## White Lake Algae: Just Give Us A Little Respect

#### Summary

Different types of algae can respond quickly to changing conditions. Cyanobacteria (=bluegreen algae) prefer the conditions that are found in lakes in the summer and fall. In White Lake, a filamentous cyanobacterial bloom started in the fall of 2017 and persisted into 2018; the bloom created conditions—particularly high pH levels--that are well-known for maintaining cyanobacterial dominance. Such blooms end only when resources (light, nutrients) run out, and then there is an abrupt (and decidedly unpleasant) end to it.

It is important to recognize that White Lake is not going off the rails every summer. The same filamentous cyanobacteria species that dominated the bloom before the May 2018 alum treatment have been present since then, but their natural inclination to dominate has been subdued.

The variety of other phytoplankton present in the lake are utilizing the same resources sunlight and nutrients—which are abundant in this shallow lake. This diversity—which has been increasing-means that they all tend to be better-behaved. And because many of the algae are very small, their growth does not result in elevated pH levels.

Sampling of rainfall indicates that it is a diffuse source of readily available nitrogen as well as phosphorus. Algae are responding to these high rainfall/nutrient events, particularly tiny, single-celled forms that are good competitors for nutrients.

The relatively rapid change in rainfall pH (2003-2013) resulted in a relatively rapid change in lake pH, as rainfall is the primary source water for the lake. There has been a substantial increase in nitrogen levels in the lake over the same period, and most of this nitrogen is coming from the rainfall. The lake is establishing a new equilibrium under these changed conditions, which indicates that there is an inherent resiliency to the system.

White Lake, and the life in it deserves our collective understanding and respect, as this is a system that is increasingly impacted by human actions. Continuing monitoring of the lake will give us insight as to how stable the system is (healthy algae vs. harmful cyanobacteria) and how it is responding to weather variability and the effects of climate change (big storms, droughts).

#### **Background Information**

Like most lakes, White Lake teems with life—and always has, even during periods when the water has been crystal clear. What varies is how much: how much phytoplankton (the algae that are suspended in the water column) and how much filamentous benthic algae (the threadlike filaments form mats which are visible in shallow areas).

Algae and cyanobacteria (blue-green algae) are referred to as primary producers, since they use the sun's energy and pigments inside their cells (chlorophylls) to produce their own energy (the process is called photosynthesis). Measuring the amount of the pigment chlorophyll <u>a</u> in a water sample is a relatively easy way to assess the amount of phytoplankton present. A more time-consuming method is to identify, count and measure the size of all of the different phytoplankton found in a sample; the results are expressed as the number of cells per liter, and as biovolume (chlorophyll <u>a</u> concentration is a proxy for biovolume). White Lake monitoring includes monthly sampling for chlorophyll <u>a</u> and monthly or bimonthly sampling for algal identifications, with samples taken at three long-term monitoring stations located along the midline of White Lake.

A simple device called a Secchi disk is used to assess water clarity; it is a relative measure of the concentration of suspended particles in the water column, including phytoplankton, sediments that get stirred up, and materials that originate in the landscape and wash into the lake in stormwater. In shallow lakes such as Lake Mattamuskeet, reduced clarity can be a result primarily of sediment resuspension; this can be the case at White Lake at times, particularly after storms, but generally turbid conditions do not last long, so that Secchi readings often correlate fairly well with chlorophyll concentrations.



Figure 1. Windy conditions the day after a large rainfall, which introduced highly-stained stormwater into the lake. The wind also churned up the bottom, resulting in the suspension of organic sediments. Photo taken February 7, 2020.



Figure 2. White Lake chlorophyll <u>a</u> means (µg/L), Secchi depths (ft), and median pH levels for the period May 2, 2018 (prior to alum treatment) through April 2020.

