

# Brainstem Monitoring in the Neurocritical Care Unit: A Rationale for Real-Time, Automated Neurophysiological Monitoring

James L. Stone<sup>1,2,3</sup> · Julian E. Bailes<sup>1</sup> · Ahmed N. Hassan<sup>2</sup> · Brian Sindelar<sup>1,4</sup> · Vimal Patel<sup>1</sup> · John Fino<sup>2</sup>

Published online: 2 August 2016

© Springer Science+Business Media New York 2016

**Abstract** Patients with severe traumatic brain injury or large intracranial space-occupying lesions (spontaneous cerebral hemorrhage, infarction, or tumor) commonly present to the neurocritical care unit with an altered mental status. Many experience progressive stupor and coma from mass effects and transtentorial brain herniation compromising the ascending arousal (reticular activating) system. Yet, little progress has been made in the practicality of bedside, noninvasive, real-time, automated, neurophysiological brainstem, or cerebral hemispheric monitoring. In this critical review, we discuss the ascending arousal system, brain herniation, and shortcomings of our current management including the neurological exam, intracranial pressure monitoring, and neuroimaging. We present a rationale for the development of nurse-friendly—continuous, automated, and alarmed—evoked potential monitoring, based upon the clinical and experimental literature, advances in the prognostication of cerebral anoxia, and intraoperative neurophysiological monitoring.

**Keywords** Neurocritical care unit · Severe traumatic brain injury · Intracranial space-occupying lesions · Transtentorial herniation · Ascending arousal system · Intracranial pressure · Real-time, automated brainstem and cerebral monitoring · Somatosensory evoked potentials · Motor evoked potentials · Neuromonitoring

## Abbreviations

AAS	Ascending arousal (reticular activating) system
BAEP	Brainstem auditory evoked potential
BR	Blink reflex
CPP	Cerebral perfusion pressure
CSF	Cerebrospinal fluid
CT	Computed tomography
EMG	Electromyography
EP	Evoked potential
ICP	Intracranial pressure
IONM	Intraoperative neurophysiological monitoring
MAP	Mean arterial pressure
MBAEP	Modified forms of the BAEP
MEP	Motor evoked potentials
NCCU	Neurocritical care unit
SOL	Space-occupying lesion
SSEP	Short latency somatosensory evoked potentials
sTBI	Severe traumatic brain injury
TcE-MEP	Transcranial electrical motor evoked potentials
TcM-MEP	Transcranial magnetic motor evoked potentials
TrSSEP	Trigeminal short latency somatosensory evoked potential
TTH	Transtentorial herniation

✉ James L. Stone  
jlstone4@gmail.com

<sup>1</sup> Department of Neurosurgery, NorthShore University HealthSystem, Evanston, IL, USA

<sup>2</sup> Departments of Neurology and Neurological Surgery, University of Illinois at Chicago, Chicago, IL, USA

<sup>3</sup> Division of Neurosurgery, Department of Surgery, Cook County Stroger Hospital, Chicago, IL, USA

<sup>4</sup> Department of Neurosurgery, University of Florida, Gainesville, FL, USA

## Introduction

Patients in the neurocritical care unit (NCCU) often have an altered level of consciousness and require sedation, analgesia, and muscle paralysis for restlessness, pain, intubation/airway management, facilitating radiological studies, and controlling intracranial pressure (ICP). The neurological examination to establish—level of consciousness, pupils, extraocular motility, and extremity responses—may be limited to brief periods of reduced or withheld medications. Often without apparent warning—or following carbon dioxide retention, a seizure, vigorous tracheal suctioning, diminished venous outflow, or ventricular system blockage—ominous findings occur such as onset of coma, pupillary changes, and motor posturing [1–6].

Lethargy, hypersomnolence, stupor, and coma secondary to space-occupying lesions (SOLs), usually indicate involvement of the ascending arousal (ascending reticular activating) system (AAS) [6–11]. SOLs are commonly associated with adjacent vasogenic edema, hyperemia, and ischemia. Compensation occurs by displacement of cerebrospinal fluid (CSF) and venous blood, and distortion of the brain parenchyma [12, 13]. As these mechanisms fail, precipitous brain tissue displacements, secondarily increased ICP, lateral midbrain shifts (uncal herniation) and/or downward herniation through the tentorial opening (transtentorial herniation, TTH) ensues. Without early detection, life threatening rostrocaudal deterioration and severe disability or death result from deep hemispheric shifts with ischemic injury, and/or secondary brainstem infarctions and hemorrhages [8–11]. Indeed, TTH is among the most emergent situations encountered in clinical medicine [4, 6, 8, 11, 14–16].

Various limitations and pitfalls exist in our present reliance on ICP, cerebral perfusion pressure (CPP), and neuroimaging [3, 6, 17, 18]. Reliable physiological or functional information is needed for more timely treatment. We advocate the use of short latency sensory and motor evoked potential modalities (EPs) in close proximity to the AAS in the upper pons, midbrain, and diencephalic region [19–22]. These monitoring tools, and possibly several others, can be automated for practical, real-time, nurse-friendly application within the NCCU.

‘Real-time assessment of global or regional brain dysfunction could help clinicians recognize early worsening, prompt specific management changes, monitor response to therapy...(and)...used as surrogate endpoints in clinical trials [23].’ Such monitoring would augment our present treatments and be utilized in patients not ordinarily considered for invasive ICP monitoring—such as somnolent patients with moderate head injuries, cerebral infarctions, and hemorrhages without rupture into the ventricle. This

review is intended to stimulate development of bedside nurse-friendly, automated electrophysiological monitoring for these challenging patients.

## The Ascending Arousal System and Transtentorial Herniation

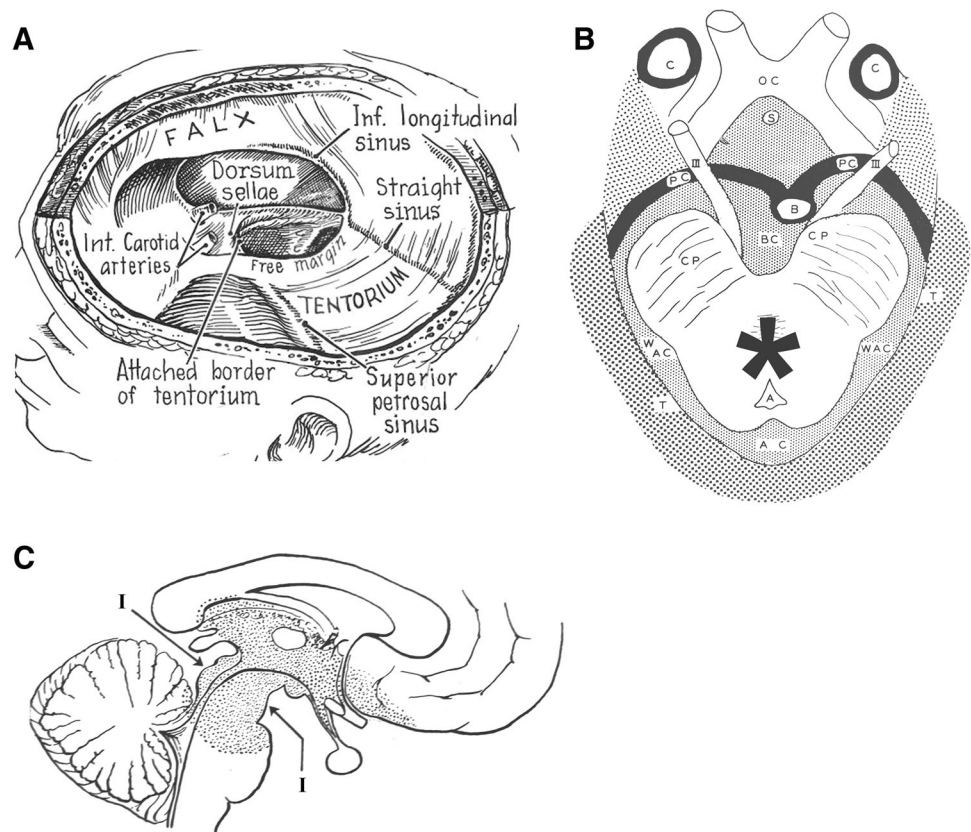
Critical structures enclosed within the incisural plane or hiatus of the tentorium cerebelli are depicted in Fig. 1a–c [4, 14, 16]. These include the midbrain nuclei and fibers of the AAS which modulates cerebral cortical activity in the maintenance of vigilance and consciousness [9, 24]. Additional causes of coma consist of bilateral or central AAS lesions, or their thalamic targets, and portions of the hypothalamus and basal forebrain (Figs. 1c, 2) [7–11, 16, 19, 25]. Upward projecting AAS nuclei and major fiber bundles such as the central tegmental tract, being aligned parallel to the long axis of the brainstem (Fig. 2), may make the system particularly vulnerable to perpendicular or lateral bending forces.

TTH is dependent upon SOL volume, rate of radial expansion, vector force, and the presence or absence of cerebral atrophy [7, 8, 26–35]. Central herniation directs the mesodiencephalic region more caudally than laterally [7, 8, 10, 11, 36–39]. Earlier stages of TTH typically show mesial displacement of the basal hemisphere at or just above the tentorium, with widening of the ipsilateral ambient cistern (Figs. 3, 4) [36]. Accompanying this displacement is transposition of the attached midbrain, whose peduncle may become compressed or ‘notched’ against the contralateral rigid tentorial edge, causing hemiparesis ipsilateral to the SOL—the Kernohan-Woltman Phenomena (Figs. 3, 4, 5) [36, 40].

About half of SOL patients with TTH have only horizontal midbrain displacement and the other half downward shift of the midbrain tectum [4, 41–43]. Stupor and coma more closely correlate with a 6- to 13-mm midline hemispheric shift than vertical descent [44]. And effacement or closure of the perimesencephalic (ambient) cisterns a worsened outcome as well [45, 46]. At times, brainstem ischemia can be a significant factor without clear evidence of mechanical herniation or brainstem hemorrhage [36, 47–52].

The sudden and unpredictable nature of TTH is likely influenced by the extreme anatomic variability of the incisural length and width, midbrain proximity to the tentorial edge (0–7 mm), and oculomotor nerve distances [14, 16, 36, 53, 54]. Preferably, surgical decompressions should be performed at the early diencephalic (drowsy or stuporous) phase; but with loss of the neurological exam, and without a clear physiological indicator, much variability remains [46, 55–59].

