

SPERM MORPHOLOGY, MOTILITY, AND CONCENTRATION IN FERTILE AND INFERTILE MEN

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ABSTRACT

Background Although semen analysis is routinely used to evaluate the male partner in infertile couples, sperm measurements that discriminate between fertile and infertile men are not well defined.

Methods We evaluated two semen specimens from each of the male partners in 765 infertile couples and 696 fertile couples at nine sites. The female partners in the infertile couples had normal results on fertility evaluation. The sperm concentration and motility were determined at the sites; semen smears were stained at the sites and shipped to a central laboratory for an assessment of morphologic features of sperm with the use of strict criteria. We used classification-and-regression-tree analysis to estimate threshold values for subfertility and fertility with respect to the sperm concentration, motility, and morphology. We also used an analysis of receiver-operating-characteristic curves to assess the relative value of these sperm measurements in discriminating between fertile and infertile men.

Results The subfertile ranges were a sperm concentration of less than 13.5×10^6 per milliliter, less than 32 percent of sperm with motility, and less than 9 percent with normal morphologic features. The fertile ranges were a concentration of more than 48.0×10^6 per milliliter, greater than 63 percent motility, and greater than 12 percent normal morphologic features. Values between these ranges indicated indeterminate fertility. There was extensive overlap between the fertile and the infertile men within both the subfertile and the fertile ranges for all three measurements. Although each of the sperm measurements helped to distinguish between fertile and infertile men, none was a powerful discriminator. The percentage of sperm with normal morphologic features had the greatest discriminatory power.

Conclusions Threshold values for sperm concentration, motility, and morphology can be used to classify men as subfertile, of indeterminate fertility, or fertile. None of the measures, however, are diagnostic of infertility. (N Engl J Med 2001;345:1388-93.)

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SEMen analysis is routinely used to evaluate the male partner in infertile couples^{1,2} and to assess the reproductive toxicity of environmental or therapeutic agents.³ Although widely used thresholds for normal semen measurements have been published by the World Health Organization (WHO),⁴⁻⁷ the available norms for sperm concentration, motility, and morphology fail to meet rigorous clinical, technical, and statistical standards. In recognition of these limitations, the nomenclature in the most recent WHO manual⁷ for semen evaluation was changed from “normal” to “reference” values. Two recent prospective studies of semen quality and fertility concluded that the current WHO reference values should be reconsidered.^{8,9}

In this study, we sought to determine values for semen measurements that best discriminate between fertile and infertile men and to evaluate the relative value of standard semen measurements in distinguishing between fertile and infertile men.

METHODS

Study Population

As part of a randomized clinical trial of intrauterine insemination and superovulation in the treatment of infertility at nine centers in the United States, we recruited infertile couples in which the female partners had normal results on fertility evaluation.¹⁰ All of these couples had been unable to conceive for at least 12 months; the mean duration of infertility was 43 months. The women were required to have regular menstrual cycles, a normal hysterosalpingogram, normal results on laparoscopy, and a luteal-phase endometrial-biopsy specimen that was histologically consistent with menstrual dating. The men were required to have some motile sperm in ejaculated semen specimens.¹⁰

Fertile men (controls) were recruited from prenatal classes at the same hospitals in which the infertile couples were recruited, as well as through local advertising. The partners of fertile men had to be pregnant or to have delivered a child within the previous two years. Fertile men were excluded only if they had a history of infertility (inability to conceive during 12 months of attempts), vasovasotomy, or varicocelelectomy.

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All the men were required to be between the ages of 20 and 55 years at the time of enrollment, and their partners were required to be between the ages of 20 and 40 years. The fertile couples were frequency-matched to the infertile couples according to the five-year age groups of both partners. Matching was performed within each clinical site, except in the case of one combination of age groups for which it was difficult to recruit participants — a male partner 20 to 25 years of age with a female partner 25 to 29 years of age. The matching of couples in this category was performed without regard to clinical site. In all, we studied 765 men from infertile couples and 696 men from fertile couples.

