

REVIEW ARTICLE

Regenerative potential of Wharton's jelly-derived mesenchymal stem cells: A new horizon of stem cell therapy

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Abstract

Umbilical cord Wharton's jelly-derived mesenchymal stem cells (WJ-MSCs) have recently gained considerable attention in the field of regenerative medicine. Their high proliferation rate, differentiation ability into various cell lineages, easy collection procedure, immuno-privileged status, nontumorigenic properties along with minor ethical issues make them an ideal approach for tissue repair. Besides, the number of WJ-MSCs in the umbilical cord samples is high as compared to other sources. Because of these properties, WJ-MSCs have rapidly advanced into clinical trials for the treatment of a wide range of disorders. Therefore, this paper summarized the current preclinical and clinical studies performed to investigate the regenerative potential of WJ-MSCs in neural, myocardial, skin, liver, kidney, cartilage, bone, muscle, and other tissue injuries.

KEYWORDS

mesenchymal stem cells, regenerative medicine, umbilical cord, Wharton's jelly

1 | INTRODUCTION

Emerging data have indicated that mesenchymal stem cells (MSCs) provided a promising tool for cell therapy and regeneration of human tissues due to their differentiation multipotency, self-renewal capability, long-term ex vivo proliferation, paracrine effects, and immunomodulatory potential (El Omar et al., 2014; Wang, Yuan, & Xie, 2018b). MSCs can be collected from adult and fetal/perinatal tissues, such as bone marrow (BM), adipose tissue, endometrial polyps, dental pulp, peripheral blood as well as the placenta, endometrium, umbilical cord blood, and Wharton's jelly (Kalaszczynska & Ferdyn, 2015; Marino et al., 2019).

MSCs derived from adult tissues, especially BM, are most frequently used for therapeutic purposes, however, their use has been

hampered because of some reasons such as their limited proliferative ability, low cell contents, and invasive and painful isolation procedure associated with significant morbidity and risk of infection (about 0.01–0.001% of total mononuclear cells; Mazini, Rochette, Amine, & Malka, 2019; Nancarrow-Lei, Mafi, Mafi, & Khan, 2017). Additionally, donor health, age, genetics, and exposure to environmental stress could potentially affect the proliferation and differentiation capacity of MSCs (Brown et al., 2019).

Unlike adult MSCs, Wharton's jelly derived mesenchymal stem cells (WJ-MSCs) have recently generated a tremendous level of interest for allogeneic or autologous application because of their beneficial properties including unlimited availability, low cost, large quantities, easy and noninvasive isolation procedure and higher proliferative capacity (Abbaszadeh, Ghorbani, Derakhshani, Movassaghpour, & Yousefi, 2020;

Kalaszczyńska & Ferdyn, 2015). WJ-MSCs are also less likely to cause graft versus host disease and appear to be nontumorigenic and do not induce teratomas after transplantation (Brown et al., 2019; El Omar et al., 2014). Additionally, these MSCs are very young cells that have affected less environmental interference (Vieira Paladino, de Moraes Rodrigues, da Silva, & Goldberg, 2019). Besides, their research does not raise any ethical concerns associated with their application in regenerative medicine. Furthermore, WJ-MSCs possess an immune-privileged status which makes them an ideal option for regenerative medical use (Marino et al., 2019).

For clinical application, WJ-MSCs should be extracted from healthy donors of full-term pregnancies over the age of 18 and who have water broken for no longer than 18 hr (Vieira Paladino et al., 2019). Furthermore, due to the higher expression of OCT-4 and DNMT1, WJ-MSCs derived from male subjects have higher and quicker differentiation potential (Balzano et al., 2019a; Balzano et al., 2019b).

Considering these advantages, in this review, we first provided a summary of the biological characteristics of WJ-MSCs. Then, we attempted to describe several recent nonclinical and clinical trials which evaluated their regenerative efficiency or safety in various disorders including neurological, cardiovascular, cutaneous, liver, and kidney diseases as well as cartilage, bone, muscle and several other tissue damages. Last, the concluding part summarized the current status, addressed existing challenges, and gave an outlook on potential future prospects.

2 | ISOLATION AND CHARACTERISTICS OF WJ-MSCs

WJ is the connective tissue of the umbilical cord which was initially described by Thomas Wharton in 1656 (Davies, Walker, & Keating, 2017). The first isolation of MSCs from the WJ portion of the human umbilical cord was achieved by McElreavey and colleagues in 1991 (McElreavey, Irvine, Ennis, & McLean, 1991).

One centimeter of the umbilical cord yields approximately $1-5 \times 10^4$ MSCs (Kalaszczyńska & Ferdyn, 2015). Currently, digestion-based methods are one of the most common techniques applied for the isolation of MSCs from the extracellular matrix of WJ by using collagenase, hyaluronidase, or other proteases (Liau, Ruszymah, Ng, & Law, 2019; Widowati et al., 2019). The doubling time of MSCs isolated by this technique is considerably longer, however, they may increase the risk of cellular damages and reduce cell viability. Alternatively, tissue explant can be used for WJ-MSCs isolation because of some benefits such as lower cost and higher cell yield. Due to the release of paracrine factors during the in vitro culture, the proliferative capacity of isolated WJ-MSCs through the explant method is also higher as compared to enzymatic protocols (Hassan, Kasem, Soukkaieh, & Aljamali, 2017; Kalaszczyńska & Ferdyn, 2015; Liau et al., 2019).

Phenotypic investigation performed by several groups evidenced that WJ-MSCs fit the minimal criteria for MSCs as proposed by the

International Society for Cellular Therapy (Dominici et al., 2006). They are spindle-shaped fibroblast-like cells expressing typical mesenchymal markers such as CD73, CD90, CD105, CD13, CD29, and CD44, while lacking expression of hematopoietic, CD45, CD34, and endothelial, CD31, markers (Corrao et al., 2013; Kalaszczyńska & Ferdyn, 2015). WJ-MSCs can also be unique options for therapeutic applications due to their primitive properties, as they display several characteristics of embryonic stem cells (ESCs), such as ESC-like antigen Tra-1-60, Tra-1-81, SSEA-1, and SSEA-4, and pluripotency genes including Oct-4, Nanog, SSEA-4, and SOX-2 (Marino et al., 2019). Moreover, WJ-MSCs are safe for allogeneic therapeutic goals because of their immune-privileged status. They present a minimal expression for major histocompatibility complex (MHC) Class I and no expression for MHC-II that protect them from Natural killer cell-mediated lysis. They also lack expression of costimulatory molecules CD40, CD80, and CD86 and have high levels of immune inhibitors such as prostaglandin E2, indoleamine-2,3-dioxygenase, and HLA-G (Weiss et al., 2008; Zhou et al., 2011).

3 | REGENERATIVE EFFECTS OF WJ-MSCs IN DIFFERENT TISSUES

Accumulating data have recently evaluated the potential contribution of WJ-MSCs in the treatment of different diseases or tissue regeneration, which are discussed in the following parts (Figure 1).

3.1 | Neural regeneration

The application of MSCs derived from human WJ has been yielded promising results in the treatment of neurological diseases. It has been established that the WJ-MSCs are capable of differentiating into neural cells (Ma et al., 2005; Peng et al., 2011). WJ-MSCs are also associated with more nerve regeneration, neuroprotection, and less inflammation following spinal cord injury (SCI) in dogs compared to adult MSCs from BM or adipose tissue (Ryu et al., 2012). In 2008, a study provided evidence that transplantation of human WJ-MSCs is an effective approach to promote the regeneration of corticospinal fibers and locomotor recovery after SCI in rats (Yang et al., 2008). Intrathecal transplantation of WJ-MSCs has demonstrated anti-inflammatory effects in SCI rat models through targeting the inflammasome complex (Mohamadi et al., 2018; Mohamadi et al., 2019). Additionally, Li et al. (2016), indicated that the recovery of motor function and integrity of the spinal cord was better in WJ-MSC transplanted rats than the control group because of reducing interleukin-1b and increasing nerve growth factor expression. It has also been concluded that the therapeutic effect of WJ-MSCs in SCI is dose-dependent and could be accelerated by repeated application (Krupa et al., 2018). Nevertheless, it has been proven by Chudickova et al. (2019) that conditioned media derived from WJ-MSCs has a superior effect than direct MSC transplantation for the treatment of SCI in rats.