The following points can be noted by comparing the graphs of chlorophyll concentrations, Secchi depths and pH levels in Figure 2:

- Chlorophyll and pH levels were quite high, and clarity was low (Secchi disk disappeared at 1.5 ft) prior to the alum treatment (which began on May 3, 2018).
- $\circ$  The alum treatment did not remove all algae from the water column, but it did remove the filamentous cyanobacteria that dominated the bloom. Once they were bound up in the floc the pH levels dropped, and more typical algae prospered. As a result, water clarity was improved over pre-treatment conditions, but it was only when temperatures dropped in November 2018 that the lake's clarity was transformed—algae were not gone, but had declined to a mean chlorophyll level of 6.3 µg/L. This was when the filamentous bottom algae mats started to appear.
- Chlorophyll levels stayed within a fairly tight range in the summer of 2019, and clarity ranged from 4-6 feet over the period July-October. In the other months of the year the Secchi disk was visible on the lake bottom. The filamentous benthic mats disappeared in the summer and did not reappear in early winter as they had the previous year.
- Early 2020: there was more variability in conditions over the January-March time frame, with above average rainfall (and one three-inch rain), so more nutrients entering the lake; and warmer temperatures (which could also stimulate algae growth). Stormy weather churned up the lake (Fig. 1) and this was reflected in higher turbidity levels. Chlorophyll and clarity were similar to what was seen in the summer of 2019.

Another way of assessing the differences is by looking at photos taken from the same spot in March of 2018-2020 (Fig. 3).



Figure 3. Photos of White Lake, taken from the same pier on March 15, 2018 (clarity 1.5 ft.), March 18, 2019, (clarity to lake bottom) and March 6, 2020 (clarity 6.5 ft). The yellow material floating on the lake surface is pine pollen.

The phytoplankton analysis done in 2018 and 2019 tells us more about what was in the lake:

- <u>Less dominance, more diversity</u>: while the biomass of phytoplankton was relatively low in April 2019, there were 71 different phytoplankton taxa found that month, indicating a healthy diversity (Appendix 1). The number of different taxa increased to 86 by August 2019. By comparison, there were more cyanobacterial taxa found in 2018, but much lower overall diversity (32 phytoplankton taxa before treatment and 29-50 taxa after the alum treatment; Spirogyra 2018).
- <u>Less abundance</u>: Lower phytoplankton densities were seen in 2019 compared to 2018 (Appendix 2).
- <u>Algal mats come and go</u>: filamentous mats were seen all around the lakeshore from November 2018 through the first half of 2019. This alga was also identified in the phytoplankton samples from August 2018 through May 2019, indicating that filaments can become suspended. They also grow attached to aquatic vegetation, creating the appearance of "furry plants".



Figure 4. The filamentous green algae *Mougeotia* sp. along the White Lake shoreline. Photo taken January 24, 2019.

## A Closer Look at Some White Lake Algae

There are many different kinds of algae found in the lake; one of the most interesting in appearance is a colony-forming golden alga called *Dinobryon*. Single cells are contained in tiny vase-shaped structures that are attached together with stalks (Fig. 5). This alga was very happy for a brief period after the May 2018 alum treatment (note the point labeled as *Dinobryon* on the chlorophyll graph in Fig. 2).





Another common algal group in White Lake are desmids, and their shapes—such as the elongation of portions of a cell seen in *Staurastrum* (Fig. 6)-help keep them afloat.





http://protist.i.hosei.ac.jp/pdb/Images/Chlorophyta/Staurastrum/Eustaurastrum/Processiformes/2\_arms/tetracerum/sp\_01.html ).

The sizes as well as shapes of phytoplankton vary considerably—the tiniest phytoplankton are referred to as picoplankton (they are roughly  $1/50^{\circ}$  of the diameter of a human hair), and one type is abundant in White Lake: the single-celled cyanobacteria *Synechococcus* (Fig. 7). In terms of cell counts it ranks high, but in terms of biomass it doesn't amount to much because of its small size.



Figure 7. A magnified view of the single-celled cyanobacterium <u>Synechococcus</u> sp. (photo by John A. Strand, downloaded from the Research Gate website on May 18, 2020).

