







ORIGINAL RESEARCH

Circulating Ectonucleotidases Signal Impaired Myocardial Perfusion at Rest and Stress

Rachel G. Kroll , BS; Corey Powell , PhD; Jun Chen, MS; Natasha T. Snider, PhD; Cynthia St. Hilaire , PhD; Akshay Reddy , BS; Judy Kim, BS; David J. Pinsky, MD; Venkatesh L. Murthy , MD, PhD; Nadia R. Sutton , MD, MPH

BACKGROUND: Ectonucleotidases maintain vascular homeostasis by metabolizing extracellular nucleotides, modulating inflammation and thrombosis, and potentially, myocardial flow through adenosine generation. Evidence implicates dysfunction or deficiency of ectonucleotidases CD39 or CD73 in human disease; the utility of measuring levels of circulating ectonucleotidases as plasma biomarkers of coronary artery dysfunction or disease has not been previously reported.

METHODS AND RESULTS: A total of 529 individuals undergoing clinically indicated positron emission tomography stress testing between 2015 and 2019 were enrolled in this single-center retrospective analysis. Baseline demographics, clinical data, nuclear stress test, and coronary artery calcium score variables were collected, as well as a blood sample. CD39 and CD73 levels were assessed as binary (detectable, undetectable) or continuous variables using ELISAs. Plasma CD39 was detectable in 24% of White and 8% of Black study participants ($P=0.02$). Of the clinical history variables examined, ectonucleotidase levels were most strongly associated with underlying liver disease and not other traditional coronary artery disease risk factors. Intriguingly, detection of circulating ectonucleotidase was inversely associated with stress myocardial blood flow (2.3 ± 0.8 mL/min per g versus 2.7 mL/min per g ± 1.1 for detectable versus undetectable CD39 levels, $P<0.001$) and global myocardial flow reserve (Pearson correlation between myocardial flow reserve and $\log(\text{CD73}) -0.19$, $P<0.001$). A subanalysis showed these differences held true independent of liver disease.

CONCLUSIONS: Vasodilatory adenosine is the expected product of local ectonucleotidase activity, yet these data support an inverse relationship between plasma ectonucleotidases, stress myocardial blood flow (CD39), and myocardial flow reserve (CD73). These findings support the conclusion that plasma levels of ectonucleotidases, which may be shed from the endothelial surface, contribute to reduced stress myocardial blood flow and myocardial flow reserve.

Key Words: adenosine ■ coronary artery calcium ■ ectonucleotidase ■ myocardial flow reserve ■ positron emission tomography

Ectonucleotidases are a family of enzymes that are implicated in the suppression of thrombosis and inflammation.¹ As tethered transmembrane proteins on endothelium and leukocytes, ectonucleotidases are expressed on and potentially shed from the plasma-lemma, with externally oriented active sites that phosphohydrolyze the terminal phosphate(s) of extracellular

nucleotides.^{2,3} Ectonucleotidases sequentially degrade nucleotides (ATP, ADP, and AMP), which are released by injured and dying cells, and also act extracellularly as signaling molecules (Figure 1).⁴ The end product of the cascade of ectonucleotidase activity is adenosine, which has been shown to have anti-inflammatory and vasorelaxant properties (primarily via the adenosine A_{2A} receptor).⁵

Correspondence to: Nadia R. Sutton, MD, MPH, Vanderbilt University Medical Center, Preston Research Building 360, 2220 Pierce Ave, Nashville, TN 37232. Email: nadia.sutton@vumc.org

This manuscript was sent to Erik B. Schelbert, MD, MS, Associate Editor, for review by expert referees, editorial decision, and final disposition.

Supplemental Material is available at <https://www.ahajournals.org/doi/suppl/10.1161/JAHA.122.027920>

For Sources of Funding and Disclosures, see page 11.

© 2023 The Authors. Published on behalf of the American Heart Association, Inc., by Wiley. This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial](https://creativecommons.org/licenses/by-nc/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

JAHA is available at: www.ahajournals.org/journal/jaha

CLINICAL PERSPECTIVE

What Is New?

- Ectonucleotidases CD39 and CD73 are responsible for extracellular metabolism of ATP and ADP to adenosine, and in this study, circulating ectonucleotidases were inversely correlated with stress myocardial blood flow and myocardial flow reserve.
- Patients with detectable CD39 and elevated CD73 were less likely to have increases in stress myocardial blood flow and myocardial flow reserve in response to regadenoson, an adenosine A_{2A} receptor agonist, implying a relationship between untethered ectonucleotidases and vasodilatory function.

What Are the Clinical Implications?

- There is growing awareness of the impact of microvascular function on patient symptoms, disease progression, and long-term outcomes; the role of ectonucleotidases in microvascular pathophysiology deserves further study.

Nonstandard Abbreviations and Acronyms

ENTPD1	ectonucleoside triphosphate diphosphohydrolase-1
HGPS	Hutchinson-Gilford progeria syndrome
MFR	myocardial flow reserve
NT5E	ecto-5'-nucleotidase

Ectonucleotidases are integral to the maintenance of vascular health, as evidenced by pathology that has been traced to abnormal ectonucleotidase production or function.^{6–10} For example, the ectonucleotidase CD39 (also known as ENTPD1 [ectonucleoside triphosphate diphosphohydrolase-1]) catabolizes extracellular ATP and ADP to AMP.^{11,12} Genetic variants of CD39 (encoded by *ENTPD1*) have been associated with an increased risk of venous thromboembolism.¹³ Individuals with inactivating mutations in the gene encoding CD73 (ecto-5'-nucleotidase [*NT5E*]), which converts AMP to adenosine, develop profound lower extremity vascular calcification. This has been attributed to reduced cAMP production, which enables activation of TNAP (tissue-nonspecific alkaline phosphatase) via alkaline phosphatase, biomineralization associated, an essential enzyme required for ectopic calcification.^{8,9,14} Similar to mutations in *NT5E*, inactivating mutations in ectonucleotide pyrophosphatase/

phosphodiesterase 1 (*ENPP1*) cause generalized arterial calcification of infancy.¹⁰

Hutchinson-Gilford progeria syndrome (HGPS) is a rare disease that results in premature death in humans attributable to stroke or cardiovascular disease, and murine models of HGPS have implicated ectonucleotidases in promoting vascular calcification in HGPS pathogenesis, as well.^{15,16} In HGPS, the expression ratio of *CD39* to *ENPP1* is doubled, leading to reduced extracellular pyrophosphate.¹⁵ Specifically inhibiting CD39 (ENTPD1) and TNAP results in preferential metabolism of ATP by ENPP1, reducing vascular calcification and increasing lifespan in an HGPS murine model.¹⁷

Although ectonucleotidases have been characterized as key regulators of vascular homeostasis, most research has focused on these enzymes in their cell surface–tethered forms. The relationship between tethered and circulating forms of ectonucleotidases and their relative contributions to human pathophysiology remains uncertain. There have been no translational human studies clarifying the relationship of circulating plasma ectonucleotidases with arterial calcification and vasodilation.^{18,19} To address this gap in knowledge, we evaluated whether circulating human plasma ectonucleotidases CD39 and CD73 are associated with traditional cardiovascular risk factors, coronary artery calcification, and myocardial flow reserve (MFR), as an indicator of capacity to vasodilate.

METHODS

This study included 529 patients presenting to the University of Michigan (Michigan Medicine) nuclear cardiology laboratory for clinically indicated nuclear positron emission tomography stress testing between June 2015 and April 2019. Study participants were prospectively enrolled and provided comprehensive written informed consent for collection of baseline demographic, clinical, and nuclear stress testing data, in addition to a 30-mL blood sample. Blood samples were drawn from existing intravenous lines or via a separate venipuncture if a line was not already in place. The clinical study was reviewed and approved by the University of Michigan institutional review board (HUM00091439). The authors had full access to all of the data in the study and assume responsibility for its integrity and the data analysis. The data that support the findings of this study are available from the corresponding author upon reasonable request. The study inclusion criteria were age ≥18 years, ability to provide consent, recent or planned (within 7 days) cardiovascular stress imaging, and preexisting, functioning intravenous or arterial catheters of sufficient size to allow blood to be drawn, or willingness for a venipuncture. Prisoners, patients

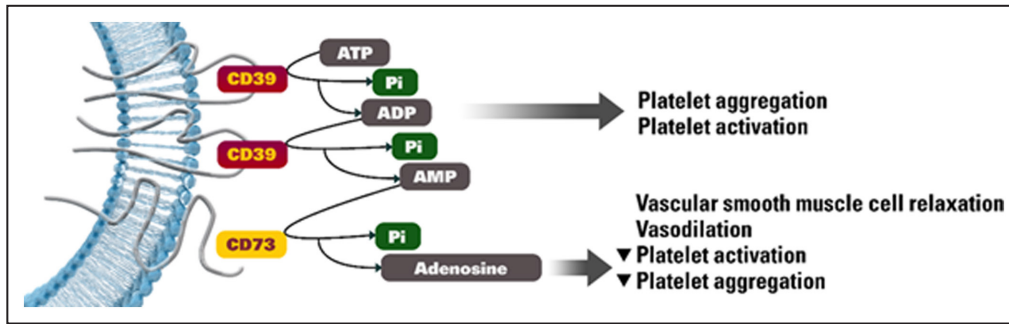


Figure 1. Catabolic activity of CD39 and CD73 produces anti-inflammatory adenosine.

