

Regulation of Gene Expression

Friday, September 5, 2025 6:11 AM

I. Overview

- Gene expression: Multistep process producing functional RNA or protein.
- Primary regulation: Transcription (DNA \rightarrow RNA) in both prokaryotes & eukaryotes.
- Eukaryotes: Additional regulation via posttranscriptional, posttranslational processes, and DNA accessibility.
- Types of genes:
 - Constitutive/housekeeping genes: Constant expression; needed for basic cellular functions.
 - Regulated genes: Expressed under specific conditions; may be cell-type specific (e.g., fibrinogen alpha chain in hepatocytes).
- Importance: Allows cellular differentiation, morphogenesis, and adaptability.

- Prokaryotes vs eukaryotes: Control mechanisms best understood in prokaryotes; many principles repeat in eukaryotes.

II. Regulatory Sequences and Molecules

- Cis-acting elements: DNA sequences on the same chromosome influencing gene expression (e.g., promoters, operators).
- Trans-acting factors: Proteins or molecules that can diffuse and regulate genes on any chromosome (e.g., transcription factors).
- Mechanism: Regulatory proteins bind DNA via structural motifs:
 - Zinc finger
 - Leucine zipper
 - Helix-turn-helix

III. Regulation of Prokaryotic Gene Expression

- Primary control: At transcription level.
- Efficiency: Regulating the first step prevents waste of

energy.

- Methods: Initiation control or premature transcription termination.

A. mRNA Transcription from Operons

- Operon: Cluster of structural genes + regulatory sequences controlling proteins in a metabolic pathway.
- Polycistronic mRNA: Single mRNA encodes multiple proteins; allows coordinated regulation.

B. Operators in Operons

- Operator: DNA segment where a repressor protein binds to block transcription.
 - Repressor bound: RNA polymerase cannot transcribe → no mRNA/protein produced.
 - Inducer present: Binds repressor → repressor releases operator → RNA polymerase initiates transcription.
- Example: Lactose (lac) operon – demonstrates both negative and positive regulation.

C. Lac Operon

- Structural genes:
 - lacZ: β -galactosidase \rightarrow lactose \rightarrow glucose + galactose.
 - lacY: Permease \rightarrow lactose transport into cell.
 - lacA: Thiogalactoside transacetylase \rightarrow acetylates lactose (function unclear).
- Regulation: Maximally expressed only when lactose is present & glucose absent.
- Regulatory elements:
 - Promoter: RNA polymerase binds.
 - Operator (O): Repressor binds here.
 - CAP site: Activated by cAMP-CAP complex \rightarrow enhances transcription.
- Repressor: LacI gene (trans-acting factor) binds operator with high affinity; LacI has its own promoter and is separate from the operon.

✓ Exam Points:

- Prokaryotic operons = polycistronic mRNA + cis-regulatory sequences + trans-acting factors.
- Lac operon: classic negative (repressor) + positive (CAP-cAMP) regulation example.
- Glucose preference → low cAMP → CAP inactive → lac operon not expressed.

C. Lac Operon Regulation

1. Only glucose available

- Lac operon repressed (turned off).
- Mechanism:
 - Repressor protein binds operator (O site) via helix-turn-helix motif.
 - RNA polymerase cannot bind promoter → transcription inhibited.
- Type of regulation: Negative regulation.

2. Only lactose available

- Lac operon induced (maximally expressed).

- Mechanism:
 - Lactose → small amount converted to allolactose (inducer).
 - Allolactose binds repressor → repressor releases operator → RNA pol can transcribe.
 - cAMP-CAP complex binds CAP site (active adenyl cyclase since glucose absent) → transcription initiation enhanced.
- Type of regulation: Positive regulation.
- Outcome: Single polycistronic mRNA → lacZ, lacY, lacA proteins synthesized.
- Note: lacI gene (repressor) is constitutive, always expressed.

3. Both glucose and lactose available

- Lac operon uninduced, transcription negligible.
- Mechanism:
 - Glucose inhibits adenyl cyclase → no cAMP →

CAP site empty.

- RNA pol cannot initiate efficiently even though operator is free.

- Term: Catabolite repression.
- Expression level: Basal (very low).

D. Tryptophan (trp) Operon

- Contains 5 structural genes → enzymes for Trp synthesis.
- Regulation: Negative control by Trp as corepressor + attenuation.

Mechanisms

1. Negative control:

- Trp binds repressor → repressor binds operator → transcription blocked.

2. Attenuation:

- Transcription starts but terminates prematurely if

Trp abundant.

- Attenuator = hairpin (stem-loop) structure in mRNA → stops transcription.
- If Trp scarce → ribosomes stall at Trp codons → hairpin does not form → transcription continues.