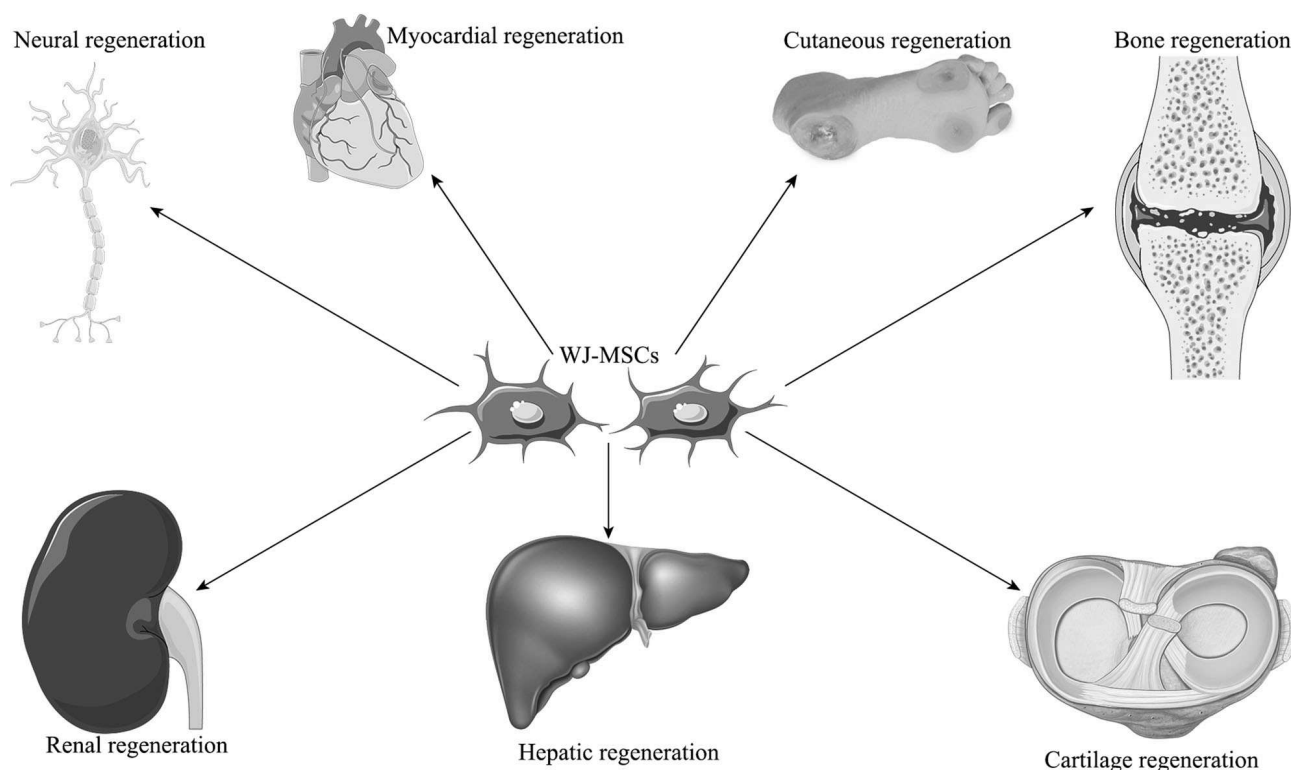


FIGURE 1 Regenerative effects of Wharton jelly mesenchymal stem cells (WJ-MSCs) in various tissues

In a study conducted by Fu et al (2006), human WJ-MSCs were successfully differentiated into dopaminergic neurons *in vitro*. Then, the administration of these differentiated MSCs into the striatum of rats with Parkinson's disease (PD) could alleviate amphetamine-induced rotation, showing their promising therapeutic effects. Transplantation of undifferentiated human WJ-MSCs was also significantly reduced amphetamine-evoked rotation in the rat model of PD (Weiss et al., 2006).

Ding et al. (2007) reported that intracerebral injection of WJ-MSCs into the stroke rat model could enhance neuroplasticity via regulating the expression of $\beta 1$ -integrin. Furthermore, neural differentiation of human WJ-MSCs could improve the recovery of neurological function after implantation in ischemic stroke rats (Zhang et al., 2017). Intracerebral transplantation of human WJ-MSCs has also neuroprotective effects on a rodent model of stroke through reducing inflammation and increasing the expression of trophic factors (Wu et al., 2018b).

Moreover, Wang et al. (2019) have recently investigated the therapeutic effect of human WJ-MSCs in a mouse model of transected sciatic nerves. The data showed that the human WJ-MSCs group had a significantly better functional recovery, quality of regenerated nerve, and higher neurotrophic factors in comparison with the nontreated and human adipose stem cell treated control groups.

In another study, Zhang et al. (2014a) focused on the potential of human WJ-MSCs in the treatment of neonatal rats with hypoxic-ischemic encephalopathy (HIE). The results indicated that intravenous transplantation of these MSCs at an early stage of HIE

significantly improved the behavior of rats and suppressed gliosis. It has also been shown that thrombin preconditioning of human WJ-MSCs could remarkably attenuate severe HIE-induced brain infarction and improve behavioral function in neonatal rats through enhancing their anti-inflammatory, antiapoptotic and antiastrogial activities (Kim et al., 2019). Another study suggested that extracellular vesicles derived from human WJ-MSCs might protect against HI-induced apoptosis in neuronal cells by delivering let-7-5p miR (Joerger-Messerli et al., 2018).

3.2 | Myocardial regeneration

The beneficial effects of WJ-MSCs in the regeneration of myocardial tissue have been shown in several studies. For instance, Lopez et al. (2013) suggested that intravenous administration of WJ-MSCs might be more beneficial than BM-MSCs as an off-the-shelf therapy for myocardial ischemia. Liu et al. (2016) found that intracoronary transplantation combined with multiple intravenous deliveries of WJ-MSCs may enhance left ventricular function, perfusion, and remodeling in the porcine model of chronic myocardial infarction. The impact of WJ-MSCs has also been evaluated in acute myocardial infarction (AMI) porcine models and followed-up to 6 weeks (Zhang et al., 2013). The transplanted MSCs were able to differentiate into cardiomyocytes, induce angiogenesis, reduce apoptosis and fibrosis, decrease infarct size, and consequently enhanced ventricular remodeling and function. Moreover, WJ-MSC therapy was shown to

attenuate interstitial fibrosis and cardiac dysfunction in a dilated cardiomyopathy rat model by decreasing tumor necrosis factor- α and transforming growth factor- β 1 (TGF- β 1) expression and ERK1/2 signaling activation (Zhang et al., 2018b). Bhuvanalakshmi, Arfuso, Kumar, Dharmarajan, and Warriar (2017) reported that epigenetic modification could enhance the differentiation of WJ-MSCs into the cardiomyocyte lineage through regulation of Wnt mediators in cardiac injury mice model. It has also been proposed that combinatory therapy with human WJ-MSCs and insulin-like growth factor-1 significantly enhanced myocardial regeneration, angiogenesis, reduced fibrosis, and improved cardiac function in a rabbit model of MI (Rabbani et al., 2018). Furthermore, WJ-MSCs modified to overexpress antifibrotic factors provided a novel strategy for protecting cardiac fibrosis in vitro (Nimsanor, Phetfong, Kitiyanant, Kamprom, & Supokawej, 2019). It has also been shown that WJ-MSCs could be a more suitable substitute of endothelial and interstitial cells in cardiovascular regenerative medicine as compared to amniotic membrane because of their higher viability, proliferation, and superior collagen deposition properties (Pu et al., 2017).

Martin-Rendon et al. (2008) compared the differentiation potential of MSCs derived from BM, umbilical cord, and cord blood into cardiomyocytes in vitro. In spite of other studies, their result indicated that WJ and cord blood MSCs did not express cardiac-specific markers either spontaneously or after treatment with 5-azacytidine and may not be able to generate an acceptable amount of cardiomyocytes in MI cell therapy without pharmacological or genetic modification.

3.3 | Cutaneous regeneration

WJ-MSCs have also been broadly studied as a promising source for skin wound healing. For example, as described by Azari et al. (2011) in their article, caprine WJ-MSCs showed complete re-epithelialization of the wounds with minimum inflammation and scar formation on Day 7. However, no complete epithelialization was observed in the untreated control group after 12 days. Administration of human WJ-MSCs has also improved wound healing in an excisional full-thickness skin murine model through enhancing re-epithelialization, vascularization, proliferation, survival, and migration of skin fibroblasts (Arno et al., 2014). An animal model of burn has also been employed to confirm the potential of WJ-MSC therapy in this pathology. Subcutaneous injection of the WJ-MSCs suppressed secondary inflammation via reducing inflammatory cytokines and, therefore, promoted burn healing process and skin repair in rat burn models (Zhang et al., 2015). Sun, Zhang, Song, Zhu, and Zhu (2019) assessed the healing effects of these MSCs in radiation-induced skin injury in rats. They found that WJ-MSCs improved wound healing quality by promoting cell proliferation, sebaceous glands regeneration, and angiogenesis. Another animal study showed that WJ-MSCs primed with poly I:C or Interferon- γ have enhanced anti-inflammatory potential in atopic dermatitis (Park et al., 2019). The impact of heterologous WJ-MSCs has also been

investigated on a nonhealing large chronic wound in a filly. In this study, the application of WJ-MSCs contributed to complete wound healing with no side events or scar tissue formation (Lanci, Merlo, Mariella, Castagnetti, & Iacono, 2019). Recently, Martin-Piedra et al. (2019) also reported that because of their epidermal differentiation capability and lack of HLA antigens, WJ-MSCs are more suitable sources for the generation of bioengineered human skin substitute than MSCs derived from adipose tissue, dental pulp, and BM. Additionally, two studies have been demonstrated that a mixture of WJ-MSCs and skin microparticles could remarkably elevate the quality of cutaneous wound regeneration in mice models (Shi, Jia, Liu, & Chen, 2015; Zhang, Hao, Liu, Fu, & Han, 2012b). WJ-MSCs administered on a decellularized amniotic membrane scaffold into the wound bed, showed reduced scar formation with hair growth (Sabapathy, Sundaram, Mankuzhy, & Kumar, 2014). Similarly, the combination of WJ-MSCs and silk fibroin scaffold exhibited significant enhancement in wound re-epithelization and reduced fibrotic scar formation (Millan-Rivero et al., 2019). Besides, the association of human WJ-MSCs and poly (vinyl alcohol) hydrogel membrane resulted in the promotion of wound healing in two dogs with chronic skin wounds where the standard treatments failed (Ribeiro et al., 2014).