CD39 catabolizes ATP and ADP, generating inorganic phosphate and AMP, the latter of which is then catabolized by CD73 to form adenosine. Adenosine is known to have vasorelaxant and antithrombotic properties. PET indicates positron emission tomography; and P_i , inorganic phosphate.

with severe psychiatric conditions, and those with advanced dementia were excluded from the study.

For patients from whom blood was able to be collected and with a sufficient sample, whole blood was collected in EDTA tubes before stress testing, centrifuged, and stored at -80°C for later analysis. Plasma was serially and numerically coded, and clinical and identifying variables were collected and managed in a secure REDCap electronic database hosted at the University of Michigan.²⁰

The primary goal of this study was to determine if circulating plasma ectonucleotidase levels (CD39 or CD73) are associated with baseline demographic or clinical variables, myocardial perfusion and flow reserve, coronary artery calcification, or age given that coronary artery calcification is known to be modulated with age.

Patients were asked to abstain from caffeine and methylxanthines for 24 hours. They were initially injected intravenously with a weight adjusted dose of rubidium-82 under resting conditions with concomitant positron emission tomography imaging (Siemens mCT, Siemens Healthineers, Knoxville, TN) in list mode. After administration of 0.4 mg of regadenoson intravenously, an additional identical dose of rubidium-82 was injected, with imaging again performed in list mode. A low dose computed tomography was obtained for attenuation correction. List mode images were unlisted into dynamic series for quantification of MBF at rest and stress and MFR as the ratio of stress to rest MBF using a 1 tissue compartment model as previously described.²¹

Individuals measuring ectonucleotidase levels and calculating the coronary artery calcium scores were blinded to the baseline demographic and clinical variables. CD73 was measured using an ELISA kit for NT5E (Abcam ab213761) designed for the quantitative measurement of human CD73 in serum and plasma, with plasma samples diluted 1:5 before assaying. CD39 was measured using an ELISA kit for human

CD39 serum and plasma (R & D systems DY4397-05), with plasma samples diluted 1:2 before assaying; 100- μL diluted plasma was loaded per well. Samples were measured in duplicate on a VERSAmax microplate reader (Molecular Devices), and means were recorded. A standard curve was used to calibrate readings from each 96-well plate.

Because of our interest in the relationship between plasma ectonucleotidases, age, and coronary calcification, study participants were categorized by age deciles, and the relationship between age and ectonucleotidase expression was determined. Study participants without prior percutaneous coronary interventions underwent coronary artery calcium scoring using the Agatston method.²² Coronary artery calcium was measured using 4DM (INVIA Solutions). Calcium in the left main, left anterior descending, left circumflex, and right coronary arteries was manually highlighted. The sum of the 4 calcium scores was calculated and reported as the total calcium score for each study participant. Study participants with prior percutaneous coronary intervention were not included in analyses including calcification given the inability to distinguish stent material from coronary calcification.

For analyses, calcium scores were categorized from 0, 1 to 99, 100 to 399, and ≥ 400 based on the Agatston value generated for each score. The intent was to determine if any relationship existed between age, plasma ectonucleotidase concentration, and coronary artery calcification. To separate aging as a biological process from exposure to disease states over time, we also examined the relationship between ectonucleotidase expression and healthy aging in study participants, excluding those with abnormal stress tests (summed stress score 0), diabetes, smoking, prior myocardial infarction, prior percutaneous coronary intervention, prior coronary artery bypass graft surgery, prior heart transplant, cirrhosis, liver transplant, and those on dialysis.

Baseline characteristics of study participants are presented as mean±SD or median (interquartile range) for continuous variables and percentages for categorical variables. Associations between pairs of categorical variables are based on a chi-squared test of independence or on a Fisher exact test. Because >75% of study participants had undetectable CD39 in the plasma, CD39 is dichotomized as CD39=0 and CD39>0. CD73 and coronary artery calcium are log-transformed because of the variation of these quantities over orders of magnitude. Associations between continuous variables and nonbinary categorical variables, such as race, are determined using 1-way ANOVA. Associations of binary variables with continuous variables are determined using t-tests. Correlations between pairs of continuous variables are based on the Pearson correlation coefficient. All analyses were performed using SPSS 27 or SPSS 28 software (IBM).

RESULTS

Baseline characteristics of the study population are presented in Table 1. Of the 529 study participants enrolled in the study, 329 (62%) were men, and the mean age was 61±12 years. Coronary artery disease risk factors were prevalent in the study population; 384 (73%) had a history of hypertension, 168 (32%) had a

history of dyslipidemia, and 199 (38%) had a history of diabetes.

Detectable CD39 ranged between 0.2 and 20377 pg/mL. Associations of CD39 (as a dichotomous variable: CD39=0 or CD39>0) with baseline demographic, clinical, and laboratory values are presented in Table 2.²³ At baseline, CD39 was more likely to be detected in White (24%) compared with Black (8%) study participants ($P=0.02$). Study participants with liver disease (35% versus 20% with no liver disease, $P<0.01$), cirrhosis (55% versus 19% with no cirrhosis, $P<0.001$), and prior liver transplant (53% versus 19% with no prior transplant, $P<0.001$) were more likely to have detectable plasma CD39. Correspondingly, detectable CD39 was associated with elevated aspartate aminotransferase (35.1±22.1 versus 29.9±18.4 IU/L, $P=0.04$), elevated bilirubin (1.1±1.5 versus 0.7±0.7 mg/dL, $P<0.01$), and lower albumin (3.9±0.6 versus 4.1±0.4 g/dL, $P<0.01$), when compared with study participants without detectable CD39.

Given the differences in detectable CD39 by race and the presence of liver disease, we further explored the relationship between race, liver disease, and detectable CD39. More White (22%) than Black (8%) study participants had liver disease. In a subanalysis excluding those with liver disease or cirrhosis, White patients were still more likely (21%) than Black patients (3%) to have detectable circulating CD39 (Table S1).

Detectable plasma CD73 ranged between 759 and 82931 pg/mL. Several baseline variables showed significant associations with log-transformed CD73 (Table 3). Log-transformed CD73 was higher in study participants with liver disease compared with study participants without liver disease (8.2±1.1 versus 7.7±0.8, $P<0.001$), study participants with cirrhosis compared with those without cirrhosis (8.8±1.2 versus 7.7±0.8, $P<0.001$), and study participants with prior liver transplant compared with those without prior liver transplant (8.8±1.2 versus 7.7±0.8, $P<0.001$). Correspondingly, log-transformed CD73 was positively associated with elevated aspartate aminotransferase (correlation coefficient=0.40, $P<0.001$), alanine transaminase (correlation coefficient=0.26, $P<0.001$), alkaline phosphatase (correlation coefficient=0.40, $P<0.001$), bilirubin (correlation coefficient=0.2, $P<0.001$), and was negatively associated with albumin (correlation coefficient=-0.28, $P<0.001$).

Because of the interest in understanding the association between detectable plasma ectonucleotidase and age, study participants were stratified by decades of age (Figure 2 and Figure 3). CD39 was undetectable in the majority of participants all age deciles, with the peak (50–59-year age category) having >40% of study participants with detectable CD39 ($P<0.001$ based on a Fisher's exact test for the overall analysis). There was no significant association between log(CD73) and age decile.

We were interested in understanding if comorbidities influenced the relationship between age and

Table 1. Baseline Characteristics of the Study Population

Characteristic	n (%) or mean±SD, n=529
Age, y	61±12
Race	
White	435 (82.2%)
Black	62 (11.7%)
Asian	6 (1.1%)
Hispanic	1 (0.2%)
Other*	19 (3.6%)
Sex (% men)	329 (62%)
Body mass index	34±9
Hypertension	384 (73%)
Dyslipidemia	168 (32%)
Diabetes	199 (38%)
Smoking (current or within 5y)	122 (23%)
Family history of coronary artery disease	117 (22%)
Prior myocardial infarction	140 (26%)
Prior percutaneous coronary intervention	100 (19%)
Prior heart transplant	81 (15%)
Liver disease	101 (19%)
Cirrhosis	55 (10%)

*Other indicates race other than White, Black, or Asian. Other races could be American Indian or Alaska Native, Native Hawaiian or Other Pacific Islander.

