander, L. M. Steffen, J. A. Nettleton, and E. Boerwinkle, "Metabolomic Patterns and Alcohol Consumption in African Americans in the Atherosclerosis Risk in Communities Study," *American Journal of Clinical Nutrition* 99, no. 6 (2014): 1470–1478, https://doi.org/10.3945/ajcn.113.074070.
- [99] H. Schröder, R. de la Torre, R. Estruch, et al., "Alcohol Consumption Is Associated With High Concentrations of Urinary Hydroxytyrosol," *American Journal of Clinical Nutrition* 90, no. 5 (2009): 1329–1335, https://doi.org/10.3945/ajcn.2009.27718.
- [100] C. Arancibia-Riveros, I. Domínguez-López, A. Tresserra-Rimbau, et al., "Total Urinary Polyphenol Excretion: A Biomarker of an Anti-Inflammatory Diet and Metabolic Syndrome Status," *American Journal of Clinical Nutrition* 117, no. 4 (2023): 814–822, https://doi.org/10.1016/j.ajcnut.2022.12.016.
- [101] T. Dorjgochoo, Y. T. Gao, W. H. Chow, et al., "Major Metabolite of F2-Isoprostane in Urine May Be a More Sensitive Biomarker of Oxidative Stress Than Isoprostane Itself," American Journal of Clinical Nutrition 96, no. 2 (2012): 405–414, https://doi.org/10.3945/ajcn.112.034918.
- [102] H. J. Thompson, J. Heimendinger, S. Sedlacek, et al., "8-Isoprostane F2alpha Excretion Is Reduced in Women by Increased Vegetable and Fruit Intake," *American Journal of Clinical Nutrition* 82, no. 4 (2005): 768–776.
- [103] J. B. Henry, Clinical Diagnosis and Management by Laboratory Methods xxivW.B. Saunders, 20th edition, 2001).
- [104] Y. Rakanita, M. R. A. A. Syamsunarno, R. K. Sinuraya, E. W. Suradji, R. Abdulah, and A. A. Suwantika, "Cost-Effectiveness of Ferrous Fumarate-Folic Acid and Ferrous Gluconate-Multivitamins in a High Prevalence Area of Iron Deficiency Anemia in Indonesia," *Therapeutics and Clinical Risk Management* 17 (2021): 1075–1081, https://doi.org/10.2147/TCRM.S328226.
- [105] Z. Z. Zhang, E. E. Lee, J. Sudderth, et al., "Glutathione Depletion, Pentose Phosphate Pathway Activation, and Hemolysis in Erythrocytes Protecting Cancer Cells From Vitamin C-Induced Oxidative Stress," *Journal of Biological Chemistry* 291, no. 44 (2016): 22861–22867.
- [106] P. L. Geltman, A. F. Meyers, S. D. Mehta, et al., "Daily Multivitamins With Iron to Prevent Anemia in High-Risk Infants: A Randomized Clinical Trial," *Pediatrics* 114, no. 1 (2004): 86–93.
- [107] F. Islami, E. M. Ward, H. Sung, et al., "Annual Report to the Nation on the Status of Cancer, Part 1: National Cancer Statistics," *JNCI: Journal of the National Cancer Institute* 113, no. 12 (2021): 1648–1669, https://doi.org/10.1093/jnci/ djab131.
- [108] L. S. Valberg, J. M. Holt, and G. M. Brown, "Sodium, Potassium, Calcium, Magnesium, Copper, and Zinc Composition of Erythrocytes in Vitamin B₁₂ Deficiency and Iron Deficiency," *Journal of Clinical Investigation* 44, no. 7 (1965): 1225–1233, https://doi.org/10.1172/JCI105228.
- [109] M. F. Lopes-Virella, K. J. Hunt, N. L. Baker, G. Virella, and VADT Group of Investigators, "High Levels of AGE-LDL, and of IgG Antibodies Reacting With MDA-Lysine Epitopes Expressed by oxLDL and MDA-LDL in Circulating Immune

- Complexes Predict Macroalbuminuria in Patients With Type 2 Diabetes," *Journal of Diabetes and its Complications* 30, no. 4 (2016): 693–699, https://doi.org/10.1016/j.jdiacomp.2016.01.012.
