

**SOAH DOCKET NO. 582-20-1895
TCEQ DOCKET NO. 2019-1156-IWD**

**IN THE MATTER OF THE
APPLICATION OF PORT OF
CORPUS CHRISTI AUTHORITY OF
NUECES COUNTY FOR TPDES
PERMIT NO. WQ0005253000**

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**BEFORE THE STATE OFFICE

OF

ADMINISTRATIVE HEARINGS**

EXHIBIT PAC-48R

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IN THE MATTER OF THE	§	BEFORE THE STATE OFFICE
APPLICATION OF PORT OF	§	
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PERMIT NO. WQ0005253000	§	ADMINISTRATIVE HEARINGS

REMAND PREFILED TESTIMONY

OF

KRISTIN NIELSEN

ON BEHALF OF

PORT ARANSAS CONSERVANCY

SUBMITTED ON FEBRUARY 2, 2022

**SOAH DOCKET NO. 582-20-1895
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REMAND PREFILED TESTIMONY OF KRISTIN NIELSEN, PH.D.

TABLE OF CONTENTS

I.	INTRODUCTION	3
II.	QUALIFICATIONS	3
III.	SUMMARY OF OPINIONS	7
IV.	OPINIONS REGARDING THE ECOLOGICALLY SENSITIVE NATURE OF THE AREA ADJACENT TO THE PROPOSED DISCHARGE.....	9
V.	OPINIONS REGARDING SALINITY CHANGES UPON DISCHARGE	10
VI.	OPINIONS REGARDING CONTAMINANTS OTHER THAN SALINITY UPON DISCHARGE.....	21
VII.	OPINIONS REGARDING THE LOCATION OF THE OUTFALL	26
VIII.	OPINIONS REGARDING MULTI-STRESSOR EFFECTS AND LATENT MORTALITY	28
IX.	CONCLUSION.....	29

LIST OF EXHIBITS

Exhibit PAC-48R KN-1	Curriculum Vitae
Exhibit PAC-48R KN-2	Proposed Harbor Island Seawater Reverse Osmosis Desalination Facility: A Prospective Evaluation of Toxicological Risk (March 2021)
Exhibit PAC-48R KN-3	Report documenting rangefinder test for the median lethal concentration (Test 1), the results of the first median lethal time test (Test 2), and all data from follow-up testing
Exhibit PAC-48R KN-4	Manuscript submitted for peer review
Exhibit PAC-48R KN-5	Referenced Literature
Exhibit PAC-48R KN-6	Historical rainfall records of Port Aransas

1 I. INTRODUCTION

2 Q. PLEASE STATE YOUR NAME.

3 A. Kristin M. Nielsen.

4 Q. PLEASE STATE YOUR ADDRESS.

5 A. My address is 15345 Beaufort Court, Corpus Christi, TX 78418.

6 Q. WHAT HAVE YOU BEEN ASKED TO DO WITH REGARD TO THIS MATTER,
7 SOAH DOCKET NO. 582-20-1895?

8 A. I have been retained by the Port Aransas Conservancy to review and evaluate the
9 application of the Port of Corpus Christi Authority of Nueces County (the Port of Corpus
10 Christi) for a water quality permit for a proposed seawater desalination facility on Harbor
11 Island in Nueces County, Texas, as well as the draft permit prepared by TCEQ. I have been
12 asked to review documents and provide my professional assessment as to the potential
13 effects of the proposed activities, specifically, the potential effects on aquatic and aquatic-
14 dependent life. I have also been asked to prepare this written testimony and to testify at the
15 hearing related to the Port of Corpus Christi's permit application and the draft permit.

16 II. QUALIFICATIONS

17 Q. PLEASE BRIEFLY DESCRIBE YOUR EDUCATIONAL BACKGROUND.

18 A. I earned a B.A. in Biology from Texas A&M University and a Ph.D. in Aquatic Toxicology
19 from the University of North Texas (UNT). My dissertation research examined the
20 maternal transfer and developmental toxicity of dietary methylmercury to early life stage
21 fish (*Pimephales promelas*). While completing my Ph.D., I also participated in research
22 conducted in support of the National Resource Damage Assessment (NRDA) for the
23 *Deepwater Horizon* oil spill, which investigated the combined toxicity of oil and intense
24 solar radiation to early life stage marine organisms in the Gulf of Mexico, including red
25 drum (*Sciaenops ocellatus*), speckled seatrout (*Cynoscion nebulosus*), red snapper
26 (*Lutjanus campechanus*), mahi mahi (*Coryphaena hippurus*), bay anchovy (*Anchoa*

1 *mitchilli*), fiddler crab (*Uca longisignalis*), and others. After graduating with my Ph.D., I
2 completed a two-year postdoctoral research fellowship in Aquatic Toxicology at UNT,
3 while also teaching Aquatic Toxicology to graduate/upper division students as an adjunct
4 faculty member. A large part of my postdoctoral research was focused on examining the
5 photo-enhanced toxicity of oil to early life stage estuarine and marine fish in the Gulf of
6 Mexico, including improving risk prediction models. These studies were funded by NOAA
7 as part of a separate Damage Assessment that I worked on (Taylor Energy Site).

8 **Q. PLEASE BRIEFLY DESCRIBE YOUR CURRENT OCCUPATION AND OTHER**
9 **RELEVANT PROFESSIONAL EXPERIENCE.**

10 **A.** I am currently an Assistant Professor at the University of Texas at Austin Marine Science
11 Institute in Port Aransas, Texas, where my lab researches how chemical and physical
12 environmental stressors (separately and in combination with one another) adversely impact
13 the health of aquatic ecosystems. Although my work incorporates levels of biological
14 organization through the whole ecosystem level, I primarily focus on how environmental
15 stressors impact fish development. My teaching responsibilities at the University of Texas
16 include Marine Environmental Science, as well as a graduate level course in Aquatic
17 Toxicology and Risk Assessment.

18 I also have prior professional experience in environmental public health and risk
19 assessment from my time in the private and government sectors. I continue my risk
20 assessment work in a research capacity, with a focus on the human health risks of dietary
21 contaminant exposure from subsistence fishing.

22 **Q. PLEASE IDENTIFY WHAT HAS BEEN MARKED AS EXHIBIT PAC-48R KN-1.**

23 **A.** This is a copy of my current Curriculum Vitae, which includes a list of my publications,
24 presentations, and research grants. My CV reflects my expertise for the opinions I am
25 providing here.