3.4 | Hepatic regeneration

The potential of WJ-MSCs to differentiate into the endodermal lineage, such as hepatocyte-like cells, makes them an attractive alternative for the treatment of liver diseases (Buyl et al., 2014; Mortezaee et al., 2015; Vojdani et al., 2015; Zhang, Lie, & Wei, 2009). The impact of WJ-MSCs has been investigated in a liver fibrosis rat model by Tsai et al. (2009). They reported that the administration of human WJ-MSCs could significantly reduce liver fibrosis by decreasing the collagen deposition, levels of serum glutamic oxaloacetic transaminase, glutamic pyruvate transaminase, as well as TGF- β 1 and increasing mesenchymal-epithelial transition factor-phosphorylated type and hepatocyte growth factor (HGF). Lin et al. (2010) also determined that intravenous injection of human WJ stem cells downregulate liver collagen and thus could alleviate liver fibrosis in rat models. In addition, WJ-MSCs have been proven to have the capacity in the enhancement of sepsis-associated liver damage in rats (Condor et al., 2016). Li et al. (2015) also loaded WJ-MSCs into poly scaffolds and administered into liver-injured mice. One month after WJ-MSC transplantation, the liver morphology significantly promoted and looked similar to a normal one. Furthermore, one report has indicated that WJ-MSC therapy is able to improve liver function and rescue the hepatotoxicity induced by D-Galactosamine in acute liver injury mice model (Ramanathan et al., 2017). Zhang et al. (2012a) found that human WJ-MSCs could be a potential source for cell therapy of acute liver failure when compared to human hepatocytes, because of their ability in stimulating endogenous liver regeneration through paracrine effects. It has been also shown that intrauterine xenotransplantation of WJ-MSCs successfully

relieved liver injury in rabbit fetuses through inducing the expressions of albumin, α -fetoprotein, hepatic nuclear Factor 4, and CYP2B6 messenger RNA (Rezaeian et al., 2018). Another study suggested that the application of praziquantel in combination with WJ-MSCs has more beneficial effects in the treatment of the *Schistosoma mansoni*-induced liver fibrosis, presumably via enhancing differentiation capacity of the transplanted WJ-MSCs to functioning liver-like cells (Hammam et al., 2016).

Nevertheless, Rengasamy et al. (2017) suggested that human BM-MSCs alleviate CCl₄-induced liver fibrosis more effectively than WJ-MSCs in rat models which could be associated with their expression of certain matrix metalloprotease and angiogenic factors.

3.5 | Renal regeneration

With a rat model of unilateral ischemia-reperfusion injury (IRI), a single intravenous administration of WJ-MSCs exerted beneficial effects on IRI-induced acute and chronic kidney injury via an endocrine mechanism (Du et al., 2012). According to the results of this study, activated Akt signal in tubular epithelial cells (TECs) resulted in a decrease of apoptosis, the elevation of proliferation, and the induction of endogenous HGF. Similarly, another animal study demonstrated that these MSCs contributed to the induction of native and foreign HGF in damaged TECs at the initial stage of acute kidney injury (AKI), which subsequently resulted in the EMT delay and mitigation of renal fibrosis (Du et al., 2013). It has also been shown that human WJ-MSCs seeded in a decellularized kidney scaffold could ameliorate renal fibrosis through decreasing EMT by the TGF- β /Smad signaling pathway after subtotal nephrectomy in rats (Hu et al., 2020). Moreover, the human WJ-MSC potential has been proven in the promotion of renal injury in streptozotocin-induced diabetic mice (Maldonado, Huang, Yang, Xu, & Ma, 2017).

Several studies also indicated that microvesicles derived from WJ-MSCs ameliorate renal IRI through various mechanisms including regulating Erk1/2 signaling, enhancing renal cell proliferation, mitigating apoptosis and inflammation, and suppressing CX3CL1 (Chen, Yan, Song, Ding, & Du, 2017; Wu et al., 2018a; Zou et al., 2014). Zhang et al. (2014b) reported that a single infusion of human WJ-MSC-microvesicles could prevent renal injury by alleviating oxidative stress in the early stage of IRI through downregulating NOX2 expression. Moreover, extracellular vesicles released from human WJ-MSCs protected against AKI via antioxidation by improving Nrf2/ARE activation in rats (Zhang et al., 2016).

3.6 | Cartilage regeneration

It has been established that human WJ-MSCs could be better sources for fibrocartilage tissue engineering compared to BM-MSCs (Wang, Tran, Seshareddy, Weiss, & Detamore, 2009). It has been shown that WJ-MSCs could be an appealing source, when combined with appropriate scaffolds, for cell therapy in cartilage disorders. Recently,

Zhang et al. (2019) revealed that coculture of human WJ-MSCs and primary articular cartilage cells in acellular cartilage extracellular matrix-oriented scaffold could successfully construct engineered hyaline articular cartilage in vitro. Moreover, a study conducted by Paduszyński et al. (2016), assessed changes in transcriptional activity of cartilaginous genes through in vitro chondrogenic differentiation of WJ-MSCs on 3D poly (L-lactide-co-glycolide, PLGA) scaffolds. Their findings suggested that the chondrogenic capacity of WJ-MSCs cultured on the PLGA scaffolds could be used as a promising cell-based cartilage repair therapy. Similarly, combining WJ-MSCs with 3D HyStem hydrogel and hybrid scaffolds promoted cartilage regeneration (Aleksander-Konert, Paduszynski, Zajdel, Dzierzewicz, & Wilczok, 2016; Shie et al., 2017). Besides, WJ-MSCs embedded into collagen hydrogel underwent enhanced chondrogenic differentiation by elevating the expression of cartilage-specific matrix proteins (Chen et al., 2013). A combination of WJ-MSCs and acellular cartilage extracellular matrix scaffold also provided better quality in repair and regeneration of cartilage in comparison with microfracture in a caprine model (Zhang et al., 2018a).

It has also been indicated that WJ-MSCs have great potential for fabricating tissue-engineered cartilage without scaffold (Liu et al., 2014). Modulation of metalloproteinase gene expression in synovium and prevention of cartilage degradation reported in 30 rabbits with osteoarthritis (OA), following intra-articular administration of WJ-MSCs into the knee joint (Saulnier et al., 2015). Cheng et al. (2019) have found that combined extracorporeal shockwave therapy and WJ-MSCs are more effective for the treatment of early OA in rat knee.

3.7 | Bone regeneration

Growing evidences have demonstrated that WJ-MSCs are potentially valuable candidates for bone regeneration as an alternative to BM-MSCs (Ansari et al., 2018; Cabrera-Perez et al., 2019; Kang et al., 2012; Lim et al., 2018). In a recent report, Bustos et al. (2017) have found that modulation of RUNX2/p57 expression via inhibition of JARID1B histone demethylase is associated with the upregulation of WJ-MSC osteoblastic potential. Likewise, RUNX2/p57, but not SP7 gene expression, has demonstrated to be strongly activated upon induction of WJ-MSC osteoblast differentiation (Sepulveda et al., 2017). Another study conducted by Shi, Zhou, Huang, Zhang, and Wang (2018) provided additional insight about the interactions of hydroxyapatite nanoparticles with WJ-MSCs and suggested their promising therapeutic potential in bone repair.

Todeschi et al. (2015) proposed that subcutaneous implantation of WJ-MSCs loaded onto scaffolds in mice with bone defects could result in bone regeneration by the enhancement of angiogenesis. Human WJ-MSCs seeded into hierarchical fibrous scaffolds were also shown as an appropriate strategy for bone tissue engineering (Canha-Gouveia et al., 2015). Moreover, human WJ-MSCs-loaded nano-hydroxyapatite/chitosan/gelatin scaffolds were shown as an ideal bone substitute (Jamalpoor, Taromi, Soleimani, Koudehi, & Asgari, 2019).

A recent systematic review conducted by Ansari et al. (2018) systematically analyzed the osteogenic induction capacity of WJ-MSCs. They concluded that WJ-MSCs are potential stem cell sources which could be used as an alternative to BM-MSCs for bone fracture healing.

3.8 | Muscular regeneration

Recently, Su et al. (2019) reported that WJ-MSCs could restore functional impairment postmuscle injury in mice via downregulating neutrophil-mediated acute inflammation and antifibrotic effects. Kwon et al. (2016) elucidated that human WJ-MSCs and human WJ-MSC-derived XCL1 protein could be used as novel therapeutic approaches for the treatment of myopathies by their antiapoptotic effects. In addition, human WJ-MSCs successfully attenuated sarcopenia in aged mice model associated with skeletal muscle cell activation, and downregulation of apoptosis and inflammation (Wang et al., 2018a). It has also been demonstrated that TGF- β 1 and ascorbic acid are effective in differentiating WJ-MSCs towards smooth muscle cells (Mesure, Huber-Villaume, Menu, & Velot, 2017). The pretreatment of WJ-MSCs with Sdf-1 was proven to play a key role in increasing their migration ability during skeletal muscle regeneration in vitro (Kowalski et al., 2017). Moreover, WJ-MSCs are associated with an increased neuro-muscular regeneration when combined with biodegradable and biocompatible biomaterials, enhancing the recovery of sensory and motor function in neurotmesis injuries (Caseiro et al., 2017).

