

# KOPANA Spring Seminar 2024

Chi Young Ok, MD

Associate Professor

Department of Hematopathology

UT MD Anderson Cancer Center

Education:

- Medical School: Yonsei University of College of Medicine
- Residency: University of Massachusetts
- Fellowship: UT MD Anderson Cancer Center

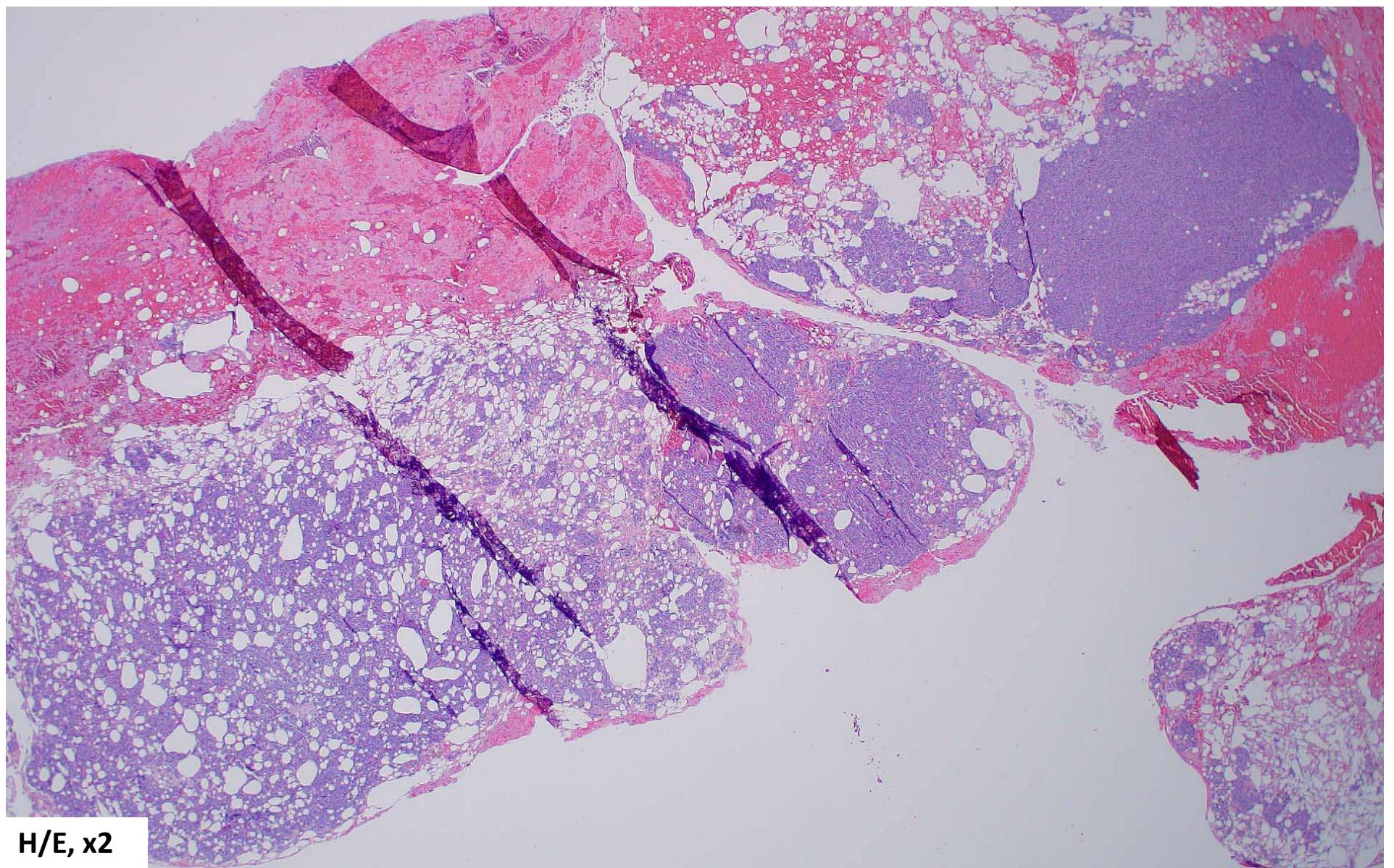
Current Position & Institution: Associate Professor,  
Department of Hematopathology, UT MD Anderson Cancer  
Center