1 **Q. IS THIS A TRUE AND CORRECT COPY OF YOUR CV?**

2 **A.** Yes.

3 *PAC offers Exhibit PAC-48R KN-1.*

4 **Q. HAVE YOU EVER TESTIFIED AS AN EXPERT WITNESS IN A TRIAL OR AN**
5 **ADMINISTRATIVE HEARING PRIOR TO THIS MATTER?**

6 **A.** No.

7 **Q. HAVE YOU PREPARED ANY ANALYSIS OF THE PORT OF CORPUS CHRISTI'S**
8 **APPLICATION, OTHER THAN YOUR TESTIMONY HERE?**

9 **A.** Yes. Prior to being asked to serve as an expert for this case, I authored a report titled

10 *Proposed Harbor Island Seawater Reverse Osmosis Desalination Facility: A Prospective*

11 *Evaluation of Toxicological Risk* that prospectively evaluated potential ecotoxicological

12 risks associated with the discharge of effluent from the Port of Corpus Christi's proposed

13 desalination facility to the Corpus Christi Ship Channel and the surrounding habitat. As the

14 report indicates, this study was requested and funded by the UTMSI Marine Science

15 Council and was conducted as part of my responsibilities as a faculty member at the

16 University of Texas. It was published in 2021.

17 **Q. PLEASE IDENTIFY WHAT HAS BEEN MARKED AS EXHIBIT PAC-48R KN-2.**

18 **A.** This is a copy of the report I just mentioned.

19 **Q. IS THIS A TRUE AND CORRECT COPY OF THAT REPORT?**

20 **A.** Yes.

21 *PAC offers Exhibit PAC-48R KN-2.*

22 **Q. DID YOU RELY ON THIS REPORT TO INFORM YOUR OPINIONS ON THIS**
23 **MATTER?**

24 **A.** Yes.

25 **Q. HAVE YOU CONDUCTED ANY OTHER STUDIES THAT ARE RELEVANT TO**
26 **THE ISSUES ADDRESSED IN YOUR TESTIMONY IN THIS MATTER?**

1 A. Yes. I conducted multiple studies investigating the survival and growth implications of
2 anthropogenic-mediated salinity stress to early life stage red drum (*Sciaenops ocellatus*).
3 While these studies are highly relevant to this issue and have informed my opinion
4 regarding the Port of Corpus Christi's application and draft permit, it should be noted that
5 the results have implications for estuarine-dependent fisheries beyond this matter. As with
6 the report I prepared for the UTMSI Marine Science Advisory Council, these studies were
7 funded entirely by UTMSI departmental funds and were conducted as part of my
8 responsibilities as a faculty member at the University of Texas.

9 **Q. PLEASE IDENTIFY THE DOCUMENT MARKED AS EXHIBIT PAC-48R KN-3.**

10 A. PAC-48R KN-3 is the report documenting the methods and results of the rangefinder test
11 for the median lethal concentration (Test 1), the results of the first median lethal time test
12 (Test 2), and all data from follow-up testing (i.e., the final median lethal concentration test
13 and the second median lethal time test).

14 **Q. IS THIS A TRUE AND CORRECT COPY OF THIS REPORT?**

15 A. Yes.

16 *PAC offers Exhibit PAC-48R KN-3.*

17 **Q. DID YOU RELY ON THIS REPORT TO INFORM YOUR OPINION ON THIS**
18 **MATTER?**

19 A. Yes.

20 **Q. HAS THIS STUDY BEEN PEER-REVIEWED?**

21 A. The manuscript is currently undergoing peer review at *Estuaries & Coasts*, the Coastal and
22 Estuarine Research Federation's scientific journal. In addition, I submitted my data to an
23 uninvolved third-party for review as a part of my University-approved Financial Conflict
24 of Interest (FCOI) Management Plan. My data was submitted to Dr. Matthew Alloy, with

1 US EPA, for preservation and review. His evaluation resulted in similar (though slightly
2 more conservative) toxicity values to those reported in the manuscript.

3 **Q. PLEASE IDENTIFY THE DOCUMENT MARKED AS EXHIBIT PAC-48R KN-4.**

4 **A.** Exhibit PAC-48R KN-4 is the manuscript currently undergoing peer review.

5 *PAC offers Exhibit PAC-48R KN-4.*

6 **Q. DID YOU RELY ON THIS MANUSCRIPT TO INFORM YOUR OPINION ON**
7 **THIS MATTER?**

8 **A.** Yes.

9 **Q. WHAT OTHER MATERIALS HAVE YOU REVIEWED, IF ANY, IN**
10 **DEVELOPING YOUR OPINIONS IN THIS MATTER?**

11 **A.** I have reviewed the Port of Corpus Christi's application, draft Permit No. WQ000523000,
12 and supplemental materials. I also have reviewed the pre-filed testimony of the Port's
13 expert witnesses. I have reviewed prior deposition testimony from Dr. Andrew Esbaugh
14 and Dr. Scott Holt. I have also reviewed TCEQ Chapter 307 Texas Surface Water Quality
15 Standards, US EPA Risk Assessment guidance and toxicity testing methods, plus a range
16 of scientific literature, the titles of which are included in Exhibit PAC-48R KN-5.

17 *PAC offers Exhibit PAC-48R KN-5.*

18 **Q. ARE THESE THE TYPES OF RESOURCES GENERALLY RELIED ON BY**
19 **EXPERTS IN YOUR FIELD?**

20 **A.** Yes.

21 III. SUMMARY OF OPINIONS

22 **Q. HAVE YOU DEVELOPED ANY OPINIONS REGARDING THE APPLICATION**
23 **OF THE PORT OF CORPUS CHRISTI OR THE DRAFT PERMIT PREPARED BY**
24 **TCEQ?**

25 **A.** Yes.

26 **Q. PLEASE SUMMARIZE THOSE OPINIONS.**

27 **A.** My overarching opinion is that the proposed activities described in the draft permit pose a
28 high degree of risk to estuarine habitats hydrologically connected to the Corpus Christi

1 Ship Channel. The permit application and the draft permit fail to include and consider the
2 toxicological and environmental data necessary to perform a robust and adequate
3 environmental impact assessment and/or ecological risk assessment. The evaluation that
4 was submitted in support of the Port's application by Dr. Fontenot is wholly inadequate
5 and its conclusions are misleading for several key reasons.