Semen Collection and Laboratory Evaluation

Written informed consent was obtained from all participants after recruitment. Semen samples were collected by masturbation at the clinical site, after the men had been asked to abstain from ejaculation for at least 48 hours before semen was collected. All semen analyses were performed manually within one hour after the sample was collected and included measurements of the volume of the ejaculate and determinations of the sperm concentration and the percentage of sperm with any evidence of flagellar movement (percentage motility). Details of these procedures have been published previously.¹¹

Two semen specimens were obtained from each of the fertile men a mean of 16 days apart; 27 fertile men submitted samples more than 30 days apart. From infertile men, up to six semen samples were obtained — two before randomization and one for each of up to four treatment cycles.¹⁰ Of these specimens, we used the two obtained closest together in time; the mean number of days between the specimen collections was 41.5. The mean values for the sperm concentrations, the percentages of motile sperm, and the percentages of sperm with normal morphologic features in the two samples were used in the analysis.

Technicians from the nine clinical sites attended training sessions in semen analysis at the central laboratory at the University of California, Davis. The proficiency of the 26 technicians was tested at the clinical sites approximately twice each year with the use of blindly coded sperm suspensions and videotapes distributed by the central laboratory.

Semen smears were stained at the clinical sites by the Papanicolaou method and shipped to the central laboratory for assessment of sperm morphology by a single technician. Sperm were classified as having normal or abnormal morphologic features according to strict criteria.⁷ Either 200 or 300 sperm were analyzed per slide. Initially, 100 sperm from each of two different locations on the slide were analyzed. If the difference between the percentage of normal sperm in the two areas was 5 percentage points or less, the mean value was calculated. If the difference was more than 5 percentage points, an additional 100 sperm were evaluated from a third location, and the median of the three values was used.

The technician who assessed sperm morphology attended a training session conducted by Dr. Thinus Kruger, who developed the criteria that are used for strict assessments of morphology.⁷ A set of 65 slides from patients in the study, scored by Dr. Kruger, was used as the standard for purposes of quality control. The percentage of sperm with normal morphologic features was known to the technician for 10 of these slides but was unknown for the other 55 standard slides. Each day, the technician scored two of the slides whose morphologic values she knew and compared her results with the standard value; she then scored two unknown slides. Approximately every two months, the mean percentage of sperm with normal morphologic features for these 55 slides was compared with the mean percentage as scored by Dr. Kruger. The two mean values never differed by more than 1 percentage point, and the Spearman rank-correlation coefficient for the two data sets was always at least 0.92.

Statistical Analysis

Data were sent to the data coordinating center at Columbia University, where computerized checks for out-of-range values and errors in logic were performed. Data that did not meet the standards were sent back to the clinical sites for verification. Analyses were per-

formed with the use of the SAS (version 6.12, SAS Institute, Cary, N.C.) and S-Plus (StatSci, Seattle) statistical packages.

Classification-and-regression-tree (CART)¹² analysis was used to estimate thresholds for each sperm measurement that would discriminate between fertile men and infertile men. The CART algorithm uses an exhaustive search of all possible divisions of participants according to one of the continuous predictor variables to identify the division that results in the greatest improvement in the goodness of fit. In the present application, two thresholds were estimated for each sperm measurement; one became the threshold between the subfertile and indeterminate ranges, and the other became the threshold between the indeterminate and fertile ranges. We used the thresholds for discrimination defined by the CART analysis to create categorical variables for the measurements. We used logistic regression to estimate the association of fertility status (using 1 to indicate infertility and 0 to indicate fertility) with the various semen measurements.

We then calculated the sensitivity and specificity of the CART-defined thresholds for classifying infertility. The analysis of receiver-operating-characteristic curves,^{13,14} which defines tradeoffs between sensitivity and specificity along the spectrum of possible thresholds, was used to test whether each semen measurement discriminated between fertile and infertile men and to assess the relative performance of the three semen measurements in making this discrimination.