Subspecialty area: Hematopathology, molecular genetic  
pathology

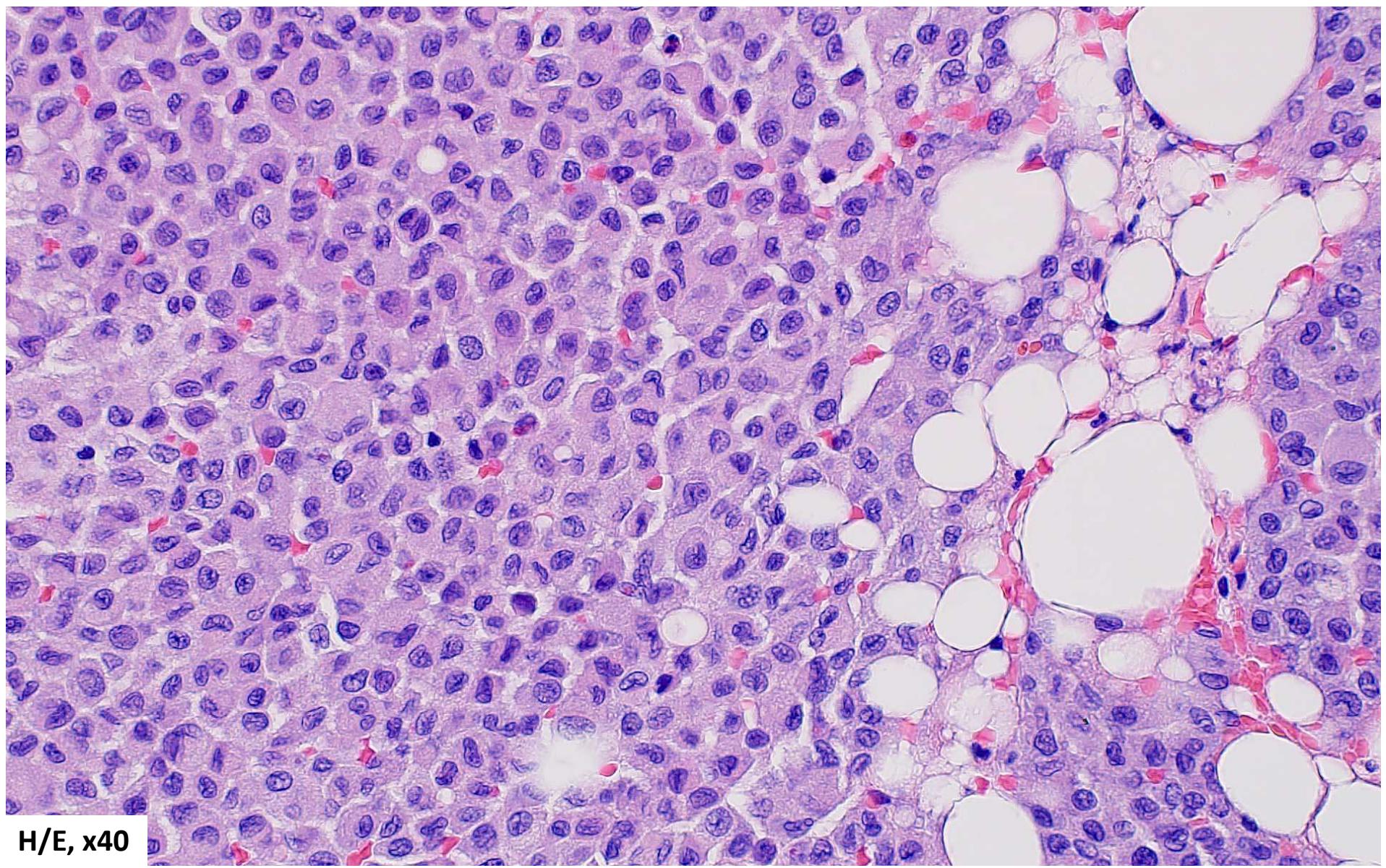


# Case 1

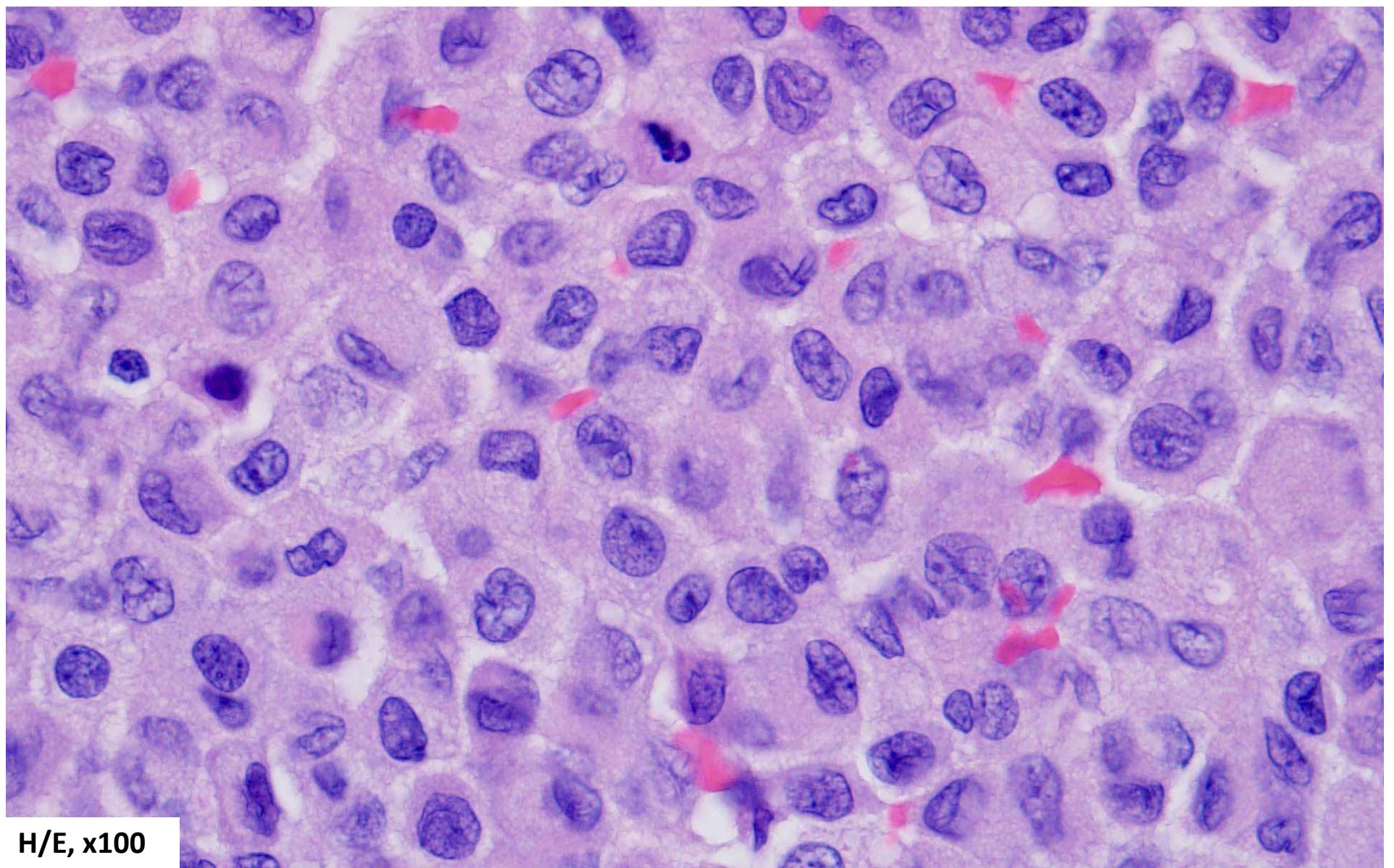
- 71 y/o man
- h/o squamous cell carcinoma at base of tongue (8 years ago)
- s/p chemoradiation, completed 8 years ago
- Visits hospital for aortic valve replacement
- Aortic valve: valve tissue with degeneration and calcification
- Atrial appendage and blood clot
  - Atrial appendage: no significant pathologic change
  - Blood clot