A series of photomicrographs was included in the 2017 White Lake Algae Report, which provides a good visual comparison of changes in phytoplankton abundance over the summer, with the photo on the right showing the dominance of the filamentous cyanobacterium <u>Planktolyngbya</u> <u>limnetica</u> (Fig. 8). After the May 2018 low-dose alum treatment, cyanobacterial filaments in samples were visibly impacted, with fragmented filaments and non-living cells, which gradually disappeared from the water column (Spirogyra 2018).



Figure 8. Algal assemblages in White Lake from June to September 2017 (magnified 100x). Photos by Leigh Stevenson, NC DWR.

#### <u>References</u>

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- LIMNOSCIENCES, 2020. 2019 White Lake Monitoring Results. Report to the Town of White Lake.
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Appendix 1. Phytoplankton taxa found in White Lake, 2019. Sample dates analyzed: April 17, May 23, July 10, and September 12. Richness averages determined from three grab samples/month.

4/17/19 Richness	0 54193737	5/23/19	0 47938678		7/10/19		9/12/19	
greens	35	40	0.47350070		53		53	
4/47/40		5/00/40			7/40/40		0/40/40	
4/17/19 Synechococcu	c cn	5/23/19 Synechococcus	sn.		7/10/19	ue en	9/12/19 Synechococcus	en
Cvanogranis f	erriginea	Cyanogranis fe	rriginea		Anhanizome	on sn	Planktolyngbya	op. crassa
Aphanocapsa	sp.	Aphanocapsa d	elicatissima		Planktolyngb	va sp.	Chroococcus ap	nanocapsoides
Raphidiopsis o	urvata	Aphanizomeno	n sp.		Jaaginema s	D.	Aphanocapsa sp	
imnolyngbya	circumcreta	Desmodesmus	sp.		Desmodesmi	us sp.	Planktolyngbya	imnetica
Merismopedia	a tenuissima	Monoraphidiur	m contortum		Monoraphidiu	m contortum	Aphanocapsa de	licatissima
Planktolyngby	a limnetica	Monoraphidiur	m arcuatus		Tetraedron co	nstrictum	Aphanizomenon	sp.
Staurastrum	americanum	Dictyosphaeriu	ım sp.		Pediastrum b	iradiatum	Cyanogranis ferri	ginea
Desmodesmus	s sp.	Tetraedron cor	nstrictum		Staurastrum f	etracerum	Chroococcus apl	nanocapsoides
Oocystis sp.		Pediastrum bir	adiatum		Staurodesmu	s sp.	Raphidiopsis cu	vata
Monoraphidiu	ım contortum	Pediastrum du	plex		Staurastrum a	piculatum	Dolichospermum	sp.
Fetraedron m	inimum	Staurastrum te	etracerum		Pediastrum te	etras	Dictyosphaerium	sp.
Mougeotia sp		Staurodesmus	sp.		Cosmarium a	chondrum	Monoaphidium a	rcuatus
Elakatothrix v	riridis	Staurastrum a	piculatum		Closterium a	utum	Ankistrodesmus	falcatus
Staurastrum 1	etracerum	Pediastrum tet	tras		Selanastrum	sp.	Desmodesmus s	.p.
Monoraphidiu	ım minutum	Cosmarium reg	nesi v. monta	anum	Ankistrodesn	nus falcatus	Tetraedron const	rictum
Pediastrum bi	radiatum	Eutetramorus	planktonica		Tetraedron ca	udatum	Ankistrodesmus	spiralis
Crucigenia tet	rapedia	Koliella sp.			Selanastrum	gracile	Staurastrum apic	ulatum
Pediastrum te	tras	Cosmarium ach	ondrum		Elakatothrix	rindis	Staurastrum tetra	cerum
Staurastrum	margaritaceum	Cosmarium pha	aseolus		Coenochloris	sp.	Staurodesmus s	p.
Staurastrum a	apiculatum	Closterium acu	tum		Sorastrum an	nericanum	Staurastrum man	garitaceum
staurodesmus	s sp.	Selanastrum sp	p.		Monoraphidiu	m convolutum	Pediastrum tetras	5
Coelastrum ca	mbricum	Ankistrodesmu	us falcatus		Staurastrum a	americanum	Pediastrum birad	liatum
Scenedesmus	acutus	Kirchneriella sp	p.		Staurastrum	nargaritaceum	Tetraedron cauda	atum v. longum
Tetraedron ca	udatum v. longum	Tetraedron cau	udatum v. Ion	gum	Crucigenia te	trapedia	Coelastrum prob	oscideum
chlorella sp.		Monoraphidiur	m minutum		Closterium ve	nus	Monoraphidium o	onvolutum
Monoaphidiu	n arcuatus	Tetraedron tri	gonum		Tetraedron m	nimum	Elakatothrix virid	lis
Ankistrodesm	us falcatus	Selanastrum g	racile		Selanastrum	bibrainum	Selanastrum sp.	
