

# The Nerve of Glaucoma!

Robert A. Schumer, MD, PhD, Steven M. Podos, MD

**C**ontemporary concepts of open angle glaucoma suggest that the current emphasis on reduction of elevated intraocular pressure could be augmented by other therapeutic approaches. In this article, we describe significant recent developments in the molecular and cellular biology and neuropharmacology of nerve damage that are likely, in coming years, to suggest new therapeutic approaches to the management of glaucoma. These developments may lead to the achievement of pharmacologic protection of the optic nerve from damage or possibly promotion of reversal of damage. We review selected studies of excitotoxins and *N*-methyl-D-aspartate receptor antagonists,  $\text{Ca}^{2+}$ -induced damage and calcium channel blockers, the intracellular messenger nitric oxide and its perturbation, free-radical damage and scavengers, nerve regeneration, and growth factors. Several basic research questions are posed, answers to which may transform our concepts of glaucoma therapy. (*Arch Ophthalmol.* 1994;112:37-44)

Clinical experience and epidemiologic research indicate that elevated intraocular pressure (IOP) is the single most common concomitant finding in primary open angle glaucoma.<sup>1-3</sup> This outcome explains the overwhelming emphasis on screening, treating, and investigating IOP.

However, evidence from the study of low-tension glaucoma and ocular hypertension suggests that elevation of IOP is neither necessary nor sufficient for the production of glaucomatous optic neuropathy. Thus, there are many patients with elevated IOP who, for unknown reasons, do not manifest glaucoma, while, conversely, there are many patients with low to normal IOP who do. In some survey studies, between one third and one half of patients with open angle glaucoma do not initially have ocular hypertension,<sup>4,7</sup> and as many as one sixth of patients with glaucomatous damage do not show elevated

IOP, even on repeated testing.<sup>7</sup> Moreover, ocular hypertension may be as much as eight to 10 times as common a condition as glaucoma.<sup>4,8</sup> Furthermore, in some patients with glaucoma, progression of field loss is apparently not related to IOP.<sup>9,10</sup> Finally, although IOP elevation is certainly a primary risk factor for glaucoma, numerous risk factors other than elevated IOP have been reported.<sup>11</sup> Mounting recognition and recent expressions of these views are part of the *zeitgeist*, as reflected in the tide of recent expert opinion.<sup>6,12-14</sup>

One implication of these observations is that there may be other causal variables involved in glaucomatous damage, and, in concert with this idea, there should be other therapies. Indeed, if elevated IOP turns out not to be the direct cause of damage, then other modalities may even supersede reduction of IOP as the primary therapeutic modality.

There have been attempts to find means of pharmacologic protection of the optic nerve other than through reduction of IOP. Thus, phenytoin, a membrane stabilizer, has been reported to reverse partially the effects of an-

*From the Department of Ophthalmology, Mount Sinai School of Medicine, New York, NY. The authors have no financial or proprietary interest in the compounds mentioned in this article. Dr Podos is a consultant to Allergan Inc, Irvine, Calif, Alcon Laboratories Inc, Fort Worth, Tex, and Pharmos Corp, New York.*

oxia on neurons<sup>15</sup> and to retard glaucomatous damage in patients.<sup>16</sup> More recently, calcium channel blockers (CCBs), which may protect against vasospasm-induced hypoperfusion of the optic nerve head blood supply, have also been studied.<sup>17-21</sup> None of these has thus far proven to be clinically useful.

Where then to turn? As we look to other related fields, such as molecular and cellular biology, neurology, and neuropharmacology, we note dramatic progress in our knowledge of nerve cells, glial cells, vascular cells, and extracellular matrix-bound cells as well as the intracellular molecular events that follow transmembrane potential changes or receptor activation. Much is now known about the biochemical pathways involved in cell injury and death following ischemia or the application of endogenously occurring cellular toxins. Moreover, in certain clinical areas, such as stroke, degenerative neurological disease, and cardiovascular medicine, direct clinical and experimental applications of this knowledge have already been undertaken.

While there are not yet examples of successful clinical application of this knowledge within the field of glaucoma, these findings from other fields give hope that pharmacologic neuroprotection or even neurorepair may one day be viable approaches to glaucoma management.<sup>22-25</sup> Studies of  $Ca^{2+}$ -induced damage, excitotoxins and *N*-methyl-D-aspartate (NMDA) receptor antagonists, the messengers nitric oxide (NO) and carbon monoxide, free-radical scavengers, nerve regeneration, and growth factors, to name some of the more prominent areas, are opening entirely new investigational perspectives of neuroprotection that may radically transform how we think about glaucoma. A sampling of recent developments in these fields provides a glimpse of what may prove to be the next generation of therapies in glaucoma.

### EXCITOTOXINS AND NMDA RECEPTORS

One area of intense recent study has been that of so-called excitotoxins,

which are excitatory neurotransmitters that, if released in excessive amount, can induce a toxic effect on target cells.<sup>26</sup> With aspartate and glutamate as their likely endogenous neurotransmitters, receptors have been located throughout the brain and layers of the retina, which are excited by the nonendogenous experimental drug NMDA. Three different classes of ionotropic NMDA receptors (NMDARs) have been defined by the differential affinities of these receptors to the glutamate- and aspartate-analogues NMDA, kainic acid, and DL- $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA).<sup>27-29</sup> These receptors have broad functions,<sup>30,31</sup> acting as neurotransmitter-gated ion channels and permitting voltage-dependent intracellular influx of calcium and sodium, the former activating "second messengers" that affect cell function. Importantly, when the excitatory transmitters are present in excess, their receptors also seem to play a major role in mediating ischemic neuronal damage.<sup>32,33</sup>

Excitatory neurotransmitters may be massively released following cellular ischemia or stress.<sup>34,35</sup> Sustained hypoxia leads to membrane depolarization, which, in turn, leads to increased synaptic glutamate release and reduced glutamate uptake, together resulting in a buildup of extracellular glutamate.<sup>36-38</sup> Excitotoxic effects result from the resulting enhanced action of the transmitter on its receptor, which leads to excess influx of  $Ca^{2+}$ , itself contributing to the postsynaptic cell's inability to regulate its own intracellular  $Ca^{2+}$  and hence to even more  $Ca^{2+}$  buildup, neuronal damage, and death.<sup>39</sup>

Activators of NMDARs, whether of endogenous origin or not, lead to neural damage within a wide range of neural structures and also lead to inner retinal degeneration.<sup>40</sup> Retinal ganglion cells possess receptors for excitatory amino acids,<sup>41</sup> and kainic acid, NMDA, and AMPA, in excess, are all toxic to ganglion cells.