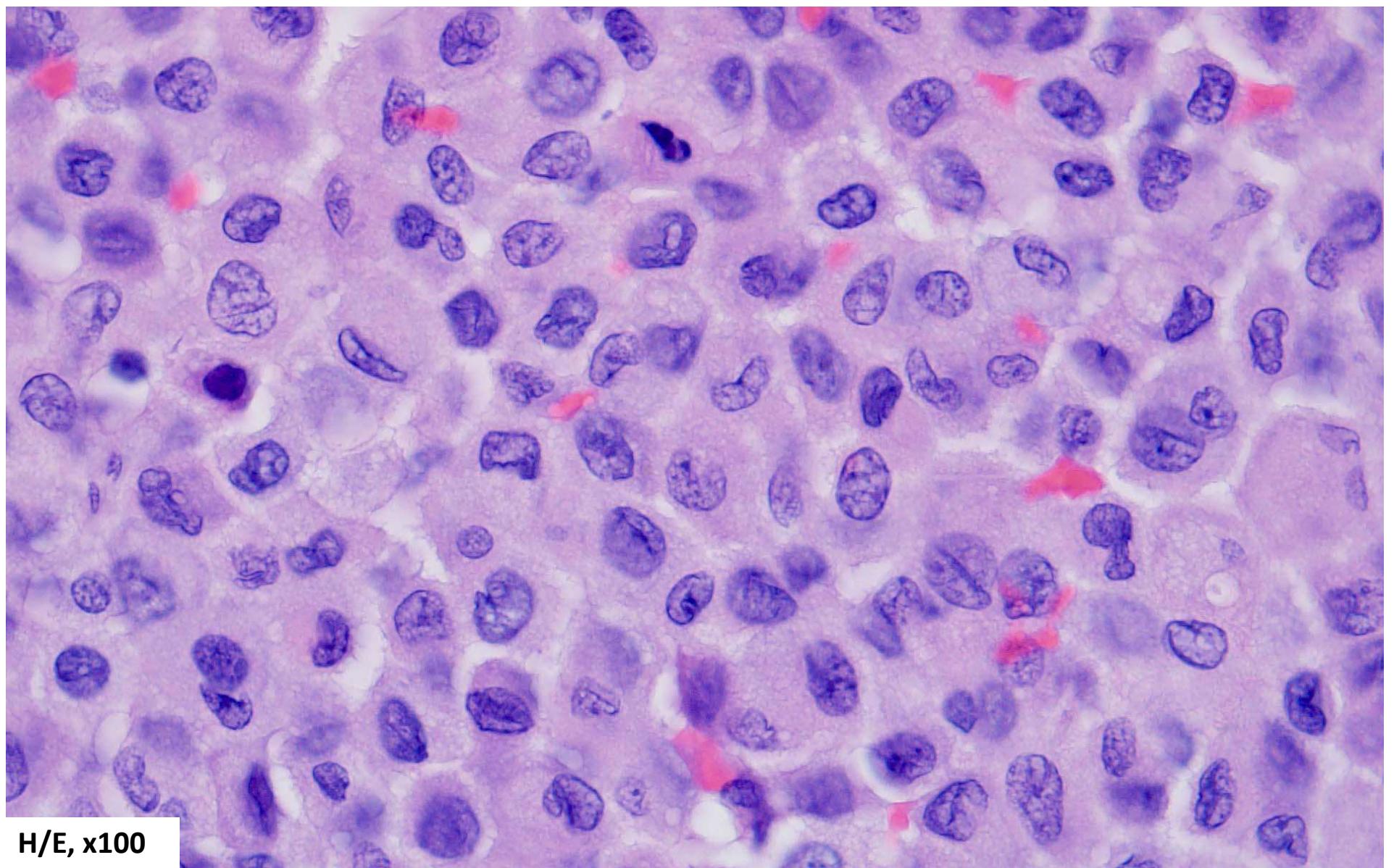
H/E, x2



H/E, x40



H/E, x100

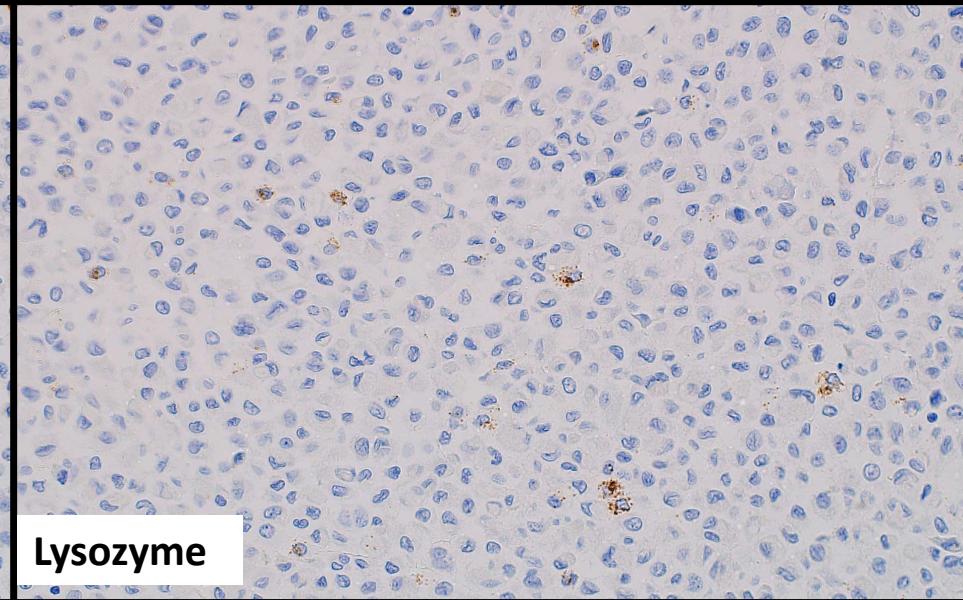