Ankistrodesm	us spiralis	Elakatothrixvi	ridis		Coelastrum p	roboscideum	Selanastrum grad	cile
Fetraedron tr	igonum	Ankistrodesmu	us spiralis		Tetraedron vi	ctoriae	Monoraphidium r	ninutum
Docidium sp.		Coenochloris st	D.		Cosmarium s	p.	Staurastrum ame	ricanum
Quadrigula la	custris	Sorastrum ame	ericanum		Microspora s	D.	Sorastrum americ	canum
Cosmarium ac	hondrum	Mougeotia sp.			Coelastrum c	ambricum	Microspora sp.	
Sorastrum am	ericanum	Monoraphidiur	m convolutum	1	Oocystis sp.		Monoraphidium o	ontortum
Cosmarium re	gnesi v. montanum	Docidium sp.			Golenkinia ra	diata	Occystis sp.	
Selanastrum (	gracile	Selanastrum B	ibrainum		Selanastrum	sp.	Cosmarium acho	ndrum
Fetraedron ca	udatum	Golenkinia rad	iata		Tetraedron in	cus	Kirchneriella sp.	
Gonatozygon	sp.	Chlamydomona	as sp.		Monoraphidiu	m minutum	Coelastrum cam	oricum
Dictyosphaeri	um sp.	Staurastrum m	argaritaceun	n	Ankistrodesn	nus spiralis	Staurastrum cha	etoceras
Coelastrum m	icroporum	Closterium gra	cile		Monoraphidiu	m arcuatus	Closterium sp.	
Quadrigula la	custris	Tetraedron mi	nimum		Kirchneriella	sp.	Gonatozygon sp.	
Staurodesmus	sp.	Cosmarium sp.			Botryococcus	sp.	Quadrigula lacus	tris
Aulacoseira g	ranulata	Sphaerozosma	sp.		Cosmarium n	gnesi v. monta	num Tetraedron trigon	um
Navicula sp.		Cyclotella sp.			Tetraedron lin	nneticum	Cosmarium regn	esi v. montanum
Cyclotella sp.		Navicula sp.			Closterium s	<b>)</b> .	Eutetramorus pla	nktonica
Achnanthidiu	n minutissima	Achnanthidium	n minutissima		Nephrocytiun	n sp.	Coenochloris sp	
Fragilaria lon	gifusiformis	Pinnularia sp.			Pleurosigma	sp.	Golenkinia radiat	a
Fragilaria sp.		Fragilaria sp.			Tetredron trig	onum	Closterium gracil	e
Synedra rump	ens	Dinobryon sert	tularia		Schroederia	etigerum	Chlorella sp.	
stauroneis sp		Centritractus b	elanophorus		Eutetramorus	planktonica	Closterium acutu	im
Melosira itali	ca	Dinobryon sp.2	2		Staurastrum	haetoceras	Pediastrum tetra	8
Pinnularia sp.		Synura sp.			Closteriopsis	longissima	Schroederia seti	gerum
Achnanthes e	xiguum	Cryptomonas s	p.		Dictyosphae	ium sp.	Tetraedron trigon	um
Synedra actin	astroides	Komma caudat	a		Pediastrum d	uplex	Coelastrum micro	oporum
Caloneis vent	ricosa	Euglena sp.			Kirchneriella	sp.	Tetrastrum hetera	acanthum
Dinobryon ser	tularia	Trachelomonas	s volvocina		Coelastrum n	nicroporum	Tetraedron limne	ticum
Centritractus	belanophorus	Phacus agilis			Kirchneriella	obesa	Pediastrum duple	эx
Dinobryon sp	2	Peridiopsis sp.			Pinnularia sp		Cosmarium sp.	
Synura sp.		Peridinium sp.			Melosira itali	ca	Pediastrum duple	эx
Chrysochromu	ilina sp.	Cystodinium sp	).		Aulacoseira	ranulata	Tetraedron minim	ıum
Chrysochromu	lina parva				Cymbella sp.		Radiococcus nin	nbatus
Cryptomonas	sp.				Cyclotella sp		Quadrigula lacus	tris
Komma cauda	ta				Navicula abr	ptum	Cosmarium regn	əsi v. montanum
Plagioselmis r	annoplanktonica				Eunotia sp.		Botryococcus sp	
Chilomonas sr					Gomphonema	sp.	Cyclotella sp.	
Euglena sp.					Nitzschia sp.		Eunotia sp.	
Trachelomona	s varians				Dinobryon se	rtularia	Urosolenia erien	sis
Euglena minu	tus				Dinobryon cy	indricum	Navicula sp.	
nacus sp.					Mallomonas	acaroides	Synedra rumpen	3
enumopsis s	у <b>р.</b>				Counterror	alatuuris	Meiosira varians	
arvoulniums	,				Cryptomonas	piatyuns	Syneara una Dinobruon culind	ricum
					Euglena sp	I CIICA d	Dinobryon sertula	aria
					Trachelomon	as varians	Mallomonas aca	roides
					Phacus agilis		Mallomonas pse	udocoronata
					Parvodinium	sp.	Cryptomonas sp.	
					Cystodinium	sp.	Cryptomonas ref	exa
					Systoainium	op.	Lepocinclis acut	a
							Trachelomonas v	arians
							Trachelomonas i	ntermedia
							Phacus agilis	
					-		Peridiniopsis sp.	
					-		Peridinium sp.	
							Parvodinium sp.	