- [110] H. Strohmaier, H. Hinghofer-Szalkay, and R. J. Schaur, "Detection of 4-Hydroxynonenal (HNE) as a Physiological Component in Human Plasma," *Journal of Lipid Mediators and Cell Signalling* 11, no. 1 (1995): 51–61.
- [111] L. Milkovic, N. Zarkovic, Z. Marusic, K. Zarkovic, and M. Jaganjac, "The 4-Hydroxynonenal-Protein Adducts and Their Biological Relevance: Are Some Proteins Preferred Targets?," *Antioxidants* 12, no. 4 (2023): 856, https://doi.org/ 10.3390/antiox12040856.
- [112] K. Mehta and V. B. Patel, "Measurement of 4-Hydroxynonenal (4-HNE) Protein Adducts by ELISA," in Redox-Mediated Signal Transduction: Methods and Protocols 1990, Springer, 2019), 43–52, https://doi.org/10.1007/978-1-4939-9463-2_4.
- [113] A. García-Hermoso, R. Ramírez-Vélez, R. M. Alfonso-Rosa, and B. del Pozo Cruz, "Cardiorespiratory Fitness, Physical Activity, Sedentary Behavior, and Circulating White Blood Cells in US Youth," Scandinavian Journal of Medicine & Science in Sports 31, no. 2 (2021): 439–445, https://doi.org/10.1111/sms.13845.
- [114] S. W. Farrell, D. Leonard, K. Shuval, et al., "Cardiorespiratory Fitness, White Blood Cell Count, and Mortality in Men and Women," *Journal of Sport and Health Science* 11, no. 5 (2022): 605–612, https://doi.org/10.1016/j.jshs.2021.10.005.
- [115] A. Ambring, M. Johansson, M. Axelsen, L. Gan, B. Strandvik, and P. Friberg, "Mediterranean-Inspired Diet Lowers the Ratio of Serum Phospholipid n-6 to n-3 Fatty Acids, the Number of Leukocytes and Platelets, and Vascular Endothelial Growth Factor in Healthy Subjects," American Journal of Clinical Nutrition 83, no. 3 (2006): 575–581.
- [116] C. Huang, W. Huang, R. Wang, and Y. He, "Ulinastatin Inhibits the Proliferation, Invasion and Phenotypic Switching of PDGF-BB-Induced VSMCs via Akt/eNOS/NO/cGMP Signaling Pathway," *Drug Design, Development and Therapy* 14 (2020): 5505–5514, https://doi.org/10.2147/DDDT.S275488.
- [117] L. Zhao, Y. Ma, Q. Li, and Y. Wang, "Ulinastatin Combined With Glutamine Improves Liver Function and Inflammatory Response in Patients With Severe Acute Pancreatitis," *American Journal of Translational Research* 14, no. 2 (2022): 918–926.
- [118] L. Wang, W. Jiao, J. Wu, J. Zhang, M. Tang, and Y. Chen, "Ulinastatin Alleviates Early Brain Injury After Intracerebral Hemorrhage by Inhibiting Necroptosis and Neuroinflammation via MAPK/NF-κB Signaling Pathway," *Acta Cirurgica Brasileira* 37, no. 3 (2022): e370301, https://doi.org/10.1590/acb370301.
- [119] X. Zhang, C. Su, S. Zhao, J. Li, and F. Yu, "Combination Therapy of Ulinastatin With Thrombomodulin Alleviates Endotoxin (LPS) Induced Liver and Kidney Injury via Inhibiting Apoptosis, Oxidative Stress and HMGB1/TLR4/NF-κB Pathway," *Bioengineered* 13, no. 2 (2022): 2951–2970, https://doi.org/10.1080/21655979.2021.2024686.
- [120] M. S. Hipp, P. Kasturi, and F. U. Hartl, "The Proteostasis Network and Its Decline in Ageing," *Nature Reviews Molecular Cell Biology* 20, no. 7 (2019): 421–435, https://doi.org/10.1038/s41580-019-0101-y.
- [121] C. Musilli, S. Paccosi, L. Pala, et al., "Characterization of Circulating and Monocyte-Derived Dendritic Cells in Obese and

- Diabetic Patients," *Molecular Immunology* 49, no. 1-2 (2011): 234–238, https://doi.org/10.1016/j.molimm.2011.08.019.