Each toxin, however, produces a specific pattern of cell death with larger or smaller retinal ganglion cells being more or less affected.<sup>42,43</sup> Of note, large retinal ganglion cells in culture are more susceptible to excitotoxic as well as hypoxic injury than are small ganglion cells,<sup>44</sup> a situation that may be analogous to glaucomatous damage.<sup>45-47</sup> Intriguingly, vitreous specimens from humans with open angle glaucoma have elevated concentrations of glutamate, but not other amino acids.<sup>48</sup>

Moreover, agents that inhibit the action of excitotoxins improve neuron viability following toxic exposure to excitotoxins.<sup>49</sup> Cell death in experimental conditions mimicking stroke is successfully and dramatically reduced by use of glutamate receptor blocking agents, such as MK-801, phencyclidine, NBQX, or D-CPP-ene.<sup>50-53</sup> For example, NBQX protects hippocampal neurons from death caused by bilateral carotid artery occlusion even when given 1 hour after the termination of ischemia.<sup>50</sup> Severe experimental cerebral ischemia produces large increases in intracellular calcium ion concentration. MK-801, along with the CCB nimodipine, reduces the influx of calcium in this model and also reduces histologic damage and improves the recovery of the electroencephalogram following reperfusion.<sup>54</sup> MK-801, which crosses the blood-brain barrier, seems to be most effective when NMDA channels are already open, ie, after excitotoxin buildup.

In the eye, the swelling and death of ganglion cells in chick embryo retinas in response to anoxic conditions is blocked by  $\gamma$ -D-glutamylglycine, an excitatory amino acid antagonist.<sup>55</sup> Retinal ganglion cells, grown in culture, or in the intact adult retina, are killed by NMDA and kainic acid, and these actions are blocked by NMDA receptor antagonists.<sup>56-58</sup> Pretreatment with the NMDA antagonist dextromethorphan, which protects against experi-

mental cerebral anoxic tissue damage,<sup>59</sup> enhances recovery of the electroretinographic (ERG) b-wave amplitude and reduces histologic alteration in an ischemic model of stroke in the rabbit retina produced by acutely elevated IOP.<sup>60</sup> MK-801 protects retinal ganglion cells in a similar model.<sup>61</sup> Can glaucomatous damage be similarly reduced?

Apart from the short-term toxic effects just mentioned, there is some evidence that in chronic, progressive conditions, such as Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis, olivopontocerebellar atrophy, and Alzheimer's disease, NMDA receptor activation is involved in pathogenesis.<sup>62-64</sup> For example, NMDA receptor antagonists prevent glutamate-induced diphosphorylation of certain enzymes and thereby facilitate dopaminergic function in animal models of Parkinson's disease.<sup>65</sup> Might the damage of glaucoma fit into the category of these kinds of degeneration?

Preliminary, but highly encouraging, clinical applications of the new understanding concerning excitotoxic effects include successful clinical treatment of stroke, transitory ischemic attack, and multi-infarct dementia.<sup>66,67</sup> Useful drug development poses a challenge for the future. Many of the NMDA antagonists, such as phencyclidine and ketamine, have significant psychotomimetic properties in humans limiting their usefulness. However, certain anticholinergic or  $\gamma$ -aminobutyric acid-like agents have been reported to protect choroidal neurons against these adverse side effects,<sup>68</sup> and the recent cloning of many of the subunits of the glutamate receptor family<sup>29,69,70</sup> will doubtless facilitate novel and improved drug development.

### CALCIUM CHANNEL BLOCKERS

By acting on vascular smooth muscle, CCBs can lead to vasodilation or re-

lief from vasospasm. Indeed, after acute ischemic stroke, there seems to be a therapeutic window, perhaps hours in length, following the onset of ischemia during which damage may be reversible. Further, there is a penumbra of viable neural tissue surrounding a region of complete infarction, within which recovery is possible. In keeping with these concepts, efforts to improve blood flow after ischemic insults from stroke<sup>71,72</sup> have been made using CCBs. Following such leads, interest in CCBs within the field of glaucoma has arisen from a desire to improve optic nerve head blood supply.

However, CCBs may also act, in part, by reducing toxic intracellular  $Ca^{2+}$  levels, by altering the metabolism of target nerve cells, or by acting on receptors in the central nervous system (CNS) that are entirely different from those found in peripheral vascular tissue or even peripheral neurons. Elevation of intracellular calcium levels is likely to be neurotoxic for a number of reasons,<sup>39</sup> including activation of catabolic enzymes, phospholipases, superoxide or other free radicals, and protein kinases, and positive feedback stimulation of the release of additional glutamate.

There are two main types of calcium channels, ie, voltage-operated channels, requiring membrane depolarization for function, and receptor-operated channels, requiring a specific ligand, such as NMDA, to bind to a receptor molecule. Within the class of voltage-operated channels, there are four main types of channel:<sup>73-75</sup> T-type, L-type, N-type, and P-type. Calcium channel blockers currently available in the United States, mostly dihydropyridines, typically affect voltage-operated calcium channels and are targeted principally against L-type calcium channels, found mostly on cardiac and smooth muscle cells, such as vascular muscle cells. Neurons tend to possess T-, N-, and P-type channels. However, high con-

servation of binding regions on the known calcium channels has suggested that modifications of existing drug structures may yield selective antagonists of the other channel subtypes that are more specific for neuronal channels and which, therefore, may be valuable in the treatment of calcium-related neurodegenerative or neurotoxic conditions.<sup>75</sup>

Indeed, one CCB, flunarizine, available in Europe, has particularly high affinity for neuronal T-type channels and low affinity for peripheral or vascular sites.<sup>76,77</sup> Migraine, a condition previously identified as having an association with low-tension glaucoma,<sup>78</sup> has been successfully treated with flunarizine.<sup>79</sup> Flunarizine has been shown to protect neural tissue from the effects of ischemia in several animal models.<sup>80,81</sup> Interestingly, flunarizine also protects cultured cerebellar cells from glutamate-induced toxic effects,<sup>82</sup> raising the possibility that this CCB may also act at receptor-operated channels, either directly or perhaps through reduction of intracellular calcium-induced glutamate release.<sup>74</sup>

The clinical utility of CCBs in chronic forms of glaucoma has only begun to be explored.<sup>17-21</sup> Naftidrofuryl oxalate, another vasodilatory agent, which is an antagonist of serotonin (5<sub>2</sub>) receptors, has also been reported to be salutary in glaucoma.<sup>83-85</sup> Findings from the cardiac and neurologic literature<sup>67</sup> indicate the need for investigating further the role of these agents in glaucoma therapy.

### NITRIC OXIDE

Nitric oxide is a short-lived compound that has been recently identified<sup>86</sup> in vascular endothelium, where it promotes vasodilation (and was previously known as endothelium-derived relaxing factor); in macrophages, where it seems to be bactericidal and tumoricidal; and in numerous different areas of

the brain,<sup>87</sup> where it may act as a neurotransmitter, although not of the conventional, synaptosomal variety.<sup>88,89</sup>

One action of glutamate on NMDA receptors is to cause Ca<sup>2+</sup> to enter the postsynaptic cell and NO to be synthesized there.<sup>35</sup> Once inside the cell, Ca<sup>2+</sup>, acting with calmodulin, activates the enzyme NO synthase, which generates NO from L-arginine.<sup>90</sup> Nitric oxide seems in turn to act by binding to the heme subunit of guanylyl cyclase, which transforms GTP into the intracellular second messenger cGMP.<sup>91</sup> This, in turn, activates a protein kinase that may phosphorylate other proteins leading to metabolic alteration of the cell through unknown mechanisms.