**Fig. 1** **a** Drawing depicting the incisural opening of the tentorium cerebelli for passage of the midbrain. Note adjacent structures (from Finney and Walker 1962 with permission) [14]. **b** Cross section of the midbrain within the incisural plane as viewed from below. *A*—aqueduct of Sylvius, *B*—basilar artery, *BC*—basal cistern, *C*—internal carotid arteries, *CP*—cerebral peduncles, *OC*—optic chiasm, *PC*—posterior cerebral arteries, *S*—stalk of pituitary, *T*—tentorium cerebelli, *WAC*—wings of the ambient cistern. Large asterisk approximates the midbrain (reticular) ascending arousal system (AAS) (adapted from Walker 1969 with permission) [16]. **c** Sagittal section, stippling represents the predominant areas most related to preservation of consciousness. ‘I’—approximates the incisural plane (from Jefferson 1958 with permission) [26]



## Intracranial Pressure (ICP) and Related Issues

‘The pathogenesis of signs and symptoms of an expanding mass lesion that causes coma is rarely a function of the increase in ICP itself, but usually results from imbalances of pressure between different (intradural) compartments leading to tissue herniation.’ [8, p. 95].

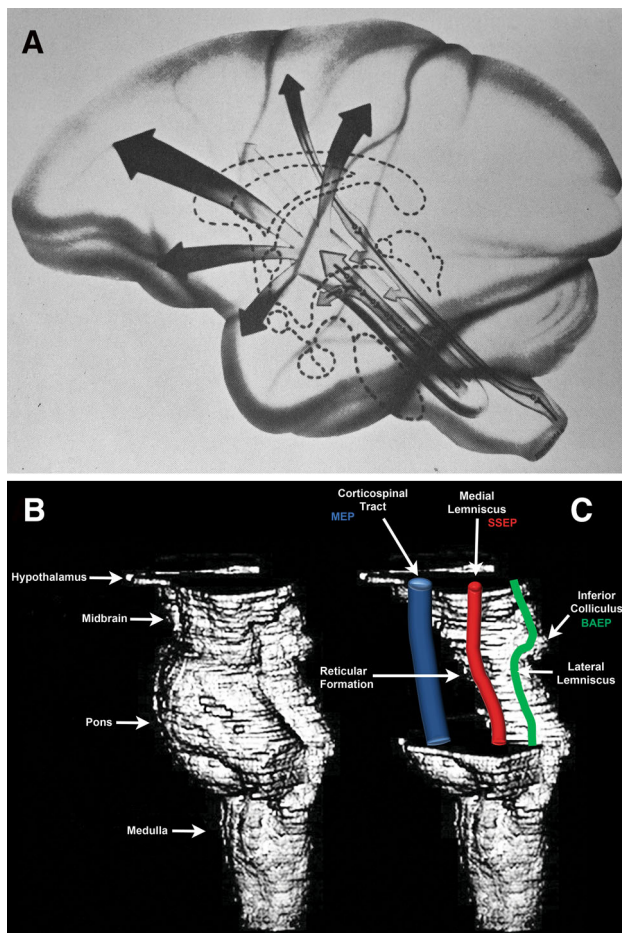
The intracranial contents largely consist of inhomogeneous brain tissue, liquid blood, and CSF [60–63]. The largest component—brain is a deformable viscoelastic structure, which exhibits properties characteristic of a solid as well as a fluid [12, 13, 61, 64–71]. This property predisposes to ‘shear stresses’ resulting in ‘pressure differentials’ or ‘gradients’ between regions of increased and decreased tension within the brain. Gradients are frequent in the vicinity of compressed or deformed brain tissue but also occur contralateral and across compartments predisposed to TTH and foramen magnum herniation [61, 64–66]. Experimental intracranial mass expansion results in the immediate appearance and elevation of such gradients, with quick reversal by decompression [28, 30–32, 34, 64, 65, 72–74]. Gradients have also been detected in SOL patients with multiple ICP monitors [17, 29, 75–82].

Due to the risk of causing clinical deterioration and TTH, ICP devices are customarily placed contralateral to

the SOL. Thus, ICP underestimates distant gradient effects, but also the effects of brain turgor and compliance adjacent to the mass lesion or deeper, or how fast deadly processes may be occurring [60, 67, 70, 71, 82–84]. Consequently, ICP devices must be suspect of measuring a local as opposed to a generalized, broadly applied value like systemic arterial pressure. In addition, midline shifts may lead to falsely low ICP values by narrowing third ventricular width, impeding CSF egress, thus transmission of ICP to the contralateral ventricular monitor [72]. Acute mass lesions in the temporal or posterior fossa have progressed to somnolence and herniation even with ICP levels recorded at 20 mm Hg [18].

Cerebral perfusion pressure (CCP) has been helpful in management of patients with increased ICP to avert hypotension [62, 84]. Either hypotension—reduced mean arterial pressure (MAP) or increased ICP (without an associated increase in MAP)—results in decreased cerebral perfusion pressure (CPP = MAP minus ICP) [83–86]. However, the CPP is critically dependent upon the ICP value and, like ICP, only can be relied upon to reflect the value near the monitor’s tip [18].

There is certainty that refractory ICP is detrimental, and an initial ICP value greater than 20 mm Hg is associated with poor outcome after severe traumatic brain injury (sTBI, Glasgow Coma Score  $\leq 8$ ) [18, 46, 62, 84]. In



**Fig. 2** **a** Classic depiction of the brainstem ascending (reticular) arousal system (AAS) activating higher central nervous system centers resulting in arousal (from Magoun 1954 with permission) [25]. **b** Model of the brainstem and hypothalamus prepared from thinly cut human material, and sub-millimeter MRI images from a number of patients with brainstem coma. **c** A ‘cutaway model’ depicting the volume of the midbrain and pons that constitutes the reticular formation of the AAS (designated with *long horizontal arrow*). Superimposed in proximity are the corticospinal tract, medial lemniscus, and lateral lemniscus/inferior colliculus—responsible for the respective evoked potential responses—MEP, SSEP, and BAEP (adapted from Parvizi and Damasio 2004 with permission) [9]

addition, ICP per se is not a useful indicator for a functional outcome [6, 62, 86]. Recent consensus holds that ICP/ CPP-directed monitoring and adherence to guidelines leads to overall improved attention and management of the sTBI patient (efficiency of care). However, due to a number of factors such as a baseline high mortality, variability in ICP devices, a lack of standards for recording ICP values, uncertainty of individual ICP threshold values, as well as the ability to effectively control ICP, outcome remains similar to those managed without ICP monitoring [17, 62, 85–94].

A large, prospective, double-blind, sTBI study analyzed secondary pupillary or motor score deterioration and

argued a critical need to identify patients at particular risk [46]. Another prospective study examined patients with a large-volume cerebral hemispheric infarction who underwent decompressive hemicraniectomy [55]. This study made a strong case that ICP monitoring was unreliable, thalamic/brainstem shift on imaging studies held more importance, and earlier recognition was essential to preserve life and a functional recovery [55]. Perhaps more reliable or different information is needed than can be presently obtained from ICP monitoring [17, 95, 96].

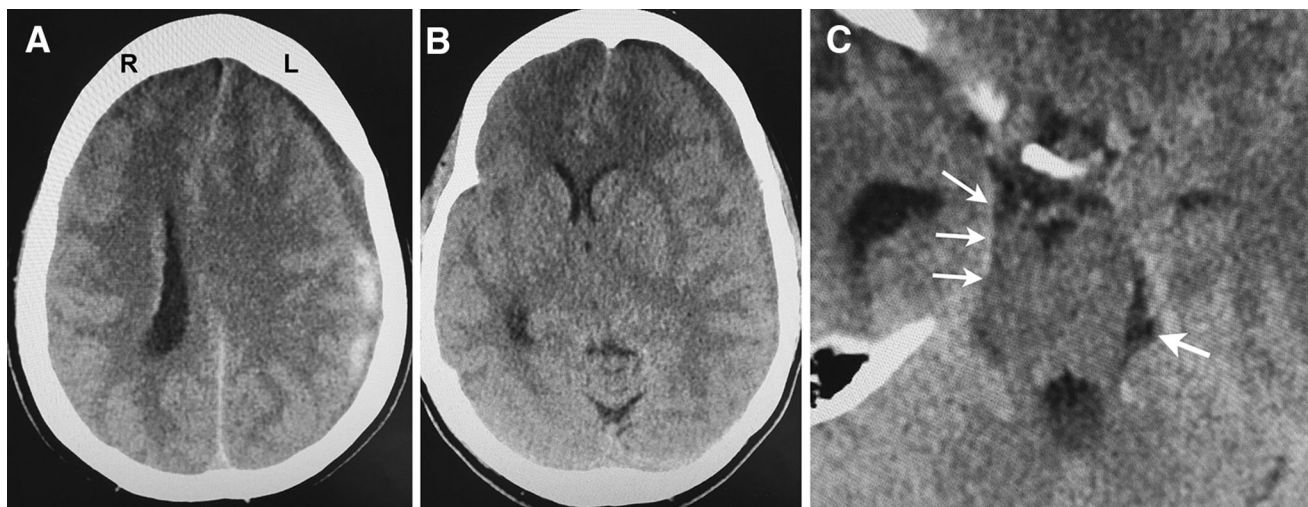
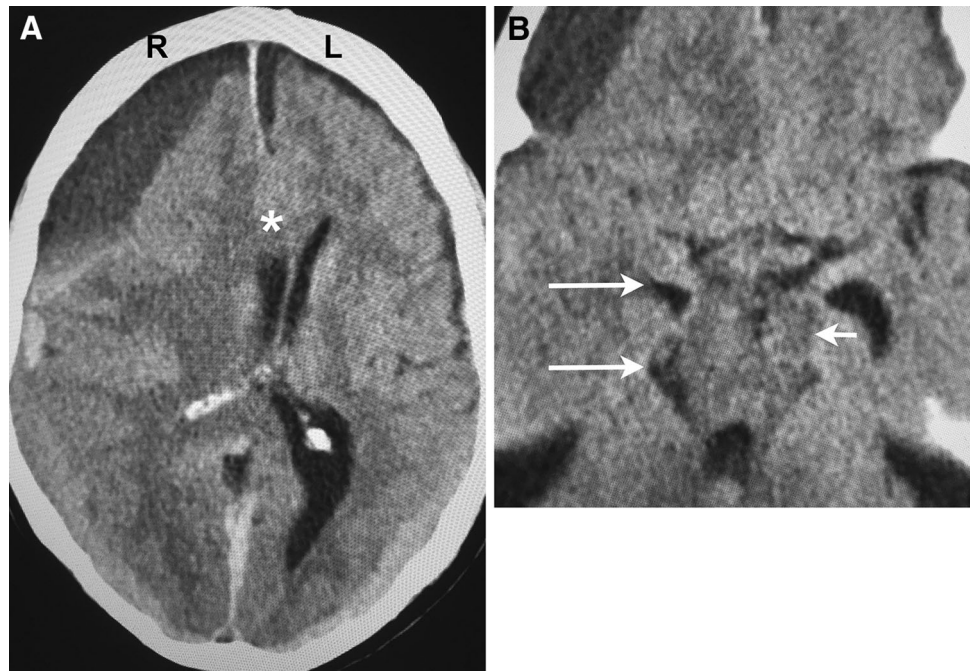
## Neuroimaging

Neuroimaging provides essential structural-anatomic detail for acute diagnosis and emergency medical and surgical treatments of hemorrhages, hydrocephalus, tumors, and infarctions [6]. However, clear imaging of brain tissue impacted by deep mass effects and the propensity for identifying early TTH is limited (see above ‘The Ascending Arousal System and Transtentorial Herniation’ section). In patients without pupillary findings or motor posturing, the ICP value usually carries more importance than the significance of nonhemorrhagic CT findings such as increased edema, midline shift, or basal cistern obliteration. In stable patients, MRI may be performed and coronal images are particularly revealing (Fig. 5b) compared to axial CT and coronal reconstructions. Unfortunately, these patients are often unstable and transporting them may be problematic [6]. Laying these patients flat and moving them into and out of scanners not infrequently leads to stubborn increases in ICP and monitor malfunction or dislodgement, including other mounted monitoring devices. We believe functional or physiological bedside information is necessary for more timely treatment of TTH.