RESULTS

The demographic characteristics of the study population are shown in Table 1. Partners in fertile couples had higher educational levels than partners in infertile couples. Infertile couples were more likely to be white, to smoke, and to consume alcohol.

There was considerable overlap between the sperm measurements for the fertile men and those for the infertile men. The mean ($\pm$ SD) sperm concentration was $67 \pm 50 \times 10^6$ per milliliter in fertile men (median, 56×10^6 per milliliter) and $52 \pm 42 \times 10^6$ per milliliter in infertile men (median, 42×10^6 per milliliter). The mean percentage of sperm with motility was 54 ± 13 percent in fertile men (median, 55 percent) and 49 ± 15 percent in infertile men (median, 55 percent). The mean percentage of sperm with normal morphologic features was 14 ± 5 percent in fertile men (median, 14 percent) and 11 ± 6 percent in infertile men (median, 10 percent).

The results of the CART analysis for each sperm measurement are shown in Table 2. The values that best defined infertility were a concentration of less than 13.5×10^6 per milliliter, less than 32 percent motility, and less than 9 percent normal morphologic features. The ranges associated with indeterminate fertility were concentrations of 13.5 to 48.0×10^6 per milliliter, 32 to 63 percent motility, and 9 to 12 percent normal morphologic features. The likelihood of infertility increased with decreasing sperm concentration, percentage with motility, or percentage with normal morphologic features (Table 2). For example, relative to a sperm concentration in the fertile range, the odds ratio for infertility was 1.5 (95 percent confidence interval, 1.2 to 1.8) for a sperm concentration in the indeterminate range and 5.3 (95 percent confidence interval, 3.3 to 8.3) for a sperm concentration in the subfertile range.

TABLE 1. CHARACTERISTICS OF INFERTILE AND FERTILE COUPLES FROM NINE REPRODUCTIVE-MEDICINE CENTERS.*

CHARACTERISTIC	MALE PARTNER		P VALUE	FEMALE PARTNER†		P VALUE
	INFERTILE COUPLES (N=765)	FERTILE COUPLES (N=696)		INFERTILE COUPLES (N=765)	FERTILE COUPLES (N=696)	
Age (yr)	34.7±4.9	33.5±5.0	<0.001	32.4±3.8	31.6±4.1	0.004
Bachelor's degree or higher (%)	55.0	64.3	<0.001	49.3	63.4	<0.001
White race (%)	91.0	85.2	<0.001	90.3	85.4	0.004
Current smoking (%)‡	17.9	12.5	0.02	12.2	8.7	0.17
Alcohol consumption (%)§	62.7	57.4	0.06	40.5	22.3	<0.001

*Plus-minus values are means ±SD. Data on age were missing for 1 female partner in a fertile couple; data on education were missing for 113 male partners in fertile couples and 114 female partners in fertile couples; data on race or ethnic group were missing for 73 male partners in fertile couples and 112 female partners in fertile couples; and data on smoking status were missing for 195 male partners in infertile couples, 31 male partners in fertile couples, 181 female partners in infertile couples, and 147 female partners in fertile couples.

†All female partners in infertile couples had normal results on fertility evaluation.

‡Current smoking was defined as smoking within one month before the questionnaire was completed.

§Alcohol consumption was defined as any alcohol consumption during the previous week.

TABLE 2. FERTILE, INDETERMINATE, AND SUBFERTILE RANGES FOR SPERM MEASUREMENTS FROM CLASSIFICATION-AND-REGRESSION-TREE ANALYSIS AND CORRESPONDING ODDS RATIOS FOR INFERTILITY.*

VARIABLE	SEMEN MEASUREMENT		
	CONCENTRATION	MOTILITY	MORPHOLOGY
	×10 ⁶ /ml	%	% normal
Fertile range	>48.0	>63	>12
Indeterminate range	13.5–48.0	32–63	9–12
Univariate odds ratio for infertility (95% CI)	1.5 (1.2–1.8)	1.7 (1.5–2.2)	1.8 (1.4–2.4)
Subfertile range	<13.5	<32	<9
Univariate odds ratio for infertility (95% CI)	5.3 (3.3–8.3)	5.6 (3.5–8.3)	3.8 (3.0–5.0)

*CI denotes confidence interval.