Further, NO may rapidly leave the cell through passive transport and may then exert toxic actions directly on adjacent cells.<sup>92</sup> The toxic effect itself may arise from the free radical (see below) nature of NO. The neurotoxic effect of NO has been demonstrated in numerous *in vitro* models.<sup>49,93,94</sup> A similar mechanism to that described for NO has been proposed for carbon monoxide, which is liberated by an enzyme similar to NO synthase, heme oxygenase, and which, like NO, acts on guanylyl cyclase to catalyze the formation of cGMP.<sup>95</sup>

Numerous inhibitors of the enzyme that synthesizes NO, NO synthase (NOS),<sup>96</sup> have been identified, and these substances have been shown to protect neurons from the toxic actions of NO.<sup>97</sup> For example, the competitive NOS inhibitor NG-monomethyl-L-arginine maintains action potential production during hypoxia in a hippocampal slice preparation.<sup>93</sup> Moreover, NMDA-induced cell damage is reduced in cultured neurons by various NOS inhibitors,<sup>98</sup> which, along with other evidence, suggests that NO may be the intracellular mediator of glutamate receptor-induced toxic effects. Also, the effects of experimental stroke in mice are attenuated not

only by NMDAR antagonists<sup>50-53</sup> but by the NOS inhibitor nitroarginine,<sup>99</sup> which suggests that interruptions of the latter steps of the excitotoxic pathway can also protect cells from damage.

Nitric oxide synthase has now been identified in widespread structures of the CNS<sup>87</sup> and the eye, including retinal photoreceptors, chorioidal nerve fibers, and some cells of the ganglion cell layer.<sup>100,101</sup> It has been found in just those neurons that were previously known to contain nicotinamide adenine dinucleotide phosphate (NADPH) diaphorase (NDP),<sup>100,102</sup> which has intrinsic NOS activity.<sup>103</sup> This is of interest because NDP-positive neurons are reported to be resistant to degeneration in conditions such as Huntington's disease, Alzheimer's disease, ischemic stroke, and NMDA-induced toxic effects,<sup>104-108</sup> and there are groups of NDP-positive cells of the inner retina that also exhibit selective resistance to excitotoxic effects.<sup>109</sup> It may be that NOS activity, possibly owing to its vasodilatory activity, is somehow protective of these cells, a function that could be ascendent under different circumstances than those in which NO is toxic to neurons.

Investigations of the role of NO in our emerging concepts of ischemia, and in models of glaucoma, are continuing to be worked out, but this important new molecule in the sequence of cytotoxic mechanisms will almost certainly play an important role in future understanding of retinal ganglion cell damage.

#### FREE-RADICAL SCAVENGERS AND ANTIOXIDANTS

Cell death following stressful ischemic or excitotoxic conditions, as outlined above, requires certain molecular events that mediate long-term neural damage. Oxygen free radicals (OFRs), which are highly reactive molecules that contain one or more unpaired electrons, are one

such particularly important class of mediator,<sup>110</sup> since they are formed by all cells, including neurons, since the high unsaturated lipid content of neuronal membranes may confer high vulnerability to OFR-induced damage, and since the NMDAR activation-induced increase in intracellular Ca<sup>2+</sup> has been shown to generate oxygen radicals.<sup>94</sup> Oxygen free radicals (such as superoxide ion or hydroperoxyl radical) are formed massively during reoxygenation following the cessation of ischemia and appear to contribute to neural damage (reperfusion injury)<sup>111-114</sup> by increasing peroxidation of fatty or nucleic acids and breaking protein cross-linking.<sup>115-119</sup>

Evidence supporting the toxic role of OFRs comes from the demonstration, soon after reperfusion, of dramatic production of OFRs<sup>120,121</sup> and fatty acid oxidation products<sup>122</sup> as well as decreased endogenous antioxidant levels.<sup>123</sup> Further, agents that scavenge OFRs (eg, vitamin E, D-mannitol), catabolize them (eg, superoxide dismutase [SOD], catalase), or reduce their formation (eg, nonsteroidal anti-inflammatory drugs, steroids) can protect against excitotoxic or ischemic injury.<sup>117,124-126</sup>

For example, following the reoxygenation phase in a transient retinal artery occlusion model of retinal ischemia, both histologic damage and large transmembrane ion fluxes, including that of Ca<sup>2+</sup>, are largely eliminated by the free radical scavengers SOD and EGB 761.<sup>127-129</sup> EGB 761, which has small molecular weight and is orally active, preserves the ERG b-wave from lipo-peroxidation-induced extinction.<sup>130</sup> Further, catalase has been shown to protect both the ERG a-wave and b-wave following transient retinal ischemia produced by elevation of IOP in rabbits.<sup>131</sup> These results suggest that oxygen radicals are an important link in the pathways of damage.

Another pathway by which OFRs may cause damage is by interfering with the ability of naturally occurring NO to (adaptively)



regulate vascular tone. In a rabbit model of acute retinal ischemia induced by elevated IOP, exogenous NO, presumably acting as a vasodilator and not as a toxic free radical, prevented ERG extinction, and the free radical scavengers SOD and catalase also protected against acute extinction of the ERG. Surprisingly, the NOS inhibitor nitro-L-arginine eliminated the protective effect of the free radical scavengers,<sup>132</sup> implying that OFRs destroy or block the action of NO<sup>133</sup> and therefore that antioxidant protection promotes protective vasodilation. In addition, certain of the toxic effects of OFRs may be shared with excitotoxins,<sup>134</sup> and OFRs may even modulate NMDAR function.<sup>135</sup> Oxygen free radicals also can cause increased excitotoxin release, further contributing to a vicious cycle.<sup>136</sup>

The damaging role of OFRs may not be limited to brief periods of hypoxia, but could also be important in neural degenerations, such as chronic glaucoma. For example, in amyotrophic lateral sclerosis, motor neuron death may be secondary to increased levels of intracellular Ca<sup>2+</sup>, leading to enzymatic production of superoxide radicals;<sup>137,138</sup> this in turn may be related to mutations in the copper/zinc superoxide dismutase gene<sup>137</sup> or, in part, to an excess of glutamate or its analogues.<sup>139,140</sup>

Possible therapeutic modalities to deter glaucomatous damage could involve the introduction of exogenous free radical scavengers<sup>126,130,141</sup> or the induction of endogenous tolerance to oxidative stress.<sup>142</sup> For example, brief pretreatment with the SOD inhibitor diethyldithiocarbamate promotes enhanced expression of SOD, which can result in reduced neuronal damage from subsequent ischemia.<sup>142</sup>

## NERVE REGENERATION AND GROWTH FACTORS

A recent panel of experts on ganglion cell growth and connectivity

concluded that "the goal of restoring vision through the regeneration of central visual pathways is a realistic one."<sup>25</sup> This optimism derives in part from the recent demonstration that axons within rat optic nerve show the surprising capability to regrow following transection.<sup>143-145</sup> Placement of a peripheral, axon-free nerve graft into the axotomy site induces the regrowth of retinal ganglion cells that may even produce functional inhibitory and excitatory synaptic contacts with CNS target sites.<sup>146</sup> It is believed that the peripheral nerve graft promotes ganglion cell growth through the action of certain growth factors.