## Neurophysiological Monitoring at the Incisural Plane: A Review of Related Clinical and Experimental Research

Neurophysiological monitoring ‘allows for real-time assessment of neurologic integrity, signal transmission, and secondary processing of sensory information... Signal changes can be measured over time... to detect responses to therapeutic interventions, recovery of brain function, or progression of injury’ [97]. For decades, somatosensory evoked potentials (EPs) have been used as indicators to assess stroke and sTBI patients [98–104]. Positive clinical correlations were found, and advantages of serial EP recordings were realized [98, 103–106]. Reviews were optimistic regarding the future use of somatosensory EPs in

**Fig. 3** Eighty-three-year-old male with dementia and falls presented with extreme lethargy and right-sided weakness. **a** Plain CT scan of the brain showed a large right, mixed-density, chronic, subdural hematoma with marked subfalcine herniation (*asterisk*) and pronounced *right to left* midline shift. **b** Basal image of the CT scan shows medial displacement of the right temporal horn (*upper arrow*) and widening of the right ambient cistern (*lower arrow*). The left midbrain cerebral peduncle appears compressed (*short arrow*) and the adjacent left temporal horn dilated. Despite timely surgical decompression, the patient later expired from medical complications

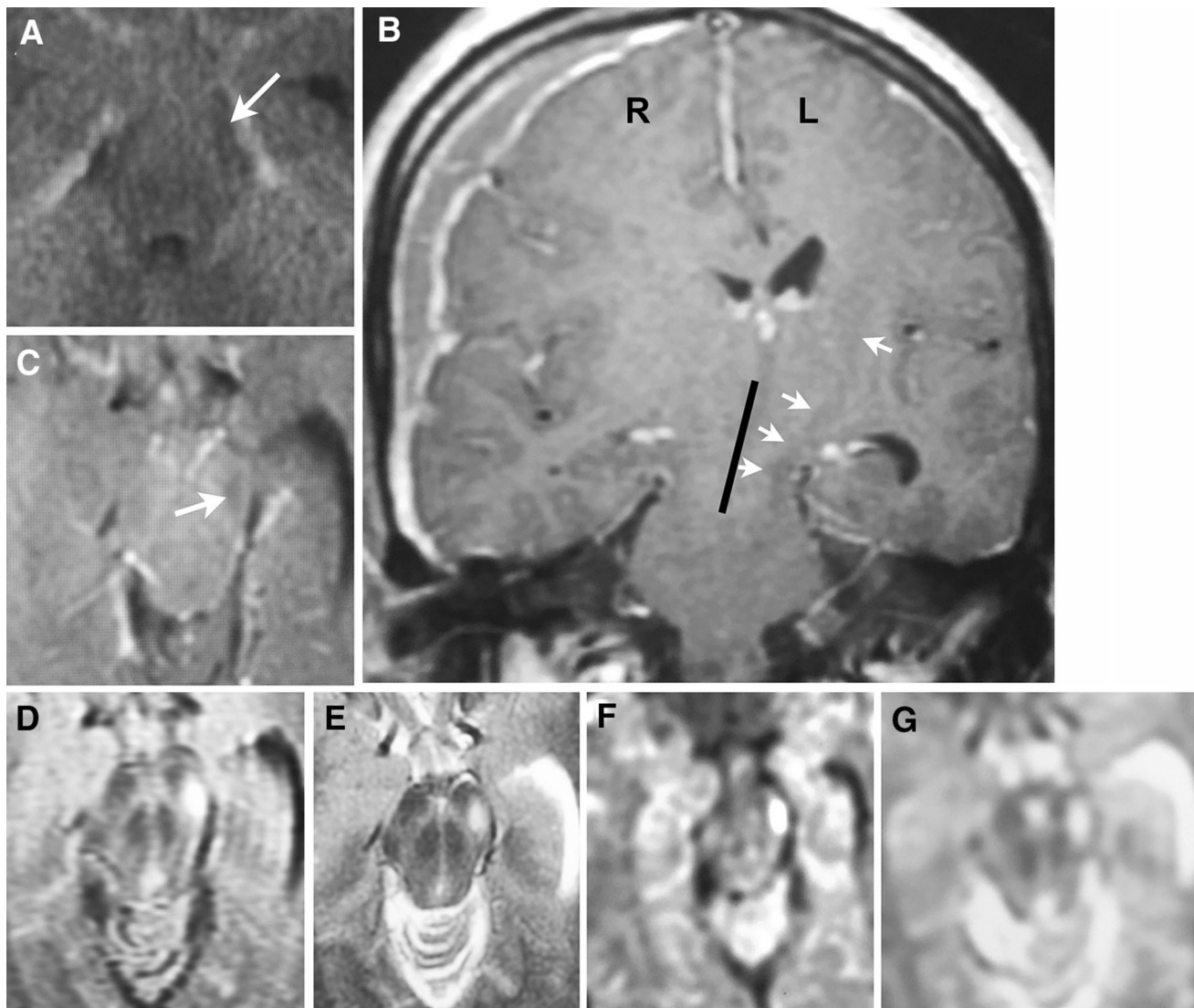


**Fig. 4** Fifty-one-year-old nurse with progressive headaches requiring aspirin, recent vomiting, and lethargy. She slept when left alone (somnolent), opened her eyes (but not fully) after moderate stimuli, was oriented to person, place and time, and followed commands. Glasgow Coma Score was deceptive at 14. She had drift of the left outstretched arm and a right Babinski sign. **a** Plain CT scan showed a large left subacute-appearing subdural hematoma, obliteration of much of the left lateral ventricle, and approximately 1.5-cm midline shift. **b, c** A trapped, dilated right temporal horn was present; and the right midbrain cerebral peduncle appeared effaced by the right

tentorial edge (*arrows*). The left ambient cistern was enlarged (*left arrow*). Due to the alteration in consciousness, mass effect, and radiographic appearance of incipient TTH, she was given mannitol and platelets, promptly taken to the operating room for intubation, and a left frontotemporoparietal craniotomy was performed. Neurological deficits resolved and she returned to work as a nurse about 6 months later

coma and TTH, but advances in neuroimaging and adoption of ICP/ CPP-directed care led to less use of EPs in the NCCU [86, 97–99, 101, 107]. More recent NCCU studies and improved EP technology have significantly confirmed and extended the usefulness of EPs in these patients [97, 100, 108–118].

In accordance with recent literature on neurocritical care, the ideal monitor provides bedside, noninvasive, real-time, user-friendly advanced data analysis to improve treatment [86, 97, 118]. We believe some of the neurophysiological modalities described in this section could be adapted to fulfill this present void in our NCCU care.



**Fig. 5** Thirty-six-year-old male was beaten up, found moderately lethargic (stuporous), only intermittently followed commands, and Glasgow Coma Score was 10. A right hemiparesis was present. **a** CT scan of the brain disclosed a large right-sided subdural hematoma with midline shift, a right basifrontal hemorrhagic contusion, and smaller bitemporal contusions. The basal cisterns showed an abundance of subarachnoid hemorrhage and a possible area of low density in the left midbrain cerebral peduncle (*arrow*). Due to the prominent subarachnoid hemorrhage as well as weakness ipsilateral to the subdural, MRI/MRA of the brain was obtained and no vascular lesions were detected. **b** MRA—*coronal* view shows the large subdural, and right to left shift with angulation of the diencephalic-midbrain region (*solid line*). An altered signal intensity (*white*

*arrows*) believed to represent insult to the left corticospinal (*pyramidal*) tract was evident in the hemisphere and upper brainstem. The midbrain was well visualized and T1 axial (MRA) image (**e**) showed a faint low density within the left midbrain peduncle (*arrow*). A very bright, white signal was noted in the axial left midbrain peduncle on Flair (**d**), T2 (**e**), DWI (**f**), and ADC (**g**) images. The findings are not considered compatible with an ischemic event but rather a non-hemorrhagic bruise or damage to the densely packed motor fibers of the corticospinal (*pyramidal*) tract. Increased lethargy and slight right arm extensor posturing prompted emergent right-sided craniotomy for evacuation of the subdural and basifrontal hematoma. After extensive rehabilitation, he remained moderately disabled due to right-sided weakness and cognitive problems

### The Blink Reflex (BR)

The blink reflex (BR), used to indicate facial nerve integrity, is largely a pontine reflex resulting in momentary, bilateral eye closure (orbicularis oculi muscles) [119, 120]. A brief stimulus applied to the supraorbital branch of the

trigeminal nerve results in an early ipsilateral oligosynaptic orbicularis oculi muscle action potential (R1-10 ms), and a later bilateral multisynaptic response (R2-20 ms) interfaces with the pontine reticular formation before synapsing in bilateral facial motor nuclei [120, 121]. In patients with STBI or cerebrovascular lesions affecting the cerebral

cortex, basal ganglia, and deep cerebral regions, R2 is absent or suppressed in the early days after the insult [119, 122–125]. Perhaps the polysynaptic connections of R2 make this waveform of increased sensitivity to cephalad or mesodiencephalic reticular influences. A study in cats of increased ICP, which led to mesodiencephalic ischemia, was associated with the disappearance of R2 and preservation of R1 [126]. Although known to be suppressed by sleep and sedatives, the BR can now be recorded under general anesthesia [127–130]. Less-studied reflexes utilize the jaw jerk (masseteric reflex) believed to invoke the mesencephalic trigeminal nucleus and could have future clinical applications [120, 128].

