# **Development of a Diffusive Gradients in Thin-Films Passive Sampling Device for PFAS (ER20-1363)** Samuel D. Hodges, Huong T. Pham, and Julian L. Fairey (University of Arkansas)

# **Introduction:**

Diffusive gradients in thin-films (DGT) passive sampling devices (PSDs) are kinetic samplers in which analytes sorb in a binding layer following diffusion through (1) a stagnant diffusive boundary layer (DBL), the thickness of which,  $\delta_{\text{DBL}}$ , varies with local hydrodynamics, and (2) a hydrogel of known thickness,  $\delta_{\text{Gel}}$ . The hydrogel restricts mass transport to molecular diffusion and, therefore, diffusion coefficients in hydrogels, D<sub>ael</sub>, are needed to determine time-weighted average PFAS concentrations. Here, a two-compartment diffusion cell (2Comp-DCell) was used to measure PFAS concentration profiles across hydrogels as a function of pH and temperature. A finitedifference model (FDM) was used to determine  $D_{PFAS-Gel}$ , informed by 2Comp-DCell tests with nitrate in which  $\delta_{DBL}$ was determined as a function of mixing speed in the Source and Sink compartments.

# **<u>Finite-Difference Model for the Two-Compartment Diffusion Cell:</u>**

D<sub>qel</sub> determinations from 2Comp-DCell tests typically rely on four assumptions: (I) Analyte diffusion in the gel is indistinguishable from that in the water,  $D_{qel} = D_{water}$ , (II) the source compartment concentration is constant over the experiment duration, C<sub>source</sub> = constant, (III) the sink compartment concentration increases linearly in time and its maximum is negligible compared to the source concentration,  $C_{Max-Sink} << C_{Source}$ , and (IV) the  $\delta_{DBL}$ is negligible, such that  $\delta_{\text{DBL}} << \delta_{\text{Gel}}$ .

At environmentally relevant PFAS concentrations, 2Comp-DCell experiments span 3–5 days and violate assumptions II and III. The FDM developed in Task 1 eliminates the need for these assumptions through implementation of a stepwise regression procedure. The handling of assumptions I and IV is described next.



# **Diffusion Boundary Layer Thickness:**

Assumption I is only necessary if  $\delta_{\text{DBL}} \not \ll \delta_{\text{Gel}}$ . If assumption IV is true,  $\delta_{\text{DBL}}$  can be ignored and both assumptions are unnecessary. Assumption IV is dependent on the hydrodynamic properties in the diffusion cell. To investigate the 2Comp-DCell hydrodynamics, tests with nitrate were completed as a function of  $\delta_{Gel}$  (0.08, 0.12, 0.16, 0.20) cm), gel type (agarose and agarose cross-linked polyacrylamide (APA)), and stir rate (100, 200, and 500 RPM). This array of 60 tests was analyzed with the FDM to determine  $\delta_{DBI}$  in the 2Comp-DCell.



### Figure 3. (above)

Sink compartment nitrate concentration profiles from 2Comp-DCell tests with agarose gel. The points are measured nitrate concentrations and lines are FDM-simulated profiles.



Figure 2. (above) Digital rendering of a 2Comp-DCell Compartment

 $> D_{Gel-APA} < D_{Gel-agarose} \rightarrow$  used agarose gel for PFAS  $> D_{Gel-APA} = 14.0 \times 10^{-6} \text{ cm}^2 \cdot \text{s}^{-1}$  $> D_{Gel-agarose} = 15.7 \times 10^{-6} \text{ cm}^2 \cdot \text{s}^{-1}$  $\succ$  At 100 RPM,  $\delta_{DBL} = 80 \ \mu m$ , total DBL distance = 160  $\mu m$  $\succ$  At 200 RPM,  $\delta_{DBL} = 50 \ \mu m$ , total DBL distance = 100  $\mu m$ > At 500 RPM,  $\delta_{\text{DBL}} \approx 0$ , used for PFAS 2Comp-DCell tests



### Figure 4. (above)

Source and sink compartment PFAS concentration profiles from 2Comp-DCell tests. The points are measured PFAS concentrations and lines are FDM simulated profiles. Tests were completed at room temperature (22 °C) and pH 7 with a gel thickness,  $\delta_{Gel}$ , of 0.12 cm, and compartment mixing speed of 500 RPM.



### Figure 5. (above)

Diffusion coefficients in agarose gel (D<sub>PFAS-Gel</sub>) for the (a) C4–C10 PFCAs, (b) C4–C8 PFSAs, and (c) 4:2- and 6:2 FTSs. The source compartment was spiked with a mixture of the 24 target PFAS at 10,000 ng•L<sup>-1</sup>. Three experiments (black circles) were completed at pH 7.0 and room temperature (22 °C) and one experiment was completed at (i) pH 7.0 and 5 °C (blue circles), (ii) pH 5.0 and 22 °C (magenta circles), and (iii) pH 9.0 and 22 °C (green circles). Error bars are the weighted residual sum of squares (WRSS) between the model fit and experimental data. The Stokes-Einstein equation was used to predict D<sub>PFAS-Gel</sub> at 5 °C (blue crosses) based on their respective values at 22 °C and pH 7. D<sub>PFAS-Gel</sub> from Fang et al. 2021<sup>1</sup> at pH 5.6, room temperature, and source compartment PFAS spiked at 2,500,000 ng•L<sup>-1</sup> (magenta triangles).

- > D<sub>PFAS-Gel</sub> decreased with increasing carbon chain length for the C4–C10 PFCAs and C4–C8 PFSAs
- > pH 5 (n=1)  $\approx$  pH 7 (n=3) < pH 9 (n=1) → negative charge on Agarose may retard PFAS diffusion
- $\succ$  Dropping temperature from 22 to 5 °C decreased D<sub>PEAS-Gel</sub> by about one-half  $\succ$  Estimates of D<sub>PFAS-Gel</sub> by Stokes-Einstein equation overpredicted D<sub>PFAS-Gel</sub> by 15–30 %
  - > Decreased temperature may produce additional restrictions of gel pore spaces

# **Future Work:**

- 2Comp-DCell tests with C11–C14 PFCAs, C9–C11 PFSAs, 8:2 FTS, PFOSA, N-MeFOSAA, and N-EtFOSAA
- 2Comp-DCell tests with nitrate to assess impact of pH and conductivity on mass transport through Agarose
- DGT-PSD binding layer selection (strong base anion and/or weak-base anion resins)
- Laboratory-scale deployments of DGT-PSDs to assess accuracy for PFAS mixtures and determine method detection limits vs. deployment time for the 24 target PFAS

# **Acknowledgements:**

The authors thank David G. Wahman (USEPA, Cincinnati, Ohio) who is a technical advisor on ER20-1363 and Levi M. Haupert (USEPA, Cincinnati, Ohio) for help with the numerical modeling.

**Reference**: <sup>1</sup> Fang et al., *Environ. Sci. Technol.*, 2021, 55, 14, 9548-9556 | **Contact**: Julian Fairey, julianf@uark.edu