4 | THERAPEUTIC APPLICATION OF WJ-MSCs IN CLINICAL TRIALS

In a controlled case clinical trial, a 16-year-old boy diagnosed with HIE received an intrathecal, intramuscular, and intravenous infusion of WJ-MSCs. The clinical outcomes progress suggested the feasibility and safety of the triple route WJ-MSC transplantations in the HIE patient (Kabatas et al., 2018). One clinical study has also been recently investigated the safety of intrathecal administration of WJ-MSCs in 43 patients with amyotrophic lateral sclerosis (Barczewska et al., 2019). No serious adverse effects were observed during the 6-month follow-up period. Recently, Hashemi et al. (2019) reported a randomized clinical trial where acellular amniotic membrane seeded with WJ-MSCs were used for 9 days, every 3 days in five patients with chronic diabetic ulcers. This treatment accelerated wound healing through a significant decrease of the wound size and time of healing (Hashemi et al., 2019). Zhang et al. (2012a, 2012c) conducted a nonrandomized controlled trial involving 45 chronic hepatitis B patients with decompensated liver cirrhosis. Thirty patients received WJ-MSC therapy and the remaining 15 patients received saline as the control. The authors revealed that treatment with WJ-MSCs is clinically safe and mediated a significant decrease in the volume of ascites and enhanced liver function. Safety and efficacy of WJ-MSC

transfusion in patients with hepatitis B virus-associated acute-on-chronic liver failure (ACLF) have been reported in an open-labeled, Phase I/II trial. No serious adverse effects were observed during this study and there was a significant increase in survival rates, improvements in liver function, and reduction in a model of end-stage liver disease scores in ACLF patients (Shi et al., 2012). Sadlik et al. (2017, 2018) presented a novel, one-stage, and minimally invasive strategy for knee cartilage regeneration in a patient using WJ-MSCs and collagen scaffold under dry arthroscopy technique. Musialek et al. (2015) treated 10 patients with AMI using WJ-MSCs, indicating the feasibility and safety of these MSCs as an off-the-shelf therapeutic agent. Likewise, a double-blind, randomized controlled trial has been performed by Gao et al. (2015) to investigate the efficacy and safety of WJ-MSCs in the treatment of patients with ST-elevation AMI. A total of 116 patients were recruited and randomly received an intracoronary injection of WJ-MSCs or placebo repeatedly, and followed-up to 18 months. It was concluded that WJ-MSCs were safe in AMI therapy and successfully improved myocardial viability and heart function.

5 | CONCLUSION AND FUTURE PERSPECTIVE

As compared with adult MSCs, WJ-MSCs have prominent implications in regenerative medicine because of their several unique properties, including multipotent differentiation ability, unlimited availability, high amount, easy and noninvasive separation process, superior proliferation, and cost-effectiveness. Additionally, their immuno-privileged status, nontumorigenic features, along with no ethical issues make them a promising candidate for tissue regeneration or repair. The increasing number of studies have been evaluated the feasibility of WJ-MSCs in various diseases, however, more clinical trials are still required to warrant the safety and efficacy of this therapeutic strategy before their routine use in the clinic. Application of the WJ-MSCs in combination with scaffolds provide a three-dimensional structure which promotes their adhesion and proliferation and subsequently enhancing the therapeutic results. Besides, more understanding of the mechanisms through which the advantageous effects of WJ-MSCs are reached can facilitate the modification of these MSCs to improve their future clinical application.

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CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

AUTHOR CONTRIBUTIONS

H. A. and F. G. wrote the manuscript. M. T. drew the schematic figure. M. D. and M. Y. wrote some parts of the manuscript and edited the

final version. A. M. and K. S. designed and supervised the study, whole correspondence during the paper submission.

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REFERENCES

- Abbaszadeh, H., Ghorbani, F., Derakhshani, M., Movassaghpour, A., & Yousefi, M. (2020). Human umbilical cord mesenchymal stem cell-derived extracellular vesicles: A novel therapeutic paradigm. *Journal of Cellular Physiology*, 235(2), 706–717. <https://doi.org/10.1002/jcp.29004>
- Aleksander-Konert, E., Paduszynski, P., Zajdel, A., Dzierzewicz, Z., & Wilczok, A. (2016). In vitro chondrogenesis of Wharton's jelly mesenchymal stem cells in hyaluronic acid-based hydrogels. *Cellular and Molecular Biology Letters*, 21, 11. <https://doi.org/10.1186/s11658-016-0016-y>
- Ansari, A. S., Yazid, M. D., Sainik, N., Razali, R. A., Saim, A. B., & Idrus, R. B. H. (2018). Osteogenic Induction of Wharton's Jelly-derived mesenchymal stem cell for bone regeneration: A systematic review. *Stem Cells International*, 2018, 2406462. <https://doi.org/10.1155/2018/2406462>
- Arno, A. I., Amini-Nik, S., Blit, P. H., Al-Shehab, M., Belo, C., Herer, E., ... Jeschke, M. G. (2014). Human Wharton's jelly mesenchymal stem cells promote skin wound healing through paracrine signaling. *Stem Cell Research & Therapy*, 5(1), 28. <https://doi.org/10.1186/scrt417>
- Azari, O., Babaei, H., Derakhshanfar, A., Nematollahi-Mahani, S. N., Poursahebi, R., & Moshrefi, M. (2011). Effects of transplanted mesenchymal stem cells isolated from Wharton's jelly of caprine umbilical cord on cutaneous wound healing: histopathological evaluation. *Veterinary Research Communications*, 35(4), 211–222. <https://doi.org/10.1007/s11259-011-9464-z>
- Balzano, F., Bellu, E., Basoli, V., Dei Giudici, S., Santaniello, S., Cruciani, S., ... Maioli, M. (2019a). Lessons from human umbilical cord: Gender differences in stem cells from Wharton's jelly. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 234, 143–148. <https://doi.org/10.1016/j.ejogrb.2018.12.028>
- Balzano, F., Campesi, I., Cruciani, S., Garroni, G., Bellu, E., Dei Giudici, S., ... Maioli, M. (2019b). Epigenetics, stem cells, and autophagy: Exploring a path involving miRNA. *International Journal of Molecular Sciences*, 20(20), 5091. <https://doi.org/10.3390/ijms20205091>
- Barczewska, M., Grudniak, M., Maksymowicz, S., Siwek, T., Oldak, T., Jezierska-Wozniak, K., ... Maksymowicz, W. (2019). Safety of intrathecal injection of Wharton's jelly-derived mesenchymal stem cells in amyotrophic lateral sclerosis therapy. *Neural Regeneration Research*, 14(2), 313–318. <https://doi.org/10.4103/1673-5374.243723>
- Bhuvanalakshmi, G., Arfuso, F., Kumar, A. P., Dharmarajan, A., & Warriar, S. (2017). Epigenetic reprogramming converts human Wharton's jelly mesenchymal stem cells into functional cardiomyocytes by differential regulation of Wnt mediators. *Stem Cell Research & Therapy*, 8(1), 185. <https://doi.org/10.1186/s13287-017-0638-7>
- Brown, C., McKee, C., Bakshi, S., Walker, K., Hakman, E., Halassy, S., ... Chaudhry, G. R. (2019). Mesenchymal stem cells: Cell therapy and regeneration potential. *Journal of Tissue Engineering and Regenerative Medicine*, 13(9), 1738–1755. <https://doi.org/10.1002/term.2914>
- Bustos, F., Sepúlveda, H., Prieto, C., Carrasco, M., Díaz, L., Palma, J., ... Palma, V. (2017). RUNX2 Induction during Differentiation of Wharton's jelly mesenchymal stem cells to osteoblasts is regulated by JARID1B histone demethylase. *Stem Cells*, 35, 2430–2441. <https://doi.org/10.1002/stem.2704>
- Buyl, K., De Kock, J., Najar, M., Lagneaux, L., Branson, S., Rogiers, V., & Vanhaecke, T. (2014). Characterization of hepatic markers in human Wharton's Jelly-derived mesenchymal stem cells. *Toxicology In Vitro*, 28(1), 113–119. <https://doi.org/10.1016/j.tiv.2013.06.014>
- Cabrera-Perez, R., Monguio-Tortajada, M., Gamez-Valero, A., Rojas-Marquez, R., Borrás, F. E., Roura, S., & Vives, J. (2019). Osteogenic commitment of Wharton's jelly mesenchymal stromal cells: Mechanisms and implications for bioprocess development and clinical application. *Stem Cell Research & Therapy*, 10(1), 356. <https://doi.org/10.1186/s13287-019-1450-3>
- Canha-Gouveia, A., Rita Costa-Pinto, A., Martins, A. M., Silva, N. A., Faria, S., Sousa, R. A., ... Neves, N. M. (2015). Hierarchical scaffolds enhance osteogenic differentiation of human Wharton's jelly derived stem cells. *Biofabrication*, 7(3):035009. <https://doi.org/10.1088/1758-5090/7/3/035009>
- Caseiro, A., Pereira, T., Ribeiro, J., Santos, J. D., Amorim, I., Luis, A., & Mauricio, A. C. (2017). Neuro-muscular regeneration using scaffolds with mesenchymal stem cells (MSCs) isolated from human umbilical cord Wharton's jelly. *Ciência & Tecnologia dos Materiais*, 29, e135–e139. <https://doi.org/10.1016/j.ctmat.2016.04.003>
- Chen, W., Yan, Y., Song, C., Ding, Y., & Du, T. (2017). Microvesicles derived from human Wharton's Jelly mesenchymal stem cells ameliorate ischemia-reperfusion-induced renal fibrosis by releasing from G2/M cell cycle arrest. *Biochemical Journal*, 474(24), 4207–4218. <https://doi.org/10.1042/bcj20170682>
- Chen, X., Zhang, F., He, X., Xu, Y., Yang, Z., Chen, L., ... Zeng, Y. (2013). Chondrogenic differentiation of umbilical cord-derived mesenchymal stem cells in type I collagen-hydrogel for cartilage engineering. *Injury* (J), 44(4), 540–549. <https://doi.org/10.1016/j.injury.2012.09.024>
- Cheng, J. H., Wang, C. J., Chou, W. Y., Hsu, S. L., Chen, J. H., & Hsu, T. C. (2019). Comparison efficacy of ESWT and Wharton's jelly mesenchymal stem cell in early osteoarthritis of rat knee. *American Journal of Translational Research*, 11(2), 586–598.
- Chudickova, M., Vackova, I., Urdzikova, L. M., Jancova, P., Kekulova, K., Rehorova, M., ... Kubinova, S. (2019). The effect of wharton jelly-derived mesenchymal stromal cells and their conditioned media in the treatment of a rat spinal cord injury. *International Journal of Molecular Sciences*, 20(18), 4516. <https://doi.org/10.3390/ijms20184516>
- Condor, J. M., Rodrigues, C. E., Sousa Moreira, R., Canale, D., Volpini, R. A., Shimizu, M. H., ... Andrade, L. (2016). Treatment with human Wharton's Jelly-derived mesenchymal stem cells attenuates sepsis-induced kidney injury, liver injury, and endothelial dysfunction. *Stem Cells Translational Medicine*, 5(8), 1048–1057. <https://doi.org/10.5966/sctm.2015-0138>
- Corrao, S., La Rocca, G., Lo Iacono, M., Zummo, G., Gerbino, A., Farina, F., & Anzalone, R. (2013). New frontiers in regenerative medicine in cardiology: The potential of Wharton's jelly mesenchymal stem cells. *Current Stem Cell Research & Therapy*, 8(1), 39–45. <https://doi.org/10.2174/1574888x11308010006>
- Davies, J. E., Walker, J. T., & Keating, A. (2017). Concise review: Wharton's Jelly: The rich, but enigmatic, source of mesenchymal stromal cells. *Stem Cells Translational Medicine*, 6(7), 1620–1630. <https://doi.org/10.1002/sctm.16-0492>
- Ding, D. C., Shyu, W. C., Chiang, M. F., Lin, S. Z., Chang, Y. C., Wang, H. J., ... Li, H. (2007). Enhancement of neuroplasticity through upregulation of beta1-integrin in human umbilical cord-derived stromal cell implanted stroke model. *Neurobiology of Disease*, 27(3), 339–353. <https://doi.org/10.1016/j.nbd.2007.06.010>
- Dominici, M., Le Blanc, K., Mueller, I., Slaper-Cortenbach, I., Marini, F., Krause, D., ... Horwitz, E. (2006). Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy*, 8(4), 315–317. <https://doi.org/10.1080/14653240600855905>
- Du, T., Cheng, J., Zhong, L., Zhao, X. F., Zhu, J., Zhu, Y. J., & Liu, G. H. (2012). The alleviation of acute and chronic kidney injury by human Wharton's jelly-derived mesenchymal stromal cells triggered by