### Upper Extremity Short Latency Somatosensory Evoked Potentials (SSEP)

The SSEP provides an ‘objective test for cortical reactivity to external stimulation—a measure of responsiveness to the outside world and *sine qua non* of consciousness’ [97]. The upper extremity SSEP is derived secondary to median nerve stimulation in the upper extremity. The signal is recorded at the brachial plexus (Erb’s point), carried in the ipsilateral posterior column, and a cervical electrode (N13) indicates spinal cord entry or the cervicomedullary region. After a synapse in the cuneatus nucleus, the tract crosses to become the contralateral medial lemniscus, which ascends the medulla, pons, and midbrain before a synapse in the ventroposterolateral nucleus of the thalamus with relay to the postcentral gyrus. Here, the major cortically generated peak is N20, recorded from the central regions of the scalp C3’ and C4’ (Fig. 2c) [99, 131–138].

SSEP responses are durable, and widely used in continuous intraoperative neurophysiological monitoring (IONM), where an N20 latency increase of 10 %, or 50 % diminished amplitude from the patient’s presurgical baseline is considered a significant change and the surgical team notified [131, 132, 139, 140]. Recently, SSEPs have become very useful in the NCCU as an important prognostic indicator after cerebral anoxia. Bilateral absence of N20 after cardiac arrest is associated with persistent vegetative state or death in all patients [112, 113, 118, 138, 141–143]. In a meta-analysis, SSEP as a single prognostic marker of both good and bad outcomes after sTBI performed better than pupillary response, Glasgow Coma Scale, CT findings, and EEG [113]. Recent work suggests amplitude values of a preserved N20 and later cortical N35 peak may relate to outcome quality, further increasing prognostic accuracy [144].

### Trigeminal Short Latency Somatosensory Evoked Potentials (TrSSEPs)

Of possible interest in regard to NCCU monitoring are the TrSSEPs [145, 146]. Electrical stimulation is applied to the lower lip, and scalp recording of a potential at 20 ms is obtained from positions just lateral to those used for upper extremity SSEPs (C5’, C6’) [147, 148]. A latency difference between sides of greater than 1 ms or diminished amplitude greater than 50 % is considered abnormal [149]. Having been used largely in the dental/maxillofacial fields under general anesthesia, TrSSEPs may prove of value in NCCU monitoring [148, 150].

### Brainstem Auditory Evoked Potentials (BAEP)

The BAEP is a subcortical response generated by the cochlear and brainstem auditory pathways. Each ear is stimulated separately in response to moderately loud, short-duration click stimuli delivered thru soft insert earphones. The waveform response is recorded from the vertex (Cz) or frontal scalp (Fz) and generally appears within 10 ms after stimulation. Wave I—Auditory nerve, Wave III—Superior olivary nucleus (pons), and Waves V and Vn—Inferior colliculus (midbrain, tentorial incisura) are measured for their latency and amplitude [151–155]. This robust response is refractory to level of consciousness, medications, general anesthesia, or muscle paralyzing agents [98, 139, 152–155]. Like the SSEP, threshold abnormalities of the major peaks during IONM are a 10 % latency increase or 50 % amplitude decrease from baseline [131, 135, 139, 152, 154, 156]. Nurse-friendly neonatal hearing screening with rapid automated interpretation utilizing BAEP Wave V is at present extensively performed [152, 157].

BAEP studies in patients with herniation syndromes have shown correlations with ICP, pupillary changes, TTH, and outcome. Timely decompressive treatments in a number of these studies resulted in rapid clinical and electrophysiological improvement [115–117, 158–168].

Faster rates of BAEP stimulation worsen BAEP abnormalities, and modified forms of the BAEP (MBAEP) may increase the diagnostic utility of the test [98, 155, 169, 170]. Notable MBAEP changes were found in normal volunteers placed in a 10°–15° downward head position to simulate increased ICP [155] and patients symptomatic of mild to moderately increased ICP with mass effects and shifts from large, slow-growing cerebral SOLs (mostly tumors) [171, 172]. In the patients, significant MBAEP changes normalized when re-tested after surgical excision [171, 172].

## Upper Extremity Motor Evoked Potentials (MEP)

Upper extremity MEPs are obtained during IONM in neurosurgical patients undergoing operations under general anesthesia, or on awake patients, in response to respective transcranial electrical (TcE-MEP) or transcranial magnetic (TcM-MEP) stimulation over the central regions (C3, C4) [173, 174]. Following such pyramidal/corticospinal tract stimulation, the signal is recorded from extremity muscles by needle or surface EMG. Although the responses are more variable than the sensory EPs, a 50 % or greater loss of amplitude is concerning [173–175]. In conscious patients, TcM-MEP has found much interest in clinical neurology to assess central motor pathways [176, 177]. TcM-MEP is delivered by a very well tolerated, locally applied cap-like coil MEP [176, 177]. Repetitive TcM-MEP techniques lead to intracortical motor facilitation and have improved the motor response [178–180].

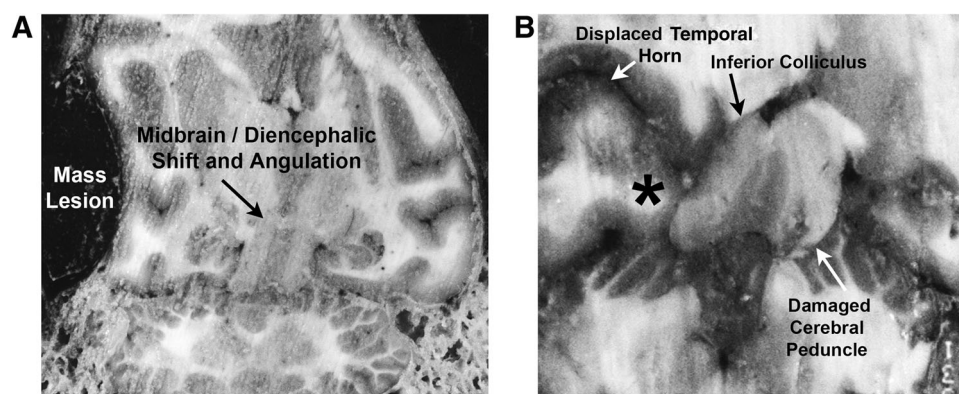
Recent clinical reports on the Kernohan notch syndrome (Figs. 3, 4, 5) have included MEP studies on patients with cerebral mass lesions and TTH [181–190]. Upper extremity TcM-MEP was performed in a small group of patients with intracerebral hematomas, altered mental status, and pyramidal tract involvement [190]. The presence of any MEP response indicated a compressed but not destroyed pyramidal tract with propensity for recovery [190].

## Herniation Research Using Multiple Modalities

A number of years ago our group studied anatomically confirmed TTH in the cat and monkey—indicated by sudden pupillary dilatation or midposition fixation—secondary to an expanding temporal extradural balloon. TTH in the cat over a

2-h period was monitored with ICP, BAEP, upper extremity SSEP, and in some MEP [191–193]. Cats showed significant BAEP/SSEP abnormalities from baseline, and the prominent warning sign just before TTH was a 30 % drop in BAEP Wave V amplitude. Complete Wave V flattening, caudal displacement of the inferior colliculus [191, 192], and a marked drop in inferior colliculus blood flow were tightly correlated with TTH [193]. A unilateral MEP loss was considered hemispheric compression, whereas bilateral MEP loss, especially if associated with significant changes in BAEP Wave V, heralded the beginning of TTH [194].

A 4-h primate model of TTH [195] was patterned after classic studies decades earlier [27, 28, 33]. Four hours was chosen to simulate the frequently encountered SOL, acute subdural hematoma with a similar delay allowing some compensatory mechanisms to occur [196]. Twelve macaque monkeys underwent gradual expansion of an extradural balloon, which at 10 % of brain volume led to the precipitous onset of bilaterally dilated or less often midposition fixed pupils (Fig. 6a, b) [195]. Two hours before TTH, a transient rise in ICP followed each inflation, as did systolic hypertension and bradycardia (Cushing reflex). ICP gave significant forewarning of TTH 1 h before (16 mm Hg,  $p < 0.05$ ), one-half hour before (23 mmHg,  $p < 0.01$ ), and at TBH (44 mm Hg,  $p < 0.001$ ). Compared to ICP, Wave V of the BAEP gave an identically significant warning (25 % amplitude depression) 1 h before TTH, further Wave V amplitude depression being a more statistically significant warning than the ICP one-half hour before ( $p < .001$ ), and a similarly significant severe Wave V flattening or loss at TTH ( $p < .001$ ) [195]. Upper extremity SSEP showed depressed amplitude one-half hour before TTH ( $p < .05$ ) and near loss or absence of SSEP at TTH ( $p < .01$ ) [195].



**Fig. 6** **a** Frontal/coronal section of the non-human primate experimental TTH study depicting the right-sided (*balloon*) mass and its effect on the hemisphere at the time of TTH. Note the diencephalic/midbrain angulation. **b** Magnified view of the displaced temporal horn and herniated right temporal lobe (*asterisk*) at the time of TTH. Note

the displaced inferior colliculus and damage to the contralateral cerebral peduncle. This overall appearance closely models the TTH situation in patients (adapted from Stone et al. 1990 with permission) [195]



## The Future

Monitoring in the NCCU (SSEP, BAEP, TcM-MEP) must be nurse friendly, preferably continuous (or nearly so), with automation of threshold breaches (similar to those used in IONM) leading to alarms. Nurses, residents, fellows, or technicians are easily taught scalp surface electrode placement for stimulation and recording (International 10-20 system). Technical requirements such as adequate noninvasive afferent signal delivery, grounding, electrode impedance, artifact rejection, advanced filtering, and other concerns have much improved in recent decades [97, 109–111, 115].

## Conclusion

This focused review of the neurocritical care and related literature has emphasized a number of shortcomings in our current ICP/neuroimaging-directed management of patients with an altered level of consciousness secondary to intracranial mass lesions. We elected to concentrate on noninvasive bedside electrophysiological monitoring tools that we believe could substantially improve patient management.

Data collected over decades on stuporous or comatose NCCU patients is highly supportive of the continuous monitoring of sensory and motor EPs, as well as the development of nurse-friendly, automated devices to monitor brain function. Structured clinical trials correlating neurophysiological changes in the early and later phases of TTH would appear to be a fertile field to investigate and develop. Improving the neurological and neurosurgical care of these challenging conditions by more timely recognition and intervention is of obvious importance, and technology to do so may well be at hand.