Table 2. Associations of CD39 (As a Dichotomous Variable: CD39=0 or CD39>0) With Baseline Demographic, Clinical, and Laboratory Values*

Variable	CD39=0 n=363	CD39>0 n=105	P value
Age, y	61.3±12.2	60.1±10.7	0.34
Sex			0.09
Men	217 (75%)	73 (25%)	
Women	146 (82%)	32 (18%)	
Race			0.02
White	298 (76%)	96 (24%)	
Black	49 (93%)	4 (8%)	
Asian	3 (60%)	2 (40%)	
Other [†]	10 (77%)	3 (23%)	
Hypertension			0.06
Yes	274 (80%)	69 (20%)	
No	89 (71%)	36 (29%)	
Dyslipidemia			0.81
Yes	247 (78%)	69 (22%)	
No	115 (77%)	35 (23%)	
Diabetes			>0.99
Yes	138 (78%)	40 (22%)	
No	225 (78%)	64 (22%)	
Smoking (current or quit within 5y)			0.19
Yes	90 (83%)	19 (17%)	
No	272 (76%)	86 (24%)	
Family history of coronary artery disease			>0.99
Yes	81 (78%)	23 (22%)	
No	274 (78%)	79 (22%)	
Prior myocardial infarction			0.51
Yes	90 (75%)	30 (25%)	
No	273 (78%)	75 (22%)	
Prior PCI			0.83
Yes	67 (76%)	21 (24%)	
No	296 (78%)	84 (22%)	
Prior CABG			>0.99
Yes	32 (78%)	9 (22%)	
No	331 (78%)	96 (23%)	
Heart transplant			0.10
Yes	47 (69%)	21 (31%)	
No	316 (79%)	84 (21%)	
Liver disease			<0.01
Yes	62 (65%)	33 (35%)	
No	292 (80%)	71 (20%)	
Cirrhosis			<0.001
Yes	23 (45%)	28 (55%)	
No	331 (81%)	76 (19%)	

(Continued)

Table 2. Continued

Variable	CD39=0 n=363	CD39>0 n=105	P value
Liver transplant			<0.001
Yes	22 (47%)	25 (53%)	
No	332 (81%)	79 (19%)	
On dialysis			0.50
Yes	9 (69%)	4 (31%)	
No	345 (78%)	100 (23%)	
Body mass index	34.8±9.7	33.1±7.5	0.052
AST	29.9±18.4	35.1±22.1	0.04
ALT	30.8±21.7	30.3±17.8	0.84
Alk Phos	97.0±66.1	101.1±51.8	0.59
Phosphate	3.52±0.83	3.47±0.68	0.71
Total protein	7.04±3.36	6.84±0.67	0.56
Albumin	4.1±0.4	3.9±0.6	<0.01
Bilirubin	0.7±0.7	1.1±1.5	<0.01
Creatinine	1.21±0.84	1.34±0.91	0.16
Glomerular filtration rate [‡]	53.8±12.2	51.4±13.4	0.11
INR	1.18±0.53	1.29±0.44	0.16

Alk Phos indicates alkaline phosphatase (IU/L); ALT, alanine aminotransferase (IU/L); AST, aspartate aminotransferase (IU/L); CABG, coronary artery bypass graft; INR, international normalized ratio; and PCI, percutaneous coronary intervention.

*Categorical variables are presented as number (%), and continuous variables are presented as mean±SD. Differences between numbers reported for a given variable and the total number of study participants because of missing data for these variables.

[†]Other indicates race other than White, Black, or Asian. Other races could be American Indian or Alaska Native, Native Hawaiian or Other Pacific Islander.

[‡]Glomerular filtration rates (mL/min per 1.73m²) reported in this study were calculated using the Modification of Diet in Renal Disease study equation (estimated glomerular filtration rate = 175×(SCr)^{-1.154}×(age)^{-0.203}×0.742 [if women]×1.212 [if Black race]), where SCr=standardized serum creatinine in mg/L and age is in years.²³

circulating ectonucleotidases. We performed a sub-analysis with 73 healthy participants. These participants did not have any stress test abnormalities, did not have diabetes, did not smoke, did not have cirrhosis, were not evaluated for liver transplant, were not on dialysis, and had no prior myocardial infarction, percutaneous coronary intervention, coronary artery bypass graft surgery, or heart transplant. Age was not significantly correlated with log-transformed CD73 ($r=-0.1$, $P=0.38$) in these participants, and there was not a significant difference in age between participants with CD39=0 (60.4) and participants with CD39>0 (60.5, $P=0.97$).

There was not a significant difference between categories of coronary artery calcium, for CD39=0 versus CD39>0 ($P=0.31$) (Table S2). The correlation between log-transformed CD73 and log-transformed coronary artery calcium was also not significant ($P=0.77$) (Table S3).

Table 3. Associations of log(CD73) With Baseline Demographic, Clinical, and Laboratory Values*

Variable	log(CD73) if variable present	log(CD73) if variable not present	P value
Sex (men)	7.8±1.0	7.7±0.7	0.38
Race			0.53
White	7.8±0.9		
Black	7.6±0.9		
Asian	7.5±0.5		
Other [†]	8.0±0.7		
Hypertension	7.7±0.9	7.9±0.9	0.1
Dyslipidemia	7.7±0.9	7.8±0.9	0.18
Diabetes	7.9±1.1	7.7±0.7	0.055
Smoking (current or quit within 5y)	7.9±1.0	7.8±0.9	0.22
Family history of coronary artery disease	7.9±0.9	7.7±0.9	0.1
Prior myocardial infarction	7.7±0.7	7.8±0.9	0.22
Prior PCI	7.8±0.8	7.8±0.9	0.96
Prior CABG	7.9±0.8	7.8±0.9	0.42
Heart transplant	7.8±0.9	7.8±0.9	0.99
Liver disease	8.2±1.1	7.7±0.8	<0.001
Cirrhosis	8.8±1.2	7.7±0.8	<0.001
Liver transplant	8.8±1.2	7.7±0.8	<0.001
On dialysis	7.8±1.0	7.8±0.9	0.88
Age	−0.08		0.09
Body mass index, kg/m ²	−0.02		0.74
AST	0.4		<0.001
ALT	0.26		<0.001
Alk Phos	0.4		<0.001
Phosphate	−0.07		0.34
Total protein	−0.01		0.90
Albumin	−0.28		<0.001
Bilirubin	0.2		<0.001
Creatinine	0.01		0.84
Glomerular filtration rate [‡]	−0.05		0.34
INR	0.08		0.21

Alk Phos indicates alkaline phosphatase (IU/L); ALT, alanine aminotransferase (IU/L); AST, aspartate aminotransferase (IU/L); CABG, coronary artery bypass graft; INR, international normalized ratio; and PCI, percutaneous coronary intervention.

*The table summarizes log(CD73) with the mean±SD for categorical variables and the Pearson correlation coefficient for log(CD73) with continuous variables.

[†]Other indicates race other than White, Black, or Asian. Other races could be American Indian or Alaska Native, Native Hawaiian or Other Pacific Islander.

[‡]Glomerular filtration rates (mL/min per 1.73 m²) reported in this study were calculated using the Modification of Diet in Renal Disease study equation (estimated glomerular filtration rate =175×(SCr)^{−1.154}×(age)^{−0.203}×0.742 [if women]×1.212 [if Black race]), where SCr indicates standardized serum cystatin C in mg/L and age is in years.²³

The relationship between plasma ectonucleotidase expression and positron emission tomography stress testing variables was determined for dichotomized

CD39 (Table 4) and log(CD73) (Table 5). Stress MBF was higher in study participants without detectable plasma CD39 compared with those with detectable CD39 (median and interquartile range, 2.49 (1.88–3.44) versus 2.33 (1.79–2.88) mL/min per g, *P*<0.001). Log(CD73) was negatively associated with MFR (correlation coefficient −0.19, *P*<0.001), suggesting that increasing log(CD73) values were associated with decreasing MFR. Decreased log(CD73) was noted in study participants with normal MFR compared with those without normal MFR (7.6±0.8 versus 8.0±1.0 pg/mL, *P*=0.002).

In a separate sensitivity analysis (data not shown), we separately excluded study participants with underlying liver disease. The major conclusions on the relationships between ectonucleotidases, age, coronary calcification, comorbidities, and stress myocardial flow parameters did not differ when study participants with underlying liver disease were excluded from the overall population.

DISCUSSION

Previously, deficient functional ectonucleotidase activity has been shown to be culpable for the development of heritable peripheral vascular disease and responsible for tuning the inflammatory response to myocardial ischemia and infarction.^{7–9,24–26} As these modulators of the extracellular purinergic cascade could potentially be shed from endothelial cells or circulating leukocytes, we queried whether ectonucleotidases CD39 and CD73 may represent circulating biomarkers. These could be assessed with traditional cardiovascular risk factors, coronary artery calcification, and potentially even predict myocardial flow and vasodilatory reserve. Here, we show that detectable CD39 was increased in White compared with Black study participants. Of the clinical history variables examined, ectonucleotidase expression was most strongly associated with underlying liver disease and not other traditional coronary artery disease risk factors. Intriguingly, detection of circulating ectonucleotidase was inversely associated with stress MBF (CD39) and MFR (CD73) (Figure 4). It will be interesting in future studies to clarify how the density or function of ectonucleotidases and A_{2A} receptors might regulate myocardial microvascular flow and resistance.