6 I am also of the opinion that discharge of effluent from the Port's proposed facility to the
7 Corpus Christi Ship Channel will result in salinity concentrations that have a high potential
8 to cause adverse effects on aquatic life, particularly on early life stage red drum and other
9 aquatic species that are sensitive to salinity stress during early development.

10 I am also concerned that the proposed location of the outfall in the Corpus Christi Ship
11 Channel is hydrologically connected to an enclosed bay system with high ecological
12 value/high productivity and with low flushing/mixing/water turnover. It is my opinion that
13 the siting of the proposal outfall in this location will lead to degradation of the critical
14 estuarine habitats that rely on the Corpus Christi Ship Channel for water exchange, as well
15 as the organisms that use the channel as a major migratory corridor.

16 Furthermore, the Port failed to consider the impacts of simultaneous exposure to multiple
17 stressors experienced by organisms in the wild, including the important role of latent
18 mortality. These factors are particularly critical to evaluate when predicting ecological risk,
19 as simultaneous exposure to multiple stressors is known to cause lethal impacts to early
20 life stage aquatic organisms at levels that may be orders of magnitude lower than laboratory
21 toxicity tests would suggest. Estuarine-dependent species that spawn in nearshore waters
22 in proximity to the Corpus Christi Ship Channel, especially those with planktonic offspring

1 that must passively drift through channel to complete development in shallow seagrass
2 nurseries in adjacent bays, are of particular concern in this respect.

3 **Q. HAVE YOU COMMUNICATED WITH OTHER TESTIFYING WITNESSES**
4 **RETAINED BY PAC AND, IF SO, WHICH ONES?**

5 **A.** Yes. I have communicated with Andrew Esbaugh, Brad Erisman, Daniel Schlenk, Greg
6 Stunz, Scott Holt, and Scott Socolofsky.

7 **Q. HAVE YOU RELIED ON THE OPINIONS, DATA, OR INFORMATION FROM**
8 **THOSE OTHER TESTIFYING WITNESSES RETAINED BY PAC IN FORMING**
9 **YOUR OPINIONS?**

10 **A.** Yes. I relied on modeling conducted by Dr. Socolofsky. I also relied on Drs. Stunz' and
11 Holt's intimate knowledge of the ecology of the Corpus Christi Ship Channel and
12 hydrologically connected habitats.

13 **IV. OPINIONS REGARDING THE ECOLOGICALLY SENSITIVE NATURE OF**
14 **THE AREA ADJACENT TO THE PROPOSED DISCHARGE**

15 **Q. YOU MENTIONED THAT THE AQUATIC HABITAT ADJACENT TO THE**
16 **PROPOSED DISCHARGE SITE IS HIGHLY PRODUCTIVE AND OF HIGH**
17 **ECOLOGICAL VALUE. PLEASE EXPLAIN WHAT YOU MEAN.**

18 **A.** This area contains some of the most well-preserved and productive estuarine habitats in the
19 nation. By productive, I mean that it generates large amounts of organic biomass to support
20 complex coastal food webs, which maintains high biological diversity and ecosystem
21 function. Seagrass beds serve as essential nursery grounds for early life stage aquatic
22 organisms, including many managed and protected fish species. When the ecosystem is
23 healthy and functions properly, humans reap the benefits through a wide range of
24 ecosystem services, such as water filtration and detoxification, enhanced coastal resiliency,
25 erosion prevention, and maintenance of biogeochemical cycles. The economic benefits of
26 tourism, recreation, and fisheries to the area also cannot be overstated here. Texas has a
27 \$3.2 billion recreational fishing industry, two thirds of which comes from red drum
28 (*Sciaenops ocellatus*) and speckled seatrout (*Cynoscion nebulosus*) fisheries, both of which

1 are estuarine-dependent species that thrive here. Though red drum can be found elsewhere
2 along the Texas Coast, our immediate area supports some of the most robust populations
3 in the nation.

4 **Q. PLEASE EXPLAIN WHY THIS IS IMPORTANT.**

5 **A.** I mentioned above that this is an area of high ecological value that (when functioning
6 correctly) provides critical nursery and feeding grounds for early life stage fish, like red
7 drum. This is important to consider because the sudden reduction in number of red drum
8 larvae that successfully reach the seagrass beds in any given year would mean lower adult
9 fish numbers are available to fisherman 3 to 5 years later. Fewer adults mean fewer
10 successful spawns the next year, further depressing fish populations through a series of
11 feedbacks. Because red drum are so long lived, adverse impacts to seasonal recruitment of
12 young can impact the health of fisheries for years. In fact, red drum populations are not
13 expected to recover from the low productivity of the 2010, 2011, 2012, and 2013 year
14 classes (from the *Deepwater Horizon* oil spill) until the year 2053.

15 There is an abundance of peer-reviewed scientific literature to show that the collapse of
16 important marine fisheries eliminates sustainable livelihoods for nearby communities.

17 **V. OPINIONS REGARDING SALINITY CHANGES UPON DISCHARGE**

18 **Q. YOU MENTIONED SALINITY OF THE DISCHARGE. HAVE YOU DEVELOPED**
19 **OPINIONS REGARDING THE POTENTIAL IMPACT OF SALINITY OF THE**
20 **PROPOSED DISCHARGE ON AQUATIC LIFE?**

21 **A.** Yes.

22 **Q. WHAT ARE YOUR OPINIONS REGARDING SALINITY IN THE PROPOSED**
23 **DISCHARGE?**

24 **A.** Global experts explicitly recommend against discharging brine into habitats with (1) high
25 ecological value, (2) high productivity, (3) high concentrations of sub-adult organisms, and
26 (4) poor flushing and long water residence times. All of these characteristics describe

1 estuaries hydrologically connected to the Gulf of Mexico by the Corpus Christi Ship
2 Channel. Neither the application nor the draft permit have demonstrated that the salinity or
3 any other chemical constituents present in the discharge will not adversely impact the
4 growth or survival of aquatic and aquatic-dependent life (particularly organisms in early
5 life stages) Modeling has shown that the outfall may easily result in salinities that are ≥ 2
6 ppt above ambient, which my research and that of others have found to be the threshold
7 above which adverse ecological effects are possible in a system of this kind. Collectively,
8 the biological and modeling data confirm that the upper range of naturally occurring
9 salinities within the Corpus Christi Ship Channel are already near the tolerance threshold
10 of native red drum larvae and other estuarine dependent species. Therefore, under frequent
11 summer drought conditions, even minimal elevations in the salinity of the channel may
12 lead to significant adverse ecological impacts. Further, the modeling done by the Port of
13 Corpus Christi does not demonstrate that this hypersaline effluent will not adversely impact
14 aquatic life beyond the mixing zone. In summary, the effluent discharged from the Port's
15 proposed facility to the Corpus Christi Ship Channel and surrounding habitat will result in
16 salinity concentrations with a high potential to cause adverse effects on aquatic life,
17 particularly on sensitive estuarine-dependent organisms, like early life stage red drum.