- [122] M. C. Flynn, G. Pernes, M. K. S. Lee, P. R. Nagareddy, and A. J. Murphy, "Monocytes, Macrophages, and Metabolic Disease in Atherosclerosis," *Frontiers in Pharmacology* 10 (2019): 666, https://doi.org/10.3389/fphar.2019.00666.
- [123] S. S. Zhang, X. J. Yang, Q. H. Ma, et al., "Leukocyte Related Parameters in Older Adults With Metabolically Healthy and Unhealthy Overweight or Obesity," *Scientific Reports* 11, no. 1 (2021): 4652, https://doi.org/10.1038/s41598-021-84367-7.
- [124] B. D. Hoit, E. A. Gilpin, A. A. Maisel, H. Henning, J. Carlisle, and J. Ross Jr., "Influence of Obesity on Morbidity and Mortality After Acute Myocardial Infarction," *American Heart Journal* 114, no. 6 (1987): 1334–1341.
- [125] H. H. Marks, "Influence of Obesity on Morbidity and Mortality," *Bulletin of the New York Academy of Medicine* 36, no. 5 (1960): 296–312.
- [126] J. M. Olefsky, "Decreased Insulin Binding to Adipocytes and Circulating Monocytes From Obese Subjects," *Journal of Clinical Investigation* 57, no. 5 (1976): 1165–1172.
- [127] I. Alvarez-Alvarez, M. A. Martinez-Gonzalez, A. Sanchez-Tainta, et al., "Adherence to an Energy-Restricted Mediterranean Diet Score and Prevalence of Cardiovascular Risk Factors in the PREDIMED-Plus: A Cross-Sectional Study," *Revista Española de Cardiología (English Edition)* 72, no. 11 (2019): 925–934, https://doi.org/10.1016/j.rec.2018.08.010.
- [128] M. Gašperlin and M. Gosenca, "Main Approaches for Delivering Antioxidant Vitamins Through the Skin to Prevent Skin Ageing," *Expert Opinion on Drug Delivery* 8, no. 7 (2011): 905–919, https://doi.org/10.1517/17425247.2011.581657.
- [129] L. Jiang, H. Su, X. Wu, et al., "Leptin Receptor-Expressing Neuron Sh2b1 Supports Sympathetic Nervous System and Protects Against Obesity and Metabolic Disease," *Nature Communications* 11, no. 1 (2020): 1517, https://doi.org/ 10.1038/s41467-020-15328-3.
- [130] J. Kim, G. Yang, Y. Kim, J. Kim, and J. Ha, "AMPK Activators: Mechanisms of Action and Physiological Activities," *Experimental & Molecular Medicine* 48, no. 4 (2016): e224, https://doi.org/10.1038/emm.2016.16.
- [131] C. M. van Stijn, J. Kim, A. J. Lusis, G. D. Barish, and R. K. Tangirala, "Macrophage Polarization Phenotype Regulates Adiponectin Receptor Expression and Adiponectin Anti-Inflammatory Response," FASEB Journal 29, no. 2 (2015): 636–649, https://doi.org/10.1096/fj.14-253831.
- [132] J. L. Mark Welch, Y. Hasegawa, N. McNulty, J. I. Gordon, and G. G. Borisy, "Spatial Organization of a Model 15-Member Human Gut Microbiota Established in Gnotobiotic Mice," Proceedings of the National Academy of Sciences of the United States of America 114, no. 43 (2017): E9105–E9114, https:// doi.org/10.1073/pnas.1711596114.
- [133] M. C. Hibberd, D. M. Webber, D. A. Rodionov, et al., "Bioactive Glycans in a Microbiome-Directed Food for Children With Malnutrition," *Nature* 625, no. 7993 (2024): 157–165, https://doi.org/10.1038/s41586-023-06838-3.
- [134] M. Idrees, A. R. Mohammad, N. Karodia, and A. Rahman, "Multimodal Role of Amino Acids in Microbial Control and Drug Development," *Antibiotics* 9, no. 6 (2020): 330, https://doi.org/10.3390/antibiotics9060330.
- [135] C. Wittenbecher, K. Mühlenbruch, J. Kröger, et al., "Amino Acids, Lipid Metabolites, and Ferritin as Potential Mediators Linking Red Meat Consumption to Type 2 Diabetes,"

- American Journal of Clinical Nutrition 101, no. 6 (2015): 1241–1250, https://doi.org/10.3945/ajcn.114.099150.