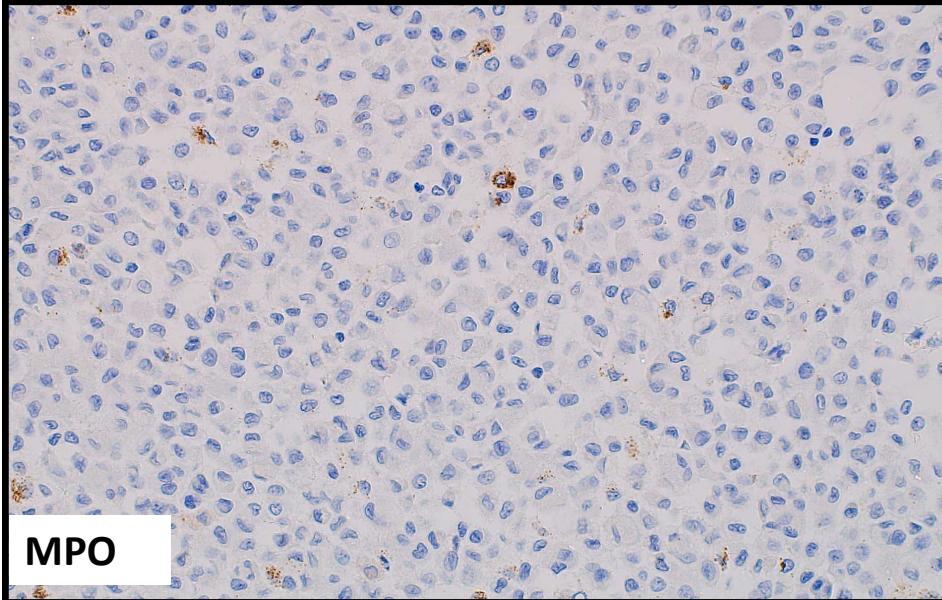
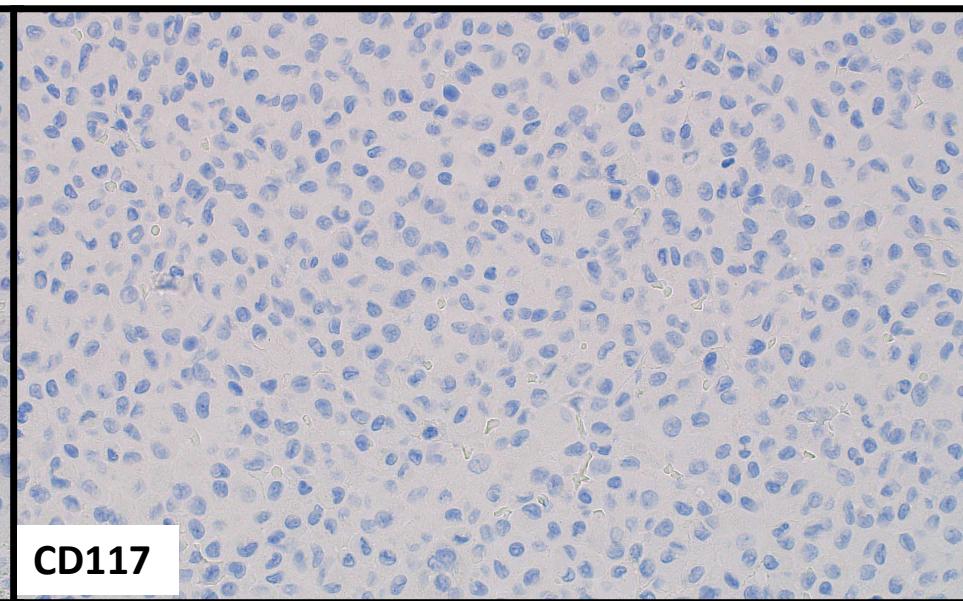
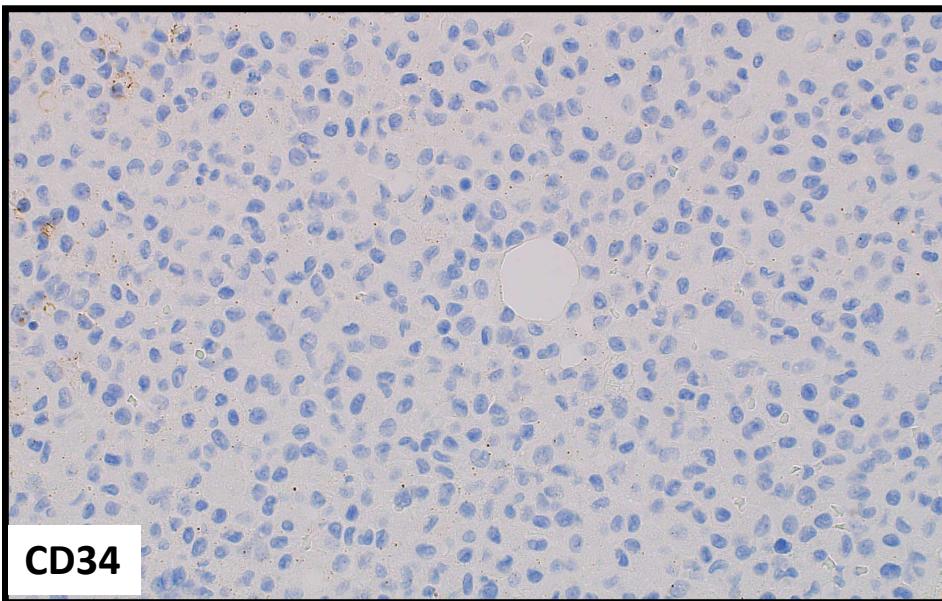