For example, brain-derived neurotrophic factor, which is found in peripheral nerves<sup>147</sup> such as those used to promote transected optic nerve regrowth,<sup>144,145</sup> enhances cultured ganglion cell survival.<sup>148,149</sup> This protein is only one member of the family of neurotrophins, growth factors that stimulate differentiation, proliferation, and survival of many different CNS and peripheral nervous system target neurons through receptor binding.<sup>150</sup> Thus, following transection of hippocampally projecting axons of the medial septum, nerve growth factor, another neurotrophin, causes a dramatic increase in survival of the axotomized nerve cells<sup>151</sup> and also improves survival of axotomized ganglion cells within the eye.<sup>152</sup> Peripheral nerves that are inserted into lesioned CNS sites and that induce extensive axon regrowth express markedly increased levels of nerve growth factor.<sup>153</sup> Study of the factors that promote differentiation and growth in this paradigm may reveal an approach to preservation or reconstitution of degenerated axons in glaucoma.

## COMMENT

It seems likely that significant parts of the rapidly unfolding excitotoxin, calcium, nitric oxide, and free radical story will be germane to our

understanding of ganglion cell function and survival under stress. It may be that neural degenerative processes, such as those reviewed herein, are at work in some or all forms of glaucoma. Numerous new questions leading to novel kinds of research arise.

Could glaucomatous optic nerve atrophy be related to ischemic effects? Do studies of acute and reversible ischemic injury have application to our understanding of glaucoma, a chronic and progressive condition? Might transient and localized but recurrent ischemic episodes (related perhaps to waking IOP spikes<sup>154</sup> or to nocturnal systemic hypotension<sup>155,156</sup>) induce cytotoxic damage? Does elevated IOP lead to excess buildup of excitotoxins? Are excitotoxins responsible for neural atrophy in glaucoma? Are free radicals generated during transient ischemic/reoxygenation cycles caused by elevations in IOP? Would inhibitors of free radicals, NOS, NMDA receptor-type channels, or voltage-operated calcium channels retard glaucomatous damage? Can we replace or augment ocular hypotensive agents for the treatment of glaucoma? The answers to these questions just may favorably modulate the nerve of glaucoma!

Accepted for publication July 30, 1993.

This study was supported in part by an unrestricted grant from Research to Prevent Blindness Inc, New York, NY.

Reprint requests to the Department of Ophthalmology, Box 1183, Mount Sinai School of Medicine, One Gustave L. Levy Place, New York, NY 10029 (Dr Podos).

## REFERENCES

1. Armaly MF, Krueger DE, Maunder L, et al. Biostatistical analysis of the Collaborative Glaucoma Study, I: summary report of the risk factors for glaucomatous visual-field defects. *Arch Ophthalmol*. 1980;98:2163-2171.
2. Perkins ES. The Bedford Glaucoma Survey, I: long-term follow-up of borderline cases. *Br J Ophthalmol*. 1975;57:179-185.