The odds of infertility increased with an increasing number of sperm measurements in the subfertile range (Table 3). In comparison with an increase by a factor of 2 to 3 in the likelihood of infertility when one sperm measurement was in the subfertile range, there was an increase by a factor of 5 to 7 in the risk of infertility when two sperm measurements were subfertile, and an increase by a factor of 16 when all three measurements were subfertile. For example, when the percentage of sperm with normal morphologic features was in the fertile range but the percentage of motile sperm and the concentration were both in the subfertile range, the odds ratio for infertility was 5.5.

The frequency distributions of fertile and infertile

men with regard to sperm concentration, motility, and morphology, divided on the basis of the thresholds determined by the CART analysis, are shown in Figure 1. There was a marked excess of infertile men with values in the subfertile ranges of these semen measurements, and a corresponding excess of fertile men with values in the fertile ranges. Approximately equal proportions of fertile and infertile men had values in the indeterminate ranges of these variables.

On the basis of the area under the receiver-operating-characteristic curve, each of the three measurements — the sperm concentration, percentage of motile sperm, and percentage of sperm with normal morphologic features — provided information that was helpful in discriminating between fertile and infertile men. The area under the curve for the percentage with normal morphologic features (0.66) was significantly greater than that for sperm concentration (0.60; $P<0.001$) and motility (0.59; $P<0.001$), whereas the areas under the curves for sperm concentration and motility were similar. The sensitivity and specificity of these measurements for identifying infertile men at various thresholds, including those defined by our CART analysis, are shown in Table 4. Lowering the threshold for indeterminate fertility reduces the sensitivity of the measures (the likelihood of correctly identifying infertile men) but increases their specificity (the likelihood of correctly identifying fertile men).

DISCUSSION

The results of this study confirm that measurements of sperm concentration, motility, and morphology all

TABLE 3. ODDS RATIOS FOR INFERTILITY FOR COMBINATIONS OF SPERM MEASUREMENTS.*

MORPHOLOGIC FEATURES	SPERM MEASUREMENT RANGE		ODDS RATIO (95% CI)
	MOTILITY	CONCENTRATION	
Fertile	Fertile	Fertile	1.0
Subfertile	Fertile	Fertile	2.9 (2.2–3.7)
Fertile	Subfertile	Fertile	2.5 (1.6–4.2)
Fertile	Fertile	Subfertile	2.2 (1.3–3.6)
Subfertile	Subfertile	Fertile	7.2 (4.3–12.2)
Subfertile	Fertile	Subfertile	6.3 (3.8–10.3)
Fertile	Subfertile	Subfertile	5.5 (3.0–10.2)
Subfertile	Subfertile	Subfertile	15.8 (8.7–29.0)

*The ranges of the sperm measurements were defined by the following thresholds derived from the classification-and-regression-tree analysis: percentage with normal morphologic features — subfertile, less than 9 percent; fertile, more than 12 percent; percent motile — subfertile, less than 32 percent; fertile, more than 63 percent; concentration — subfertile, less than 13.5×10^6 per milliliter; fertile, more than 48.0×10^6 per milliliter. The reference group for the odds ratios is the men with all three measurements in the fertile range. CI denotes confidence interval.

provide useful information for diagnosing male infertility. Sperm morphology, as measured according to strict criteria, appears to be the most informative semen measurement for discriminating between fertile and infertile men. However, none of the measures, alone or in combination, can be considered diagnostic of infertility.