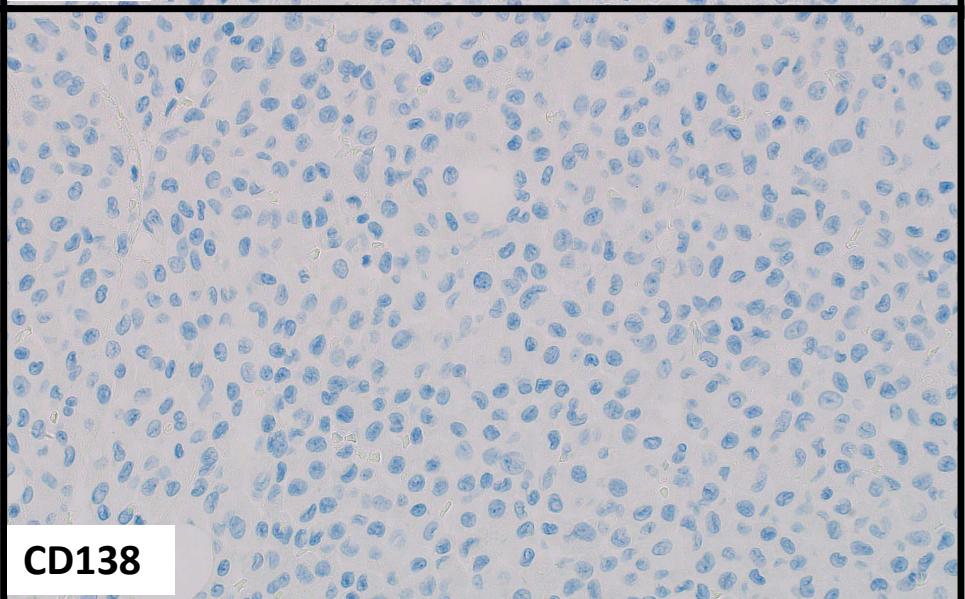
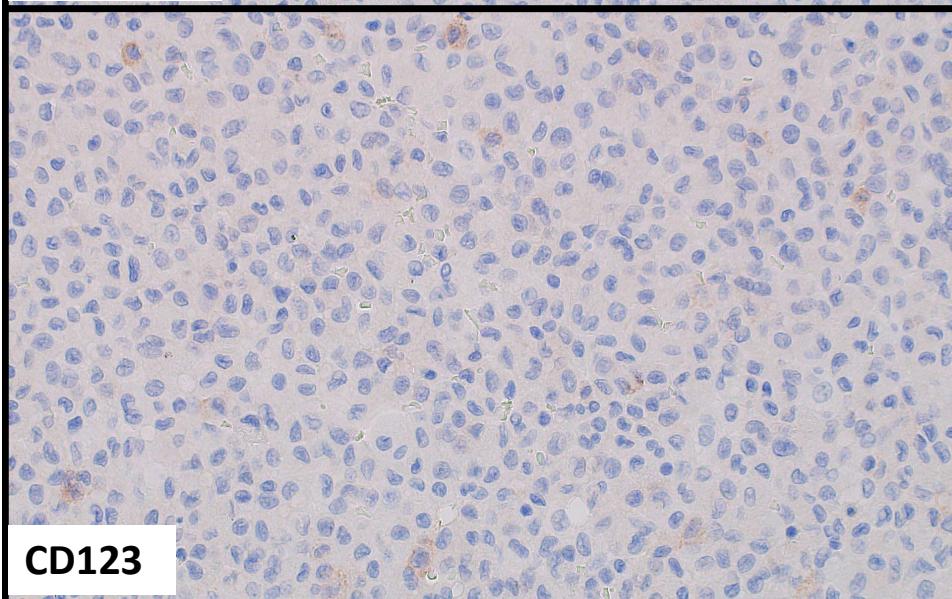
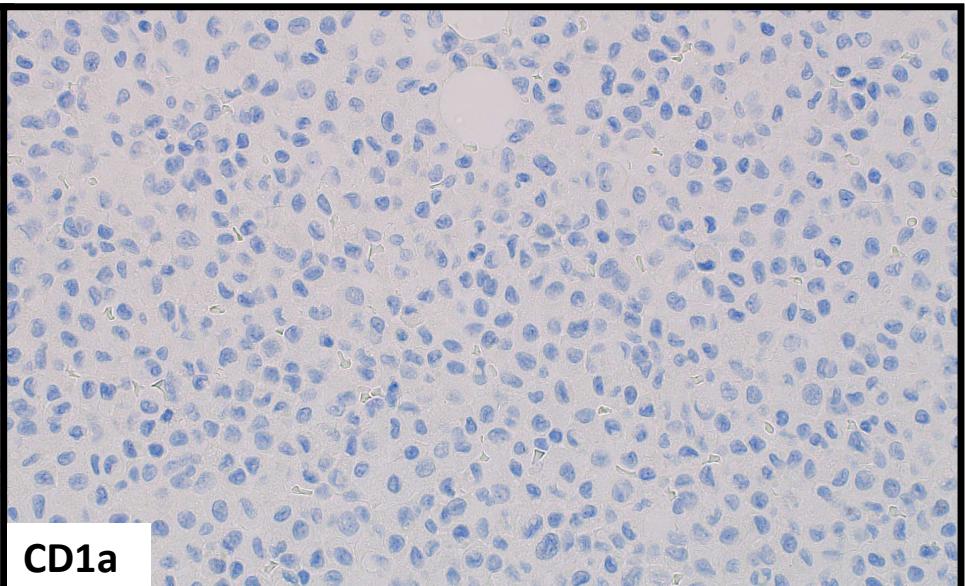
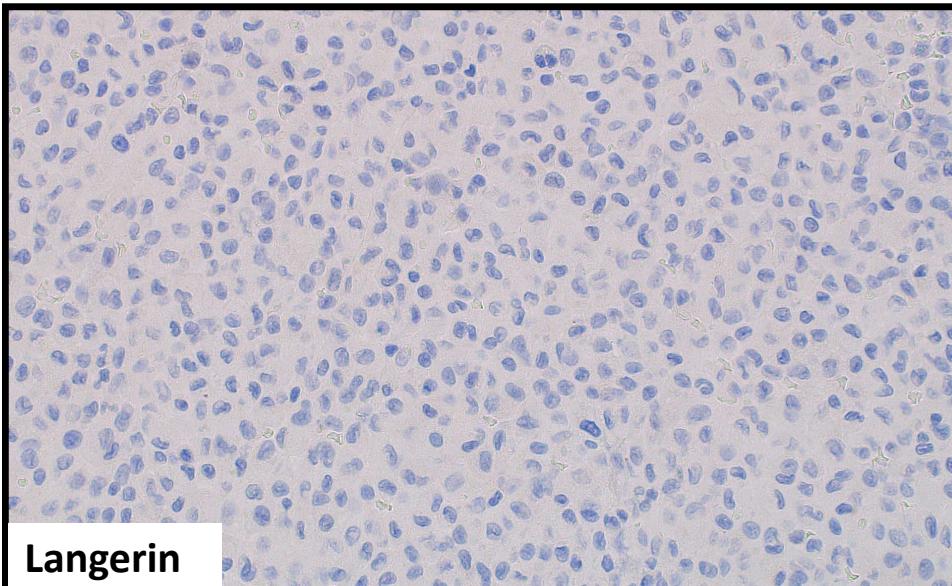
H/E, x100