3. Kitazawa Y, Horie T, Aoki S, et al. Untreated ocular hypertension. *Arch Ophthalmol*. 1977; 95:1180-1184.
4. Hollows FC, Graham PA. Intraocular pressure, glaucoma and glaucoma suspects in a defined population. *Br J Ophthalmol*. 1966;50:570-586.
5. Kahn HA, Milton RC. Alternative definitions of open-angle glaucoma: effect on prevalence and association in the Framingham Eye Study. *Arch Ophthalmol*. 1980;98:2172-2177.
6. Sommer A. Intraocular pressure and glaucoma. *Am J Ophthalmol*. 1989;107:186-188.
7. Sommer A, Tielsch JM, Katz J, et al. Relationship between intraocular pressure and primary open angle glaucoma among white and black Americans. *Arch Ophthalmol*. 1991;109:1090-1095.
8. Bankes JLK, Perkins ES, Tsolakis S, et al. Bedford Glaucoma Survey. *BMJ*. 1968;1:791-796.
9. Schulzer M, Drance SM, Carter CJ, Brooks DE, Douglas GR, Lau W. Biostatistical evidence for two distinct chronic open angle glaucoma populations. *Br J Ophthalmol*. 1990;74:196-200.
10. Chauhan BC, Drance SM. The relationship between intraocular pressure and visual field progression in glaucoma. *Graefes Arch Clin Exp Ophthalmol*. 1992;30:521-526.
11. Wilson MR. Epidemiological features of glaucoma. *Int Ophthalmol Clin*. 1990;30:153-160.
12. Anderson DR. Glaucoma: the damage caused by pressure. XLVI Edward Jackson memorial lecture. *Am J Ophthalmol*. 1989;108:485-495.
13. Drance SM. Bowman Lecture. Glaucoma: changing concepts. *Eye*. 1992;6:337-345.
14. Van Buskirk ME, Cioffi GA. Glaucomatous optic neuropathy. *Am J Ophthalmol*. 1992;113:447-452.
15. Bassett AL, Bigger JT Jr, Hoffman BF. 'Protective' action of diphenylhydantoin on canine Purkinje fibers during hypoxia. *J Pharmacol Exp Ther*. 1970;173:336-343.
16. Becker B, Stamper RL, Asseff C, Podos SM. Effect of diphenylhydantoin on glaucomatous field loss: a preliminary report. *Trans Am Acad Ophthalmol Otolaryngol*. 1972;76:412-422.
17. Kitazawa Y, Shirai H, Go FJ. The effect of Ca<sup>2+</sup>-antagonist on visual field in low-tension glaucoma. *Graefes Arch Clin Exp Ophthalmol*. 1989; 227:408-412.
18. Gasser P, Flammer J, Guthauser U, Mahler F. Do vasospasms provoke ocular disease? *Angiology*. 1990;41:213-220.
19. Serle JB, Schmidt KG, Mittag TW, Schumer RA, Podos SM, Camras CB. Nifedipine and ocular blood flow. *Invest Ophthalmol Vis Sci*. 1992; 33(suppl):1279.
20. Netland PA, Chaturvedi N, Dreyer EB. Calcium channel blockers in the management of low-tension and open-angle glaucoma. *Am J Ophthalmol*. 1993;15:608-613.
21. Piiltz JR, Bose S, Grunwald JE, Petrig BL, Riva CE. Effect of nimodipine on automated threshold perimetry, spatial contrast sensitivity and macular blood flow in normal tension glaucoma and controls. *Invest Ophthalmol Vis Sci*. 1993;34(suppl):1287.
22. Levy JC. La recherche dans le glaucome vers une neuroprotection. In: Alain Bechetolle, ed. *Normal Pressure Glaucomas*. Angers, France: Japperenard; 1990:213-221.
23. Neufeld AH. Protection of the optic nerve in glaucoma. In: Drance SM, Van Buskirk EM, Neufeld AH, eds. *Pharmacology of Glaucoma*. Baltimore, Md: Williams & Wilkins; 1992:292-300.
24. Bresnick GH. Excitotoxins: A possible new mechanism for the pathogenesis of ischemic retinal damage. *Arch Ophthalmol*. 1989;107:339-341.
25. Shatz CJ, O'Leary DDM. Repair and replacement to restore sight. *Arch Ophthalmol*. 1993; 111:472-477.
26. Choi DW, Hartley DM. Calcium and glutamate-induced cortical neuronal death. In: Waxman SG, ed. *Molecular and Cellular Approaches to the Treatment of Neurological Disease*. New York, NY: Raven Press; 1993.
27. Watkins JC, Evans RH. Excitatory amino acid transmitters. *Ann Rev Pharmacol Toxicol*. 1981; 21:165-204.
28. Watkins JC, Krosggaard-Larsen P, Honore T. Structure-activity relationships in the development of excitatory amino acid receptor agonists and competitive antagonists. *Trends Pharmacol Sci*. 1990;11:25-33.
29. Sommer B, Seeburg PH. Glutamate receptor channels: novel properties and new clones. *Trends Pharmacol Sci*. 1992;13:291-296.
30. Stevens CF. NMDA receptors: on to molecular mechanisms. *Nature*. 1992;358:18-19.
31. Daw NW, Stein PSG, Fox K. The role of NMDA receptors in information processing. *Annu Rev Neurosci*. 1993;16:207-222.
32. Albers GW, Goldberg MP, Choi DW. Do NMDA antagonists prevent neuronal injury? Yes. *Arch Neurol*. 1992;49:418-420.