- ischemia-reperfusion injury via an endocrine mechanism. *Cytotherapy*, 14(10), 1215–1227. <https://doi.org/10.3109/14653249.2012.711471>
- Du, T., Zou, X., Cheng, J., Wu, S., Zhong, L., Ju, G., ... Xia, S. (2013). Human Wharton's jelly-derived mesenchymal stromal cells reduce renal fibrosis through induction of native and foreign hepatocyte growth factor synthesis in injured tubular epithelial cells. *Stem Cell Research & Therapy*, 4(3), 59. <https://doi.org/10.1186/scrt215>
- El Omar, R., Beroud, J., Stoltz, J. F., Menu, P., Velot, E., & Decot, V. (2014). Umbilical cord mesenchymal stem cells: The new gold standard for mesenchymal stem cell-based therapies? *Tissue engineering. Part B, Reviews*, 20(5), 523–544. <https://doi.org/10.1089/ten.TEB.2013.0664>
- Fu, Y. S., Cheng, Y. C., Lin, M. Y., Cheng, H., Chu, P. M., Chou, S. C., ... Sung, M. S. (2006). Conversion of human umbilical cord mesenchymal stem cells in Wharton's jelly to dopaminergic neurons in vitro: Potential therapeutic application for Parkinsonism. *Stem Cells*, 24(1), 115–124. <https://doi.org/10.1634/stemcells.2005-0053>
- Gao, L. R., Chen, Y., Zhang, N. K., Yang, X. L., Liu, H. L., Wang, Z. G., ... Hu, X. (2015). Intracoronary infusion of Wharton's jelly-derived mesenchymal stem cells in acute myocardial infarction: Double-blind, randomized controlled trial. *BMC Medicine*, 13, 162. <https://doi.org/10.1186/s12916-015-0399-z>
- Hammam, O. A., Elkhaif, N., Attia, Y. M., Mansour, M. T., Elmazar, M. M., Abdelsalam, R. M., ... El-Khatib, A. S. (2016). Wharton's jelly-derived mesenchymal stem cells combined with praziquantel as a potential therapy for *Schistosoma mansoni*-induced liver fibrosis. *Scientific Reports*, 6, 21005. <https://doi.org/10.1038/srep21005>
- Hashemi, S. S., Mohammadi, A. A., Kabiri, H., Hashempoor, M. R., Mahmoodi, M., Amini, M., & Mehrabani, D. (2019). The healing effect of Wharton's jelly stem cells seeded on biological scaffold in chronic skin ulcers: A randomized clinical trial. *Journal of Cosmetic Dermatology*, 18(6), 1961–1967. <https://doi.org/10.1111/jocd.12931>
- Hassan, G., Kasem, I., Soukkaieh, C., & Aljamali, M. (2017). A simple method to isolate and expand human umbilical cord derived mesenchymal stem cells: Using explant method and umbilical cord blood serum. *International Journal of Stem Cells*, 10(2), 184–192. <https://doi.org/10.15283/ijsc17028>
- Hu, D., Zhang, D., Liu, B., Liu, Y., Zhou, Y., Yu, Y., ... Wei, G. (2020). Human ucMSCs seeded in a decellularized kidney scaffold attenuate renal fibrosis by reducing epithelial-mesenchymal transition via the TGF- β /Smad signaling pathway. *Pediatric Research*, 87(2), 1–10. <https://doi.org/10.1038/s41390-019-0736-6>
- Jamaloopour, Z., Taromi, N., Soleimani, M., Koudehi, M. F., & Asgari, A. (2019). In vitro interaction of human Wharton's jelly mesenchymal stem cells with biomimetic 3D scaffold. *Journal of Biomedical Materials Research. Part A*, 107(6), 1166–1175. <https://doi.org/10.1002/jbm.a.36608>
- Joerger-Messerli, M. S., Oppliger, B., Spinelli, M., Thomi, G., di Salvo, I., Schneider, P., & Schoeberlein, A. (2018). Extracellular vesicles derived from Wharton's jelly mesenchymal stem cells prevent and resolve programmed cell death mediated by perinatal hypoxia-ischemia in neuronal cells. *Cell Transplantation*, 27(1), 168–180. <https://doi.org/10.1177/0963689717738256>
- Kabatas, S., Civelek, E., Inci, C., Yalcinkaya, E. Y., Gunel, G., Kir, G., ... Karaoz, E. (2018). Wharton's Jelly-derived mesenchymal stem cell transplantation in a patient with hypoxic-ischemic encephalopathy: A pilot study. *Cell Transplantation*, 27(10), 1425–1433. <https://doi.org/10.1177/0963689718786692>
- Kalaszczynska, I., & Ferdyn, K. (2015). Wharton's jelly derived mesenchymal stem cells: Future of regenerative medicine? Recent findings and clinical significance. *BioMed Research International*, 2015, 430847. <https://doi.org/10.1155/2015/430847>
- Kang, B. J., Ryu, H. H., Park, S. S., Koyama, Y., Kikuchi, M., Woo, H. M., ... Kweon, O. K. (2012). Comparing the osteogenic potential of canine mesenchymal stem cells derived from adipose tissues, bone marrow, umbilical cord blood, and Wharton's jelly for treating bone defects. *Journal of Veterinary Science*, 13(3), 299–310.
- Kim, Y. E., Sung, S. I., Chang, Y. S., Ahn, S. Y., Sung, D. K., & Park, W. S. (2019). Thrombin preconditioning enhances therapeutic efficacy of human wharton's jelly-derived mesenchymal stem cells in severe neonatal hypoxic ischemic encephalopathy. *International Journal of Molecular Sciences*, 20(10), 2477. <https://doi.org/10.3390/ijms20102477>
- Kowalski, K., Kolodziejczyk, A., Sikorska, M., Plackiewicz, J., Cichosz, P., Kowalewska, M., ... Brzoska, E. (2017). Stem cells migration during skeletal muscle regeneration—The role of Sdf-1/Cxcr4 and Sdf-1/Cxcr7 axis. *Cell Adhesion & Migration*, 11(4), 384–398. <https://doi.org/10.1080/19336918.2016.1227911>
- Krupa, P., Vackova, I., Ruzicka, J., Zaviskova, K., Dubisova, J., Koci, Z., ... Jendelova, P. (2018). The effect of human mesenchymal stem cells derived from Wharton's jelly in spinal cord injury treatment is dose-dependent and can be facilitated by repeated application. *International Journal of Molecular Sciences*, 19(5), 1503. <https://doi.org/10.3390/ijms19051503>
- Kwon, S., Ki, S. M., Park, S. E., Kim, M. J., Hyung, B., Lee, N. K., ... Chang, J. W. (2016). Anti-apoptotic effects of human Wharton's jelly-derived mesenchymal stem cells on skeletal muscle cells mediated via secretion of XCL1. *Molecular Therapy*, 24(9), 1550–1560. <https://doi.org/10.1038/mt.2016.125>
- Lanci, A., Merlo, B., Mariella, J., Castagnetti, C., & Iacono, E. (2019). Heterologous Wharton's jelly derived mesenchymal stem cells application on a large chronic skin wound in a 6-month-old Filly. *Frontiers in Veterinary Science*, 6, 9. <https://doi.org/10.3389/fvets.2019.00009>
- Li, C., Chen, X., Qiao, S., Liu, X., Liu, C., Zhu, D., ... Wang, Z. (2016). Effects of Wharton's jelly cells of the human umbilical cord on acute spinal cord injury in rats, and expression of interleukin-1 β and nerve growth factor in spinal cord tissues. *Artificial Cells, Nanomedicine, and Biotechnology*, 44(5), 1254–1258. <https://doi.org/10.3109/21691401.2015.1019671>
- Li, P., Zhang, J., Liu, J., Ma, H., Liu, J., Lie, P., ... Wei, X. (2015). Promoting the recovery of injured liver with poly (3-hydroxybutyrate-co-3-hydroxyvalerate-co-3-hydroxyhexanoate) scaffolds loaded with umbilical cord-derived mesenchymal stem cells. *Tissue engineering. Part A*, 21(3–4), 603–615. <https://doi.org/10.1089/ten.TEA.2013.0331>
- Liau, L. L., Ruszymah, B. H. I., Ng, M. H., & Law, J. X. (2019). Characteristics and clinical applications of Wharton's jelly-derived mesenchymal stromal cells. *Current Research in Translational Medicine*, 68, 5–16. <https://doi.org/10.1016/j.retram.2019.09.001>
- Lim, J., Razi, Z. R. M., Law, J. X., Nawi, A. M., Idrus, R. B. H., Chin, T. G., ... Ng, M. H. (2018). Mesenchymal stromal cells from the maternal segment of human umbilical cord is ideal for bone regeneration in allogenic setting. *Journal of Tissue Engineering and Regenerative Medicine*, 15(1), 75–87. <https://doi.org/10.1007/s13770-017-0086-6>
- Lin, S. Z., Chang, Y. J., Liu, J. W., Chang, L. F., Sun, L. Y., Li, Y. S., ... Chiou, T. W. (2010). Transplantation of human Wharton's jelly-derived stem cells alleviates chemically induced liver fibrosis in rats. *Cell Transplantation*, 19(11), 1451–1463. <https://doi.org/10.3727/096368910x514198>
- Liu, S., Hou, K. D., Yuan, M., Peng, J., Zhang, L., Sui, X., ... Guo, Q. (2014). Characteristics of mesenchymal stem cells derived from Wharton's jelly of human umbilical cord and for fabrication of non-scaffold tissue-engineered cartilage. *Journal of Bioscience and Bioengineering*, 117(2), 229–235. <https://doi.org/10.1016/j.jbiosc.2013.07.001>
- Liu, C. B., Huang, H., Sun, P., Ma, S. Z., Liu, A. H., Xue, J., ... Zhang, X. Z. (2016). Human umbilical cord-derived mesenchymal stromal cells improve left ventricular function, perfusion, and remodeling in a porcine model of chronic myocardial ischemia. *Stem Cells Translational Medicine*, 5(8), 1004–1013. <https://doi.org/10.5966/sctm.2015-0298>
- Lopez, Y., Lutjemeier, B., Seshareddy, K., Trevino, E. M., Hageman, K. S., Musch, T. I., ... Weiss, M. L. (2013). Wharton's jelly or bone marrow mesenchymal stromal cells improve cardiac function following myocardial infarction for more than 32 weeks in a rat model: A preliminary report. *Current Stem Cell Research & Therapy*, 8(1), 46–59.