We had hypothesized that declining age-related arterial endothelial ectonucleotidase expression, possibly associated with higher detectable circulating ectonucleotidase expression, would be associated with increasing age, coronary artery calcification, comorbidities associated with coronary artery disease, and indicators of microvascular function. Our hypotheses were based on prior observations of increased

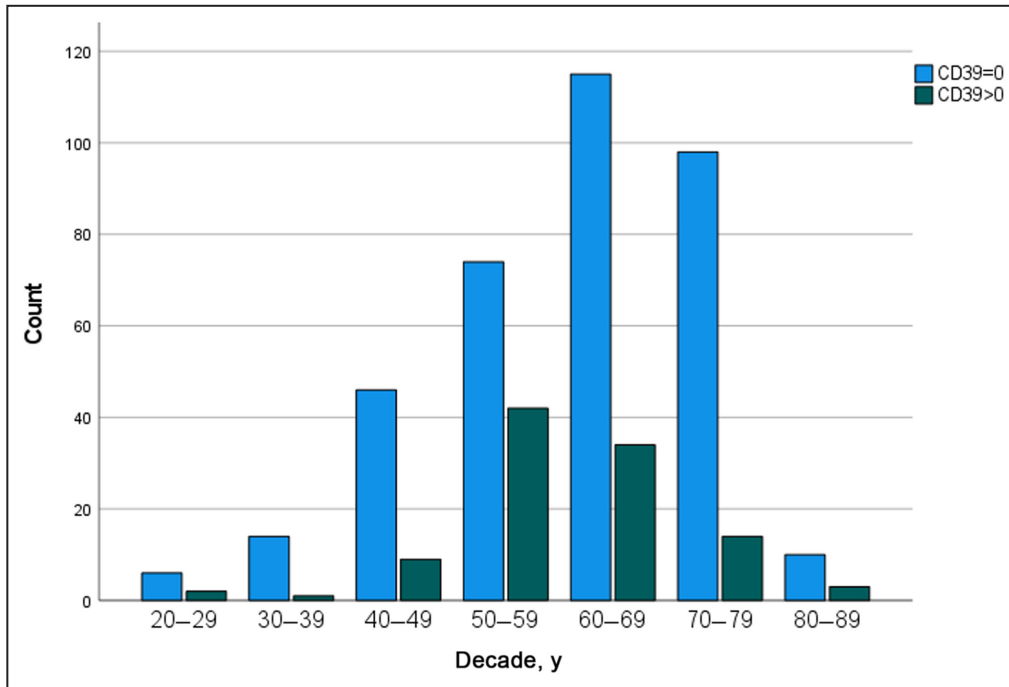


Figure 2. Decade of age and the presence of detectable plasma CD39.
* $P < 0.001$ based on Fisher exact test.

expression of CD39 in study participants aged 65 to 85 years and decreased CD39 and CD73 mRNA in centenarians (Table S4).^{27,28} The authors concluded that CD39 and CD73 gene expression may increase in middle age, but healthy aging was associated with

lower levels.²⁷ In this study, we found some evidence that circulating CD39 was more likely to be detectable in the 50- to 59-year age group compared with other adult age deciles, though no such pattern was observed for circulating CD73. We conclude that while

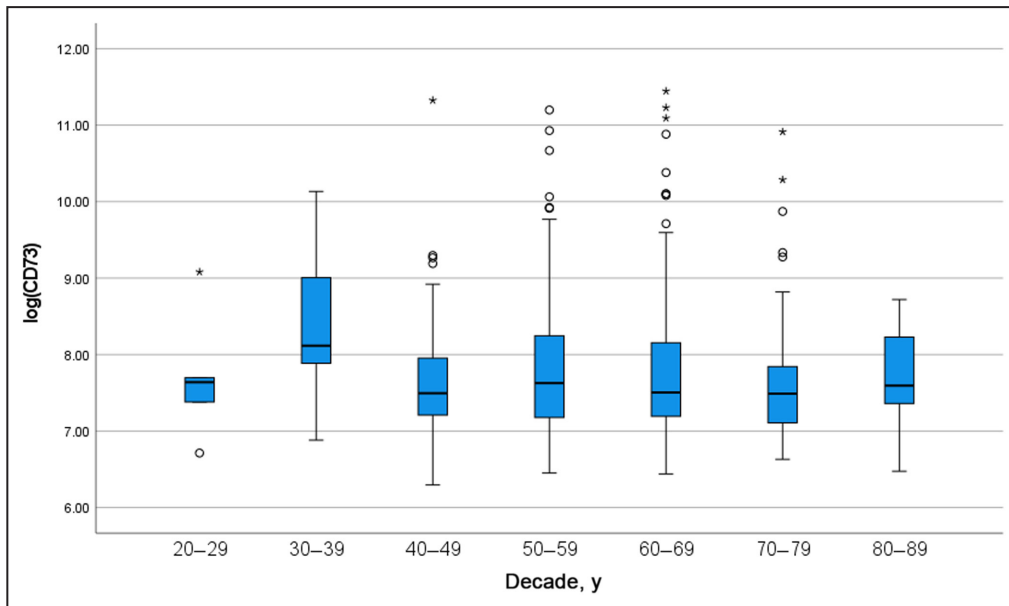


Figure 3. Decade of age and plasma log(CD73).
Box plots represent median and 25th and 75th percentiles. Circles represent data points that are >1.5 and <3 times below or above the box limits. Stars represent points that are >3 times below the lower or above the upper box limits. $P = 0.08$ based on ANOVA; n per age category: 20 to 29 (n=6), 30 to 39: (n=15), 40 to 49 (n=55), 50 to 59: (n=113), 60 to 69: (n=147), 70 to 79 (n=114), 80 to 89 (n=13).

Downloaded from <http://ahajournals.org> by on September 6, 2023

Table 4. Dichotomized Plasma CD39 and Positron Emission Tomography Stress Test Variables.

Variable	Median (IQR) for CD39=0 n=363	Median (IQR) for CD39>0 n=105	P value
Summed stress score	0 (0–6)	0 (0–7)	0.6
Summed difference score	0 (0–2)	0 (0–2)	0.45
Rest global defect	2 (0–8)	2 (0–6)	0.62
Stress global defect	1 (0–11)	2 (0–11)	0.58
Reversibility	0 (0–3)	0 (0–3)	0.65
TID ratio	1.02 (0.98–1.05)	1.00 (0.97–1.05)	0.28
Rest myocardial blood flow (mL/min-g)	1.17 (0.88–1.55)	1.12 (0.84–1.44)	0.14
Stress myocardial blood flow (mL/min-g)	2.49 (1.88–3.44)	2.33 (1.79–2.88)	0.02
Myocardial flow reserve*	2.19 (1.68–2.63)	2.03 (1.63–2.66)	0.37
Variable	Variable present	Variable not present	P value
	n (%)	n (%)	
Normal myocardial flow reserve			0.13
Yes	173 (80%)	43 (20%)	
No	115 (73%)	43 (27%)	

Mann–Whitney *U* test used for median and interquartile ranges. IQR indicates interquartile range; and TID ratio, transient ischemic dilation ratio.

*Myocardial flow reserve (where <2.0 abnormal and ≥2.0 normal).

there may be a hyperbolic relationship between circulating CD39 and CD73 and age, the relationship is likely subtle and possibly more influenced by cellular ectonucleotidase expression, rather than circulating plasma ectonucleotidases.

In this study, we found no significant association between circulating plasma CD39 or CD73 and any traditional cardiovascular risk factors such as hypertension, hyperlipidemia, diabetes, or smoking. We did

observe that White study participants were more likely to have detectable circulating CD39 than Black study participants. Differential ectonucleotidase expression by race has not been previously reported to our knowledge; whether this represents a haplotype association or other demographic, environmental, or structural interactions remains to be determined. Counter to our hypotheses, an association between circulating ectonucleotidases and traditional cardiovascular risk factors and stress testing parameters such as summed stress score or stress global defect was not present in this study.

Our study findings may be compared with those of Jalkanen et al.^{2,29} In prior studies, investigators evaluated plasma ATP and ADP levels and serum CD39 and CD73 activity in 226 study participants with stable peripheral artery disease admitted for nonurgent imaging.²⁹ Serum ATP, ADP, and CD73 values were higher in study participants with atherosclerosis of the peripheral vessels than in controls without clinically evident peripheral artery disease. Subsequently, Jalkanen et al went on to describe that CD73 expression was elevated in the vasa vasorum of developing plaques, but expression was lost in mature occlusive plaques removed at the time of peripheral vascular endarterectomy, thus implicating that elevated serum CD73 activity was elevated because of shedding of CD73 from diseased arterial walls.²

The regulation and shedding of ectonucleotidases remains incompletely explored. Human *ENTPD1* (which encodes the ectonucleotidase CD39) promoter polymorphisms have been described that increase transcriptional activity by 8-fold,¹³ while interleukin-4

Table 5. Log-Transformed CD73 and Positron Emission Tomography Stress Test Variables

Variable	Correlation coefficient	P value	
Summed stress score	–0.02	0.67	
Summed difference score	–0.02	0.69	
Rest global defect	–0.02	0.66	
Stress global defect	–0.03	0.54	
Reversibility	–0.03	0.60	
TID ratio	0.04	0.39	
Rest MBF	0.1	0.056	
Stress MBF (mL/(min-per g))	–0.05	0.38	
MFR* (mL/min-per g)	–0.19	<0.001	
Variable	Variable is present (mean±SD for log(CD73))	Variable is not present (mean±SD log(CD73))	P value
Normal MFR*	7.6±0.8	8.0±1.0	0.002

MBF indicates myocardial blood flow; MFR, myocardial flow reserve; and TID ratio, transient ischemic dilation ratio.

*Myocardial flow reserve (using <2.0 abnormal and ≥2.0 normal).