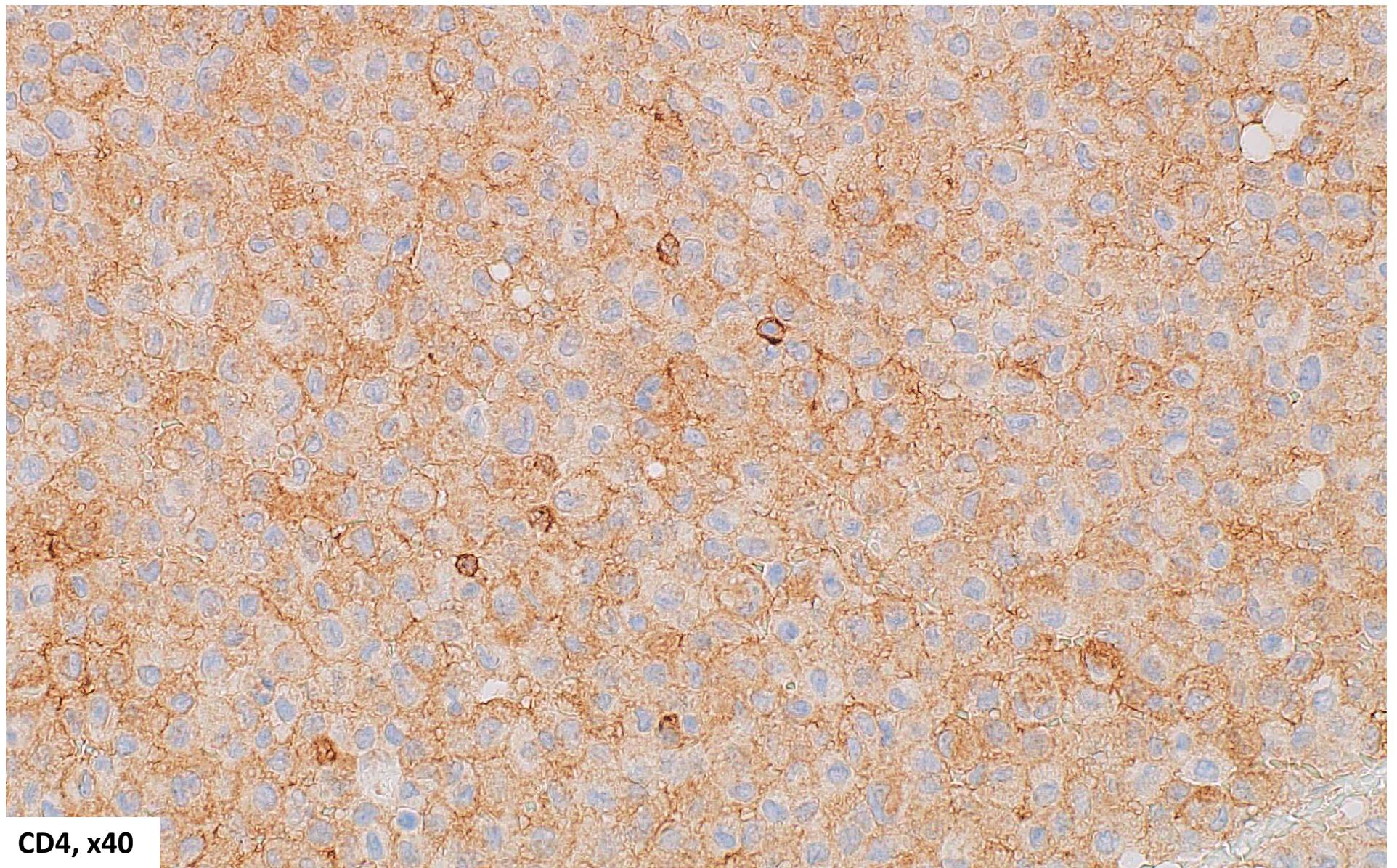


# Differential diagnosis

- Langerhans cell histiocytosis
- Myeloid sarcoma
- Histiocytic sarcoma



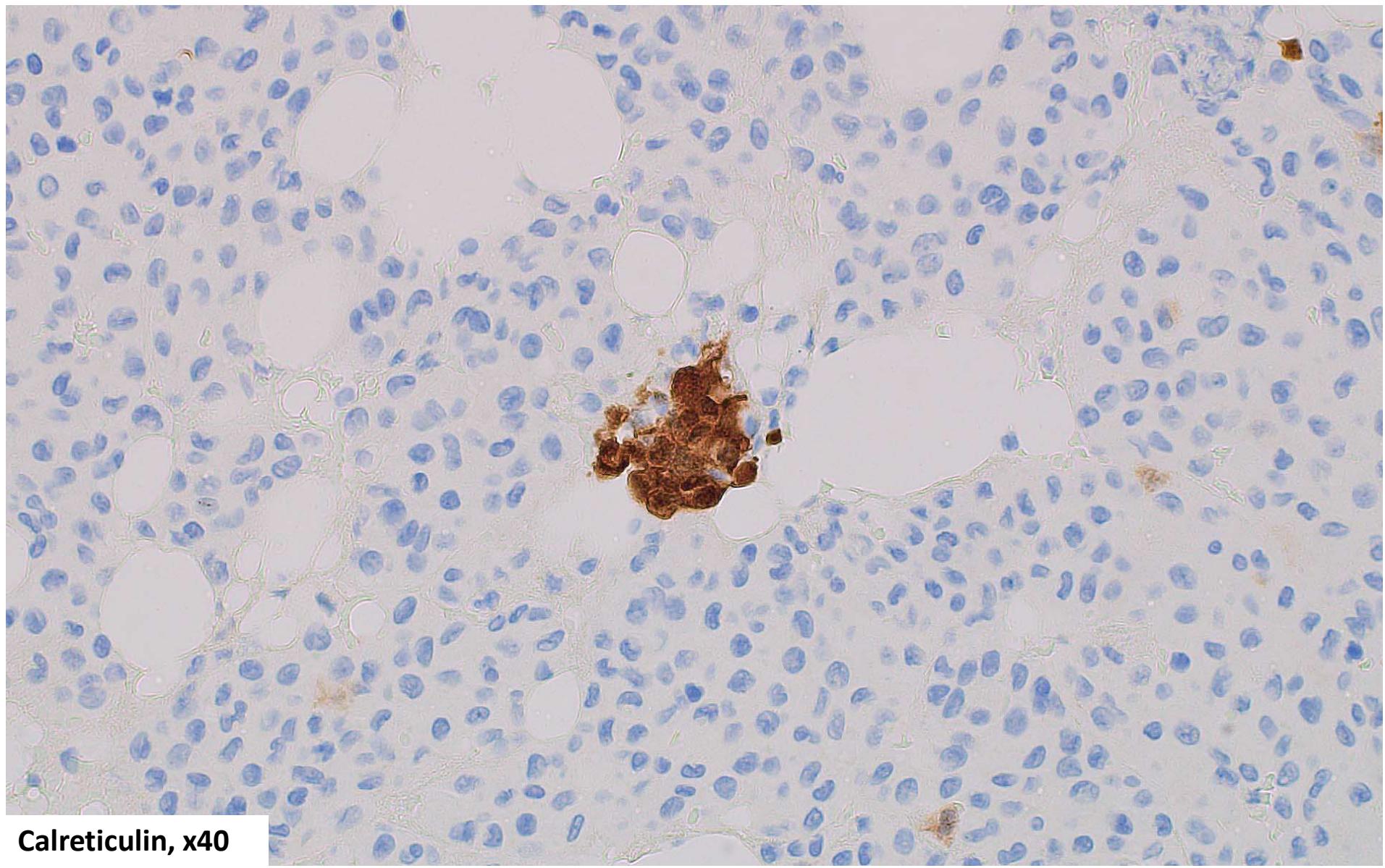




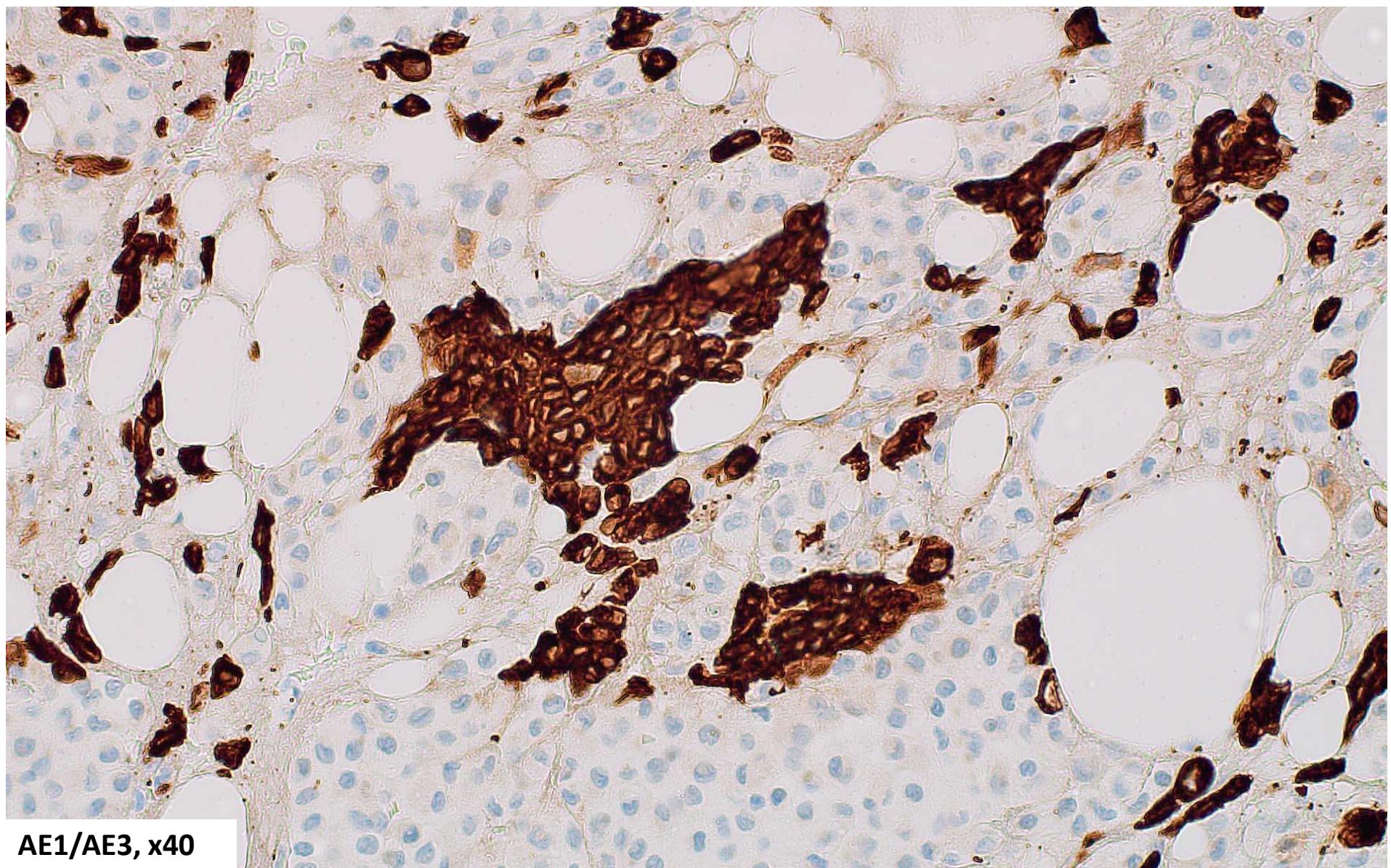
**CD4, x40**

# Differential diagnosis

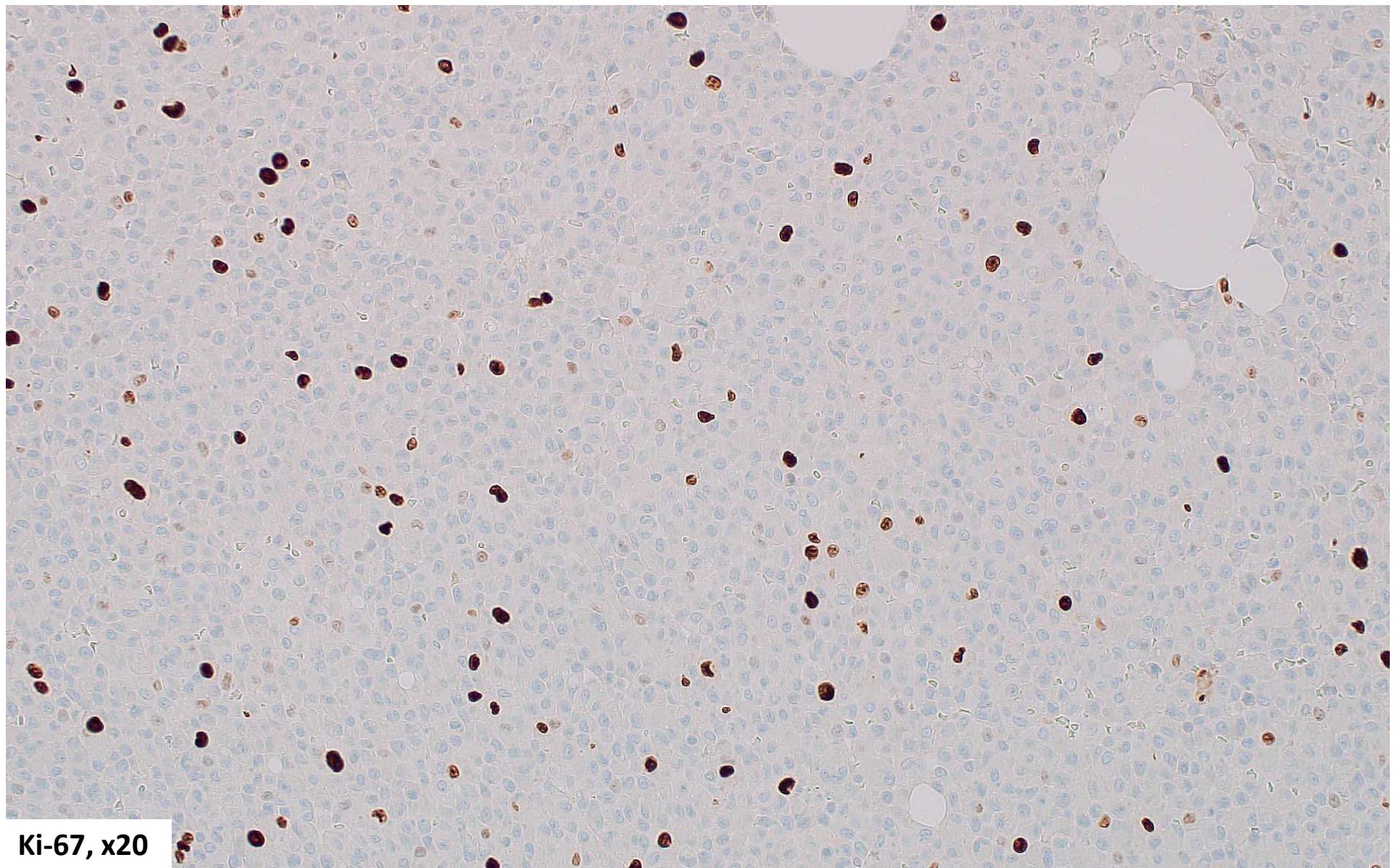
- ~~Langerhans cell histiocytosis~~
- ~~Myeloid sarcoma~~
- ~~Histiocytic sarcoma~~



Calreticulin, x40



AE1/AE3, x40



Ki-67, x20

# Diagnosis

- Mesothelial/monocytic incidental cardiac excrescences (MICE)

# Mesothelial/monocytic incidental cardiac excrencences

- Synonyms
  - Nodular mesothelial hyperplasia (NMH) by Rosai/Dehner
  - **Nodular histiocytic mesothelial hyperplasia (NHMH)** by JKC Chan
  - Nodular histiocytic aggregates
  - Nodular histiocytic hyperplasia
- Benign and reactive nature
- Pathogenesis
  - unknown
- Can occur in any age, gender or race group

# Mesothelial/monocytic incidental cardiac excrescences (MICE)

- Usual sites
  - Mesothelial-lined surfaces: peritoneum, pericardium, pleura, hernia sac and spermatic cord