- [136] C. M. Rebholz, Z. Zheng, M. E. Grams, et al., "Serum Metabolites Associated With Dietary Protein Intake: Results From the Modification of Diet in Renal Disease (MDRD) Randomized Clinical Trial," *American Journal of Clinical Nutrition* 109, no. 3 (2019): 517–525, https://doi.org/10.1093/ajcn/nqy202.
- [137] H. Chappus-McCendie, L. Chevalier, C. Roberge, and M. Plourde, "Omega-3 PUFA Metabolism and Brain Modifications During Aging," Progress in Neuro-Psychopharmacology and Biological Psychiatry 94 (2019): 109662, https://doi.org/10.1016/j.pnpbp.2019.109662.
- [138] C. L. Kien, J. Y. Bunn, R. Stevens, et al., "Dietary Intake of Palmitate and Oleate Has Broad Impact on Systemic and Tissue Lipid Profiles in Humans," *American Journal of Clinical Nutrition* 99, no. 3 (2014): 436–445, https://doi.org/10.3945/ajcn.113.070557.
- [139] S. Velmurugan, J. M. Gan, K. S. Rathod, et al., "Dietary Nitrate Improves Vascular Function in Patients With Hypercholesterolemia: A Randomized, Double-Blind, Placebo-Controlled Study," *American Journal of Clinical Nutrition* 103, no. 1 (2016): 25–38, https://doi.org/10.3945/ajcn.115.116244.
- [140] R. L. Prior, "Fruits and Vegetables in the Prevention of Cellular Oxidative Damage," 3 supplement, *American Journal of Clinical Nutrition* 78, 570S–578S, https://doi.org/10.1093/ajcn/78.3.570S.
- [141] S. Shab-Bidar, T. R. Neyestani, A. Djazayery, et al., "Improvement of Vitamin D Status Resulted in Amelioration of Biomarkers of Systemic Inflammation in the Subjects With Type 2 Diabetes," *Diabetes/Metabolism Research and Reviews* 28, no. 5 (2012): 424–430, https://doi.org/10.1002/dmrr.2290.
- [142] M. Razavi, M. Jamilian, M. Samimi, et al., "The Effects of Vitamin D and Omega-3 Fatty Acids Co-Supplementation on Biomarkers of Inflammation, Oxidative Stress and Pregnancy Outcomes in Patients With Gestational Diabetes," Nutrition & Metabolism 14, no. 1 (2017): 80, https:// doi.org/10.1186/s12986-017-0236-9.
- [143] A. Hill, S. Wendt, C. Benstoem, et al., "Vitamin C to Improve Organ Dysfunction in Cardiac Surgery Patients-Review and Pragmatic Approach," *Nutrients* 10, no. 8 (2018): 974, https://doi.org/10.3390/nu10080974.
- [144] A. Kawashima, A. Sekizawa, K. Koide, et al., "Vitamin C Induces the Reduction of Oxidative Stress and Paradoxically Stimulates the Apoptotic Gene Expression in Extravillous Trophoblasts Derived From First-Trimester Tissue," Reproductive Sciences 22, no. 7 (2015): 783–790, https://doi.org/10.1177/1933719114561561.
- [145] R. Kumar and S. I. Rizvi, "Vitamin C Improves Inflammatory-Related Redox Status in Hyperlipidemic Rats," *Indian Journal of Clinical Biochemistry* 38, no. 4 (2023): 512–518, https://doi.org/10.1007/s12291-022-01070-8.
- [146] M. D. Defagó, N. R. Perovic, M. A. Valentich, P. G. R. Marquez, and A. B. Actis, "Omega-3 and Omega-6 Salivary Fatty Acids as Markers of Dietary Fat Quality: A Cross-Sectional Study in Argentina," *Acta Odontologica Latinoamericana*: AOL 31, no. 2 (2018): 97–103.
- [147] S. Saboori, S. Shab-Bidar, J. R. Speakman, E. Yousefi Rad, and K. Djafarian, "Effect of Vitamin E Supplementation on Serum C-Reactive Protein Level: A Meta-Analysis of Randomized Controlled Trials," *European Journal of Clinical Nutrition* 69, no. 8 (2015): 867–873, https://doi.org/10.1038/ejcn.2014.296.