18 **Q. WHAT IS THE BASIS FOR YOUR OPINION?**

19 **A.** My opinion is based on my experience as an ecotoxicologist, in both a research and risk
20 assessment context. I have performed a large number of studies with early life stage
21 fisheries species in the Gulf of Mexico, including multiple stressor studies and salinity
22 testing with red drum and other native fish and invertebrate species. I also research coupled
23 human and natural systems in subsistence fishing communities, and I have professional

1 experience in environmental public health and risk assessment that also contributed to my
2 opinions. Further, I relied on the opinions of other relevant experts (previously identified).

3 **Q. YOU PREVIOUSLY MENTIONED TOXICITY TESTS ON EARLY LIFE STAGE**
4 **RED DRUM. PLEASE EXPLAIN HOW YOU CONDUCTED THESE TOXICITY**
5 **TESTS.**

6 **A.** I performed two types of single stressor toxicity tests with embryonic and larval red drum.

7 The first type was designed to find the median lethal concentration (LC50), which is the
8 concentration that will cause 50% of larvae to die at pre-set timepoints. That test also allows
9 us to predict two other values, called a NOEC (no observable effect concentration) and
10 LOEC (lowest observable effect concentrations). The LOEC is the lowest treatment
11 concentration that had a statistically significant effect, and the NOEC is the concentration
12 just below the LOEC (in which any effects were not yet statistically significant). The
13 second type of test was designed to find the median lethal time (LT50), which is the length
14 of time required to kill 50% of embryo-larval red drum that drift into full strength brine
15 (i.e., an LT50 test). Both tests counted only mortality that occurred while the test was
16 running, so latent mortality is not accounted for (i.e., the values likely underestimate risk
17 to a high degree).

18 I performed both types of tests in duplicate. The first round of testing consisted of
19 rangefinder tests, which is standard practice in my field. To perform a rangefinder for an
20 LC50 test, you start with a very broad range of concentrations and then you use the results
21 of that test to refine your final study design to focus on a narrow range of concentrations
22 at which you see effects start to occur. This second step is key to generating toxicity values
23 with a high degree of certainty, because you can avoid extrapolating across large gaps in
24 concentrations (e.g., 5 ppt or 10 ppt increments). The second round of testing I conducted

1 consisted of this type of refined test. We also doubled the number of replicates we tested
2 in each concentration to increase our statistical power even further.

3 For both LC50 tests, survival was evaluated at nominal test hours 24, 48, and 72. We did
4 not run the tests out for the standard 96 hours used by EPA, because embryo-larval red
5 drum have used up their yolk sac by 96 hours, so mortality counts would be confounded
6 by starvation at the last time point. This is standard practice for embryo-larval red drum
7 tests, including those conducted for National Resource Damage Assessments. The rationale
8 is that at test hour 72, a rapidly developing red drum will be developmentally similar to an
9 EPA estuarine model fish at test hour 96.

10 **Q. WHAT WERE THE RESULTS OF THE TOXICITY TESTS?**

11 **A.** The results of the LC50 rangefinder studies showed that 50% of red drum would not hatch
12 at test hour 24 as expected if the salinity is around 49 ppt. At test hour 48, a salinity of
13 around 43.8 ppt would be lethal to 50% of larvae. At test hour 72, half of the larvae will
14 die in water with a salinity of about 40.4 ppt.

15 Using this information, we repeated the test to focus on a narrower range of salinities that
16 were close to the LC50s from the first test, with treatments that increased by only 2 ppt
17 (instead of the 5 ppt increments used in Test 1). Results of this second round of tests agreed
18 excellently with our first round of results, despite the fact that we used eggs from Texas
19 Parks and Wildlife (spawned in 25 ppt tanks) for the first test, and eggs from UTMSI
20 (spawned at 35 ppt) for the second test.

21 Follow-up testing showed that 50% of red drum embryos will fail to hatch in water with a
22 salinity of 50.8 ppt. Twenty-four hours later (test hour 48), 50% of larvae are expected to
23 die in water with a salinity of 44.8 ppt. At 48 hours post hatch (test hour 72), a salinity of
24 37.7 ppt is expected to kill 50% of larvae. The LOEC for all timepoints (i.e., the

1 concentration at which adverse effects on hatch and survival first starts to occur) was 37
2 ppt. This means that the only treatment that did not cause significant lethality to larvae was
3 the 35 ppt control treatment (i.e., the NOEC for salinity is essentially the salinity they were
4 spawned in).

5 During both rounds of LT50 testing, we observed significant lethality at every timepoint
6 evaluated, including the first timepoint (4 minutes for round 1 and 10 min for round 2).
7 This means that significant effects on the survival of larval red drum drifting through the
8 ZID will begin sometime between 0 and 4 minutes, with 50% mortality between 47.7 and
9 55.4 minutes.

10 **Q. DID YOU CONDUCT ANY MORE FOLLOW-UP TESTS THAT YOU HAVE NOT**
11 **ALREADY MENTIONED?**

12 **A.** I have not yet done any follow-up tests, but I did perform confirmatory statistical analyses
13 in R using probit models and (where appropriate) post hoc comparisons via estimated
14 marginal means. I also used a Tukey adjustment to control for the increased rate of error
15 inherent in multiple comparisons. Using this approach produced toxicity values that were
16 either the same, or lower than my initial values.

17 **Q. WHAT WERE THE RESULTS OF THESE CONFIRMATORY STATISTICAL**
18 **ANALYSES?**

19 **A.** The new run showed that 50% of red drum embryos will fail to hatch around 46.6 ppt, as
20 compared to my original run at 50.4 ppt. At test hour 48, the probit model predicts that 50%
21 of larvae will be dead at a salinity of 43.3 ppt, as compared with my original value of 44.8
22 ppt. Finally, at test hour 72, 50% of the larvae will be dead at a salinity around 37.9 ppt,
23 which is almost identical to my previous value of 37.7 ppt. The LOEC for all timepoints
24 remained at 37 ppt.