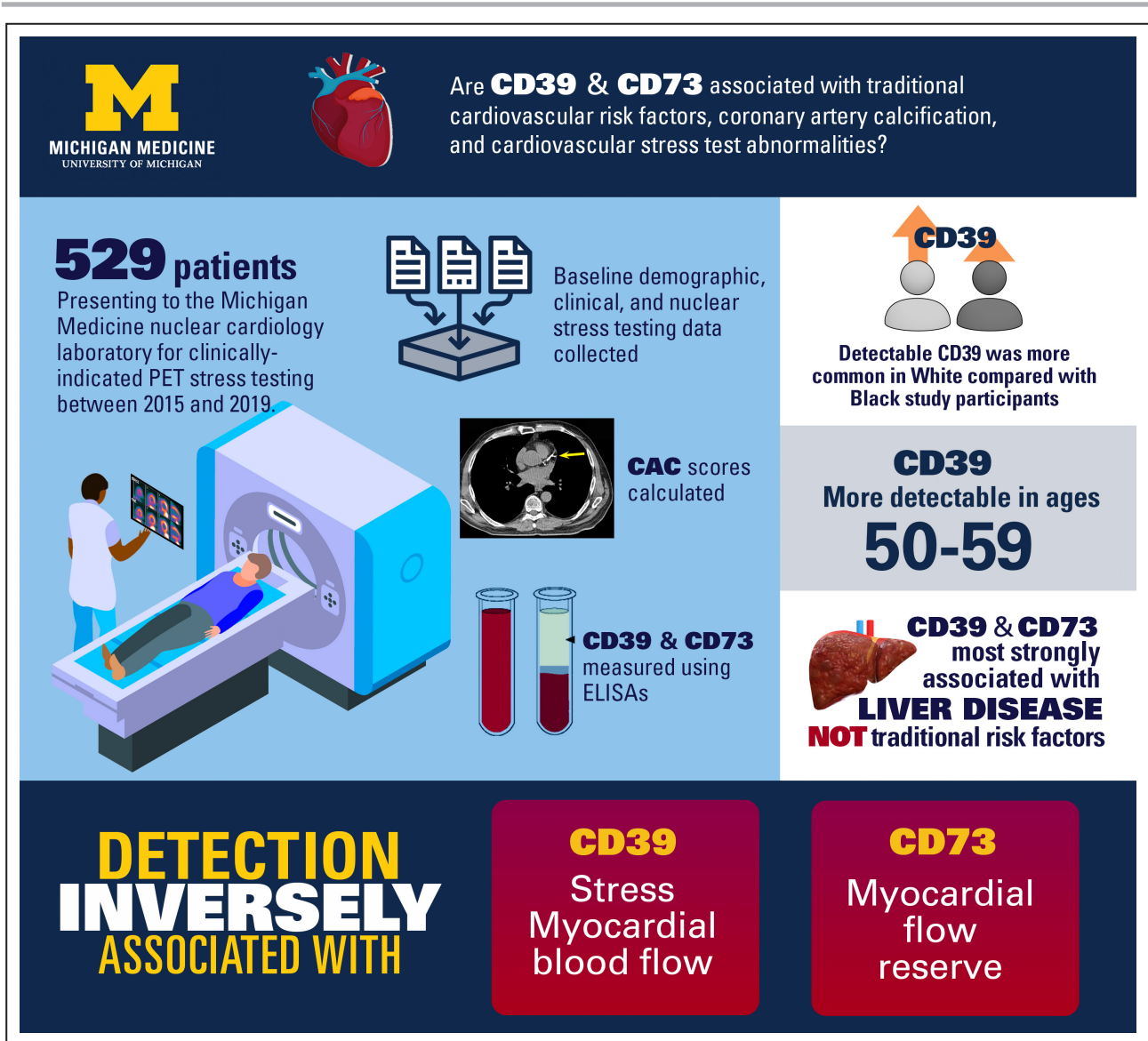


Figure 4. Summary of the key study findings.

Study findings support an inverse relationship between plasma ectonucleotidases, stress myocardial blood flow (CD39), and myocardial flow reserve (CD73). CAC indicates coronary artery calcium; and PET, positron emission tomography.

inhibits *ENTPD1* transcription through the phosphorylation of STAT6 (signal transducer and activator of transcription 6) and the upregulation of GATA3 (GATA binding protein 3), GF11 (growth factor independent 1 transcriptional repressor), and YY1 (Yin Yang 1).³⁰ Soluble, circulating CD39 has been described,^{31,32} with contribution from circulating microparticles.^{33–35} Prior studies suggest increased extracellular CD39 release in response to increased intracellular cyclic AMP.^{19,36} CD73 is a GPI (glycosylphosphatidylinositol)-anchored ectonucleotidase that becomes soluble when the GPI anchor is shed by the action of phosphatidylinositol-specific phospholipase; CD73 sheds variably in response to stimuli between lymphocytes and endothelial

cells.³ Ectonucleotidase shedding from the cell surface also occurs in response to shear stress.³⁷ Kanthi et al previously described reduced CD39 expression (in an animal model) in atheroprone disturbed flow vascular regions.³⁸ Soluble CD39³⁵ and CD73^{29,39} maintain ectonucleotidase enzymatic activity. Taken together, we hypothesize that higher detectable circulating ectonucleotidase expression may reflect endothelial dysfunction and pathological shedding. Whether this is an actively mediated or passive process requires further investigation.

Striking evidence links genetic defects in CD73 to profound peripheral arterial calcification.⁸ In the absence of functional CD73, reduced local adenosine

disinhibits tissue nonspecific alkaline phosphatase activity, resulting in inorganic phosphate production, breakdown of pyrophosphate (an endogenous inhibitor of calcification), and ultimately results in increased vascular calcification.^{8,9,40} Based on these data, which are, to date and to our knowledge the largest and only biomarker study attempting to characterize the relationship between circulating ectonucleotidases and cardiovascular disease, we do not believe that circulating plasma ectonucleotidases play any appreciable role on the development or inhibition of coronary artery calcification.

In this study, we noted an association between detectable circulating ectonucleotidase and study participants with underlying liver disease. This was a single-center study with an active liver transplantation program; select patients undergoing liver transplant evaluations undergo positron emission tomography stress testing as part of their pre-liver transplantation evaluation. This population may have been over-represented in our study population compared with the general population.

Prior studies have demonstrated that adenosine (via ectonucleotidase activity) modulates the liver's response to injury and is involved in the regulation of liver fibrosis.^{41–45} The findings described in this manuscript are consistent with prior reports of increased ectonucleotidase expression in the setting of liver disease and fibrosis.^{46–49} Increased CD39 transcript and protein expression in response to liver injury is dependent upon Sp1 transcription factor activation.⁵⁰ During the pathologic process of liver fibrosis, hepatic stellate cells and portal fibroblasts differentiate into hepatic myofibroblasts.⁵¹ This process is associated with CD73 transcriptional upregulation, dependent upon SP1 (specificity protein 1) and small mothers against decapentaplegic promoter elements.⁵¹ In addition, Snider et al detected higher levels of the NT5E-2 splice variant in study participants with cirrhosis and hepatocellular carcinoma, implying a link between the encoded CD73 isoform and subsequent diseases characterized by abnormal tissue growth, including fibrosis-induced cirrhosis and hepatic cancer.^{42,52,53} Mice with a hepatocyte-specific deletion of *NT5E* develop spontaneous age-dependent liver disease.⁵³ In addition to supporting liver homeostasis, CD73 is a critical regulator of hepatocyte responses to different forms of stress, following alcoholic, chemically induced, and ischemic injury.^{42,43,54,55} Therefore, it is not surprising that we observed the highest serum levels of CD73 (and its catalytic partner enzyme CD39) in patients with liver disease.

A notable observation from this study was the inverse relationship between circulating ectonucleotidases and MFR and stress MBF. Adenosine is known to be vasodilatory, and the extracellular concentration

depends on cellular release, cellular reuptake, and purinergic metabolism by ectonucleotidases.^{56,57} Specifically, adenosine is generated by sequential phosphohydrolysis of ATP and ADP to AMP to adenosine by ectonucleotidases.⁵⁸ Adenosine binds to specific adenosine receptor types (and subtypes) and is known to regulate myocardial and coronary function.⁵⁹ While coronary flow is regulated by multiple adenosine receptors, the A_{2A} adenosine receptor is thought to be the predominant receptor subtype responsible for coronary blood flow and dilates in both endothelial-dependent and -independent manners.^{59,60}

One may expect that higher cell surface-tethered ectonucleotidases would result in adenosine production and higher MFR. Here, we found that circulating ectonucleotidases were associated with lower MFR and stress MBF. Based on prior studies, we attribute the detection of plasma ectonucleotidases in study participants with these flow characteristics to shedding of ectonucleotidases from endothelium, possibly attributable to endothelial dysfunction.^{2,29,38,61} While this study does not prove causation (eg, that reduced local endothelial ectonucleotidase expression causes impaired MBF—possibly microvascular), it does raise a plausible hypothesis that local impaired adenosine production could contribute to impaired MBF, even in the absence of atherosclerotic, obstructive coronary artery disease.^{62,63} The vasodilator used in this study was regadenoson, a selective adenosine A_{2A} receptor agonist that mimics the effects of adenosine.^{64,65} We do not believe the administration of regadenoson would have impacted the findings of this study, and this does not rule out a possible role for the adenosine A_{2A} receptor in the link between detection of circulating ectonucleotidase and MBF and MFR.

Limitations of the current study are that study participants were recruited from the stress testing laboratory at the University of Michigan (a single center) and may not be generalizable to other populations or geographical locations. The majority of the study population were men and White, a reflection of the demographic characteristics of the population referred for stress testing and recruited in a consecutive fashion. Ectonucleotidase expression was measured from plasma samples and may not be reflective of circulating cell-bound leukocyte ectonucleotidase expression or of endothelial ectonucleotidase expression. Ectonucleotidase expression rather than activity was determined, though this is likely more reflective of what would be feasible if applied to clinical practice. This study remains subject to the limitations of a human biomarker study, providing hypotheses-generating observations and does not prove causality.

Much remains to be determined to elucidate the relationship between ectonucleotidases, adenosine receptors, and myocardial microvascular function.