- Ma, L., Feng, X. Y., Cui, B. L., Law, F., Jiang, X. W., Yang, L. Y., ... Huang, T. H. (2005). Human umbilical cord Wharton's jelly-derived mesenchymal stem cells differentiation into nerve-like cells. *Chinese Medical Journal*, 118(23), 1987–1993.
- Maldonado, M., Huang, T., Yang, L., Xu, L., & Ma, L. (2017). Human umbilical cord Wharton jelly cells promote extra-pancreatic insulin formation and repair of renal damage in STZ-induced diabetic mice. *Cell Communication and Signaling*, 15(1), 43. <https://doi.org/10.1186/s12964-017-0199-5>
- Marino, L., Castaldi, M. A., Rosamilio, R., Ragni, E., Vitolo, R., Fulgione, C., ... Sella, C. (2019). Mesenchymal stem cells from the Wharton's jelly of the human umbilical cord: Biological properties and therapeutic potential. *International Journal of Stem Cells*, 12(2), 218–226. <https://doi.org/10.15283/ijsc18034>
- Martin-Piedra, M. A., Alfonso-Rodriguez, C. A., Zapater, A., Durand-Herrera, D., Chato-Astrain, J., Campos, F., ... Garzon, I. (2019). Effective use of mesenchymal stem cells in human skin substitutes generated by tissue engineering. *European Cells & Materials*, 37, 233–249. <https://doi.org/10.22203/eCM.v037a14>
- Martin-Rendon, E., Sweeney, D., Lu, F., Girdlestone, J., Navarrete, C., & Watt, S. M. (2008). 5-Azacytidine-treated human mesenchymal stem/progenitor cells derived from umbilical cord, cord blood and bone marrow do not generate cardiomyocytes in vitro at high frequencies. *Vox Sanguinis*, 95(2), 137–148. <https://doi.org/10.1111/j.1423-0410.2008.01076.x>
- Mazini, L., Rochette, L., Amine, M., & Malka, G. (2019). Regenerative capacity of adipose derived stem cells (ADSCs), comparison with mesenchymal stem cells (MSCs). *International Journal of Molecular Sciences*, 20(10), 2523. <https://doi.org/10.3390/ijms20102523>
- McElreavey, K. D., Irvine, A. I., Ennis, K. T., & McLean, W. H. (1991). Isolation, culture and characterisation of fibroblast-like cells derived from the Wharton's jelly portion of human umbilical cord. *Biochemical Society Transactions*, 19(1), 29s. <https://doi.org/10.1042/bst019029s>
- Mesure, B., Huber-Villaume, S., Menu, P., & Velot, E. (2017). Transforming growth factor-beta 1 or ascorbic acid are able to differentiate Wharton's jelly mesenchymal stem cells towards a smooth muscle phenotype. *Biomedical Materials and Engineering*, 28(s1), S101–S105. <https://doi.org/10.3233/bme-171630>
- Millan-Rivero, J. E., Martinez, C. M., Romecin, P. A., Aznar-Cervantes, S. D., Carpes-Ruiz, M., Cenis, J. L., ... Garcia-Bernal, D. (2019). Silk fibroin scaffolds seeded with Wharton's jelly mesenchymal stem cells enhance re-epithelialization and reduce formation of scar tissue after cutaneous wound healing. *Stem Cell Research & Therapy*, 10(1), 126. <https://doi.org/10.1186/s13287-019-1229-6>
- Mohamadi, Y., Mousavi, M., Moogahi, S. M. H. N., Abolhassani, F., Ijaz, S., & Hassanzadeh, G. (2018). Effect of Wharton's Jelly derived mesenchymal stem cells on the expression of NLRP3 receptor and neuroinflammation in experimental spinal cord injury. *Journal of Clinical and Diagnostic Research*, 12(10), AF01–AF04. <https://doi.org/10.7860/JCDR/2018/37321.12168>
- Mohamadi, Y., Noori Moghahi, S. M. H., Mousavi, M., Borhani-Haghighi, M., Abolhassani, F., Kashani, I. R., & Hassanzadeh, G. (2019). Intrathecal transplantation of Wharton's jelly mesenchymal stem cells suppresses the NLRP1 inflammasome in the rat model of spinal cord injury. *Journal of Chemical Neuroanatomy*, 97, 1–8. <https://doi.org/10.1016/j.jchemneu.2019.01.011>
- Mortezaee, K., Minai, B., Sabbaghziarani, F., Ragerdi Kashani, I., Hassanzadeh, G., Pasbakhsh, P., ... Latifpour, M. (2015). Retinoic acid as the stimulating factor for differentiation of Wharton's Jelly-mesenchymal stem cells into hepatocyte-like cells. *Avicenna Journal of Medical Biotechnology*, 7(3), 106–112.
- Musialek, P., Mazurek, A., Jarocha, D., Tekieli, L., Szot, W., Kostkiewicz, M., ... Majka, M. (2015). Myocardial regeneration strategy using Wharton's jelly mesenchymal stem cells as an off-the-shelf 'unlimited' therapeutic agent: Results from the Acute Myocardial Infarction First-in-Man Study. *Advances in Interventional Cardiology*, 11(2), 100–107. <https://doi.org/10.5114/pwki.2015.52282>
- Nancarrow-Lei, R., Mafi, P., Mafi, R., & Khan, W. (2017). A systemic review of adult mesenchymal stem cell sources and their multilineage differentiation potential relevant to musculoskeletal tissue repair and regeneration. *Current Stem Cell Research & Therapy*, 12(8), 601–610. <https://doi.org/10.2174/1574888x12666170608124303>
- Nimsanor, N., Phetfong, J., Kitiyanant, N., Kamprom, W., & Supokawej, A. (2019). Overexpression of anti-fibrotic factors ameliorates anti-fibrotic properties of Wharton's jelly derived mesenchymal stem cells under oxidative damage. *Bioscience trends*, 13(5), 411–422. <https://doi.org/10.5582/bst.2019.01191>
- Paduszynski, P., Aleksander-Konert, E., Zajdel, A., Wilczok, A., Jelonek, K., Witek, A., & Dzierzewicz, Z. (2016). Changes in expression of cartilaginous genes during chondrogenesis of Wharton's jelly mesenchymal stem cells on three-dimensional biodegradable poly (L-lactide-co-glycolide) scaffolds. *Cellular and Molecular Biology Letters*, 21, 14. <https://doi.org/10.1186/s11658-016-0012-2>
- Park, A., Park, H., Yoon, J., Kang, D., Kang, M. H., Park, Y. Y., ... Yu, J. (2019). Priming with Toll-like receptor 3 agonist or interferon-gamma enhances the therapeutic effects of human mesenchymal stem cells in a murine model of atopic dermatitis. *Stem Cell Research & Therapy*, 10(1), 66. <https://doi.org/10.1186/s13287-019-1164-6>
- Peng, J., Wang, Y., Zhang, L., Zhao, B., Zhao, Z., Chen, J., ... Lu, S. (2011). Human umbilical cord Wharton's jelly-derived mesenchymal stem cells differentiate into a Schwann-cell phenotype and promote neurite outgrowth in vitro. *Brain Research Bulletin*, 84(3), 235–243. <https://doi.org/10.1016/j.brainresbull.2010.12.013>
- Pu, L., Meng, M., Wu, J., Zhang, J., Hou, Z., Gao, H., ... Li, Y. (2017). Compared to the amniotic membrane, Wharton's jelly may be a more suitable source of mesenchymal stem cells for cardiovascular tissue engineering and clinical regeneration. *Stem Cell Research & Therapy*, 8(1), 72. <https://doi.org/10.1186/s13287-017-0501-x>
- Rabbani, S., Soleimani, M., Sahebjam, M., Imani, M., Haeri, A., Ghaseddin, A., ... Ahmadi Tafti, S. H. (2018). Simultaneous delivery of Wharton's jelly mesenchymal stem cells and insulin-like growth factor-1 in acute myocardial infarction. *Iranian Journal of Pharmaceutical Research: IJPR*, 17(2), 426–441.
- Ramanathan, R., Rupert, S., Selvaraj, S., Satyanesan, J., Vennila, R., & Rajagopal, S. (2017). Role of human Wharton's jelly derived mesenchymal stem cells (WJ-MSCs) for rescue of d-galactosamine induced acute liver injury in mice. *Journal of Clinical and Experimental Hepatology*, 7(3), 205–214. <https://doi.org/10.1016/j.jceh.2017.03.010>
- Rengasamy, M., Singh, G., Fakharuzi, N. A., Siddikuzzaman, Balasubramanian, S., Swamynathan, P., ... Majumdar, A. S. (2017). Transplantation of human bone marrow mesenchymal stromal cells reduces liver fibrosis more effectively than Wharton's jelly mesenchymal stromal cells. *Stem Cell Research & Therapy*, 8(1), 143. <https://doi.org/10.1186/s13287-017-0595-1>
- Rezaeian, L., Hosseini, S. E., Dianatpur, M., Edalatmanesh, M. A., Tanideh, N., Mogheiseh, A., & Tamadon, A. (2018). Intrauterine xenotransplantation of human Wharton jelly-derived mesenchymal stem cells into the liver of rabbit fetuses: A preliminary study for in vivo expression of the human liver genes. *Iranian Journal of Basic Medical Sciences*, 21(1), 89–96. <https://doi.org/10.22038/ijbms.2017.24501.6098>
- Ribeiro, J., Pereira, T., Amorim, I., Caseiro, A. R., Lopes, M. A., Lima, J., ... Luís, A. L. (2014). Cell therapy with human MSCs isolated from the umbilical cord wharton jelly associated to a PVA membrane in the treatment of chronic skin wounds. *International Journal of Medical Sciences*, 11(10), 979–987. <https://doi.org/10.7150/ijms.9139>
- Ryu, H. H., Kang, B. J., Park, S. S., Kim, Y., Sung, G. J., Woo, H. M., ... Kweon, O. K. (2012). Comparison of mesenchymal stem cells derived from fat, bone marrow, Wharton's jelly, and umbilical cord blood for