**Acknowledgments** This work is dedicated to Dr. Robert A. Moody, a modern pioneer in the aggressive approach to cerebral decompression and ICP monitoring. In 1975, Dr. Moody established an experimental brain herniation laboratory at Chicago's Cook County Hospital Hektoen Laboratory with the foresight to utilize the recently developed evoked potential responses. There, the senior author was stimulated to conduct experimental work. Special thanks to Dr. Jerome B. Posner of New York City, noted authority on 'Stupor and Coma' who graciously reviewed a draft of the manuscript, which he found of much interest, and met with the senior author to review the figures. We remain particularly grateful to Dr. John R. Hughes, our longtime mentor in clinical neurophysiology at the University of Illinois in Chicago (UIC). Dr. Gleb Gorelick, neuroradiologist at Advocate Illinois Masonic Hospital in Chicago assisted in interpretation of the MRI images, and Dr. Ankit Mehta, from the UIC Department of Neurosurgery, reviewed the manuscript and provided input. Finally, we thank our active clinical neurophysiology and intraoperative monitoring teams at both the UIC and Evanston/North Shore University Hospital for their continued enthusiasm and support.

## Compliance with Ethical Standards

**Conflict of interest** James L. Stone: Natus/Biologic provided evoked potential instrumentation and some technical support of work presented in this review. Patent issued to inventor JLS on 01/12/2014 and assigned and owned by the University of Illinois. Title: NonInvasive, Bedside Intra-Cranial Pressure And Brain Shift/Herniation Monitoring Unit Utilizing Early OnSet Auditory Evoked Responses. A University of Illinois start-up cooperation (Remote Vital Monitoring) has been formed and is in the early stages of development. Outside funding has not been obtained. JLS has 53 % stock ownership in this corporation. John Fino: A University of Illinois start-up corporation (Remote Vital Monitoring) has been formed and is in the early stages of development. Outside funding has not been secured. JF is a minor shareholder (5 % stock) in this corporation. Julian E. Bailes, Ahmed N. Hassan, Brian Sindelar, and Vimal Patel declare that they have no conflict of interest.

## References

- Ropper AH, Gress DR, Diringner MN, Green DM, Mayer SA, Bleck TP. Introduction to critical care in neurology and neurosurgery, management of intracranial hypertension and mass effect, electrophysiologic monitoring in the neurological intensive care unit. In: Ropper AH, Gress DR, Diringner MN, Green DM, Mayer SA, Bleck TP, editors. Neurological and neurosurgical intensive care. 4th ed. Philadelphia: Lippincott Williams & Wilkins; 2004. p. 3–11, 26–51, 129–53.
- Wijdicks EFM. Agitation and pain, intracranial pressure, monitoring devices, and diagnostic procedures. In: Wijdicks EFM, editor. The clinical practice of critical care neurology. 2nd ed. New York: Oxford University Press. 2003. p. 25–37, 107–81.
- LeRoux PD, Levine JM, Kofke WA, editors. Monitoring in neurocritical care. Philadelphia: Elsevier-Saunders; 2013.
- Andrews BT. the recognition and management of cerebral herniation syndromes. In: Loftus C, editor. Neurosurgical emergencies. 2nd ed. New York: Thieme; 2008. p. 34–44.
- Smith NJ, van Gils M, Prior P. Neurophysiological monitoring during intensive care and surgery. New York: Elsevier; 2006. p. 125–45.
- Stevens RD, Shoykhet M, Cadena R. Emergency neurological life support: intracranial hypertension and herniation. Neurocrit Care. 2015;23(Suppl 2):S76–82.
- Plum F, Posner JB, editors. The diagnosis of stupor and coma. 3rd ed. Philadelphia: F.A. Davis Co.; 1980.
- Posner JB, Saper CB, Schiff ND, Plum F. Plum and Posner's diagnosis of stupor and coma. 4th ed. New York: Oxford University Press; 2007.
- Parvizi J, Damasio AR. Neurochemical correlates of brainstem coma. Brain. 2003;126:1524–36.
- McNealy DE, Plum F. Brainstem dysfunction with supratentorial mass lesions. Arch Neurol. 1962;7:10–32.
- Wijdicks EFM. The comatose patient. 2nd ed. New York: Oxford University Press; 2014. p. 183–4.
- Davson H, Segal MB. Physiology of the CSF and blood-brain barriers. Boca Raton: CRC Press; 1996. p. 708–12.
- Hakim S, Venegas JG, Burton JD. The physics of the cranial cavity, hydrocephalus and normal pressure hydrocephalus. Surg Neurol. 1976;5:187–210.
- Finney LA, Walker AE. Transtentorial herniation. Springfield: Charles C. Thomas; 1962.
- Maramattom BV, Wijdicks EFM. Uncal herniation. Arch Neurol. 2005;62:1932–5.

16. Walker AE. Brain herniations. In: Vinken PJ, Bruyn GW, editors. *Handbook of clinical neurology. Disturbances of nervous function, vol. 1.* Amsterdam: North Holland; 1969. p. 550–73.
17. Hawthorne C, Piper I. Monitoring of intracranial pressure in patients with traumatic brain injury. *Front Neurol* (July). 2014; 5 (Article 121):1–16.
18. Robertson C, Rangel-Castilla L. Critical care management of traumatic brain injury. In: Winn HR, editor. *Youmans neuro-logical surgery. Trauma, vol. 4.* Philadelphia: Elsevier; 2014. p. 3397–423.
19. Brodal P. *The central nervous system. Structure and function.* 2nd ed. New York: Oxford University Press; 1998. p. 421–43.
20. Haines DE. *Neuroanatomy. An atlas of structures, sections, and systems.* 6th ed. Philadelphia: Lippincott Williams & Wilkins; 2004.
21. Kretschmann HJ, Weinrich W. Neurofunctional systems. 3D reconstructions with correlated neuroimaging. *Stuttgart: Thieme, 1998:42–63, 79–88.*
22. Woolsey TA, Hanaway J, Gado MH. *The brain atlas. A visual guide to the human central nervous system.* 3rd ed. Hoboken: John; 2008. p. 183, 189, 197, 201.
23. Wijdicks EFM, Rabinstein AA. Critical care neurology: five new things. *Neurol Clin Pract.* 2011;1:34–40.
24. Sherman SM, Guillery RW. Functional connections of cortical areas. A new view from the thalamus. *Cambridge: MIT Press; 2013. p. 4–6, 80–2.*
25. Magoun HW. The ascending reticular activating system and wakefulness. In: Delafresnaye JF, editor. *Brain mechanisms and consciousness—a symposium.* Oxford: Blackwell; 1954. p. 1–20.
26. Jefferson G. The reticular formation and clinical neurology. In: Jasper HH, Proctor LD, Knighton RS, Noshay WC, Costello RT, editors. *Reticular formation of the brain.* Henry Ford Hospital international symposium. Boston: Little Brown & Co; 1958. p. 729–44.
27. Reid WL, Cone WV. The mechanism of fixed dilatation of the pupil resulting from ipsilateral cerebral compression. *JAMA.* 1939;112:2030–4.
28. Jennett WB, Stern WE. Tentorial herniation, the midbrain and the pupil. *J Neurosurg.* 1960;17:598–609.
29. Kaufmann GE, Clark K. Continuous simultaneous monitoring of intraventricular and cervical subarachnoid cerebrospinal fluid pressure to indicate development of cerebral or tonsillar herniation. *J Neurosurg.* 1970;33:145–50.
30. Langfitt TW. Pathophysiology of increased ICP. In: Brock M, Dietz H, editors. *Intracranial pressure, experimental and clinical aspects.* Berlin: Springer; 1972. p. 361–4.
31. Langfitt TW, Weinstein JD, Kassell NF, Simeone FA. Transmission of increased intracranial pressure. I. Within the craniospinal axis. *J Neurosurg.* 1964;21:989–97.
32. Rosenwasser RH, Kleiner LI, Krzeminski JP, et al. Intracranial pressure monitoring in the posterior fossa: a preliminary report. *J Neurosurg.* 1989;71:503–5.
33. Stern WE. Studies in experimental brain swelling and compression. *J Neurosurg.* 1959;16:676–704.
34. Langfitt TW, Weinstein JD, Kassell NF, Gagliardi LJ. Transmission of increased intracranial pressure. II. Within the supratentorial space. *J Neurosurg.* 1964;21:998–1005.
35. Friede RL, Roessmann U. The pathogenesis of secondary mid-brain hemorrhages. *Neurology.* 1966;16:1210–6.
36. Fisher CM. Brain herniation: a revision of classical concepts. *Can J Neurol.* 1995;22:83–91.
37. Plum F, Posner JB. *The diagnosis of stupor and coma.* 1st ed. Philadelphia: F.A. Davis Co.; 1966.
38. Plum F, Posner JB. *The diagnosis of stupor and coma.* 2nd ed. Philadelphia: F.A. Davis Co.; 1972.
39. Wijdicks EFM, Miller GM. MR imaging of progressive downward herniation of the diencephalon. *Neurology.* 1997;48:1456–9.
40. Kernohan JW, Woltman HW. Incisura of the crus due to contralateral brain tumor. *Arch Neurol Psychiatry.* 1929;21:274–87.
41. Ropper AH. Syndrome of transtentorial herniation: is vertical displacement necessary? *J Neurol Neurosurg Psychiatry.* 1993;56:932–5.
42. Feldmann E, Gandy SE, Becker R, et al. MRI demonstrates descending transtentorial herniation. *Neurology.* 1988;38:697–701.
43. Reich JB, Sierra J, Camp W, et al. Magnetic resonance imaging measurements and clinical changes accompanying transtentorial and foramen magnum herniation. *Ann Neurol.* 1993;33:159–70.
44. Ropper AH. Lateral displacement of the brain and level of consciousness in patients with an acute hemispherical mass. *N Engl J Med.* 1986;314:953–8.
45. Ross DA, Olsen WWI, Ross AM, Andrews BT, Pitts LH. Brain shift, level of consciousness, and restoration of consciousness in patients with acute intracranial hematoma. *J Neurosurg.* 1989;71:498–502.
46. Ananda A, Morris GF, Juul N, Marshall SB, Marshall LF. The frequency, antecedent events, and causal relationships of neurologic worsening following severe head injury. *Acta Neurochir (Suppl).* 1999;73:99–102.
47. Narayan RK. Brain stem blood flow, pupillary response, and outcome in patients with severe head injuries. *Neurosurgery.* 1999;44:948 (comments).
48. Hussain SI, Cordero-Tumangday C, Goldenberg FD, Wollman R, Frank JI, Rosengart AJ. Brainstem ischemia in acute herniation syndrome. *J Neurol Sci.* 2008;268:190–2.
49. Ritter AM, Muizelaar JP, Barnes T, et al. Brain stem blood flow, pupillary response, and outcome in patients with severe head injuries. *Neurosurgery.* 1999;44:941–8.
50. Ropper AH. A preliminary study of the geometry of brain displacement and level of consciousness with acute intracranial masses. *Neurology.* 1992;39:622–7.
51. Ropper AH, Shafraan B. Brain edema after stroke: clinical syndrome and intracranial pressure. *Arch Neurol.* 1984;41:26–9.
52. Wijdicks EFM. Acute brainstem displacement without uncal herniation and posterior cerebral artery injury. *J Neurol Neurosurg Psychiatry.* 2008;79:744.
53. Adler DE, Milhorat TH. The tentorial notch: anatomical variation, morphometric analysis, and classification in 100 human autopsy cases. *J Neurosurg.* 2002;96:1103–12.
54. Sunderland S. The tentorial notch and complications produced by herniations of the brain through that aperture. *Br J Surg.* 1958;45:422–38.
55. Poca MA, Benejam B, Sahuquillo J, et al. Monitoring intracranial pressure in patients with malignant cerebral artery infarction: is it useful? *J Neurosurg.* 2010;112:648–57.
56. Kim BS, Jallo J. Intracranial pressure monitoring and management of raised intracranial pressure. In: Loftus C, editor. *Neurosurgical emergencies.* 2nd ed. New York: Thieme; 2008. p. 11–26.
57. Li LM, Timofeev I, Czosnyka M, Hutchinson PJ. Review article: the surgical approach to the management of increased intracranial pressure after traumatic brain injury. *Anesth Analg.* 2010;111:736–48.
58. Murray LS, Teasdale GM, Murray GD, Miller DJ, Picard JD, Shaw MD. Head injuries in four British neurosurgical centres. *Br J Neurosurg.* 1999;13:564–9.
59. Van der Meer C, van Lindert E, Petru R. Late decompressive craniectomy as rescue treatment for refractory high intracranial pressure in children and adults. *Acta Neurochir (Suppl).* 2012;114:305–10.