Several different approaches have been used to identify standards for normal semen measurements. Some focus on infertile couples, comparing those who conceive with those who do not.^{15,16} Others have compared fertile men with infertile men^{17–19} or have followed couples after they discontinued the use of contraception.^{8,9}

Studies focusing on infertile couples undergoing treatment — i.e., those comparing couples who conceive with those who do not conceive^{15,16} — are limited by the inclusion of infertile couples only; in order to define the fertile ranges of semen measurements, fertile men must also be evaluated. Other reports have involved follow-up of couples who have discontinued their use of contraception.^{8,9} Although this approach

Figure 1. Percentage of Men from Infertile and Fertile Couples with Values in the Subfertile, Indeterminate, and Fertile Ranges for Sperm Concentration (Panel A), Percentage of Motile Sperm (Panel B), and Percentage of Sperm with Normal Morphologic Features (Panel C), as Defined by Classification-and-Regression-Tree Analysis.

Arrows indicate the thresholds between subfertile and indeterminate ranges (left) and indeterminate and fertile ranges (right).

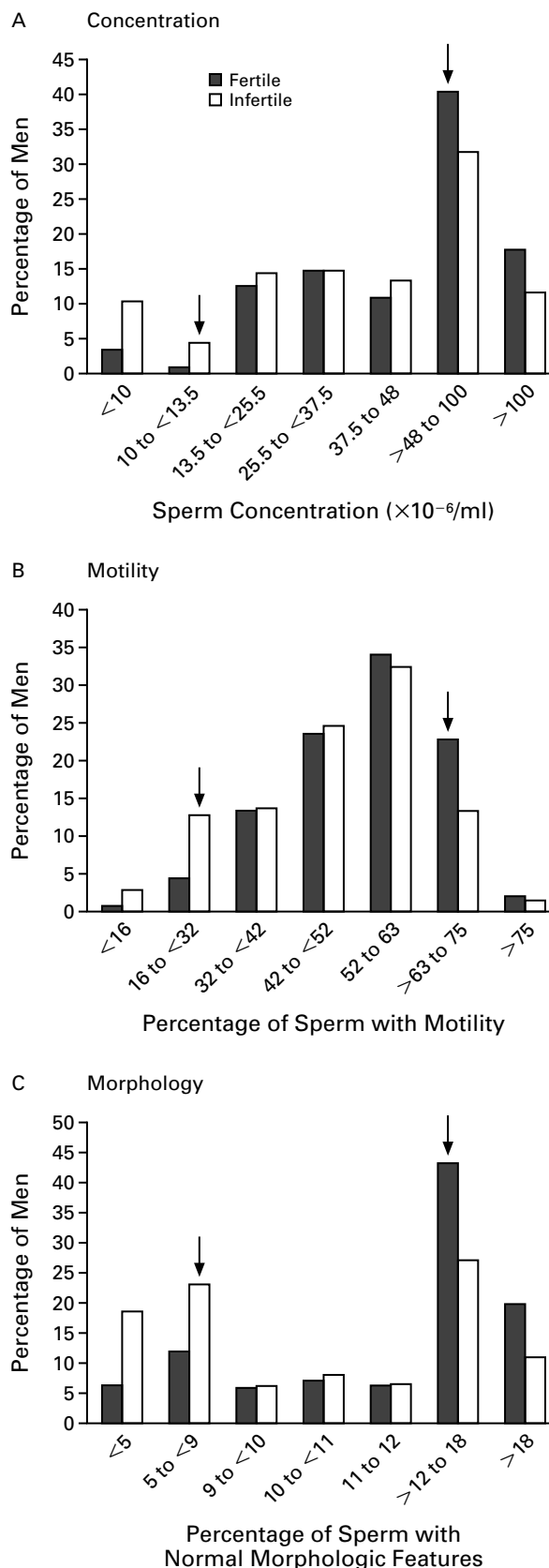


TABLE 4. SENSITIVITY AND SPECIFICITY OF SPERM MEASUREMENTS FOR IDENTIFYING INFERTILE MEN AT VARIOUS THRESHOLDS.