Appendix 2. A comparison of White Lake physio-chemical data and algae data for the month of July, from 2013 to 2019 (data for 2013 and 2017 from NC DEQ, and 2018-2019 from LIMNOSCIENCES, and Spirogyra Diversified Environmental Services).

	7/15/2013	7/20/2017	7/12/2018	7/10/2019
Mean Temperature (C)	28.6	30.4	29.2	29.0
Water Clarity, Measured as Secchi Depth (m)	1.25	1.5	1.75	1.5
Turbidity (NTU)	4.3	3.0		1.9
Mean Algae Abundance, Measured as Chlorophyll <u>a</u> Concentration (μg/L)	27.7	9.6	6	8.5
Mean # Algal Cells/ml			150,643	38,001
# of Algae Taxa			50	78
Mean Algal Biovolume (mm3/m3)			18,306	
pH Range (std. units)	8.0-8.3	6.6-6.8	6.5-6.9	6.5-6.6
Dissolved Oxygen, Mean % Saturation	103	92.5	94	93.5
Mean Total Nitrogen (mg/L)	0.41	0.61	0.70	0.62
Mean Total Phosphorus (mg/L)	0.02	0.02	0.02	<0.02
Number of Samples	3	3	7	6

# A Comparison of White Lake Water Quality Data for July, From 2013-2019