60. Zweifel C, Hutchinson P, Czoska M. Intracranial pressure, chapter 4. In: Matta BF, Menon DK, Smith M, editors. Core topics in neuroanaesthesia and neurointensive care. New York: Cambridge University Press; 2011. p. 45–62.
61. Neff S, Subramaniam RP. Monro–Kellie doctrine. *J Neurosurg.* 1996;85:1195.
62. Chestnut R, Videtta W, Vespa P, Le Roux P. Intracranial pressure monitoring: fundamental considerations and rationale for monitoring. *Neurocrit Care.* 2014;21:S64–84.
63. Beaumont A. Intracranial pressure and cerebral blood flow monitoring. Chapter 8. In: Torbey MT, editor. Neurocritical care. New York: Cambridge University Press; 2009. p. 109–18.
64. Schulman K, Marmarou A, Shapiro K. Brain tissue pressure and focal pressure gradients. In: McLaurin RL, editor. Head injuries. Second Chicago symposium on neural trauma. New York: Grune & Stratton; 1976. p. 279–87.
65. Sklar FH, Elashvili I. The pressure-volume function of brain elasticity. Physiological considerations and clinical applications. *J Neurosurg.* 1977;47:670–9.
66. Wolfa CE, Luerssen TG, Bowman RM, Putty TK. Brain tissue pressure gradients created by expanding frontal epidural mass lesion. *J Neurosurg.* 1996;84:642–7.
67. Donnelly BR, Medige J. Shear properties of human brain tissue. *J Biomech Eng.* 1997;119:423–32.
68. Sack I, Beierbach B, Hamhaber U, Klatt D, Braun J. Non-invasive measurement of brain viscoelasticity using magnetic resonance elastography. *NMR Biomed.* 2008;21:265–71.
69. Fehlner A, Papaoglou S, McGarry MD, et al. Cerebral multi-frequency MR elastography by remote excitation of intracranial shear waves. *NMR Biomed.* 2015;28:1426–32.
70. Reulen HJ, Graham R, Spatz M, Klatzo I. Role of pressure gradients and bulk flow in dynamics of vasogenic brain edema. *J Neurosurg.* 1977;46:24–35.
71. Rekaté HL. Brain turgor (Kb): intrinsic property of the brain to resist distortion. *Pediatr Neurosurg.* 1992;18:257–62.
72. Miller JD, Leech PJ, Pickard JD. Volume–pressure response in various experimental and clinical conditions. In: Lundberg N, Ponten U, Brock M, editors. Intracranial pressure II. New York: Springer; 1975. p. 97–100.
73. Nakatani S, Ommaya AK. A critical rate of cerebral compression. In: Brock M, Dietz H, editors. Intracranial pressure, experimental and clinical aspects. Berlin: Springer; 1972. p. 144–8.
74. Symon L, Pasztor E, Branston NM, Dorsch WC. Effect of supratentorial space-occupying lesions on regional intracranial pressure and local cerebral blood flow: an experimental study in baboons. *J Neurol Neurosurg Psychiatry.* 1974;37:617–26.
75. Weaver DD, Winn HR, Jane JA. Differential intracranial pressure in patients with unilateral mass lesions. *J Neurosurg.* 1982;56:660–5.
76. Bekar A, Taskapilioglu O, Yilmazlar S, Ender K, Aksoy K. Is supratentorial pressure difference clinically relevant? Analysis of 55 consecutive cases by bilateral intracranial pressure monitoring. *Neurol Res.* 2008;30:465–70.
77. Marshall LF, Zovickian J, Ostrup R, Seelig J. Multiple simultaneous recordings of ICP in patients with acute mass lesions. In: Miller JD, Teasdale GM, Rowan JO, Galbraith SL, Mendelow AD, editors. Intracranial pressure VI. New York: Springer; 1986. p. 184–6.
78. Mindermann T, Reinhardt H, Gratzl O. Significant lateralization of supratentorial ICP after blunt head trauma. *Acta Neurochir.* 1992;116:60–1.
79. Mindermann T, Gratzl O. Interhemispheric pressure gradients in severe head trauma in humans. *Acta Neurochir Suppl.* 1998;71:56–8.
80. Sahuquillo J, Poca MA, Arribas M, et al. Interhemispheric supratentorial intracranial pressure gradients in head-injured patients: are they clinically important. *J Neurosurg.* 1999;90:16–26.
81. Chambers IR, Kane PJ, Signorini DF, Jenkins A, Mendelow AD. Bilateral ICP monitoring: it's importance in detecting the severity of secondary insults. *Acta Neurochir Suppl.* 1988;71:42–3.
82. Zaccchetti L, Magnoni S, DiCorte F, Zanier ER, Stocchetti N. Accuracy of intracranial pressure monitoring: systematic review and meta-analysis. *Crit Care.* 2015;19:420.
83. Marmarou A, Schulman K, LaMorgese J. Compartmental analysis of compliance and outflow resistance of the cerebrospinal fluid system. *J Neurosurg.* 1975;43:523–34.
84. Helbok R, Olson DM, Le Roux PD, Vespa P. Intracranial pressure and cerebral perfusion pressure monitoring in non-TBI patients: special considerations. *Neurocrit Care.* 2014;21:S85–94.
85. Olson DM, Batjer HH, Abdulkadir K, Hall CE. Measuring and monitoring ICP in Neurocritical care: results from a National Practice Survey. *Neurocrit Care.* 2014;20:15–20.
86. Le Roux P, Menon DK, Citerio G, et al. The international multidisciplinary consensus conference on multimodality monitoring in neurocritical care: evidentiary tables: a statement for healthcare professionals from the Neurocritical Care Society and the European Society of Intensive Care Medicine. *Neurocrit Care.* 2014;21 Suppl 2:S297–361.
87. Lane PL, Skoretz TG, Doig G, Girotti MJ. Intracranial pressure monitoring and outcomes after traumatic brain injury. *Can J Surg.* 2000;43:442–8.
88. Qiang Y, Xing W, Yirui S, Jian Y, Zhiqi L, Zhuoying D, Ying M, et al. Impact of intracranial pressure monitoring on mortality in patients with traumatic brain injury: a systematic review and meta-analysis. *J Neurosurg.* 2015;122:574–87.
89. Chestnut RM, Temkin N, Carney N, Dikmen S, Rondina C, Videtta W, et al. A trial of intracranial pressure monitoring in traumatic brain injury. *N Engl J Med.* 2012;367:2471–81.
90. Cremer OL, van Dijk GW, van Wensen E, et al. Effect of intracranial pressure monitoring and targeted intensive care on functional outcome after severe head injury. *Crit Care Med.* 2005;33:2207–13.
91. Shafti S, Diaz-Arrastia R, Madden C, Gentilello L. Intracranial pressure monitoring in brain-injured patients is associated with worsening of survival. *J Trauma.* 2008;64:335–40.
92. Compagnone C, Murray GD, Teasdale GM, et al. The management of patients with intradural post-traumatic mass lesions: a multicenter survey of current approaches to surgical management in 729 patients coordinated by the European Brain Injury Consortium. *Neurosurgery.* 2005;57:1183–92.
93. Leech P, Childs C, Evans J, Johnson N, Protheroe R, King A. Transfer times for patients with extradural and subdural haematomas to neurosurgery in Greater Manchester. *Br J Neurosurg.* 2007;21:11–5.
94. Levi L, Guilburd J, Soustiel J, Svirni G, Constantinescu M, Zaaroor M. Why mortality is still high with modern care of 613 evacuated mass lesions presented as severe head injuries 1999–2009. *Acta Neurochir Suppl.* 2012;114:301–4.
95. Al-Tamimi YZ, Helmy A, Bavetta S, Price SJ. Assessment of zero drift in the Codman intracranial pressure monitor: a study from 2 neurointensive care units. *Neurosurgery.* 2009;64:94–9.
96. Gelabert-Gonzalez M, Ginesta-Galan V, Sernamito-Garcia R, Allut AG, Bandin-Dieguez J, Rumbo RM. The Camino intracranial pressure device in clinical practice. Assessment in a 1000 cases. *Acta Neurochir.* 2006;148:435–41.
97. Koenig MA, Kaplan PW. Invited review: clinical applications for EPs in the ICU. *J Clin Neurophysiol.* 2015;32:472–80.