Understanding the molecular underpinnings of impaired MBF (with or without obstructive coronary artery disease) is critical given that the risk of death for those with myocardial infarction with nonobstructive coronary artery disease is >10% within 5 years.⁶⁶ Adenosine can provoke vasodilation in a nonendothelial-dependent fashion, though in the vessel wall, endogenous adenosine production stems from the enzymatic activity of membrane-bound ectonucleotidases, primarily expressed on the endothelium.^{67,68} Our findings, in concert with prior studies, strongly imply that pathological shedding of endothelial ectonucleotidases and the potential resultant reduction in local adenosine are linked with MBF.

ARTICLE INFORMATION

Received December 2, 2022; accepted March 20, 2023.

Affiliations

Division of Cardiovascular Medicine, Department of Medicine, Michigan Medicine, Ann Arbor, MI (R.G.K., J.C., A.R., J.K., D.J.P., V.L.M., N.R.S.); Consulting for Statistics, Computing, and Analytics Research, University of Michigan, Ann Arbor, MI (C.P.); Department of Cell Biology and Physiology, University of North Carolina at Chapel Hill, Chapel Hill, NC (N.T.S.); Division of Cardiology, Departments of Medicine and Bioengineering, Vascular Medicine Institute, University of Pittsburgh, Pittsburgh, PA (C.S.H.); Department of Molecular & Integrative Physiology, University of Michigan, Ann Arbor, MI (D.J.P.); Division of Cardiovascular Medicine, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN (N.R.S.); and Department of Biomedical Engineering, Vanderbilt University, Nashville, TN (N.R.S.).

Acknowledgments

The authors thank Steve Alvey for assistance with graphic design.

Sources of Funding

This study was funded by the National Institutes of Health (NIH K76AG064426 and P30-AG024824 to N.R.S., R01DK110355 to N.T.S., R01HL142932 to C.S.H., R01HL150392 to D.J.P.), the American Heart Association Innovative Project Award (20IPA35260111, to C.S.H.), the Michigan Biology of Cardiovascular Aging Program (N.R.S.), the University of Michigan Undergraduate Research Opportunity Program (N.R.S.), the University of Michigan Frankel Cardiovascular Center (V.L.M.), and the Michigan Institute for Clinical & Health Research (CTSA: UL1RR024986 to V.L.M.).

Disclosures

N.T. Snider has received research funding from Vincerox Pharma and Taysha Gene Therapies, unrelated to this work. Dr Sutton serves as an advisor for and/or has received honoraria for speaking from Abbott, Cordis, Philips, Shockwave, and Zoll Medical, unrelated to this work. Dr Murthy has stock in General Electric and Cardinal Health, has stock options and serves as scientific advisor for Ionetix, and receives research grants to his institution from Siemens, unrelated to this work. The remaining authors have no disclosures to report.

Supplemental Material

Tables S1–S4
Reference⁶⁹

REFERENCES

- Hyman MC, Petrovic-Djergovic D, Visovatti SH, Liao H, Yanamadala S, Bouis D, Su EJ, Lawrence DA, Broekman MJ, Marcus AJ, et al. Self-regulation of inflammatory cell trafficking in mice by the leukocyte surface apyrase CD39. *J Clin Invest*. 2009;119:1136–1149. doi: 10.1172/JCI36433
- Jalkanen J, Hollmen M, Jalkanen S, Hakovirta H. Regulation of CD73 in the development of lower limb atherosclerosis. *Purinergic Signal*. 2017;13:127–134. doi: 10.1007/s11302-016-9545-0
- Airas L, Niemela J, Salmi M, Puurunen T, Smith DJ, Jalkanen S. Differential regulation and function of CD73, a glycosyl-phosphatidylinositol-linked 70-kD adhesion molecule, on lymphocytes and endothelial cells. *J Cell Biol*. 1997;136:421–431. doi: 10.1083/jcb.136.2.421
- Petrovic-Djergovic D, Hyman MC, Ray JJ, Bouis D, Visovatti SH, Hayasaki T, Pinsky DJ. Tissue-resident ecto-5' nucleotidase (CD73) regulates leukocyte trafficking in the ischemic brain. *J Immunol*. 2012;188:2387–2398. doi: 10.4049/jimmunol.1003671
- Wilson RF, Wyche K, Christensen BV, Zimmer S, Laxson DD. Effects of adenosine on human coronary arterial circulation. *Circulation*. 1990;82:1595–1606. doi: 10.1161/01.CIR.82.5.1595
- Sutton NR, Bouis D, Mann KM, Rashid IM, McCubbrey AL, Hyman MC, Goldstein DR, Mei A, Pinsky DJ. CD73 promotes age-dependent accretion of atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2020;40:61–71. doi: 10.1161/ATVBAHA.119.313002
- Sutton NR, Hayasaki T, Hyman MC, Anyanwu AC, Liao H, Petrovic-Djergovic D, Badri L, Baek AE, Walker N, Fukase K, et al. Ectonucleotidase CD39-driven control of postinfarction myocardial repair and rupture. *JCI Insight*. 2017;2:e89504. doi: 10.1172/jci.insight.89504
- St Hilaire C, Ziegler SG, Markello TC, Brusco A, Groden C, Gill F, Carlson-Donohoe H, Lederman RJ, Chen MY, Yang D, et al. NT5E mutations and arterial calcifications. *N Engl J Med*. 2011;364:432–442. doi: 10.1056/NEJMoa0912923
- Moorhead WJ III, Chu CC, Cuevas RA, Callahan J, Wong R, Regan C, Boufford CK, Sur S, Liu M, Gomez D, et al. Dysregulation of FOXO1 (Forkhead box O1 protein) drives calcification in arterial calcification due to deficiency of CD73 and is present in peripheral artery disease. *Arterioscler Thromb Vasc Biol*. 2020;40:1680–1694. doi: 10.1161/ATVBAHA.119.313765
- Rutsch F, Ruf N, Vaingankar S, Toliat MR, Suk A, Hohne W, Schauer G, Lehmann M, Roscioli T, Schnabel D, et al. Mutations in ENPP1 are associated with 'idiopathic' infantile arterial calcification. *Nat Genet*. 2003;34:379–381. doi: 10.1038/ng1221
- Baek AE, Sutton NR, Petrovic-Djergovic D, Liao H, Ray JJ, Park J, Kanthi Y, Pinsky DJ. Ischemic cerebroprotection conferred by myeloid lineage-restricted or global CD39 transgene expression. *Circulation*. 2017;135:2389–2402. doi: 10.1161/CIRCULATIONAHA.116.023301
- Kanthi YM, Sutton NR, Pinsky DJ. CD39: interface between vascular thrombosis and inflammation. *Curr Atheroscler Rep*. 2014;16:425. doi: 10.1007/s11883-014-0425-1
- Maloney JP, Branchford BR, Brodsky GL, Cosmic MS, Calabrese DW, Aquilante CL, Maloney KW, Gonzalez JR, Zhang W, Moreau KL, et al. The ENTPD1 promoter polymorphism -860 a > G (rs3814159) is associated with increased gene transcription, protein expression, CD39/NTPDase1 enzymatic activity, and thromboembolism risk. *FASEB J*. 2017;31:2771–2784. doi: 10.1096/fj.201600344R
- Ziegler SG, Ferreira CR, MacFarlane EG, Riddle RC, Tomlinson RE, Chew EY, Martin L, Ma CT, Sergienko E, Pinkerton AB, et al. Ectopic calcification in pseudoxanthoma elasticum responds to inhibition of tissue-nonspecific alkaline phosphatase. *Sci Transl Med*. 2017;9:eaa11669.
- Villa-Bellosta R, Rivera-Torres J, Osorio FG, Acin-Perez R, Enriquez JA, Lopez-Otin C, Andres V. Defective extracellular pyrophosphate metabolism promotes vascular calcification in a mouse model of Hutchinson-Gilford progeria syndrome that is ameliorated on pyrophosphate treatment. *Circulation*. 2013;127:2442–2451. doi: 10.1161/CIRCULATIONAHA.112.000571
- Ullrich NJ, Gordon LB. Hutchinson-Gilford progeria syndrome. *Handb Clin Neurol*. 2015;132:249–264.
- Villa-Bellosta R. ATP-based therapy prevents vascular calcification and extends longevity in a mouse model of Hutchinson-Gilford progeria syndrome. *Proc Natl Acad Sci USA*. 2019;116:23698–23704. doi: 10.1073/pnas.1910972116
- Schneider E, Rissiek A, Winzer R, Puig B, Rissiek B, Haag F, Mittrucker HW, Magnus T, Tolosa E. Generation and function of non-cell-bound CD73 in inflammation. *Front Immunol*. 2019;10:1729. doi: 10.3389/fimmu.2019.01729
- Baek AE, Kanthi Y, Sutton NR, Liao H, Pinsky DJ. Regulation of ectopyrase CD39 (ENTPD1) expression by phosphodiesterase III (PDE3). *FASEB J*. 2013;27:4419–4428. doi: 10.1096/fj.13-234625
- Harris PA, Taylor R, Thielke R, Payne J, Gonzalez N, Conde JG. Research electronic data capture (REDCap)—a metadata-driven