SPERM MEASUREMENT	SENSITIVITY SPECIFICITY	
	percent	
Concentration		
10.0×10 ⁶ /ml	10.2	96.8
13.5×10 ⁶ /ml*	14.8	96.1
48.0×10 ⁶ /ml†	57.1	58.0
60.0×10 ⁶ /ml	67.7	45.0
Percentage of motile sperm		
25	8.1	97.7
32*	15.3	95.1
63†	84.6	26.2
75	98.6	2.7
Percentage with normal morphologic features		
5	18.6	93.8
9*	43.3	81.4
11†	60.8	65.3
15	75.6	41.0

*This value was defined by classification-and-regression-tree analysis as the threshold between the subfertile range and the indeterminate range.

†This value was defined by classification-and-regression-tree analysis as the threshold between the indeterminate range and the fertile range.

has the advantage of prospectively defining couples as fertile and infertile, it requires large samples, since only 8 to 9 percent of couples are infertile.²⁰ Moreover, in an unknown proportion of infertile couples, the woman is infertile. Two recent studies that used this approach concluded that a reevaluation of the existing standards for normal semen was needed,^{8,9} but neither study derived new standards.

A comparison of semen measurements between fertile and infertile men, which was our approach, was used in the 1950s by MacLeod and Gold.^{17,21,22} In these earlier studies, however, modern methods of semen evaluation were not used, and data were obtained from male partners in infertile couples regardless of the fertility status of the female partners. Nevertheless, the minimal standards for sperm concentration (20×10⁶ per milliliter) and motility (40 percent motile cells) reported by MacLeod and Gold are near the values we derived. The standard for morphology (60 percent with normal morphologic features) cannot be compared with our results because a different scoring system was used.

In our study, sperm morphology was assessed by a single person with extensive training and substantial experience, and reliability was monitored on an ongoing basis. The application of our results in clinical laboratories would require the training of technicians and the implementation of tools for continuous calibration. The subfertile and fertile ranges for morphol-

ogy (less than 9 percent and more than 12 percent with normal morphologic features, respectively) might appear to be so close that they would be hard to distinguish. With the system of training for technicians and the calibration methods we used, however, there were only 2 of the 65 quality-assurance slides for which the assessment of the percentage of normal sperm spanned the range from less than 9 percent to more than 12 percent in 14 readings during the course of the study. Although we assessed morphology according to strict criteria, our results do not necessarily imply that this method is superior to other approaches. Nevertheless, our results do provide a reference value for sperm morphology that is missing from the current WHO manual for semen evaluation.⁷

Instead of a single value for each semen measurement that presumably distinguishes between "normal" and "abnormal," we estimated the best two values that allow for the delineation of three groups — fertile, indeterminate, and subfertile. We believe that this classification system is clinically meaningful²³ and is appropriate to what is, biologically, a continuous function.

Our data suggest that caution must be used in interpreting the significance of any given subfertile or indeterminate semen measurement. Although low values for each measurement increase the likelihood that a male factor contributes to infertility, there was substantial overlap in the frequency distributions in our study. Thus, values for sperm concentration, motility, or morphology that are in the subfertile range do not exclude the possibility of normal fertility.

To facilitate recruitment, we defined fertility as pregnancy within the previous two years rather than current pregnancy. Even though we attempted to match the fertile and infertile couples according to age and geographic location, there were demographic differences between the two groups. There were also differences between the two groups in the interval between the collection of the two semen specimens. In addition, despite normal results on fertility evaluation of the female partner, there may have been unrecognized subclinical female factors contributing to the infertility of the infertile couples. Finally, confirmation of the validity of these thresholds for semen measurements in an independent sample of fertile and infertile men is needed.

Notwithstanding these limitations, our data from a large group of couples with well-documented fertility or infertility provide clinical standards for semen measurements that may be useful for diagnosing male-factor infertility and for distinguishing between subfertile, indeterminate, and fertile ranges. These thresholds can be applied in clinical practice and research, provided that there is strict quality control.

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APPENDIX

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