25 **Q. DO THE RESULTS FROM THESE NEW ANALYSES IMPACT YOUR OPINION**
26 **IN ANY WAY?**

1 A. They do not change my opinion. For the initial analyses, all p values were < 0.001 (anything
2 less than 0.05 is significant, but the lower the number, the more undeniable the
3 significance). We also performed two rounds of testing, with eggs from two different
4 broodstock, and got nearly identical results. The second time, we used twice the replication
5 that Parson's sub-contractor used for their testing (they used the minimum required by the
6 EPA) and salinity increments between treatments of only 2 ppt, to avoid having to
7 extrapolate to any meaningful extent. The one added advantage of this second approach is
8 that it controls for the random error that is associated with multiplicity, so these results
9 essentially eliminate any uncertainty about the values.

10 **Q. DO LC50 AND LT50 VALUES HAVE ANY LIMITATIONS THAT YOU BELIEVE**
11 **ARE IMPORTANT TO YOUR OPINION?**

12 A. Yes.

13 **Q. WHAT ARE THOSE LIMITATIONS?**

14 A. It is important to note that the toxicity values from the LC50 and LT50 test do not account
15 for latent mortality (i.e., death that occurs in the hours or days following exposure). Based
16 on low salinity tolerance of this species during early development (as demonstrated by our
17 tests and by others), it is likely that latent mortality will be a significant factor affecting the
18 survival of larval red drum exposed to hypersaline conditions in the CCSC. Moreover,
19 these tests considered only salinity stress, and do not account for additional stress from
20 contamination, mechanical forces, intense solar radiation, hypoxia, or other co-stressors
21 that can interact to cause lethality at much lower exposure concentrations than lab studies
22 can predict.

23 **Q. WHY DID YOU USE EARLY LIFE STAGE RED DRUM?**

1 A. In the context of predicting and evaluating potential damage to local estuarine ecosystems
2 from human-caused environmental change in the Gulf of Mexico, early life stage red drum
3 are essentially the gold standard of study organisms.

4 The reason I specifically focus on early development is because organisms are known to
5 be more sensitive to environmental stress during early development than juveniles or adults
6 (and thus, are more important drivers of ecological risk). It is also a widely accepted
7 principle that physical and chemical stressors that adversely impact the reproductive
8 success and/or survival of early life stage fish have the most severe effects on populations
9 as a whole, so evaluating risks on developing fish is key to understanding what will happen
10 to total fish numbers in the long term.

11 While juvenile and adult red drum are known to be fairly tolerant of high salinity, early
12 developmental stages are very sensitive to changes in salinity outside of their normal range.
13 Since red drum drift through the Ship Channel during this incredibly sensitive stage, they
14 are expected to be an important driver of risk, unlike the inland silverside (the fish species
15 that the Port used for testing that is spawned in shallow bays and is widely known to be
16 euryhaline at hatch).

17 Finally, red drum are known to be especially sensitive to multiple stressor effects, including
18 both chemical and physical types of stress. These are also the reasons that NOAA chose to
19 use early life stage red drum for toxicity testing (including results of multi-stressor toxicity
20 studies) that was used to develop broader conclusions about the extent of damage caused
21 to nearshore marine ecosystems in the Gulf of Mexico by the *Deepwater Horizon* oil spill.

22 The reason people don't use them more commonly is merely a product of availability (very
23 few places can successfully spawn red drum in captivity) and the sensitivity of early life

1 stages (which makes it hard to successfully transport them long distances). In the
2 immediate area, we are exceptionally fortunate to have two local facilities with long and
3 successful histories of spawning wild caught red drum in captivity (UTMSI and Texas
4 Parks & Wildlife). Thus, my lab is able to use these organisms for regionally relevant
5 toxicity testing while maintaining high baseline survival. This species is regionally
6 important in many respects, including the following key points:

7 (1) They are critical predators that play an important role in maintaining the structure and
8 function of our local estuarine ecosystems

9 (2) They are highly relevant to all of the aquatic habitats that are hydrologically connected
10 to the Corpus Christi Ship Channel (i.e., nearshore waters, the channel itself, connected
11 bay systems), and thus, the proposed facility.

12 (3) Their economic value to Port Aransas and Corpus Christi (and coastal Texas as a whole)
13 cannot be overstated.

14 (4) Due to their long lifespan, the impacts of below average survival and recruitment can
15 persist for multiple decades.

16 **Q. DOES YOUR LAB REGULARLY USE EARLY LIFE STAGE RED DRUM FOR**
17 **TOXICITY TESTS?**

18 **A.** Yes. My lab uses early life stage red drum for a wide variety of toxicity testing, including
19 studies examining the toxicity of oil, perfluorinated compounds, and physical stressors
20 (e.g., intense ultraviolet radiation and salinity), both separately and in combination with
21 one another.

22 **Q. YOU MENTIONED THAT YOU USED RED DRUM BROODSTOCK FROM TWO**
23 **DIFFERENT SOURCES. DOES THAT INFLUENCE YOUR OPINION?**

24 **A.** The red drum broodstock at UTMSI were not being spawned the week my lab planned to
25 perform the rangefinder tests, as the tanks were not yet up to the temperature necessary to

1 induce spawning (our aquaculture experts manipulate tank temperatures very gradually to
2 avoid stressing the fish). Texas Parks & Wildlife (Flour Bluff location) spawning tanks
3 were already up to temperature, so we used embryos from their facility to perform
4 preliminary testing. This is a very standard practice for those of us working with red drum
5 at UTMSI. In fact, many of the red drum eggs used for *Deepwater Horizon* toxicity testing
6 were provided by the same Texas Parks & Wildlife facility.

7 By the time we were ready to perform our refined (final) tests, the spawning tanks at
8 UTMSI were up to temperature and producing high quality spawns. To avoid the potential
9 for transport stress to affect results, we used embryos produced at our facility for final
10 testing. It is relevant how similar the test findings were, given that eggs from two different
11 broodstock populations were used, especially since the Texas Parks & Wildlife tanks were
12 at 25 ppt (Test 1) and 28 ppt (Test 2), but the UTMSI embryos were spawned at 35 ppt.
13 Thus, the degree of alignment between the preliminary and final testing further confirms
14 the broad applicability of the toxicity values we generated for early life stage red drum
15 exposed to hypersaline conditions.

16 **Q. ARE THE METHODS YOU FOLLOWED THE TYPES OF METHODS**
17 **ACCEPTED BY PROFESSIONALS IN YOUR FIELD?**

18 **A.** Yes.