- treating spinal cord injuries in dogs. *Journal of Veterinary Medical Science*, 74(12), 1617–1630.
- Sabapathy, V., Sundaram, B., V. M. S., Mankuzhy, P., & Kumar, S. (2014). Human Wharton's Jelly Mesenchymal Stem Cells plasticity augments scar-free skin wound healing with hair growth. *PLoS One*, 9(4):e93726. <https://doi.org/10.1371/journal.pone.0093726>
- Sadlik, B., Jaroslowski, G., Gladysz, D., Puszkarz, M., Markowska, M., Pawelec, K., ... Oldak, T. (2017). Knee cartilage regeneration with umbilical cord mesenchymal stem cells embedded in collagen scaffold using dry arthroscopy technique. *Advances in Experimental Medicine and Biology*, 1020, 113–122. https://doi.org/10.1007/5584_2017_9
- Sadlik, B., Jaroslowski, G., Puszkarz, M., Blasiak, A., Oldak, T., Gladysz, D., & Whyte, G. P. (2018). Cartilage repair in the knee using umbilical cord Wharton's jelly-derived mesenchymal stem cells embedded onto collagen scaffolding and implanted under dry arthroscopy. *Arthroscopy Techniques*, 7(1), e57–e63. <https://doi.org/10.1016/j.jeats.2017.08.055>
- Saulnier, N., Viguier, E., Perrier-Groult, E., Chenu, C., Pillet, E., Roger, T., ... Boulocher, C. (2015). Intra-articular administration of xenogeneic neonatal mesenchymal stromal cells early after meniscal injury down-regulates metalloproteinase gene expression in synovium and prevents cartilage degradation in a rabbit model of osteoarthritis. *Osteoarthritis Cartilage*, 23(1), 122–133. <https://doi.org/10.1016/j.joca.2014.09.007>
- Sepulveda, H., Aguilar, R., Prieto, C. P., Bustos, F., Aedo, S., Lattus, J., ... Montecino, M. (2017). Epigenetic signatures at the RUNX2-P1 and Sp7 gene promoters control osteogenic lineage commitment of umbilical cord-derived mesenchymal stem cells. *Journal of Cellular Physiology*, 232(9), 2519–2527. <https://doi.org/10.1002/jcp.25627>
- Shi, S., Jia, S., Liu, J., & Chen, G. (2015). Accelerated regeneration of skin injury by co-transplantation of mesenchymal stem cells from Wharton's jelly of the human umbilical cord mixed with microparticles. *Cell Biochemistry and Biophysics*, 71(2), 951–956. <https://doi.org/10.1007/s12013-014-0292-y>
- Shi, M., Zhang, Z., Xu, R., Lin, H., Fu, J., Zou, Z., ... Wang, F.-S. (2012). Human mesenchymal stem cell transfusion is safe and improves liver function in acute-on-chronic liver failure patients. *Stem Cells Translational Medicine*, 1, 725–731. <https://doi.org/10.5966/sctm.2012-0034>
- Shi, X., Zhou, K., Huang, F., Zhang, J., & Wang, C. (2018). Endocytic mechanisms and osteoinductive profile of hydroxyapatite nanoparticles in human umbilical cord Wharton's jelly-derived mesenchymal stem cells. *International Journal of Nanomedicine*, 13, 1457–1470. <https://doi.org/10.2147/ijn.S155814>
- Shie, M. Y., Chang, W. C., Wei, L. J., Huang, Y. H., Chen, C. H., Shih, C. T., ... Shen, Y. F. (2017). 3D printing of cytocompatible water-based light-cured polyurethane with hyaluronic acid for cartilage tissue engineering applications. *Materials (Basel)*, 10(2), 136. <https://doi.org/10.3390/ma10020136>
- Su, W. H., Wang, C. J., Fu, H. C., Sheng, C. M., Tsai, C. C., Cheng, J. H., & Chuang, P. C. (2019). Human umbilical cord mesenchymal stem cells extricate bupivacaine-impaired skeletal muscle function via mitigating neutrophil-mediated acute inflammation and protecting against fibrosis. *International Journal of Molecular Sciences*, 20(17), 4312. <https://doi.org/10.3390/ijms20174312>
- Sun, J., Zhang, Y., Song, X., Zhu, J., & Zhu, Q. (2019). The healing effects of conditioned medium derived from mesenchymal stem cells on radiation-induced skin wounds in rats. *Cell Transplantation*, 28(1), 105–115. <https://doi.org/10.1177/0963689718807410>
- Todeschi, M. R., El Backly, R., Capelli, C., Daga, A., Patrone, E., Introna, M., ... Mastrogiacomio, M. (2015). Transplanted umbilical cord mesenchymal stem cells modify the in vivo microenvironment enhancing angiogenesis and leading to bone regeneration. *Stem Cells and Development*, 24(13), 1570–1581. <https://doi.org/10.1089/scd.2014.0490>
- Tsai, P. C., Fu, T. W., Chen, Y. M., Ko, T. L., Chen, T. H., Shih, Y. H., ... Fu, Y. S. (2009). The therapeutic potential of human umbilical mesenchymal stem cells from Wharton's jelly in the treatment of rat liver fibrosis. *Liver Transplantation*, 15(5), 484–495. <https://doi.org/10.1002/lt.21715>
- Vieira Paladino, F., de Moraes Rodrigues, J., da Silva, A., & Goldberg, A. C. (2019). The immunomodulatory potential of Wharton's jelly mesenchymal stem/stromal cells. *Stem Cells International*, 2019, 3548917. <https://doi.org/10.1155/2019/3548917>
- Vojdani, Z., Khodabandeh, Z., Jaberipour, M., Hosseini, A., Bahmanpour, S., & Talei-Khozani, T. (2015). The influence of fibroblast growth factor 4 on hepatogenic capacity of Wharton's jelly mesenchymal stromal cells. *Romanian Journal of Morphology and Embryology*, 56(3), 1043–1050.
- Wang, Q. Q., Jing, X. M., Bi, Y. Z., Cao, X. F., Wang, Y. Z., Li, Y. X., ... Hu, J. (2018a). Human umbilical cord Wharton's jelly derived mesenchymal stromal cells may attenuate sarcopenia in aged mice induced by hindlimb suspension. *Medical Science Monitor*, 24, 9272–9281. <https://doi.org/10.12659/msm.913362>
- Wang, A. Y. L., Loh, C. Y. Y., Shen, H. H., Hsieh, S. Y., Wang, I. K., Chuang, S. H., & Wei, F. C. (2019). Topical Application of human Wharton's jelly mesenchymal stem cells accelerates mouse sciatic nerve recovery and is associated with upregulated neurotrophic factor expression. *Cell Transplantation*, 28(12), 1560–1572. <https://doi.org/10.1177/0963689719880543>
- Wang, L., Tran, I., Seshareddy, K., Weiss, M. L., & Detamore, M. S. (2009). A comparison of human bone marrow-derived mesenchymal stem cells and human umbilical cord-derived mesenchymal stromal cells for cartilage tissue engineering. *Tissue engineering. Part A*, 15(8), 2259–2266. <https://doi.org/10.1089/ten.tea.2008.0393>
- Wang, M., Yuan, Q., & Xie, L. (2018b). Mesenchymal stem cell-based immunomodulation: Properties and clinical application. *Stem Cells International*, 2018, 3057624. <https://doi.org/10.1155/2018/3057624>
- Weiss, M. L., Anderson, C., Medicetty, S., Seshareddy, K. B., Weiss, R. J., VanderWerff, I., ... McIntosh, K. R. (2008). Immune properties of human umbilical cord Wharton's jelly-derived cells. *Stem Cells*, 26(11), 2865–2874. <https://doi.org/10.1634/stemcells.2007-1028>
- Weiss, M. L., Medicetty, S., Bledsoe, A. R., Rachakatla, R. S., Choi, M., Merchav, S., ... Troyer, D. (2006). Human umbilical cord matrix stem cells: Preliminary characterization and effect of transplantation in a rodent model of Parkinson's disease. *Stem Cells*, 24(3), 781–792. <https://doi.org/10.1634/stemcells.2005-0330>
- Widowati, W., Gunanegara, R. F., Rizal, R., Widodo, W. S., Amalia, A., Wibowo, S. H. B., ... Chiuman, L. (2019). Comparative analysis of Wharton's jelly mesenchymal stem cell (WJ-MSCs) isolated using explant and enzymatic methods. *Journal of Physics: Conference Series*, 1374, 012024. <https://doi.org/10.1088/1742-6596/1374/1/012024>
- Wu, X., Yan, T., Wang, Z., Wu, X., Cao, G., Zhang, C., ... Wang, J. (2018a). Micro-vesicles derived from human Wharton's Jelly mesenchymal stromal cells mitigate renal ischemia-reperfusion injury in rats after cardiac death renal transplantation. *Journal of Cellular Biochemistry*, 119(2), 1879–1888. <https://doi.org/10.1002/jcb.26348>
- Wu, K. J., Yu, S. J., Chiang, C. W., Lee, Y. W., Yen, B. L., Tseng, P. C., ... Wang, Y. (2018b). Neuroprotective action of human Wharton's jelly-derived mesenchymal stromal cell transplants in a rodent model of stroke. *Cell Transplantation*, 27(11), 1603–1612. <https://doi.org/10.1177/0963689718802754>
- Yang, C. C., Shih, Y. H., Ko, M. H., Hsu, S. Y., Cheng, H., & Fu, Y. S. (2008). Transplantation of human umbilical mesenchymal stem cells from Wharton's jelly after complete transection of the rat spinal cord. *PLoS One*, 3(10):e3336. <https://doi.org/10.1371/journal.pone.0003336>
- Zhang, S., Chen, L., Liu, T., Zhang, B., Xiang, D., Wang, Z., & Wang, Y. (2012a). Human umbilical cord matrix stem cells efficiently rescue acute liver failure through paracrine effects rather than hepatic differentiation. *Tissue engineering. Part A*, 18(13–14), 1352–1364. <https://doi.org/10.1089/ten.TEA.2011.0516>
- Zhang, Y., Hao, H., Liu, J., Fu, X., & Han, W. (2012b). Repair and regeneration of skin injury by transplanting microparticles mixed with Wharton's jelly and MSCs from the human umbilical cord. *The*