- [148] S. Rajaram, "The Effect of Vegetarian Diet, Plant Foods, and Phytochemicals on Hemostasis and Thrombosis," 3 supplement, *American Journal of Clinical Nutrition* 78, 552S–558S, https://doi.org/10.1093/ajcn/78.3.552S.
- [149] M. C. Playdon, J. N. Sampson, A. J. Cross, et al., "Comparing Metabolite Profiles of Habitual Diet in Serum and Urine," *American Journal of Clinical Nutrition* 104, no. 3 (2016): 776–789, https://doi.org/10.3945/ajcn.116.135301.
- [150] M. Fenech, A. el-Sohemy, L. Cahill, et al., "Nutrigenetics and Nutrigenomics: Viewpoints on the Current Status and Applications in Nutrition Research and Practice," *Journal of Nutrigenetics and Nutrigenomics* 4, no. 2 (2011): 69–89, https://doi.org/10.1159/000327772.
- [151] C. Cordonnier, G. le Bihan, J. G. Emond-Rheault, A. Garrivier, J. Harel, and G. Jubelin, "Vitamin B₁₂ Uptake by the Gut Commensal Bacteria Bacteroides Thetaiotaomicron Limits the Production of Shiga Toxin by Enterohemorrhagic Escherichia coli," *Toxins* 8, no. 1 (2016): 14, https:// doi.org/10.3390/toxins8010014.
- [152] S. Zhao, Z. P. Wang, X. Wen, et al., "Synthesis of Vitamin B₁₂-Antibiotic Conjugates With Greatly Improved Activity Against Gram-Negative Bacteria," *Organic Letters* 22, no. 16 (2020): 6632–6636, https://doi.org/10.1021/acs.orglett.0c02403.
- [153] N. Yanaka, T. A. Koyama, S. Komatsu, E. Nakamura, M. Kanda, and N. Kato, "Vitamin B6 Suppresses NFkappaB Activation in LPS-Stimulated Mouse Macrophages," *International Journal of Molecular Medicine* 16, no. 6 (2005): 1071–1075.
- [154] J. M. Miller, M. J. Binnicker, S. Campbell, et al., "A Guide to Utilization of the Microbiology Laboratory for Diagnosis of Infectious Diseases: 2018 Update by the Infectious Diseases Society of America and the American Society for Microbiology," *Clinical Infectious Diseases* 67, no. 6 (2018): 813–816, https://doi.org/10.1093/cid/ciy584.
- [155] R. S. Verma and A. C. Antony, "Immunoreactive Folate-Binding Proteins From Human Saliva. Isolation and Comparison of Two Distinct Species," *Biochemical Journal* 286, no. 3 (1992): 707–715, https://doi.org/10.1042/bj2860707.
- [156] M. Blakeley, A. Sobczyńska-Malefora, and G. Carpenter, "The Origins of Salivary Vitamin A, Vitamin B12 and Vitamin D-Binding Proteins," *Nutrients* 12, no. 12 (2020): 3838, https://doi.org/10.3390/nu12123838.
- [157] B. Zhao, L. Zhu, M. Ye, et al., "Oxidative Stress and Epigenetics in Ocular Vascular Aging: An Updated Review," Molecular Medicine 29, no. 1 (2023): 28, https://doi.org/10.1186/s10020-023-00624-7.
- [158] H. Zeng, X. Lu, H. Sun, and H. Wang, "Editorial: Epigenetics of Metabolism, Immunology and Aging," Frontiers in Genetics 14 (2023): 1135889, https://doi.org/10.3389/fgene.2023.1135889.
- [159] S. Hamsanathan, T. Anthonymuthu, D. Prosser, et al., "A Molecular Index for Biological Age Identified From the Metabolome and Senescence-Associated Secretome in Humans," Aging Cell 23, no. 4 (2024): e14104, https://doi.org/10.1111/acel.14104.
- [160] M. Sarkar, N. Niranjan, and P. K. Banyal, "Mechanisms of Hypoxemia," *Lung India* 34, no. 1 (2017): 47–60, https://doi.org/10.4103/0970-2113.197116.
- [161] M. Boulton and A. Al-Rubaie, "Neuroinflammation and Neurodegeneration Following Traumatic Brain Injuries," *Anatomical Science International* 100, no. 1 (2025): 3–14, https://doi.org/10.1007/s12565-024-00778-2.