33. Westbrook GL. Glutamate receptors and excitotoxicity. In: Waxman SG, ed. *Molecular and Cellular Approaches to the Treatment of Neurological Disease*. New York, NY: Raven Press; 1993.
34. Shimada N, Graf R, Rosner G, Heiss WD. Differences in ischaemia-induced accumulation of amino acids in the cat cortex. *Stroke*. 1990;21: 1445-1451.
35. Beckman JS. The double-edged role of nitric oxide in brain function and superoxide-mediated injury. *J Dev Physiol*. 1991;15:53-59.
36. Hansen AJ. Effect of anoxia on ion distribution in the brain. *Physiol Rev*. 1985;65:101-148.
37. Drejer J, Benveniste H, Diemer NH, Schousboe A. Cellular origin of ischemia-induced glutamate release from brain tissue in vivo and in vitro. *J Neurochem*. 1985;45:145-151.
38. Benveniste H, Drejer J, Schousboe A, Diemer NH. Elevation of the extracellular concentrations of glutamate and aspartate in rat hippocampus during transient cerebral ischemia monitored by intracerebral microdialysis. *J Neurochem*. 1984;43:1369-1374.
39. Choi DW. Glutamate neurotoxicity and diseases of the nervous system. *Neuron*. 1988;1:623-634.
40. Olney JW. Glutamate-induced retinal degeneration in neonatal mice. *J Neuropathol Exp Neurol*. 1969;28:455-474.
41. Zhang D, Sucher NJ, Lipton SA. Heterogeneity of AMPA and kainate receptors in rat retinal ganglion cells: subunits, flips, and flops. *Invest Ophthalmol Vis Sci*. 1993;34(suppl):1333.
42. Tung NN, Morgan IG, Ehrlich D. A quantitative analysis of the effects of excitatory neurotoxins on retinal ganglion cells in the chick. *Vis Neurosci*. 1990;4:217-223.
43. Sesma MA, Price MT, Olney JW. Sensitivity of retinal ganglion cell subtypes to specific excitotoxins. *Invest Ophthalmol Vis Sci*. 1991;32 (suppl):1263.
44. Caprioli J, Kitano S. Large retinal ganglion cells are more susceptible to excitotoxic and hypoxic injury than small cells. *Invest Ophthalmol Vis Sci*. 1993;34(suppl):1429.
45. Quigley HA, Sanchez RM, Dunkelberger GR, L'Hernault NL, Baginski TA. Chronic glaucoma selectively damages large optic nerve fibers. *Invest Ophthalmol Vis Sci*. 1987;28:913-920.
46. Dandona L, Hendrickson A, Quigley HA. Selective effects of experimental glaucoma on axonal transport by retinal ganglion cells to the dorsal lateral geniculate nucleus. *Invest Ophthalmol Vis Sci*. 1991;32:1593-1599.
47. Glovinsky Y, Quigley HA, Pease ME. Foveal ganglion cell loss is size dependent in experimental glaucoma. *Invest Ophthalmol Vis Sci*. 1993; 34:395-400.
48. Dreyer EB, Lipton SA. A proposed role for excitatory amino acids in glaucoma visual loss. *Invest Ophthalmol Vis Sci*. 1993;34(suppl): 1504.
49. Lustig HS, von Brauchitsch KL, Chan J, Greenberg DA. Ethanol and excitotoxicity in cultured cortical neurons: differential sensitivity of *N*-methyl-D-aspartate and sodium nitroprusside toxicity. *J Neurochem*. 1992;59:2193-2200.
50. Judge ME, Sheardown MJ, Jacobsen P, Honore T. Protection against post-ischemic behavioral pathology by the alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) antagonist 2,3-dihydroxy-6-nitro-7-sulfamoylbenzo(f) quinoxaline (NBQX) in the gerbil. *Neurosci Lett*. 1991;133:291-294.
51. Pai KS, Ravindranath V. Toxicity of *N*-acetylaspartylglutamate and its protection by NMDA and non-NMDA receptor antagonists. *Neurosci Lett*. 1991;126:49-51.
52. Diemer NH, Jorgensen MB, Johansen FF, Sheardown M, Honore T. Protection against ischemic hippocampal CA1 damage in the rat with a new non-NMDA antagonist, NBQX. *Acta Neurol Scand*. 1992;86:45-49.
53. Bullock R, Graham DI, Chen MH, Lowe D, McCulloch J. Focal cerebral ischaemia in the cat: pretreatment with a competitive NMDA receptor antagonist, D-CPP-ene. *J Cereb Blood Flow Metab*. 1990;10:668-674.
54. Uematsu D, Araki N, Greenberg JH, Sladky J, Reivich M. Combined therapy with MK-801 and nimodipine for protection of ischemic brain damage. *Neurology*. 1991;41:88-94.
55. David P, Lusky M, Teichberg VI. Involvement of excitatory neurotransmitters in the damage produced in chick embryo retinas by anoxia and extracellular high potassium. *Exp Eye Res*. 1988; 46:657-662.
56. Lucas DR, Newhouse JP. The toxic effect of sodium L-glutamate on the inner layers of the retina. *Arch Ophthalmol*. 1957;58:193-201.
57. Sucher NJ, Aizenman E, Lipton SA. *N*-methyl-D-aspartate antagonists prevent kainate neurotoxicity in rat retinal ganglion cells in vitro. *J Neurosci*. 1991;11:966-971.
58. Siliprandi R, Canella R, Carmignoto G, et al.