19 **Q. HAVE THESE SAME METHODOLOGIES BEEN THE BASIS OF PUBLISHED**
20 **PEER-REVIEWED STUDIES BEFORE?**

21 **A.** Yes. The underlying study design is the same or similar to most other early life stage red
22 drum studies conducted by my lab and others, as well as many other peer-reviewed studies
23 involving estuarine/marine species that are not typical EPA testing models but have been
24 found to be incredibly important for evaluating risk to valuable and unique ecosystems in

1 the Gulf of Mexico for National Resource Damage Assessments. These include fiddler crab
2 zoea, blue crab zoea, mahi mahi, speckled seatrout, anchovy, red snapper, cobia, and others.

3 **Q. DO YOU KNOW WHAT TYPE OF SPECIES THE PORT RELIED ON FOR THEIR**
4 **TESTING?**

5 Yes. The Port chose two species – mysid shrimp and inland silverside.

6 **Q. DO YOU HAVE ANY OPINIONS ABOUT THE SPECIES THE PORT RELIED ON?**

7 **A.** It is a widely accepted principal in toxicology that early life stage organisms are much more
8 sensitive to the adverse effects of environmental stress than juvenile or adult stages. The
9 salinity tolerance demonstrated by early life stage organisms in estuaries is extremely
10 variable. Some species have a remarkably high ability to cope with extreme salinity
11 fluctuations from day one, like sheepshead minnow (*Cyprinodon variegatus*), mysid
12 shrimp (*Mysidopsis bahia*), and inland silverside (*Menidia beryllina*) while others require
13 very specific salinities during certain windows to complete development.

14 Still, the Port selected mysid shrimp (*Mysidopsis bahia*) and inland silverside (*Menidia*
15 *beryllina*) as organisms for salinity testing. To start, mysid shrimp are known to be the most
16 dominant Mysidacea in the Laguna Madre, which regularly reaches 70 ppt during the arid
17 summer months. Their occurrence and survival under these conditions in the Corpus Christi
18 area has been documented in the peer reviewed literature for at least four decades, making
19 them an inappropriate toxicity model for this particular project.

20 The inland silverside is an equally poor choice for this project because of life history
21 differences that lead to high salinity stress tolerance during the earliest stages of
22 development. These organisms spawn in shallow estuarine waters that naturally experience
23 a wide range of salinities, so even newly hatched animals must be highly tolerant to salinity
24 stress in order to survive. Many organisms that spawn in tidal rivers, brackish water, or
25 other shallow estuarine habitats demonstrate the same tolerance immediately upon hatch.

1 It is worth noting that the US EPA standard operating procedures for the inland silverside
2 test contained in publication *Short-term Methods for Estimating the Chronic Toxicity of*
3 *Effluents and Receiving Waters to Marine and Estuarine Organisms* (EPA-821-R-02-014)
4 explicitly states that the inland silverside can tolerate a wide range of salinities, *from 0 ppt*
5 *to 58 ppt*.

6 In contrast to the above-mentioned and highly tolerant organisms, estuarine-dependent
7 species that spawn in nearshore waters (e.g., red drum and speckled seatrout) experience
8 gradual changes in salinity across a relatively narrow range during their earliest
9 developmental stages, so their salinity tolerance in the days following hatch is very low.
10 Along with the findings of others, testing performed in my lab shows that newly hatched
11 red drum are very intolerant of salinity fluctuations in early development (as are speckled
12 seatrout), regardless of spawning salinity. Additionally, larvae becoming increasingly
13 tolerant as they proceed through subsequent developmental stages and settle in seagrass
14 beds. In summary, the Port's species are an inappropriate and suspect choice for evaluating
15 ecological risk from hypersaline discharge.

16 **Q. ARE FISH THE ONLY CLASS OF ORGANISM THAT YOU ARE CONCERNED**
17 **ABOUT?**

18 **A.** No. Every class of organisms has members that are much less tolerant of salinity changes
19 than others. This even includes phytoplankton, which form the base of the estuarine food
20 web in this bay complex. Research has shown that hypersaline brine from desalination
21 plants kills more sensitive phytoplankton species, leaving fewer species of highly tolerant
22 phytoplankton. The decrease in phytoplankton diversity near multiple desalination
23 facilities has been shown to trigger harmful algal blooms.

1 VI. OPINIONS REGARDING CONTAMINANTS OTHER THAN SALINITY UPON
2 DISCHARGE

3 Q. YOU MENTIONED CONTAMINANTS OTHER THAN SALINITY. HAVE YOU
4 DEVELOPED OPINIONS REGARDING THE POTENTIAL IMPACT OF
5 CONTAMINANTS OTHER THAN SALINITY ON AQUATIC LIFE AS A RESULT
6 OF THE DISCHARGE?

7 A. Yes.

8 Q. WHAT ARE YOUR OPINIONS REGARDING CONTAMINANTS OTHER THAN
9 SALINITY?

10 A. My opinion is that neither the application nor the draft permit have demonstrated that there
11 will be no contaminants in the discharge, nor has it been demonstrated that those
12 contaminants in the discharge will not adversely impact aquatic and aquatic-dependent life.

13 Q. PLEASE EXPLAIN.

14 The Port claims that the intake water is not located near chemical source areas, yet the
15 Corpus Christi area is a major source of both municipal and industrial contamination,
16 including a variety of constituents from contaminant classes, such as metals,
17 polychlorinated biphenyls (PCBs), per- and polyfluoroalkyl substances (PFAS),
18 organochlorine pesticides (OC pesticides), and polycyclic aromatic hydrocarbons (PAHs).
19 Many of these contaminants can be transported long distances through normal
20 environmental processes, so they are certainly of potential concern in nearby coastal
21 systems, especially in sediments.

22 Harbor Island has been regularly exposed to point source industrial pollution. It was once
23 the largest import port for crude oil for decades. Exxon used the port to import foreign
24 sources of crude for its Humble refinery that once operated in Ingleside. The *Deepwater*
25 *Horizon* oil spill, which released millions of barrels of oil into the Gulf of Mexico and oiled
26 thousands of miles of shoreline, is another major source of potential contamination that is
27 highly relevant to this area.