- methodology and workflow process for providing translational research informatics support. *J Biomed Inform.* 2009;42:377–381. doi: 10.1016/j.jbi.2008.08.010
21. Poitrasson-Riviere A, Moody JB, Renaud JM, Hagio T, Arida-Moody L, Murthy VL, Ficaro EP. Effect of iterations and time of flight on normal distributions of (82)Rb PET relative perfusion and myocardial blood flow. *J Nucl Cardiol.* 2022;29:2612–2623. doi: 10.1007/s12350-021-02775-8
 22. Agatston AS, Janowitz WR, Hildner FJ, Zusmer NR, Viamonte M Jr, Detrano R. Quantification of coronary artery calcium using ultrafast computed tomography. *J Am Coll Cardiol.* 1990;15:827–832. doi: 10.1016/0735-1097(90)90282-T
 23. National Kidney Foundation MDRD Study Equation. Accessed November 24, 2022: <https://www.kidney.org/content/mdrd-study-equation>.
 24. Borg N, Alter C, Gorldt N, Jacoby C, Ding Z, Steckel B, Quast C, Bonner F, Friebe D, Temme S, et al. CD73 on T cells orchestrates cardiac wound healing after myocardial infarction by purinergic metabolic reprogramming. *Circulation.* 2017;136:297–313. doi: 10.1161/CIRCULATIONAHA.116.023365
 25. Bonner F, Borg N, Burghoff S, Schrader J. Resident cardiac immune cells and expression of the ectonucleotidase enzymes CD39 and CD73 after ischemic injury. *PLoS One.* 2012;7:e34730. doi: 10.1371/journal.pone.0034730
 26. Kohler D, Eckle T, Faigle M, Grenz A, Mittelbronn M, Laucher S, Hart ML, Robson SC, Muller CE, Eltzhig HK. CD39/ectonucleoside triphosphate diphosphohydrolase 1 provides myocardial protection during cardiac ischemia/reperfusion injury. *Circulation.* 2007;116:1784–1794. doi: 10.1161/CIRCULATIONAHA.107.690180
 27. Fang F, Yu M, Cavanagh MM, Hutter Saunders J, Qi Q, Ye Z, Le Saux S, Sultan W, Turgano E, Dekker CL, et al. Expression of CD39 on activated T cells impairs their survival in older individuals. *Cell Rep.* 2016;14:1218–1231. doi: 10.1016/j.celrep.2016.01.002
 28. Crooke A, Martinez-Henandez J, Martinez-Lopez J, Cruz-Jentoft A, Huete-Toral F, Pintor J. Low expression of CD39 and CD73 genes in centenarians compared with octogenarians. *Immun Ageing.* 2017;14:11. doi: 10.1186/s12979-017-0094-3
 29. Jalkanen J, Yegutkin GG, Hollmen M, Aalto K, Kiviniemi T, Salomaa V, Jalkanen S, Hakovirta H. Aberrant circulating levels of purinergic signaling markers are associated with several key aspects of peripheral atherosclerosis and thrombosis. *Circ Res.* 2015;116:1206–1215. doi: 10.1161/CIRCRESAHA.116.305715
 30. Fang F, Cao W, Mu Y, Okuyama H, Li L, Qiu J, Weyand CM, Goronzy JJ. IL-4 prevents adenosine-mediated immunoregulation by inhibiting CD39 expression. *JCI Insight.* 2022;7:e157509. doi: 10.1172/jci.insight.157509
 31. Yegutkin GG, Wieringa B, Robson SC, Jalkanen S. Metabolism of circulating ADP in the bloodstream is mediated via integrated actions of soluble adenylate kinase-1 and NTPDase1/CD39 activities. *FASEB J.* 2012;26:3875–3883. doi: 10.1096/fj.12-205658
 32. Goueli SA, Hsiao K. Monitoring and characterizing soluble and membrane-bound ectonucleotidases CD73 and CD39. *PLoS One.* 2019;14:e0220094. doi: 10.1371/journal.pone.0220094
 33. Banz Y, Beldi G, Wu Y, Atkinson B, Usheva A, Robson SC. CD39 is incorporated into plasma microparticles where it maintains functional properties and impacts endothelial activation. *Br J Haematol.* 2008;142:627–637. doi: 10.1111/j.1365-2141.2008.07230.x
 34. Schmelzle M, Splith K, Andersen LW, Kornek M, Schuppan D, Jones-Bamman C, Nowak M, Toxavidis V, Salhanick SD, Han L, et al. Increased plasma levels of microparticles expressing CD39 and CD133 in acute liver injury. *Transplantation.* 2013;95:63–69. doi: 10.1097/TP.0b013e318278d3cd
 35. Visovatti SH, Hyman MC, Bouis D, Neubig R, McLaughlin VV, Pinsky DJ. Increased CD39 nucleotidase activity on microparticles from patients with idiopathic pulmonary arterial hypertension. *PLoS One.* 2012;7:e40829. doi: 10.1371/journal.pone.0040829
 36. Liao H, Hyman MC, Baek AE, Fukase K, Pinsky DJ. cAMP/CREB-mediated transcriptional regulation of ectonucleoside triphosphate diphosphohydrolase 1 (CD39) expression. *J Biol Chem.* 2010;285:14791–14805. doi: 10.1074/jbc.M110.116905
 37. Yegutkin G, Bodin P, Burnstock G. Effect of shear stress on the release of soluble ecto-enzymes ATPase and 5'-nucleotidase along with endogenous ATP from vascular endothelial cells. *Br J Pharmacol.* 2000;129:921–926. doi: 10.1038/sj.bjp.0703136
 38. Kanthi Y, Hyman MC, Liao H, Baek AE, Visovatti SH, Sutton NR, Goonewardena SN, Neral MK, Jo H, Pinsky DJ. Flow-dependent expression of ectonucleotide tri(d)phosphohydrolase-1 and suppression of atherosclerosis. *J Clin Invest.* 2015;125:3027–3036. doi: 10.1172/JCI79514
 39. Yegutkin GG, Auvinen K, Rantakari P, Hollmen M, Karikoski M, Grennan R, Elima K, Jalkanen S, Salmi M. Ecto-5'-nucleotidase/CD73 enhances endothelial barrier function and sprouting in blood but not lymphatic vasculature. *Eur J Immunol.* 2015;45:562–573. doi: 10.1002/eji.201444856
 40. Sutton NR, Hofmann Bowman MA. Reining in peripheral arterial calcification. *Arterioscler Thromb Vasc Biol.* 2020;40:1614–1616. doi: 10.1161/ATVBAHA.120.314665
 41. Wang S, Gao S, Zhou D, Qian X, Luan J, Lv X. The role of the CD39-CD73-adenosine pathway in liver disease. *J Cell Physiol.* 2021;236:851–862. doi: 10.1002/jcp.29932
 42. Snider NT, Griggs NW, Singla A, Moons DS, Weerasinghe SV, Lok AS, Ruan C, Burant CF, Conjeevaram HS, Omary MB. CD73 (ecto-5'-nucleotidase) hepatocyte levels differ across mouse strains and contribute to mallory-denk body formation. *Hepatology.* 2013;58:1790–1800. doi: 10.1002/hep.26525
 43. Peng Z, Fernandez P, Wilder T, Yee H, Chiriboga L, Chan ES, Cronstein BN. Ecto-5'-nucleotidase (CD73)-mediated extracellular adenosine production plays a critical role in hepatic fibrosis. *Nucleosides Nucleotides Nucleic Acids.* 2008;27:821–824. doi: 10.1080/15257770802146403
 44. Rothweiler S, Feldbrugge L, Jiang ZG, Cszmadia E, Longhi MS, Vaid K, Enjiyo K, Popov YV, Robson SC. Selective deletion of ENTPD1/CD39 in macrophages exacerbates biliary fibrosis in a mouse model of sclerosing cholangitis. *Purinergic Signal.* 2019;15:375–385. doi: 10.1007/s11302-019-09664-3
 45. Peng ZW, Rothweiler S, Wei G, Ikenaga N, Liu SB, Sverdlow DY, Vaid KA, Longhi MS, Kuang M, Robson SC, et al. The ectonucleotidase ENTPD1/CD39 limits biliary injury and fibrosis in mouse models of sclerosing cholangitis. *Hepatol Commun.* 2017;1:957–972. doi: 10.1002/hep4.1084
 46. Pagani F, Panteghini M. 5'-Nucleotidase in the detection of increased activity of the liver form of alkaline phosphatase in serum. *Clin Chem.* 2001;47:2046–2048. doi: 10.1093/clinchem/47.11.2046
 47. Dixon TF, Purdom M. Serum 5-nucleotidase. *J Clin Pathol.* 1954;7:341–343. doi: 10.1136/jcp.7.4.341
 48. Sunderman FW Jr. The clinical biochemistry of 5'-nucleotidase. *Ann Clin Lab Sci.* 1990;20:123–139.
 49. Belfield A, Goldberg DM. Application of a continuous spectrophotometric assay for 5'nucleotidase activity in normal subjects and patients with liver and bone disease. *Clin Chem.* 1969;15:931–939. doi: 10.1093/clinchem/15.10.931
 50. Hart ML, Gorzolla IC, Schittenhelm J, Robson SC, Eltzhig HK. SP1-dependent induction of CD39 facilitates hepatic ischemic preconditioning. *J Immunol.* 2010;184:4017–4024. doi: 10.4049/jimmunol.0901851
 51. Fausther M, Sheung N, Saiman Y, Bansal MB, Dranoff JA. Activated hepatic stellate cells upregulate transcription of ecto-5'-nucleotidase/CD73 via specific SP1 and SMAD promoter elements. *Am J Physiol Gastrointest Liver Physiol.* 2012;303:G904–G914. doi: 10.1152/ajpgi.00015.2012
 52. Snider NT, Altschuler PJ, Wan S, Welling TH, Cavalcoli J, Omary MB. Alternative splicing of human NT5E in cirrhosis and hepatocellular carcinoma produces a negative regulator of ecto-5'-nucleotidase (CD73). *Mol Biol Cell.* 2014;25:4024–4033. doi: 10.1091/mbc.e14-06-1167
 53. Alcedo KP, Rouse MA, Jung GS, Fu D, Minor M, Willcockson HH, Greene KG, Snider NT. CD73 maintains hepatocyte metabolic integrity and mouse liver homeostasis in a sex-dependent manner. *Cell Mol Gastroenterol Hepatol.* 2021;12:141–157. doi: 10.1016/j.jcmgh.2021.01.016
 54. Hart ML, Much C, Gorzolla IC, Schittenhelm J, Kloor D, Stahl GL, Eltzhig HK. Extracellular adenosine production by ecto-5'-nucleotidase protects during murine hepatic ischemic preconditioning. *Gastroenterology.* 2008;135:1739–1750.e3. doi: 10.1053/j.gastro.2008.07.064
 55. Peng Z, Borea PA, Varani K, Wilder T, Yee H, Chiriboga L, Blackburn MR, Azzena G, Resta G, Cronstein BN. Adenosine signaling contributes to ethanol-induced fatty liver in mice. *J Clin Invest.* 2009;119:582–594. doi: 10.1172/JCI37409