- N*-methyl-D-aspartate-induced neurotoxicity in the adult rat retina. *Vis Neurosci*. 1992;8:567-573.
59. Steinberg GK, Kunis D, Saleh J, DeLaPaz R. Protection after transient focal cerebral ischemia by the *N*-methyl-D-aspartate antagonist dextrorphan is dependent upon plasma and brain levels. *J Cereb Blood Flow Metab*. 1991;11:1015-1024.
  60. Yoon YH, Marmor MF. Dextromethorphan protects retina against ischemic injury in vivo. *Arch Ophthalmol*. 1989;107:409-411.
  61. Lam TT, Stojack K, Siew E, Chu R, Tso MOM. Ameliorative effect of a *N*-methyl-D-aspartate receptor antagonist MK801 on retinal ischemia. *Invest Ophthalmol Vis Sci*. 1993;34(suppl):1431.
  62. Beal MF, Ferrante RJ, Swartz KJ, Kowall NW. Chronic quinolinic acid lesions in rats closely resemble Huntington's disease. *J Neurosci*. 1991;11:1649-1659.
  63. Appel SH. Excitotoxic neuronal cell death in amyotrophic lateral sclerosis. *Trends Neural Sci*. 1993;16:3-5.
  64. Young AB. Role of excitotoxins in hereditodegenerative neurologic diseases. In: Waxman SG, ed. *Molecular and Cellular Approaches to the Treatment of Neurological Disease*. New York, NY: Raven Press; 1993.
  65. Greenamyre JT, O'Brien CF. *N*-methyl-D-aspartate antagonists in the treatment of Parkinson's disease. *Arch Neurol*. 1991;48:977-981.
  66. Porsche-Wiebkling E. New *N*-methyl-D-aspartate antagonists for the treatment of stroke. *Drug Devel Res*. 1989;17:367-375.
  67. Harrison MJG. Protection against ischaemia: the basis of acute stroke therapy. *Curr Opin Neurol Neurosurg*. 1992;5:33-38.
  68. Olney JW, Labruyere J, Wang G, Wozniak DF, Price MT, Sesma MA. NMDA antagonist neurotoxicity: mechanism and prevention. *Science*. 1991;254:1515-1518.
  69. Hollmann M, O'Shea-Greenfield A, Rogers SW, Heinemann S. Cloning by functional expression of a member of the glutamate receptor family. *Nature*. 1989;342:643-648.
  70. Ishii T, Moriyoshi K, Sugihara H, et al. Molecular characterization of the family of the *N*-methyl-D-aspartate receptor subunits. *J Biol Chem*. 1993;268:2836-2843.
  71. Toni D, Frontoni M, Argentino C, Sacchetti ML, De Michele M, Fieschi C. Update on calcium antagonists in cerebrovascular diseases. *J Cardiovasc Pharmacol*. 1991;18(suppl 8):S10-S14.
  72. Nuglisich J, Karkhoutly C, Menzel HD, Rossberg C, Kriegstein J. Protective effect of nimodipine against ischaemic neuronal damage in rat hippocampus without changing postischaemic cerebral blood flow. *J Cereb Blood Flow Metab*. 1990;10:654-659.
  73. Bean BP. Classes of calcium channels in vertebrate cells. *Ann Rev Physiol*. 1989;51:367-384.
  74. Pauwels PJ, Leysen JE, Janssen PAJ. Ca<sup>++</sup> and Na<sup>+</sup> channels involved in neuronal cell death: protection by flunarizine. *Life Sci*. 1991;48:1881-1893.
  75. Catterall WA, Striessnig J. Receptor sites for Ca<sup>2+</sup> channel antagonists. *Trends Pharmacol Sci*. 1992;13:256-262.
  76. Peters T, Wilffert PB, Vanhoutte PM, Van Zwieteren PA. Calcium channels in the brain as targets for the calcium-channel modulators used in the treatment of neurological disorders. *J Cardiovasc Pharmacol*. 1991;18(suppl 8):S1-S5.
  77. Akaike N, Kostyuk PG, Osipchuk YV. Dihydropyridine-sensitive low threshold calcium channels in isolated rat hypothalamic neurones. *J Physiol (Lond)*. 1989;412:181-191.
  78. Phelps CD, Corbett JJ. Migraine and low-tension glaucoma: a case-control study. *Invest Ophthalmol Vis Sci*. 1985;26:1105-1108.
  79. Schmidt R, Oestreich W. Flunarizine in migraine prophylaxis: the clinical experience. *J Cardiovasc Pharmacol*. 1991;18(suppl 8):S21-S26.
  80. Alps BJ, Calder C, Hass WK, Wilson AD. Comparative protective effects of nicardipine, flunarizine, lidoflazine and nimodipine against ischaemic injury in the hippocampus of the Mongolian gerbil. *Br J Pharmacol*. 1988;93:877-883.
  81. De Ryck M, Van Reempts J, Borgers M, Wauquier A, Janssen PAJ. Photochemical stroke model: flunarizine prevents sensorimotor deficits after neocortical infarcts in rats. *Stroke*. 1989;20:1383-1390.
  82. Watanabe Y, Shibuya T. Possible mechanism of sustained neuronal death induced by excessive glutamate and endogenous glutamate release: its protection by flunarizine and presence of glia cells in cultured cerebellar granule cells. *Clin Neuropharmacol*. 1992;15(suppl 1, pt A):132A-133A.
  83. Mermoud A, Faggioni R, Van Melle GD. Double-blind study in the treatment of normal tension glaucoma with nafidrofuryl. *Ophthalmologica*. 1990;201:145-151.
  84. Mermoud A, Faggioni R. Le traitement du glaucome à pression normale avec un antagoniste des recepteurs S2 de la Sérotonine, le nafidrofuryl (Praxilen). *Klin Monatsbl Augenheilkd*. 1991;198:332-334.
  85. Beati D, Mermoud A, Faggioni R. Effet du Nafidrofuryl (Praxilen) dans le glaucome primaire à angle ouvert (GPAO): étude prospective en double insu. *Klin Monatsbl Augenheilkd*. 1992;200:407-408.
  86. Bredt DS, Snyder SH. Nitric oxide: a novel neuronal messenger. *Neuron*. 1992;8:3-11.
  87. Bredt DS, Hwang PM, Snyder SH. Localization of nitric oxide synthase indicating a neural role for nitric oxide. *Nature*. 1990;347:768-770.
  88. O'Dell TJ, Hawkins RD, Kandel ER, Arancio O. Tests of the roles of two diffusible substances in long-term potentiation: evidence for nitric oxide as a possible early retrograde messenger. *Proc Natl Acad Sci U S A*. 1991;88:11285-11289.
  89. Dawson TM, Dawson VL, Snyder SH. A novel neuronal messenger molecule in brain: the free radical, nitric oxide. *Ann Neurol*. 1992;32:297-311.
  90. Mayer B, Klatt P, Bohme E, Schmidt K. Regulation of neuronal nitric oxide and cyclic GMP formation by Ca<sup>2+</sup>. *J Neurochem*. 1992;59:2024-2029.
  91. Garthwaite J, Garthwaite G, Palmer RM, Moncada S. NMDA receptor activation induces nitric oxide synthesis from arginine in rat brain slices. *Eur J Pharmacol*. 1989;172:413-416.
  92. Chao CC, Hu S, Molitor TW, Shaskan EG, Peterson PK. Activated microglia mediate neuronal cell injury via a nitric oxide mechanism. *J Immunol*. 1992;149:2736-2741.
  93. Wallis RA, Panizzon K, Wasterlain CG. Inhibition of nitric oxide synthase protects against hypoxic neuronal injury. *Neuroreport*. 1992;3:645-648.
  94. Halliwell B. Reactive oxygen species and the central nervous system. *J Neurochem*. 1992;59:1609-1623.
  95. Verma A, Hirsch DJ, Glatt CE, Ronnett GV, Snyder SH. Carbon monoxide: a putative neural messenger. *Science*. 1993;259:381-384.
  96. Knowles RG, Palacios M, Palmer RM, Moncada S. Formation of nitric oxide from L-arginine in the central nervous system: a transduction mechanism for stimulation of the soluble guanylate cyclase. *Proc Natl Acad Sci U S A*. 1989;86:5159-5162.
  97. Moncada S, Palmer RMJ, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev*. 1991;43:109-142.
  98. Dawson VL, Dawson TM, London ED, Bredt DS, Snyder SH. Nitric oxide mediates glutamate neurotoxicity in primary cortical cultures. *Proc Natl Acad Sci U S A*. 1991;88:6368-6371.
  99. Nowicki JP, Duval D, Poignet H, Scatton B. Nitric oxide mediates neuronal death after focal cerebral ischemia in the mouse. *Eur J Pharmacol*. 1991;204:339-340.
  100. Dawson TM, Bredt DS, Fotuhi M, Hwang PM, Snyder SH. Nitric oxide synthase and neuronal NADPH diaphorase are identical in brain and peripheral tissues. *Proc Natl Acad Sci U S A*. 1991;88:7797-7801.
  101. Geyer O, Podos SM, Mittag TW. Nitric oxide synthase: distribution and biochemical properties of the enzyme in the bovine eye. *Invest Ophthalmol Vis Sci U S A*. 1993;34(suppl):826.
  102. Bredt DS, Glatt CE, Hwang PM, Fotuhi M, Dawson TM, Snyder SH. Nitric oxide synthase protein and mRNA are discretely localized in neuronal populations of the mammalian CNS together with NADPH diaphorase. *Neuron*. 1991;7:615-624.
  103. Hope BT, Michael GJ, Knigge KM, Vincent SR. Neuronal NADPH diaphorase is a nitric oxide synthase. *Proc Natl Acad Sci U S A*. 1991;88:2811-2814.
  104. Koh JY, Peters S, Choi DW. Neurons containing NADPH-diaphorase are selectively resistant to quinolinic toxicity. *Science*. 1986;234:73-76.
  105. Ferrante RJ, Kowall NW, Beal MF, Martin JB, Bird ED, Richardson EP Jr. Morphologic and histochemical characteristics of a spared subset of striatal neurons in Huntington's disease. *J Neuropathol Exp Neurol*. 1987;46:12-27.
  106. Uemura Y, Kowall NW, Beal MF. Selective sparing of NADPH-diaphorase-somatostatin-neuropeptide Y neurons in ischemic gerbil striatum. *Ann Neurol*. 1990;27:620-625.
  107. Beal MF, Swartz KJ, Finn SF, Mazurek MF, Kowall NW. Neurochemical characterization of excitotoxic lesions in the cerebral cortex. *J Neurosci*. 1991;11:147-158.
  108. Unger JW, Lange W. NADPH-diaphorase-positive cell populations in the human amygdala