1 The pollutants commonly released by industrial facilities and oil spills will preferentially
2 partition to sediments (which have not been evaluated by the Port) due to their physico-
3 chemical properties. Because oil contamination is widely known to remain in marine
4 sediments for many decades, it is not reasonable to suggest that sediment pollution is not a
5 potential concern at the intake site. In fact, the World Health Organization (WHO)
6 explicitly states that there is “a significant potential for anthropogenic contamination of
7 source waters, particularly seawater and estuarine waters” to be present in effluent from
8 Reverse Osmosis facilities, resulting in “a significant potential for contamination by
9 petroleum hydrocarbons, particularly in regions where there is substantial oil extraction
10 activity” (e.g., the Corpus Christi area).

11 Data from NOAA’s National Status and Trends Program’s ecotoxicological database
12 support these conclusions. The database provides sediment and tissue monitoring data for
13 a subset of contaminants in Corpus Christi, Aransas, and Copano Bays through 2006
14 (before the *Deepwater Horizon* spill). I evaluated these data, which indicate that a number
15 of contaminants are already at or near levels of potential concern in these bays, particularly
16 those associated with crude oil. Furthermore, tissue contamination was typically higher
17 than contamination in nearby sediments, indicating that the pollution is bioavailable and is
18 entering the food web. The Port has made no effort to show that the proposed facility will
19 not add to the existing level of contamination via redistribution of contamination from the
20 intake site to the outfall (which will occur continuously).

21 Chemicals added during the desalination process may also react with constituents present
22 in the aquatic environment to yield highly toxic halogenated by-products. Chronic
23 exposure to mixtures of contaminants of concern that exert sublethal affects (e.g., reduce

1 reproductive capacity/biomass, developmental abnormalities) are typically expected to
2 drive ecotoxicological risk for sensitive receptors at sites of this nature, rather than acute
3 mortality from single contaminant exposure. Neither the application nor the draft permit
4 provide enough information to support the Port's position that the facility will not
5 remobilize contamination from the intake site into habitats that are hydrologically
6 connected to the Corpus Christi Ship Channel.

7 **Q. WHAT IS THE BASIS FOR THIS OPINION?**

8 **A.** My opinion is based on my experience as an ecotoxicologist, in both a research and risk
9 assessment context, particularly the study I conducted entitled, *Proposed Harbor Island*
10 *Seawater Reserve Osmosis Desalination Facility: A Prospective Evaluation of the*
11 *Ecotoxicological Risk.*

12 **Q. WHAT ASSUMPTIONS DID YOU RELY ON IN THIS STUDY?**

13 **A.** Since this proposed facility is not already permitted and operating, there are no site-specific
14 quantitative data from any releases that have already occurred. Therefore, my evaluation
15 of the risks posed by contaminants relied on surrogate data. For the exposure assessment,
16 I reviewed a combination of peer-reviewed scientific journal articles, reports from various
17 government agencies, technical reports, permitting documents, and NOAA's National
18 Status and Trends Program's ecotoxicological database (which contains sediment and
19 tissue monitoring data for Corpus Christi, Aransas, and Copano Bays).

20 **Q. ARE YOU AWARE OF ANY ENVIRONMENTAL SAMPLING CONDUCTED BY**
21 **THE PORT OF CORPUS OF CHRISTI TO DETERMINE CURRENT LEVELS OF**
22 **CONTAMINATION AT THE INTAKE SITE?**

23 **A.** Yes. In supplemental materials dated June 25, 2021, the Port of Corpus Christi released the
24 results of two water quality samples it says Parsons collected near the location of the
25 proposed intake. No sediment sampling was conducted.

1 **Q. DO YOU HAVE ANY OPINIONS REGARDING THE PORT’S ENVIRONMENTAL**
2 **SAMPLING?**

3 **A.** Yes. The Tischler - Garza memo included in the supplemental materials references the two
4 water samples, saying they were to be “collected over a relatively short time period
5 (approximately one week). In fact, the samples were collected over the course of two
6 consecutive days in early June 2021. Texas Administrative Code Rule 307.9 requires that
7 samples used “to determine standards attainment in ambient water must be representative
8 in terms of location, seasonal variations and hydrological conditions.” Based on the sample
9 size and collection window alone, the Port cannot meet these requirements. The code also
10 goes on to state that “sample results that are used to assess standards attainment must not
11 include samples that are collected during extreme hydrological conditions such as high-
12 flows and flooding immediately after heavy rains.” This is critical to the interpretation of
13 the Port’s sampling results, as the historical record shows that these samples were collected
14 during a period of abnormally high rainfall in Port Aransas. See Exhibit PAC-48R KN-6.

15 *PAC offers Exhibit PAC-48R KN-6.*

16 Thus, my opinion is that the two water samples that the Port collected are in no way
17 representative of the typical water quality or contaminant concentrations present at the
18 intake site, so they cannot be used to evaluate standards attainment or to predict the
19 concentration of contaminants that will be present in effluent. Nonetheless, the Port
20 proceeded to calculate average predicted contaminant concentrations in effluent under a
21 40% recovery scenario using these values. It is noteworthy that in spite of their insufficient
22 data collection, the Port still shows that concentrations of contaminants will be higher in
23 effluent than in intake water (Tables reporting sampling results from intake location in
24 Industrial Wastewater Application Technical Report).

1 You cannot confidently evaluate risk using two biased-low water sampling values collected
2 during abnormal hydrological conditions. Nonetheless, the Port chose to perform all of its
3 risk calculations using these values. Other standard risk assessment practices that Dr.
4 Fontenot chose to forego in his so-called ecological risk assessment are evaluations of the
5 role of sediment contamination in exposure and risk, and a discussion of uncertainty
6 surrounding his conclusions.

7 **Q. YOU SAID THE PORT DID NOT CONDUCT ANY ASSESSMENT ON SEDIMENT.
8 IS THIS IMPORTANT TO YOUR OPINIONS?**

9 **A.** Yes.

10 **Q. IN WHAT WAY?**

11 **A.** Despite the fact that the Port's Process Design Basis and Narrative states that the intake
12 will pull in millions of gallons of sediment and other suspended matter (e.g., phytoplankton
13 and other small organisms, organic carbon) every day, along with intake water, no sediment
14 sampling was conducted. The Port's choice to omit this information is particularly
15 troubling for many reasons, the first being that the vast majority of contaminants in the
16 marine environment are typically found in the sediment. The Port attempts to justify this
17 omission by saying that the "intake surface water from the Gulf of Mexico is not located
18 near chemical source areas and will not contain appreciable suspended solids/sediments.