56. Layland J, Carrick D, Lee M, Oldroyd K, Berry C. Adenosine: physiology, pharmacology, and clinical applications. *JACC Cardiovasc Interv.* 2014;7:581–591. doi: [10.1016/j.jcin.2014.02.009](https://doi.org/10.1016/j.jcin.2014.02.009)
57. Chen JF, Eltzschig HK, Fredholm BB. Adenosine receptors as drug targets—what are the challenges? *Nat Rev Drug Discov.* 2013;12:265–286. doi: [10.1038/nrd3955](https://doi.org/10.1038/nrd3955)
58. Chakrabarti A, Goldstein DR, Sutton NR. Age-associated arterial calcification: the current pursuit of aggravating and mitigating factors. *Curr Opin Lipidol.* 2020;31:265–272. doi: [10.1097/MOL.0000000000000703](https://doi.org/10.1097/MOL.0000000000000703)
59. Hein TW, Wang W, Zoghi B, Muthuchamy M, Kuo L. Functional and molecular characterization of receptor subtypes mediating coronary microvascular dilation to adenosine. *J Mol Cell Cardiol.* 2001;33:271–282. doi: [10.1006/jmcc.2000.1298](https://doi.org/10.1006/jmcc.2000.1298)
60. Teng B, Qin W, Ansari HR, Mustafa SJ. Involvement of p38-mitogen-activated protein kinase in adenosine receptor-mediated relaxation of coronary artery. *Am J Physiol Heart Circ Physiol.* 2005;288:H2574–H2580. doi: [10.1152/ajpheart.00912.2004](https://doi.org/10.1152/ajpheart.00912.2004)
61. Roy C, Tabiasco J, Caillon A, Delneste Y, Merot J, Favre J, Guihot AL, Martin L, Nascimento DC, Ryffel B, et al. Loss of vascular expression of nucleoside triphosphate diphosphohydrolase-1/CD39 in hypertension. *Purinergic Signal.* 2018;14:73–82. doi: [10.1007/s11302-017-9597-9](https://doi.org/10.1007/s11302-017-9597-9)
62. Pepine CJ, Anderson RD, Sharaf BL, Reis SE, Smith KM, Handberg EM, Johnson BD, Sopko G, Bairey Merz CN. Coronary microvascular reactivity to adenosine predicts adverse outcome in women evaluated for suspected ischemia results from the National Heart, Lung and Blood Institute WISE (Women's ischemia syndrome evaluation) study. *J Am Coll Cardiol.* 2010;55:2825–2832. doi: [10.1016/j.jacc.2010.01.054](https://doi.org/10.1016/j.jacc.2010.01.054)
63. Talukder MA, Morrison RR, Ledent C, Mustafa SJ. Endogenous adenosine increases coronary flow by activation of both A2A and A2B receptors in mice. *J Cardiovasc Pharmacol.* 2003;41:562–570. doi: [10.1097/00005344-200304000-00008](https://doi.org/10.1097/00005344-200304000-00008)
64. Al Jaroudi W, Iskandrian AE. Regadenoson: a new myocardial stress agent. *J Am Coll Cardiol.* 2009;54:1123–1130. doi: [10.1016/j.jacc.2009.04.089](https://doi.org/10.1016/j.jacc.2009.04.089)
65. Gao Z, Li Z, Baker SP, Lasley RD, Meyer S, Elzein E, Palle V, Zablocki JA, Blackburn B, Belardinelli L. Novel short-acting A2A adenosine receptor agonists for coronary vasodilation: inverse relationship between affinity and duration of action of A2A agonists. *J Pharmacol Exp Ther.* 2001;298:209–218.
66. Reynolds HR, Maehara A, Kwong RY, Sedlak T, Saw J, Smilowitz NR, Mahmud E, Wei J, Marzo K, Matsumura M, et al. Coronary optical coherence tomography and cardiac magnetic resonance imaging to determine underlying causes of myocardial infarction with nonobstructive coronary arteries in women. *Circulation.* 2021;143:624–640. doi: [10.1161/CIRCULATIONAHA.120.052008](https://doi.org/10.1161/CIRCULATIONAHA.120.052008)
67. Mustafa SJ, Morrison RR, Teng B, Pelleg A. Adenosine receptors and the heart: role in regulation of coronary blood flow and cardiac electrophysiology. *Handb Exp Pharmacol.* 2009;161–188. doi: [10.1007/978-3-540-89615-9_6](https://doi.org/10.1007/978-3-540-89615-9_6)
68. Mubagwa K, Mullane K, Flameng W. Role of adenosine in the heart and circulation. *Cardiovasc Res.* 1996;32:797–813. doi: [10.1016/S0008-6363\(96\)00140-X](https://doi.org/10.1016/S0008-6363(96)00140-X)
69. Chen YH, Lu HI, Lo CM, Li SH. CD73 promotes tumor progression in patients with esophageal squamous cell carcinoma. *Cancers (Basel).* 2021;13:3982. doi: [10.3390/cancers13163982](https://doi.org/10.3390/cancers13163982)

Supplemental Material

Table S1. Counts and percentages of White and Black study participants with detectable CD39, only including participants without liver disease or cirrhosis*

Race of study participant	CD39 = 0 n = 279	CD39 > 0 n = 67
White	235 (79%)	64 (21%)
Black	44 (94%)	3 (6%)

*p = 0.026

Table S2. Dichotomized CD39 and Coronary Artery Calcium Score*

CAC score category	CD39 = 0 n = 298	CD39 > 0 n = 80
0 (n = 182)	150 (82%)	32 (18%)
1-99 (n = 50)	36 (72%)	14 (28%)
100-399 (n = 58)	46 (79%)	12 (21%)
400+ (n = 88)	66 (75%)	22 (25%)

*p = 0.31 based on Chi-squared test comparing CAC categories

CAC = coronary artery calcium

Table S3. Log-transformed CD73 and Coronary Artery Calcium Score*

CAC score category	Log-transformed CD73
0 (n = 178)	7.7 ± 0.8
1-99 (n = 48)	7.8 ± 1.1
100-399 (n = 59)	7.8 ± 1.0
400+ (n = 87)	7.8 ± 1.0
Overall (n = 372)	7.8 ± 0.9

*p = 0.77 based on ANOVA

CAC = coronary artery calcium

Table S4. Major findings of previous studies linking CD39/CD73 levels to age or disease state

Author(s) & Year published	Ectonucleotidase & directional change with age/disease	Methods of Quantification	Age categories (in years)	Major comorbidities in study population
Fang et al., 2016 ²⁰	CD39; increase with age	CD4+ T-cell CD39 mRNA transcript levels evaluated by qPCR	older: $65 \leq x \leq 85$ younger: $20 \leq x \leq 35$	None
Crooke et al., 2017 ²¹	CD39 and CD73; higher levels in older adults compared to middle and younger adults and centenarians	CD39 and CD73 mRNA transcript levels evaluated via qPCR	centenarian: $x \geq 100$ older: $x = 79^*$ middle-aged: $x = 45^*$ young: $x = 23^*$	None
Jalkanen et al., 2015 ²²	CD39 and CD73; lower CD39 activity associated with higher incidence disease; higher CD73 activity associated	nucleotidase serum activity levels were assessed using TLC and quantified via scintillation β -	older: $x \geq 60$	Stable peripheral artery disease of the lower extremities

	with increased atherosclerosis	counting		
Friedman et al., 2009 ⁴⁰	CD39; lower levels associated with higher incidence of disease	CD39 mRNA transcript levels of SNP variations evaluated via qPCR, both in the presence and absence of Crohn's disease	N/A	Crohn's disease
Guzman-Flores et al., 2015 ⁴¹	CD39 and CD73; CD39 expression increased with age and CD73 expression decreased with age and disease (T2D & obesity), in lymphocytes	CD39 and CD73 expression levels in total lymphocytes measured via flow cytometry	N/A	Type II diabetes, obesity
Chen et al., 2021 ⁶⁹	CD73; higher CD73 expression associated with worse diseased-state	CD73 expression levels were reported via an immunoreactive score calculated based on immunohistochemical staining	older: $x \geq 60$	Esophageal squamous cell carcinoma, esophagectomy

		results		
--	--	---------	--	--

*median ages of participants in this age bracket

SNP = single nucleotide polymorphism; TLC = thin-layer chromatography; T2D = type II diabetes