Kacey Diagnostics Quick Reference For Culture & Sensitivity Study

1. **INOCULATION OF CULTURE BI-PLATE**

   1) Remove the plate from refrigerator & pre-heat in the incubator @ 37 degrees for 15-20 minutes prior to inoculating the Kacey Multichrome bi-plate.
   
   2) Using a sterile inoculating loop (20uL) for liquids and a Kacey Sterile Swab for solids, inoculate the sample onto both sides of the Kacey Multichrome Bi-plate, by utilizing a zigzag streaking motion.
   
   3) Replace clear lid and place Kacey Multichrome Bi-plate in the incubator *upside down* (inverted position) and incubate at 37°C +/- 2 degrees C for no less than 24 hours. The plate should be examined after 24 hours, but no later than 48 hours after incubation.

2. **TRANSFER TO MULLER HINTON PLATE USING KACEY “WST” TUBES**

   1) Taking a Kacey Sterile Swab or Loop carefully dab into the three (3) different places containing the bacteria on the MultiChrome Bi-plate.
   
   2) Immediately place the sterile Kacey Swab or Loop containing the bacteria into the Kacey Working Solution tube (WST which contains 1.0 ml of sterile 0.085% saline solution). Mix the Kacey Swab or Loop with a gentle twirling motion while in the WST tube for approximately 3-5 seconds.
   
   3) Compare sample to the Kacey Turbidity Standard (white cap) KTS. Sample should match in turbidity, if not add more WST sol (green top) or more sample to match KTS tube. (see enclosed Turbidity Card).

3. **INOCULATION OF MULLER HINTON (MH) PLATE**

   1) Using the Kacey Swab or loop containing the new diluted WST & Turbidity adjusted dilution streak the 1st time the entire periphery of the Mueller Hinton plate making a 360° circle.
   
   2) Streak a 2nd time again the Mller Hinton Plate with a fresh sample of the WST dilution using wide broad strokes starting at the 9 to 3 o’clock position.
   
   3) Streak a 3rd time again the Mller Hinton plate with a fresh sample of the WST dilution using wide broad strokes starting at the 11 to 5 o’clock position.
   
   4) Remove “Sensi-Ring” from the foil pouch with tweezers at the inner tab & place the “Sensi-Ring” facing down onto the MH plate tapping down in non-disk areas, label the specimen to be incubated.
   
   5) Place the MH Inoculated plate into an incubator *upside down*, set timer & incubate for 24 hours. Remove & read inhibition zones with Kacey Clear Acetate Reader.

Quick reference only. Refer to detailed instruction manual for additional information.

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#### Kacey Diagnostics

**MICROBIAL ZONE INHIBITION INTERPRETIVE CHART**

For Gram Positive, Gram Negative, Ear, UTI and Skin/Wound Sensi-Rings™

<table>
<thead>
<tr>
<th>Antimicrobial Agent Name</th>
<th>Sensi Code</th>
<th>Disc Potency</th>
<th>Resistant</th>
<th>Intermediate</th>
<th>Susceptible</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amikacin</td>
<td>AK30</td>
<td>30ug</td>
<td>≤14</td>
<td>7-9</td>
<td>≥17</td>
</tr>
<tr>
<td>Amoxicillin</td>
<td>A30</td>
<td>30ug</td>
<td>≤13</td>
<td>14-17</td>
<td>≥18</td>
</tr>
<tr>
<td>Augmentin-(Clavamox)</td>
<td>AUG30</td>
<td>30ug</td>
<td>≤13</td>
<td>14-17</td>
<td>≥18</td>
</tr>
<tr>
<td>Azithromycin-(Zithromax)</td>
<td>ATH12</td>
<td>15ug</td>
<td>≤13</td>
<td>14-17</td>
<td>≥18</td>
</tr>
<tr>
<td>Cefazidime</td>
<td>CA230</td>
<td>30ug</td>
<td>≤14</td>
<td>15-17</td>
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</tr>
<tr>
<td>Cefepoxide</td>
<td>CPD10</td>
<td>15ug</td>
<td>≤17</td>
<td>18-20</td>
<td>≥21</td>
</tr>
<tr>
<td>Cephalaxin-(Keflex)</td>
<td>CFX30</td>
<td>30ug</td>
<td>≤14</td>
<td>15-17</td>
<td>≥18</td>
</tr>
<tr>
<td>Ciprofloxacin -(Cipro)</td>
<td>CPS</td>
<td>5ug</td>
<td>≤15</td>
<td>16-20</td>
<td>≥18</td>
</tr>
<tr>
<td>Clindamycin -(CliniCaps)</td>
<td>CD2</td>
<td>2ug</td>
<td>≤14</td>
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</tr>
<tr>
<td>Cotrimoxazole (trimethoprim/sulfa)</td>
<td>TS25</td>
<td>30ug</td>
<td>≤10</td>
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</tr>
<tr>
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