Key difference vs eukaryotes:

- Attenuation possible in prokaryotes because transcription & translation occur simultaneously.
- Eukaryotes have spatial separation → no attenuation.

E. Coordination of Transcription and Translation

I. Stringent Response

- Triggered by amino acid starvation in *E. coli*.
- Mechanism:
 - Uncharged tRNA binds ribosome → activates RelA enzyme → synthesizes ppGpp (alarmone).
 - ppGpp binds RNA pol → inhibits rRNA & tRNA synthesis.

- mRNA for ribosomal proteins also partially inhibited.
- mRNA for amino acid biosynthesis enzymes not inhibited.
- Purpose: Prevents wasteful ribosome production; prioritizes amino acid synthesis.

2. Regulatory r-proteins

- Excess ribosomal proteins (r-proteins) inhibit their own operons.
- Mechanism:
 - Specific r-protein binds Shine-Dalgarno (SD) sequence → blocks small ribosomal subunit → translation inhibited.
 - r-protein also binds rRNA (higher affinity).
 - Balances rRNA and r-protein synthesis → ensures proper ribosome assembly.

✓ Exam Points

- Lac operon: Negative (repressor) + Positive (CAP-cAMP).

- Catabolite repression: Glucose inhibits lac operon via low cAMP.
- Trp operon: Repressible, negative control + attenuation.
- Stringent response: ppGpp → inhibits rRNA synthesis during amino acid scarcity.
- r-protein feedback: Prevents overproduction; coordinates with rRNA.

IV. Regulation of Eukaryotic Gene Expression

Eukaryotes have complex genomes and a nuclear membrane, requiring multiple regulatory mechanisms.

- Transcription is still the primary site of regulation.
- No operons in eukaryotes → use alternate strategies for coordinated regulation.
- Regulation occurs at multiple levels:
 - Transcriptional → transcription factors + cis-elements
 - Posttranscriptional → mRNA processing & stability

- Translational / Posttranslational → protein stability, processing, targeting

A. Coordinate Regulation

- Needed for simultaneous expression of functionally related genes, often on different chromosomes.
- Mechanism:
 - Specific transcription factor (STF) = trans-acting protein
 - Binds cis-acting regulatory sequences on each gene
 - STF has:
 - DNA-binding domain (DBD) → binds DNA
 - Transcription-activation domain (TAD) → recruits coactivators (e.g., histone acetyltransferases) + general transcription factors → RNA pol binds promoter → transcription
- Combinatorial control: Effect depends on protein composition of the complex

Examples of Coordinate Regulation

1. Galactose circuit (yeast)

- Genes for galactose metabolism are on different chromosomes.
- Gal4 (STF) binds UASGal (upstream activating sequence).
- Gal80 inhibits Gal4 in absence of galactose.
- Gal3 binds Gal80 when galactose present → Gal4 activates transcription.
- Glucose inhibits Gal4 expression → prevents galactose use.

2. Hormone response system

- Hormone response elements (HREs) = DNA sequences that bind trans-acting proteins
- Hormones: bind intracellular receptors (steroids) or cell-surface receptors (peptides)

a. Intracellular (nuclear) receptors

- Examples: steroid hormones, vitamin D, thyroid hormone, retinoic acid
- Domains: DNA-binding, transcriptional activation, ligand-binding
- Mechanism: hormone binds receptor →

conformational change → dimerization → binds HRE (e.g., GRE) → recruits coactivators → transcription

- Can activate or repress target genes; allows coordinate expression across chromosomes

b. Cell-surface receptors

- Examples: insulin, glucagon, epinephrine
- Mechanism: hormone binds G-protein-coupled receptor → cAMP → protein kinase A → phosphorylates CREB (STF) → binds CRE (cis-element) → transcription of target genes (e.g., gluconeogenesis enzymes like PEPCK, glucose-6-phosphatase)

B. Messenger RNA Processing and Use

Eukaryotic mRNA undergoes processing before translation:

1. 5' capping → stability & ribosome recognition
2. 3' polyadenylation → stability, translation regulation
3. Splicing → remove introns

1. Alternative Splicing

- Generates tissue-specific protein isoforms from same pre-mRNA
- Mechanisms: exon skipping, intron retention, alternative splice donor/acceptor sites
- Example: Tropomyosin (TM) → tissue-specific isoforms
- Fact: >90% of human genes undergo alternative splicing

2. Alternative Polyadenylation (APA)

- Pre-mRNAs may have multiple cleavage/polyadenylation sites
- APA → different 3' UTRs or coding sequences → affects translation & localization
- Example: IgM → secreted vs membrane-bound forms

Note: Alternative splicing + APA + alternative transcription start sites → ~20,000–25,000 genes → >100,000 proteins