98. Chiappa KH. Brain Stem Auditory Evoked Potentials: Methodology & Interpretation. In: Chiappa KH editor. Evoked potentials in clinical medicine. 3rd ed. Philadelphia: Raven Press; 1997 a:157–282.
99. Chiappa KH. Short-latency somatosensory evoked potentials: methodology. In: Chiappa KH, editor. Evoked potentials in clinical medicine. 3rd ed. Philadelphia: Raven Press; 1997. p. 283–339.
100. De Sousa LCA, Colli BO, Piza MRT, da Csota SS, Ferez M, Lavrador MAS. Auditory brainstem response: prognostic value in patients with a score of 3 on the Glasgow Coma Scale. *Otol Neurotol*. 2007;28:426–8.
101. Stone JL, Ghaly RF, Hughes JR. Evoked potentials in head injury and states of increased intracranial pressure. *J Clin Neurophysiol*. 1988;5:135–60.
102. Lutschg J, Pfenninger J, Ludin HP, Vassella F. Brain-stem auditory evoked potentials and early somatosensory evoked potentials in neurointensively treated comatose children. *Am J Dis Child*. 1983;137:421–6.
103. Newlon PG, Greenberg RP. Evoked potentials in severe head injury. *J Trauma*. 1984;24:61–6.
104. Narayan RK, Greenberg RP, Miller JD, et al. Improved confidence of outcome prediction in severe head injury: a comparative analysis of the clinical examination, multimodal evoked potentials, CT scanning, and intracranial pressure. *J Neurosurg*. 1981;54:751–62.
105. Tsubokawa T, Nishimoto H, Yamamoto T, Kitamura M, Katayama Y, Moriyasu N. Assessment of brainstem damage by the auditory brainstem response in acute severe head injury. *J Neurol Neurosurg Psychiatry*. 1980;43:1005–11.
106. Seelig JM, Greenberg RP, Becker DP, Miller JD, Choi SC. Reversible brain-stem dysfunction following acute traumatic subdural hematoma: a clinical and electrophysiological study. *J Neurosurg*. 1981;55:516–23.
107. Stone JL, Ghaly RF, Hughes JR, Nagao S. Sensory evoked potentials in head injury. In: Stone JL, editor. Head injury and its complications. Costa Mesa: PMA Publishing; 1993. p. 95–131.
108. Su TY, Xiao SY, Haupt WF, et al. Parameters and grading of evoked potentials: prediction of unfavorable outcome in patients with severe stroke. *J Clin Neurophysiol*. 2010;27:25–9.
109. Amantini A, Fossi S, Grippo A, et al. Continuous EEG-SEP monitoring in severe brain injury. *Clin Neurophysiol*. 2009;39:85–93.
110. Carrerea E, Emerson RG, Claassen J. Evoked Potentials. In: Le Roux PD, Levine JM, Kofke WA, editors. Monitoring in neurocritical care. Philadelphia: Elsevier; 2013. p. 236–45.
111. Fossi S, Amantini A, Grippo A, et al. Continuous EEG-SEP monitoring of severely brain injured patients in NICU: methods and feasibility. *Neurophysiol Clin*. 2006;36:195–205.
112. Haupt WF, Pawlik G, Thiel A. Initial and serial evoked potentials in cerebrovascular critical care patients. *J Clin Neurophysiol*. 2006;23:389–94.
113. Carter BG, Butt W. Are somatosensory evoked potentials the best predictor of outcome after severe brain injury? A systematic review. *Intensive Care Med*. 2005;31:765–75.
114. Cruccu G, Aminoff MJ, Curio G, et al. Recommendations for the clinical use of somatosensory-evoked potentials. *Clin Neurophysiol*. 2008;119:1705–19.
115. Guerit JM, Amantini A, Amodio P, et al. Consensus on the use of neurophysiological tests in the intensive care unit (ICU): electroencephalogram (EEG), evoked potentials (EP), and electroneuromyography (ENMG). *Clin Neurophysiol*. 2009;39:71–83.
116. Guerit JM. Medical technology assessment EEG and evoked potentials in the intensive care unit. *Neurophysiol Clin*. 1999;29:301–17.
117. Guerit JM. Evoked potentials in severe brain injury. In: Laureys S, editor. Progress in brain research, vol. 150, Elsevier BV. 2005; Chapter 29:415–426.
118. Stocchetti N, Le Roux P, Vespa P, et al. Clinical review: neuromonitoring—an update. *Crit Care*. 2013;17:201–13.
119. Esteban A. A neurophysiological approach to brainstem reflexes. Blink reflex. *Neurophysiol Clin*. 1999;29:7–38.
120. Valls-Sole J. The blink reflex and other cranial nerve reflexes. In: Aminoff MJ, editor. Aminoff's electrodiagnosis in clinical neurology. 6th ed. New York: Elsevier-Saunders; 2012. p. 421–35.
121. Preston DC, Shapiro BE. Blink reflex. Electromyography and neuromuscular disorders. Clinical-electrophysiologic correlations. 2nd ed. Philadelphia: Elsevier; 2005. p. 59–64.
122. Lyon LW, Kimura J, McCormick WF. Ocularis oculi reflexes in coma: clinical, electrophysiological and pathological correlation. *J Neurol Neurosurg Psychiatry*. 1972;35:582–8.
123. Fisher MA, Shahani BT, Young RR. Assessing segmental excitability after acute rostral lesions: II. The blink reflex. *Neurology*. 1979;29:45–50.
124. Klug M, Csecsei G, Rap ZM. Evoked potentials and blink reflex in acute midbrain syndromes. Clinical and experimental findings. In: Villani R, Papo I, Giovanelli M, Gaini SM, Tomei G, editors. Advances in neurotraumatology. Excerpta medica. Amsterdam: Oxford-Princeton; 1983. p. 207–9.
125. Mabin D, Mimassi N, L'Azou D, Besson G, Tea S, Le Mevel JC. Prognostic value of the blink reflex in 49 severe trauma comas. In: Villani R, Papo I, Giovanelli M, Gaini SM, Tomei G, editors. Advances in neurotraumatology. Excerpta medica. Amsterdam: Oxford-Princeton; 1983. p. 217–9.
126. Csecsei G, Klug N, Rap ZM. Effect of increased intracranial pressure on the blink reflex in cats. *Acta Neurochir*. 1983;68:85–92.
127. Ayta S, Sohaoglu M, Uluduz D, et al. Auditory evoked blink reflex in peripheral facial paresis. *J Clin Neurophysiol*. 2015;32:34–8.
128. Lederman RJ. Cranial nerve conduction studies. The blink reflex and jaw jerk. In: Levin KH, Luders HO, editors. Comprehensive clinical neurophysiology. Philadelphia: W. B. Saunders; 2000. p. 107–11.
129. Semih A, Sohtaoglu M, Uluduz D, et al. Auditory evoked blink reflex in peripheral facial paresis. *J Clin Neurophysiol*. 2015;32:34–8.
130. Deletis V, Urriza J, Ulkatan S, Fernandez-Conejero I, Lesser J, Misita D. The feasibility of recording blink reflexes under general anesthesia. *Muscle Nerve*. 2009;39:642–6.
131. Husain AM. Neurophysiologic intraoperative monitoring. In: Ebersole JS, editor. Current practice of clinical electroencephalography. 4th ed. Philadelphia: Wolters Kluwer; 2014. p. 488–542.
132. Nuwer MR, Packwood JW. Somatosensory evoked potential monitoring with scalp and cervical recording. In: Nuwer MR, editor. Handbook of clinical neurophysiology. Intraoperative monitoring of neural function, vol. 8. Amsterdam: Elsevier; 2008. p. 180–9.
133. American Clinical Neurophysiology Society. Guidelines: guidelines on short-latency auditory and short-latency somatosensory evoked potentials. *J Clin Neurophysiol*. 2006; 23(2):157–79.
134. Aminoff MJ, Eisen A. Sensory evoked potentials. In: Aminoff MJ, editor. Aminoff's electrodiagnosis in clinical neurology. 6th ed. New York: Elsevier-Saunders; 2012. p. 581–601.