19 This statement is patently untrue, as the Port's Process Design Basis and Narrative states
20 that along with intake water, millions of gallons of sediment and other suspended matter
21 (e.g., phytoplankton and other small organisms, organic carbon) will be taken up every day.

22 In fact, in Table 2: *50 MGD Desalination Facility Design Basis Source Water and Effluent*
23 *Constituent Concentrations*, the Port shows that the facility will take in such large
24 quantities of suspended solids (including associated contaminants) that even after
25 removing and disposing of 5.1 million gallons of solids, the effluent water will still have

1 twice as many suspended solids as when the process started (The Port's supplement
2 material states that total suspended solids in source water will be 7.4 mg/L, while effluent
3 will contain 15 mg/L of suspended solids at the outfall under a 50% recovery scenario).
4 Consequently, the Port should understand the need to evaluate the potential risk associated
5 with remobilization of sediment contamination from the intake to the outfall, rather than
6 dismissing sediment as a source contribution. This is especially true given that the Port
7 states in its own materials that suspended solids will be concentrated and discharged at the
8 outfall, thereby continuously adding any associated pollutants to the baseline level of
9 contamination present near the outfall.

10 **Q. PLEASE SUMMARIZE YOUR OPINIONS REGARDING CONTAMINANTS**
11 **OTHER THAN SALINITY.**

12 **A.** In summary, desalination will take up and concentrate contamination from both water and
13 sediments at the intake site, which will be continuously released at the outfall day after day,
14 resulting in accumulation near the outfall (above and beyond any existing pollution).
15 Therefore, organisms getting hit with hypersaline water will simultaneously have to deal
16 with a plume of concentrated contamination, which will result in multi-stressor effects
17 (which causes organisms to die at much lower salinity/contaminant concentrations than
18 those predicted by laboratory toxicity studies). The Port has not provided representative or
19 reliable samples that would show the intake water will not harm aquatic life in the area
20 surrounding the proposed outfall, and the Port has not conducted any sediment sampling
21 near the intake site. The Port's evaluation misrepresents and underestimates actual risk to
22 receptors in the area.

23 VII. OPINIONS REGARDING THE LOCATION OF THE OUTFALL

24 **Q. DO YOU HAVE ANY OPINION REGARDING THE LOCATION OF THE**
25 **PROPOSED OUTFALL?**

1 A. Yes.

2 **Q. WHAT IS YOUR OPINION?**

3 A. The shallow protected bays that are hydrologically connected to the Gulf of Mexico via the
4 Corpus Christi Ship Channel are surrounded by a nearly continuous band of barrier islands
5 that prevent flushing and mixing with oceanic waters. While this minimizes wave action
6 and allows for the formation of wetlands and seagrass beds, it also results in remarkably
7 long water turnover times (up to a year) in these bays, and predisposes them to contaminant
8 accumulation, eutrophication, marine heat waves, hypersalinity and hypoxia. For these
9 reasons, experts from around the globe consistently warn against discharging desalination
10 effluents into enclosed bay systems and coastal lagoons, which typically have (1) high
11 ecological value, (2) high productivity, (3) high concentrations of sub-adult organisms, and
12 (4) poor flushing and long water residence times.

13 **Q. YOU MENTIONED HYPOXIA. PLEASE EXPLAIN.**

14 A. One of the Port's experts, Dr. Furnans, co-authored a paper that evaluated "whether
15 additional salt loads from desalination would lead to enhanced hypoxia in Corpus Christi
16 Bay (the result was affirmative)." In his publication, the authors warn engineers against
17 modeling "using depth-averaged methods" for shallow bays (like those that are
18 hydrologically connected to the Gulf of Mexico via the Ship Channel), as the "additional
19 salt load of a desalination plant" may not vertically mix as expected. Although the Port's
20 overly simplistic CORMIX models do not show the effect predicted in Dr. Furnans' 2011
21 publication, the improved modeling performed by Dr. Socolofsky confirms the likelihood
22 of a persistent density current (a cause of hypoxia) at the outfall. From an ecological risk
23 perspective, Dr. Furnans and his co-authors raise another important point, emphasizing that
24 "understanding the possible development, transport and fate of thin hypersaline layers is

1 critical to predicting the development of hypoxia and associated ecological impacts” from
2 desalination brine.

3 Simultaneous exposure to multiple stressors (such as hypoxia and hypersalinity) causes
4 lethal impacts to early life stage aquatic organisms at exposure levels orders of magnitude
5 below those that would normally be required to cause lethality in laboratory studies.

6 **VIII. OPINIONS REGARDING MULTI-STRESSOR EFFECTS AND LATENT**
7 **MORTALITY**

8 **Q. YOU PREVIOUSLY MENTIONED MULTI-STRESSOR EFFECTS AND THE**
9 **POTENTIAL FOR LATENT MORTALITY. DO YOU HAVE OPINIONS**
10 **REGARDING MULTI-STRESSOR EFFECTS RELATED TO THE PORT’S**
11 **PROPOSED FACILITY?**

12 **A.** Yes.

13 **Q. DO YOU HAVE ANY OPINIONS REGARDING THE POTENTIAL FOR LATENT**
14 **MORTALITY RELATED TO THE PORT’S PROPOSED FACILITY?**

15 **A.** Yes.

16 **Q. WHAT ARE YOUR OPINIONS REGARDING MULTI-STRESSOR EFFECTS**
17 **AND LATENT MORTALITY?**

18 As I previously mentioned, multi-stressor effects and the potential for latent mortality are
19 critical to evaluate the effects that the proposed facility will have on estuarine-dependent
20 species that spawn in nearshore waters proximal to the Corpus Christi Ship Channel,
21 especially those with planktonic offspring that must passively drift through the channel to
22 complete development in shallow seagrass nurseries in adjacent bays. Unfortunately,
23 laboratory testing is not capable of completely replicating the multitude of simultaneous
24 stressors that organisms face under “real-world” conditions, because they are focused on
25 controlling for all but only one (or some limited number) of variables. This does not mean
26 that laboratory tests are unreliable, obviously, it only means that those results must be
27 considered in context of other stressors.

1 the proposed discharge presents a likelihood of significant adverse impacts to aquatic life
2 and to aquatic ecosystems on which coastal communities rely.

3 **Q. DOES THIS CONCLUDE YOUR TESTIMONY?**

4 **A.** Yes, though I reserve my right to supplement this testimony if I learn of information that
5 causes me to change any of my opinions stated here.