- [162] C. A. Smith, K. L. H. Carpenter, P. J. Hutchinson, P. Smielewski, and A. Helmy, "Candidate Neuroinflammatory Markers of Cerebral Autoregulation Dysfunction in Human Acute Brain Injury," *Journal of Cerebral Blood Flow* & Metabolism 43, no. 8 (2023): 1237–1253, https://doi.org/ 10.1177/0271678X231171991.
- [163] A. R. Folsom, "Fibrinolytic Factors and Atherothrombotic Events: Epidemiological Evidence," supplement 1, Annals of Medicine 32, 85–91.
- [164] W. R. Wilkerson and D. C. Sane, "Aging and Thrombosis," Seminars in Thrombosis and Hemostasis 28, no. 6 (2002): 555–568.
- [165] T. Ageeva, A. Rizvanov, and Y. Mukhamedshina, "NF-κB and JAK/STAT Signaling Pathways as Crucial Regulators of Neuroinflammation and Astrocyte Modulation in Spinal Cord Injury," Cells 13, no. 7 (2024): 581, https://doi.org/10.3390/ cells13070581.
- [166] S. S. Shen-Orr, D. Furman, B. A. Kidd, et al., "Defective Signaling in the JAK-STAT Pathway Tracks With Chronic Inflammation and Cardiovascular Risk in Aging Humans," *Cell Systems* 3, no. 4 (2016): 374–384.e4, https://doi.org/10.1016/j.cels.2016.09.009.
- [167] F. He, W. Ge, K. Martinowich, et al., "A Positive Autoregulatory Loop of Jak-STAT Signaling Controls the Onset of Astrogliogenesis," *Nature Neuroscience* 8, no. 5 (2005): 616–625.
- [168] J. Pammer, H. Rossiter, M. Bilban, et al., "PIWIL-2 and piR-NAs Are Regularly Expressed in Epithelia of the Skin and Their Expression Is Related to Differentiation," *Archives of Dermatological Research* 312, no. 10 (2020): 705–714, https://doi.org/10.1007/s00403-020-02052-7.
- [169] D. Aprile, D. Patrone, G. Peluso, and U. Galderisi, "Multipotent/ Pluripotent Stem Cell Populations in Stromal Tissues and Peripheral Blood: Exploring Diversity, Potential, and Therapeutic Applications," *Stem Cell Research & Therapy* 15, no. 1 (2024): 139, https://doi.org/10.1186/s13287-024-03752-x.
- [170] A. C. Maritim, R. A. Sanders, and J. B. Watkins 3rd, "Diabetes, Oxidative Stress, and Antioxidants: A Review," *Journal of Biochemical and Molecular Toxicology* 17, no. 1 (2003): 24–38, https://doi.org/10.1002/jbt.10058.
- [171] U. Singh, S. Devaraj, and I. Jialal, "Vitamin E, Oxidative Stress, and Inflammation," *Annual Review of Nutrition* 25 (2005): 151–174.
- [172] J. P. Castro, T. Jung, T. Grune, and W. Siems, "4-Hydroxynonenal (HNE) Modified Proteins in Metabolic Diseases," Free Radical Biology and Medicine 111 (2017): 309–315, https://doi.org/10.1016/j.freeradbiomed.2016.10.497.
- [173] M. J. Duryee, L. W. Klassen, C. S. Schaffert, et al., "Malondial-dehyde-Acetaldehyde Adduct Is the Dominant Epitope After MDA Modification of Proteins in Atherosclerosis," *Free Radical Biology and Medicine* 49, no. 10 (2010): 1480–1486, https://doi.org/10.1016/j.freeradbiomed.2010.08.001.
- [174] H. J. Lin, S. T. Chen, H. Y. Wu, et al., "Urinary Biomarkers of Oxidative and Nitrosative Stress and the Risk for Incident Stroke: A Nested Case-Control Study From a Community-Based Cohort," *International Journal of Cardiology* 183 (2015): 214–220, https://doi.org/10.1016/j.ijcard.2015.01.043.
- [175] E. Zillessen, F. Frerichs, U. Herzfeld, R. Bader, and W. Hunstein, "Iron Absorption After Oral Application of a Combination of Fe2+-Succinate and Multivitamins (Author's Transl)," Arzneimittelforschung 27, no. 8 (1977): 1606–1608.