135. Husain AM. Evoked potentials overview. In: Ebersole JS, editor. *Current practice of clinical electroencephalography*. 4th ed. Philadelphia: Wolters Kluwer; 2014. p. 442–87.
136. Emerson RG, Adams DC. Intraoperative monitoring by evoked potential techniques. In: Aminoff MJ, editor. *Aminoff's electrodiagnosis in clinical neurology*. 6th ed. New York: Elsevier-Saunders; 2012. p. 651–70.
137. Nuwer MR. Somatosensory evoked potential monitoring with scalp and cervical recording. Intraoperative neurophysiologic monitoring. In: Galloway GM, Nuwer MR, Lopez JR, Zamel KM, editors. *New York: Cambridge University Press*; 2010. p. 63–74.
138. Chiappa KH, Hill RA. Short-latency somatosensory evoked potentials: interpretation. In: Chiappa KH, editor. *Evoked potentials in clinical medicine*. 3rd ed. Philadelphia: Raven Press; 1997. p. 341–423.
139. Moller AR. Monitoring brainstem auditory and somatosensory evoked potentials. In: *Intraoperative neurophysiological monitoring*. 2nd ed. Totowa: Humana; 2006. p. 85–144.
140. Toleikis JR. Intraoperative monitoring using somatosensory evoked potentials: a position statement by the American Society of Neurophysiological Monitoring. *J Clin Monit Comput*. 2005;19:241–58.
141. Fischer C, Luaute J. Evoked potentials for the prediction of vegetative state in the acute stage of coma. *Neuropsychol Rehab*. 2005;15:372–80.
142. Luaute J, Cotton F, Jemaire JJ, et al. Let live or let die after traumatic coma. Scrutinizing somatosensory evoked potentials. *Neurology: clinical practice*, American Academy of Neurology Enterprises, Baltimore: Wolters Kluwer; 2012. p. 24–32.
143. Robinson LR, Micklesen PJ, Tirschwell DL, Lew HL. Predictive value of somatosensory evoked potentials for awakening from coma. *Crit Care Med*. 2003;31:960–7.
144. Cruse D, Norton L, Gofton T, Young GB, Owen AM. Positive prognostication from median-nerve somatosensory evoked potentials. *Neurocrit Care*. 2014;21:238–44.
145. Landi A, Zanusso M, Curri D, et al. Trigeminal evoked potentials in patients undergoing percutaneous micro-compression of the Gasserian Ganglion. *Stereotact Funct Neurosur*. 1991;56: 28–36.
146. Soustiel JF, Hafner H, Guilburd JN, Zaaroor M, Levi L, Feinsod M. A physiological coma scale: grading of coma by combined use of brain-stem trigeminal and auditory evoked potentials and the Glasgow Coma Scale. *Electroencephalogr Clin Neurophysiol*. 1993;87:277–83.
147. Kawaguchi J, Matsuura N, Kasahara M, Ichinohe T. Cervical sympathetic block prolongs the latency and reduces the amplitude of trigeminal somatosensory evoked potentials. *J Clin Neurophysiol*. 2015;32:39–43.
148. Malcharek MJ, Landgraf J, Hennig G, et al. Recordings of long-latency trigeminal somatosensory-evoked responses under general anesthesia. *Clin Neurophysiol*. 2011;122:1048–54.
149. Hasegawa A. The changes of the trigeminal somatosensory evoked potentials (TSEPs) after block anesthesia to the mandibular foramen. *Aichi Gakuin Dent Soc*. 2005;43:47–53.
150. Bennett AJ, Wastell DG, Barker GR, et al. Trigeminal somatosensory evoked potentials. A review of the literature as applicable to oral dysesthesias. *Int J Oral Maxillofac Surg*. 1987;16:408–15.
151. Burkard RF, Don M, Eggermont JJ. Auditory evoked potentials. Basic principals and clinical application. Philadelphia: Lippincott Williams & Wilkins; 2007.
152. Hall JW. *New handbook of auditory evoked responses*. Boston: Allyn and Bacon; 2007. p. 212–80,434–40.
153. Legatt AD. Brainstem auditory evoked potentials: methodology, interpretation, and clinical application. In: Aminoff MJ, editor. *Aminoff's electrodiagnosis in clinical neurology*. 6th ed. New York: Elsevier-Saunders; 2012. p. 519–52.
154. Legatt AD. Mechanisms of intraoperative brainstem auditory evoked potential changes. *J Clin Neurophysiol*. 2002;19: 396–408.
155. Stone JL, Calderon-Arnulphi M, Watson KS, Patel K, Mander NS, Suss N, Fino J, Hughes JR. Brainstem auditory evoked potentials—a review and modified studies in healthy subjects. *J Clin Neurophysiol*. 2009;26:167–75.
156. Simon M. Neurophysiologic tests in the operating room. In: Simon M, editor. *Intraoperative neurophysiology*. New York: Demos Medical; 2010. p. 2–44.
157. Sininger YS. The use of auditory brainstem response in screening for hearing loss and audiometric threshold prediction. In: Burkard R, Don M, Eggermont JJ, editors. *Auditory evoked potentials. Basic principles and clinical application*. Philadelphia: Lippincott Williams & Wilkins; 2007. p. 254–74.
158. Krieger D, Adams HP, Schwarz S, Rieke K, Aschoff A, Hacke W. Prognostic and clinical relevance of pupillary responses, intracranial pressure monitoring, and brainstem auditory evoked potentials in comatose patients with acute supratentorial lesions. *Crit Care Med*. 1993;21:1944–50.
159. Krieger D, Jauss M, Schwarz S, Hacke W. Serial somatosensory and brainstem auditory evoked potentials in monitoring of acute supratentorial mass lesions. *Crit Care Med*. 1995;23:1123–31.
160. Nagao S, Sunami N, Tsutsui T, et al. A diagnosis of uncal herniation by auditory brainstem response. *Neurol Med Chir (Tokyo)*. 1984;24:396–400.
161. Nagao S, Kuyama H, Honma Y, et al. Prediction and evaluation of brainstem function by auditory brainstem responses in patients with uncal herniation. *Surg Neurol*. 1987;27:81–6.
162. Wang WP, Qui MD, Ren HJ, Zhang XH. Relations of intracranial pressure, creatine kinase and brainstem auditory evoked potential in patients with traumatic brain edema. *Chin Med J (Engl)*. 1994;107:205–8.
163. Bosch-Blancafort B, Olesti-Marco M, Poch-Puig JM, Rubio-Garcia E, Nogues-Bara P, Iglesias-Berenguer J. Predictive value of brain-stem auditory evoked potentials in children with post-traumatic coma produced by diffuse brain injury. *Childs Nerv Sys*. 1995;11:400–5.
164. Kawahara N, Sasaki M, Mii K, Takakura K. Reversibility of cerebral function assessed by somatosensory evoked potentials and its relation to intracranial pressure—report of six cases with severe head injury. *Neurol Med Chir (Tokyo)*. 1991;31:264–71.
165. Nagata K, Tazawa T, Mizukami M, Araki G. Application of brainstem auditory evoked potentials to evaluation of cerebral herniation. In: Nodar RH, Barber C, editors. *Evoked potentials*, vol. 2. Boston: Butterworths; 1984. p. 183–93.
166. Nagao S, Sunami N, Tsutsui T, Honma Y, Doi A, Nishimoto A. Serial observations of brain stem responses in central transtentorial herniation. *Surg Neurol*. 1982;17:355–7.
167. Rogowski M, Michalska BI. The importance of brain stem evoked potentials in the diagnosis of neurosurgical patients. *Neurol Neurochir Pol*. 2001;35:667–79.
168. Wang WP, Qiu MD, Ren HJ, Zhang XH. Relations of intracranial pressure, creatine kinase and brainstem auditory evoked potential in patients with traumatic brain edema. *Chin Med J (Engl)*. 1994;107:205–8.
169. Ackley RS, Herzberger-Kimball L, Burns S, Balew SD. Auditory brainstem response testing: Stimulus rate revisited. 2006. <http://www.AudiologyOnline.com>. Accessed 12 Dec 2015, p. 1–7.
170. Stone JL, Ghaly RF, Subramanian KS, Roccaforte P, Hughes JR. Modified auditory brain-stem responses (MABR): part I: rationale and normative study. *Clin Electroencephalogr*. 1987;18: 218–26.

171. Stone JL, Fino J, Vannemreddy P, Charbel F. Modified brainstem auditory evoked responses in patients with non-brainstem compressive cerebral lesions. *Acta Neurochir Suppl.* 2012; 114:81–5.
172. Stone JL, Fino J, Patel K, Calderon-Arnulphi M, Suss N, Hughes JR. Modified brain stem auditory evoked potentials in patients with intracranial mass lesions. *Clin EEG Neurosci.* 2012;43: 291–302.
173. Legatt AD, Emerson RG, Epstein CM, et al. ACNS Guideline: transcranial electrical stimulation motor evoked potential monitoring. *J Clin Neurophysiol.* 2016;33:42–50.
174. Journee HL, Motor EP. Physiology, risks and specific anesthetic effects. In: Nuwer MR, editor. *Handbook of clinical neurophysiology. Intraoperative monitoring of neural function*, vol. 8. Amsterdam: Elsevier B.V; 2008. p. 218–34.
175. Tanaguchi M, Cedzich C, Schramm J. Modification of electrical stimulation technique for motor evoked potential monitoring under general anesthesia: technical description. *Neurosurgery.* 1993;32:219–26.
176. Galloway GM, Dias BR, Brown JL, Henry CM, Brooks DA, Buggie EW. Transcranial magnetic stimulation—may be useful as a preoperative screen of motor tract function. *J Clin Neurophysiol.* 2013;30:386–9.
177. Zamel KM. Magnetic stimulation. In: Galloway GM, Nuwer MR, Lopez JR, Zamel KM, editors. *Intraoperative neurophysiological monitoring*. New York: Cambridge University Press; 2010. p. 19–32.
178. Egg-Olofsson KE. Transcranial magnetic stimulation. In: Holmes GL, Moshe SL, Jones Jr HR, editors. *Clinical neurophysiology of infancy, childhood, and adolescence*. Philadelphia: Elsevier; 2006. p. 819–25.
179. Gugino LD, Aaglio LS, Edmonds HL, Gonzalez AA. Magnetic cortical stimulation techniques. In: Nuwer MR, editor. *Handbook of clinical neurophysiology. Intraoperative monitoring of neural function*, vol. 8. Amsterdam: Elsevier B.V; 2008. p. 282–318.
180. Malcolm MP, Paxton RJ. High-frequency repetitive transcranial magnetic stimulation effects on motor intracortical neurophysiology: a sham-controlled investigation. *J Clin Neurophysiol.* 2015;32:428–33.
181. Binder DK, Lyon R, Manley GT. Transcranial motor evoked potential recording in a case of Kernohan's notch syndrome: case report. *J Neurosurg.* 2004;54:999–1003.
182. Carrasco R, Pascual JM, Navas M, Martinez-Florez P, Manzanares-Soler R, Sola RG. Kernohan–Woltman notch phenomenon caused by an acute subdural hematoma. *J Clin Neurosci.* 2009;16:1628–31.
183. Kwon HG, Lee DG, Choi BY, Chang CH, Kim SH, Jang SH. Recovery of the corticospinal tract after injury by transtentorial herniation—a case report. *NeurRehab.* 2011;29:243–6.
184. Milhorat TM, Marion DW. Comments on electrophysiology of Kernohan's notch. *Neurosurgery.* 2004;54:1002–3.
185. Moon KS, Lee JK, Joo SP, et al. Kernohan's notch phenomenon in chronic subdural hematoma: MRI findings. *J Clin Neurosci.* 2007;14:989–92.
186. Namura S, Kang Y, Matsuda I, Kamijyo Y. Magnetic resonance imaging of sequelae of temporal lobe herniation secondary to traumatic acute subdural hematoma: Kernohan's notch and posterior cerebral artery territory infarctions contralateral to the supratentorial lesion—case report. *Neurol Med Chir (Tokyo).* 1997;37:32–5.
187. Uesugi S, Suchiro E, Nakayama H, Suzuki M. Diffusion-weighted magnetic resonance imaging in a case of Kernohan's notch phenomenon. *Acta Neurochir.* 2010;152:1809–10.
188. Yeo SS, Jang SH. Corticospinal tract recovery in a patient with traumatic transtentorial herniation. *Neural Reg Res.* 2013;8: 469–73.
189. Yoo WK, Kim DS, Kwon YH, Jang SH. Kernohan's notch phenomena demonstrated by diffusion tensor imaging and transcranial magnetic stimulation. *J Neurol Neurosurg Psychiatry.* 2008;79:1295–7.
190. Suzuki A, Nishimura S, Yasui N. Electrophysiological assessment of patients of supratentorial intracerebral hematoma, In: *Brain Hemorrhage'95. Proceeding of the 1st annual meeting of intracerebral hemorrhage*, Nagoya, Japan; 1995. p. 43–9.
191. Nagao S, Roccaforte P, Moody RA. Acute intracranial hypertension and auditory brain-stem responses. Part 1: changes in the auditory brain-stem and somatosensory evoked responses in intracranial hypertension in cats. *J Neurosurg.* 1979;51:669–76.
192. Nagao S, Roccaforte P, Moody RA. Acute intracranial hypertension and auditory brain-stem responses. Part 2: the effects of brain-stem movement on the auditory brain-stem responses due to transtentorial herniation. *J Neurosurg.* 1979;51:846–51.
193. Nagao S, Sunami N, Tsutsui T, et al. Acute intracranial hypertension and brain-stem blood flow. *J Neurosurg.* 1984;60: 566–71.
194. Nagao S, Roccaforte P, Moody RA. Effect of brain compression on pyramidal tract responses in the cat. *Exp Neurol.* 1981;73:107–17.
195. Stone JL, Ghaly RF, Subramanian KS, Roccaforte P, Kane J. Transtentorial brain herniation in the monkey: analysis of brainstem auditory and somatosensory evoked potentials. *Neurosurgery.* 1990;26:26–31.
196. Seelig JM, Becker DP, Miller JD, et al. Traumatic acute subdural hematoma: major mortality reduction in comatose patients treated within four hours. *N Eng J Med.* 1981;304:1511–8.