3. mRNA Editing

- Base changes after mRNA processing → alters protein sequence
- Example: Apolipoprotein B (apo B)
 - Liver → full-length apo B-100 → VLDL
 - Intestine → cytosine (C) → uracil (U) → stop codon → shorter apo B-48 → chylomicrons

✓ Exam Points

- Coordinate regulation: STF + cis-elements → multiple genes on different chromosomes
- Galactose circuit: Gal4 (activator) + Gal80/Gal3 regulation
- Hormone responses: intracellular (steroids) vs cell-surface (peptides → cAMP → CREB)
- mRNA regulation: alternative splicing, APA, editing → protein diversity

4. Messenger RNA (mRNA) Stability

- The lifespan of an mRNA in the cytosol determines how much protein can be produced.
- Key examples: iron metabolism and RNA interference (RNAi).

a. Iron Metabolism

- Transferrin (Tf): plasma protein transporting iron.
- Tf binds transferrin receptors (TfR) → internalization → iron delivery.
- TfR mRNA: contains iron-responsive elements (IREs) in 3'-UTR → stem-loop structures.
- Iron regulatory proteins (IRPs) bind IREs:
 - Low iron: IRPs bind 3'-IRE → stabilize TfR mRNA → more TfR made.
 - High iron: IRPs dissociate → mRNA degraded → TfR decreased.
- Ferritin mRNA: has a 5'-IRE
 - Low iron: IRPs bind → block translation → less ferritin

- High iron: IRPs dissociate → ferritin synthesized to store excess iron
- Heme synthesis enzyme (ALA synthase 2) also contains 5'-IRE → regulated similarly.

b. RNA Interference (RNAi)

- Gene silencing by mRNA degradation or translation repression.
- Mediated by microRNAs (miRNAs) (~22 nt, noncoding).
- Processing steps:
 1. pri-miRNA → pre-miRNA by Drosha (nucleus)
 2. Export to cytoplasm → Dicer → short double-stranded miRNA
 3. Guide strand associates with RISC → hybridizes to 3'-UTR of target mRNA
 4. Outcome: translation repression or degradation (via Argonaute/Slicer)
- siRNA (short interfering RNA) can also trigger RNAi →

therapeutic potential

Example of RNAi therapy:

- Patisiran (2018) → treats hATTR amyloidosis
- siRNA inhibits abnormal TTR protein → prevents amyloid deposition

S. mRNA Translation Regulation

- Phosphorylation of eukaryotic initiation factor eIF-2 → inhibits translation initiation
- Prevents GDP→GTP exchange → blocks translation
- Kinases activated by:
 - Amino acid starvation
 - Heme deficiency in erythroblasts
 - Viral double-stranded RNA
 - Misfolded proteins in rough ER

C. Regulation through DNA Variation

Eukaryotic gene expression is influenced by DNA accessibility, gene copy number, and arrangement.

1. Access to DNA

- DNA + histone & nonhistone proteins → chromatin
- Euchromatin: transcriptionally active, decondensed
- Heterochromatin: transcriptionally inactive, condensed
- Histone modifications (acetylation, methylation, phosphorylation)
 - Reduce positive charge → loosen DNA → allow transcription factor access
- DNA methylation (CpG islands in promoters)
 - Hypomethylation → active genes
 - Hypermethylation → silenced genes
- These are epigenetic changes → heritable, no DNA sequence change

2. Gene Copy Number

- More gene copies → more protein product

- Example: Methotrexate resistance
 - DHFR gene amplification → more enzyme → survival despite drug

3. DNA Arrangement

- Immunoglobulin production in B cells:
 - Heavy/light chains = variable + constant regions
 - Variable region = somatic recombination of V, D, J segments → antibody diversity
- Pathologic rearrangements: chromosome translocations → disease

4. Mobile DNA Elements

- Transposons (Tn): mobile DNA segments
 - Move via transposase:
 - Direct transposition: cut & paste
 - Replicative transposition: copy inserted elsewhere
 - Retrotransposons: RNA intermediate → reverse transcription → new DNA copy

- Significance:
 - Contributes to genome variation
 - Can alter gene expression → cause disease (e.g., hemophilia A, Duchenne muscular dystrophy)
 - In bacteria: Tn on plasmids → antibiotic resistance

✓ Exam Points

- mRNA stability: iron metabolism (TfR/ferritin), RNAi (miRNA, siRNA)
- Translation regulation: eIF-2 phosphorylation → environmental stress response
- DNA-level regulation: chromatin accessibility, gene copy number, rearrangement, mobile DNA elements
- Epigenetics: histone modifications + DNA methylation → heritable gene expression control

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