- [176] V. Seshadri Reddy, P. Duggina, M. Vedhantam, M. Manne, N. Varma, and S. Nagaram, "Maternal Serum and Fetal Cord-Blood Ischemia-Modified Albumin Concentrations in Normal Pregnancy and Preeclampsia: A Systematic Review and Meta-Analysis," *Journal of Maternal-Fetal & Neonatal Medicine* 31, no. 24 (2018): 3255–3266, https://doi.org/ 10.1080/14767058.2017.1368480.
- [177] J. Mialet-Perez and E. Belaidi, "Interplay Between Hypoxia Inducible Factor-1 and Mitochondria in Cardiac Diseases," *Free Radical Biology and Medicine* 221 (2024): 13–22, https://doi.org/10.1016/j.freeradbiomed.2024.04.239.
- [178] V. H. Haase, "Hypoxic Regulation of Erythropoiesis and Iron Metabolism," *American Journal of Physiology-Renal Physiology* 299, no. 1 (2010): F1–13, https://doi.org/10.1152/ajprenal.00174.2010.
- [179] N. R. Henig and D. J. Pierson, "Mechanisms of Hypoxemia," Respiratory Care Clinics of North America 6, no. 4 (2000): 501–521
- [180] K. A. Solari and E. A. Hadly, "Evolution for Extreme Living: Variation in Mitochondrial Cytochrome c Oxidase Genes Correlated With Elevation in Pikas (Genus Ochotona)," *Integrative Zoology* 13, no. 5 (2018): 517–535, https://doi.org/10.1111/1749-4877.12332.
- [181] R. S. Rogers, H. Wang, T. J. Durham, et al., "Hypoxia Extends Lifespan and Neurological Function in a Mouse Model of Aging," *PLoS Biology* 21, no. 5 (2023): e3002117, https://doi.org/10.1371/journal.pbio.3002117.
- [182] Y. Cao, S. Cao, R. L. Ge, H. Bao, Y. Mou, and W. Ji, "Brain-Aging Related Protein Expression and Imaging Characteristics of Mice Exposed to Chronic Hypoxia at High Altitude," Frontiers in Aging Neuroscience 15 (2023): 1268230, https://doi.org/10.3389/fnagi.2023.1268230.
- [183] C. Coronel-Oliveros, V. Medel, G. A. Whitaker, et al., "Elevating Understanding: Linking High-Altitude Hypoxia to Brain Aging Through EEG Functional Connectivity and Spectral Analyses," *Network Neuroscience* 8, no. 1 (2024): 275–292, https://doi.org/10.1162/netn_a_00352.
- [184] K. M. Cochrane, J. A. Hutcheon, and C. D. Karakochuk, "Iron-Deficiency Prevalence and Supplementation Practices Among Pregnant Women: A Secondary Data Analysis From a Clinical Trial in Vancouver, Canada," *Journal of Nutrition* 152, no. 10 (2022): 2238–2244, https://doi.org/10.1093/jn/nxac135.
- [185] Y. Xing, F. Xuan, K. Wang, and H. Zhang, "Aging Under Endocrine Hormone Regulation," Frontiers in Endocrinology 14 (2023): 1223529, https://doi.org/10.3389/fendo.2023.1223529.
- [186] R. Sharifi, M. F. Tabarzadi, P. Choubsaz, et al., "Evaluation of Serum and Salivary Iron and Ferritin Levels in Children With Dental Caries: A Meta-Analysis and Trial Sequential Analysis," *Children* 8, no. 11 (2021): 1034, https://doi.org/ 10.3390/children8111034.
- [187] K. S. Schin and U. Clever, "Ferritin-Uptake by Salivary Glands of Chironomus tentans and Its Intracellular Localization," Experimental Cell Research 49, no. 1 (1968): 208–211.
- [188] N. Jagannathan, C. Thiruvengadam, P. Ramani, P. Premkumar, A. Natesan, and H. J. Sherlin, "Salivary Ferritin as a Predictive Marker of Iron Deficiency Anemia in Children," *Journal of Clinical Pediatric Dentistry* 37, no. 1 (2012): 25–30.
- [189] J. H. Kang, Y. H. Lee, and H. S. Kho, "Clinical Factors Affecting Salivary Transferrin Level, a Marker of Blood Contamination in Salivary Analysis," *BMC Oral Health* 18, no. 1 (2018): 49, https://doi.org/10.1186/s12903-018-0510-x.