- International Journal of Lower Extremity Wounds*, 11(4), 264–270. <https://doi.org/10.1177/1534734612463577>
- Zhang, J., La, X., Fan, L., Li, P., Yu, Y., Huang, Y., ... Xing, Y. (2015). Immunosuppressive effects of mesenchymal stem cell transplantation in rat burn models. *International Journal of Clinical and Experimental Pathology*, 8(5), 5129–5136. Retrieved from. <http://www.embase.com/search/results?subaction=viewrecord&from=export&id=L615600994>
- Zhang, Y. N., Lie, P. C., & Wei, X. (2009). Differentiation of mesenchymal stromal cells derived from umbilical cord Wharton's jelly into hepatocyte-like cells. *Cytotherapy*, 11(5), 548–558. <https://doi.org/10.1080/14653240903051533>
- Zhang, Z., Lin, H., Shi, M., Xu, R., Fu, J., Lv, J., ... Wang, F. S. (2012c). Human umbilical cord mesenchymal stem cells improve liver function and ascites in decompensated liver cirrhosis patients. *Journal of Gastroenterology and Hepatology*, 27(Suppl 2), 112–120. <https://doi.org/10.1111/j.1440-1746.2011.07024.x>
- Zhang, Y., Liu, S., Guo, W., Hao, C., Wang, M., Li, X., ... Guo, Q. (2019). Coculture of hWJMSCs and pACs in oriented scaffold enhances hyaline cartilage regeneration in vitro. *Stem Cells International*, 2019, 5130152. <https://doi.org/10.1155/2019/5130152>. 5130152–11.
- Zhang, Y., Liu, S., Guo, W., Wang, M., Hao, C., Gao, S., ... Guo, Q. (2018a). Human umbilical cord Wharton's jelly mesenchymal stem cells combined with an acellular cartilage extracellular matrix scaffold improve cartilage repair compared with microfracture in a caprine model. *Osteoarthritis and Cartilage*, 26(7), 954–965. <https://doi.org/10.1016/j.joca.2018.01.019>
- Zhang, W., Liu, X. C., Yang, L., Zhu, D. L., Zhang, Y. D., Chen, Y., & Zhang, H. Y. (2013). Wharton's jelly-derived mesenchymal stem cells promote myocardial regeneration and cardiac repair after miniswine acute myocardial infarction. *Coronary Artery Diseases*, 24(7), 549–558. <https://doi.org/10.1097/MCA.0b013e3283640f00>
- Zhang, L., Wang, L. M., Chen, W. W., Ma, Z., Han, X., Liu, C. M., ... Zhang, X. H. (2017). Neural differentiation of human Wharton's jelly-derived mesenchymal stem cells improves the recovery of neurological function after transplantation in ischemic stroke rats. *Neural Regeneration Research*, 12(7), 1103–1110. <https://doi.org/10.4103/1673-5374.211189>
- Zhang, X., Zhang, Q., Li, W., Nie, D., Chen, W., Xu, C., ... Tu, W. (2014a). Therapeutic effect of human umbilical cord mesenchymal stem cells on neonatal rat hypoxic-ischemic encephalopathy. *Journal of Neuroscience Research*, 92(1), 35–45. <https://doi.org/10.1002/jnr.23304>
- Zhang, C., Zhou, G., Chen, Y., Liu, S., Chen, F., Xie, L., ... Ma, L. (2018b). Human umbilical cord mesenchymal stem cells alleviate interstitial fibrosis and cardiac dysfunction in a dilated cardiomyopathy rat model by inhibiting TNFalpha and TGFbeta1/ERK1/2 signaling pathways. *Molecular Medicine Reports*, 17(1), 71–78. <https://doi.org/10.3892/mmr.2017.7882>
- Zhang, G., Zou, X., Huang, Y., Wang, F., Miao, S., Liu, G., ... Zhu, Y. (2016). Mesenchymal stromal cell-derived extracellular vesicles protect against acute kidney injury through anti-oxidation by enhancing Nrf2/ARE activation in rats. *Kidney and Blood Pressure Research*, 41(2), 119–128. <https://doi.org/10.1159/000443413>
- Zhang, G., Zou, X., Miao, S., Chen, J., Du, T., Zhong, L., ... Zhu, Y. (2014b). The anti-oxidative role of micro-vesicles derived from human wharton-jelly mesenchymal stromal cells through NOX2/gp91(phox) suppression in alleviating renal ischemia-reperfusion injury in rats. *PLoS One*, 9(3): e92129. <https://doi.org/10.1371/journal.pone.0092129>
- Zhou, C., Yang, B., Tian, Y., Jiao, H., Zheng, W., Wang, J., & Guan, F. (2011). Immunomodulatory effect of human umbilical cord Wharton's jelly-derived mesenchymal stem cells on lymphocytes. *Cellular Immunology*, 272(1), 33–38. <https://doi.org/10.1016/j.cellimm.2011.09.010>
- Zou, X., Zhang, G., Cheng, Z., Yin, D., Du, T., Ju, G., ... Zhu, Y. (2014). Microvesicles derived from human Wharton's Jelly mesenchymal stromal cells ameliorate renal ischemia-reperfusion injury in rats by suppressing CX3CL1. *Stem Cell Research & Therapy*, 5(2), 40. <https://doi.org/10.1186/scrt428>

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