- and temporal cortex: neuroanatomy, peptidergic characteristics and aspects of aging and Alzheimer's disease. *Acta Neuropathol.* 1992; 83:636-646.
109. Sagar SM. NADPH-diaphorase reactive neurons of the rabbit retina: differential sensitivity to excitotoxins and unusual morphologic features. *J Comp Neurol.* 1990;300:309-319.
  110. Kontos HA. Oxygen radicals in CNS damage. *Chem Biol Interact.* 1989;72:229-255.
  111. Granger DN, Hollwarth ME, Parks DA. Ischemia-reperfusion injury: role of oxygen-derived free radicals. *Acta Physiol Scand.* 1986;548(suppl): 47-63.
  112. Juarez CP, Tso MOM, van Heuven WAJ, Hayreh MS, Hayreh SS. Experimental retinal vascular occlusion, II: a clinicopathologic correlative study of simultaneous occlusion of central retinal vein and artery. *Int Ophthalmol.* 1986;9:77-87.
  113. Faberowski N, Stefannsson E, Davidson RC. Local hypothermia protects the retina from ischemia. *Invest Ophthalmol Vis Sci.* 1989;30: 2309-2313.
  114. Hearse DJ. Reperfusion-induced injury: a possible role for oxidant stress and its manipulation. *Cardiovasc Drugs Ther.* 1991;5:225-236.
  115. Chan PH, Yurko M, Fishman RA. Phospholipid degradation and cellular edema induced by free radicals in brain cortical slices. *J Neurochem.* 1982;38:525-531.
  116. Braughler JM, Hall ED. Central nervous system trauma and stroke. I: biochemical considerations for oxygen radical formation and lipid peroxidation. *Free Radic Biol Med.* 1989;6:289-301.
  117. Hall ED, Braughler JM. Central nervous system trauma and stroke, II: physiological and pharmacological evidence for involvement of oxygen radicals and lipid peroxidation. *Free Radic Biol Med.* 1989;6:303-313.
  118. Jesberger JA, Richardson JS. Oxygen free radicals and brain dysfunction. *Int J Neurosci.* 1991; 57:1-17.
  119. Chan PH, Yurko M, Fishman RA. Phospholipid degradation and cellular edema induced by free radicals in brain cortical slices. *J Neurochem.* 1982;38:525-531.
  120. Wei EP, Kontos HA. Oxygen radicals in cerebral ischemia. *Physiologist.* 1987;30:122.
  121. Kirsch JR, Phelan AM, Lange DG, Traystman RJ. Free radicals detected in brain during reperfusion from global ischemia. *Federation Proc.* 1987;46:799.
  122. Kurihara M. Role of monoamines in experimental spinal cord injury in rats: relationship between  $\text{Na}^+$ - $\text{K}^+$ -ATPase and lipid peroxidation. *J Neurosurg.* 1985;62:743-749.
  123. Saunders RD, Dugan LL, Demediuk P, Means ED, Horrocks LA, Anderson DK. Effects of methylprednisolone and the combination of  $\alpha$ -tocopherol and selenium on arachidonic acid metabolism and lipid peroxidation in traumatized spinal cord tissue. *J Neurochem.* 1987;49: 24-31.
  124. Davis RJ, Bulkley GB, Traystman RJ. Role of oxygen-free radicals in focal brain ischemia. *Federation Proc.* 1987;46:799.
  125. Cerchiari EL, Hoel TM, Safar P, Sclabassi RJ. Protective effects of combined superoxide dismutase and deferoxamine on recovery of cerebral blood flow and function after cardiac arrest in dogs. *Stroke.* 1987;18:869-878.
  126. Hara H, Kogure K. Prevention of hippocampus neuronal damage in ischemic gerbils by a novel lipid peroxidation inhibitor (quinazoline derivative). *J Pharmacol Exp Ther.* 1990;255:906-913.
  127. Szabo ME, Droy-Lefaix MT, Doly M, Carre C, Braquet P. Ischemia and reperfusion-induced histologic changes in the rat retina: demonstration of a free radical-mediated mechanism. *Invest Ophthalmol Vis Sci.* 1991;32:1471-1478.
  128. Szabo ME, Droy-Lefaix MT, Dolly M, Braquet P. Ischaemia- and reperfusion-induced  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  shifts in rat retina: effects of two free radical scavengers, SOD and EGB 761. *Exp Eye Res.* 1992;55:39-45.
  129. Szabo ME, Droy-Lefaix MT, Doly M, Braquet P. Modification of ischemia/reperfusion-induced ion shifts ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ ) by free radical scavengers in the rat retina. *Ophthalmic Res.* 1993;25:1-9.
  130. Droy-Lefaix MT, Bonhomme B, Doly M. Protective effect of Ginkgo biloba extract (EGB 761) on free radical-induced changes in the electroretinogram of isolated rat retina. *Drugs Exp Clin Res.* 1991;17:571-574.
  131. Nayak MS, Kita M, Marmor MF. Protection of rabbit retina from ischemic injury by superoxide dismutase and catalase. *Invest Ophthalmol Vis Sci.* 1993;34:2018-2022.
  132. Veriac S, Tissie G, Bonne C. Oxygen free radicals adversely affect the regulation of vascular tone by nitric oxide in the rabbit retina under high intraocular pressure. *Exp Eye Res.* 1993; 56:85-88.
  133. Gryglewski RJ, Palmer RMJ, Moncada S. Superoxide anion is involved in the breakdown of endothelium-derived vascular relaxing factor. *Nature.* 1986;320:454-456.
  134. Sun AY, Cheng Y, Bu Q, Oldfield F. The biochemical mechanisms of the excitotoxicity of kainic acid: free radical formation. *Mol Chem Neuropathol.* 1992;17:51-63.
  135. Aizenman E, Hartnett KA, Reynolds IJ. Oxygen free radicals regulate NMDA receptor function via a redox modulatory site. *Neuron.* 1990;5: 841-846.
  136. Pellegrini-Giampietro DE, Cherici G, Alesiani M, Carla V, Moroni F. Excitatory amino acid release and free radical formation may cooperate in the genesis of ischemia-induced neuronal damage. *J Neurosci.* 1990;10:1035-1041.
  137. Rosen DR, Siddique T, Patterson D, et al. Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature.* 1993;362:59-62.
  138. McNamara JO, Fridovich I. Did radicals strike Lou Gehrig? *Nature.* 1993;362:20-21.
  139. Rothstein JD, Martin LJ, Kuncl RW. Decreased glutamate transport by the brain and spinal cord in amyotrophic lateral sclerosis. *N Engl J Med.* 1992;326:1464-1468.
  140. Choi DW. Amyotrophic lateral sclerosis and glutamate: too much of a good thing? *N Engl J Med.* 1992;326:1493-1495.
  141. Kinoshita A, Yamada K, Kohmura E, Hayakawa T. Human recombinant superoxide dismutase protects primary cultured neurons against hypoxic injury. *Pathobiology.* 1991;59:340-344.
  142. Ohtsuki T, Matsumoto M, Kuwabara K, et al. Influence of oxidative stress on induced tolerance to ischemia in gerbil hippocampal neurons. *Brain Res.* 1992;599:246-252.
  143. Villegas-Pérez MP, Vidal-Sanz M, Bray GM, Aguayo AJ. Influences of peripheral nerve grafts on the survival and regrowth of axotomized retinal ganglion cells in adult rats. *J Neurosci.* 1988; 8:265-280.
  144. Aguayo AJ, Rasminsky M, Bray GM, et al. Degenerative and regenerative responses of injured neurons in the central nervous system of adult mammals. *Philos Trans R Soc Lond Biol.* 1991;331:337-343.
  145. Bray GM, Villegas-Pérez MP, Vidal-Sanz M, Carter DA, Aguayo AJ. Neuronal and nonneuronal influences on retinal ganglion cell survival, axonal regrowth, and connectivity after axotomy. *Ann N Y Acad Sci.* 1991;633:214-228.
  146. Kierstead SA, Rasminsky M, Fukuda Y, Carter DA, Aguayo AJ, Vidal-Sanz M. Electrophysiological responses in hamster superior colliculus evoked by regenerating retinal axons. *Science.* 1989;246:255-258.
  147. Barker P, Acheson A, Pareek S, Miller FD, Murphy RA. Detection of brain-derived neurotrophic factor (BDNF)-like biological activity and mRNA in sciatic nerve fibroblasts and Schwann cells. *Soc Neurosci Abstr.* 1990;16: 1136.
  148. Johnson JE, Barde Y-A, Schwab M, Thoenen H. Brain-derived neurotrophic factor supports the survival of cultured rat retinal ganglion cells. *J Neurosci.* 1986;6:3031-3038.
  149. Thanos S, Bähr M, Barde Y-A, Vanselow J. Survival and axonal elongation of adult rat retinal ganglion cells. *Eur J Neurosci.* 1989;1:19-26.
  150. Ebendal T. Function and evolution in the NGF family and its receptors. *J Neurosci Res.* 1992; 32:461-470.
  151. Kromer LF. Nerve growth factor treatment after brain injury prevents neuronal death. *Science.* 1987;235:214-216.
  152. Carmignoto G, Maffei L, Candeo P, Canella R, Comelli C. Effect of NGF on the survival of rat retinal ganglion cells following optic nerve section. *J Neurosci.* 1989;9:1263-1272.
  153. Messersmith DJ, Fabrizio M, Mocchielli I, Kromer LF. Effects of sciatic nerve transplants after fimbria-fornix lesion: examination of the role of nerve growth factor. *Brain Res.* 1991;557:293-297.
  154. Wilensky JT, Asrani SG, Baca L, Holck D, Viana M. Waking IOP spikes. *Invest Ophthalmol Vis Sci.* 1993;34(suppl):1383.
  155. Graham SL, Drance SM, Wijsman K, Mikelberg FS, Douglas GR. Nocturnal hypotension in glaucoma patients. *Invest Ophthalmol Vis Sci.* 1993; 34(suppl):1286.
  156. Hayreh SS, Zimmermann MB, Podhajsky P, Alward WL. The role of nocturnal hypotension in ocular and optic nerve ischemic disorders. *Invest Ophthalmol Vis Sci.* 1993;34(suppl):994.