    - MICE: endocardial involvement of NHMH
  - Lung, urinary bladder, endometrium and endocardium
- Almost always incidental finding
- Gross finding
  - Brown-tan soft tissue excrescence
  - Size range: <1 mm to 3 cm

# Mesothelial/monocytic incidental cardiac excrencences (MICE)

- Microscopic finding
  - Biphasic pattern composed of mildly to moderately pleomorphic cells (epithelioid, polygonal or ovoid cells with reniform or contorted nuclei, inconspicuous nucleoli with or without nuclear grooves and abundant cytoplasm) with distinct margins in cohesive clusters, and a lesser component of scattered, hyperchromatic and cuboidal cells
  - Lymphocytes and plasma cells can be seen, but no eosinophils
  - Mesothelial cells can be absent: entrapped?
- Immunohistochemistry
  - Histiocytes: CD4, CD68, CD163
  - Mesothelial cells: calretinin, WT-1, D2-40, CK5/6 and cytokeratin

**Table 1**

Common differential diagnoses of nodular histiocytic/mesothelial hyperplasia.

Differential Diagnosis (DDx)	DDx Features	NHMH
Neuroendocrine tumor	Lesional cells are keratin-positive, synaptophysin- and/or chromogranin- positive.	Constituting cells are of histiocytic and mesothelial origin, synaptophysin- and chromogranin negative
Hematolymphoid malignancy	Histomorphology and immunoprofile is dependent on the classification of leukemia/lymphoma	Constituting cells are of histiocytic and mesothelial origin.
Malignant mesothelioma	Bulky disease with stromal invasion, complex architecture, and necrosis may be seen, proliferating cells are of mesothelial origin, loss of BAP1 immunoexpression [12]	Usually microscopic lesion without stromal invasion, nodular growth, mixed with histiocytic component, intact BAP1 expression
Well-differentiated papillary mesothelioma	Bland mesothelial cells arranged in a predominantly papillary pattern with or without focal glandular/tubular/nesting pattern. PAX8 immunohistochemistry shows at least partial staining in most cases [13]	Predominant nodular growth pattern mixed with histiocytic component, PAX8-negative



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Review

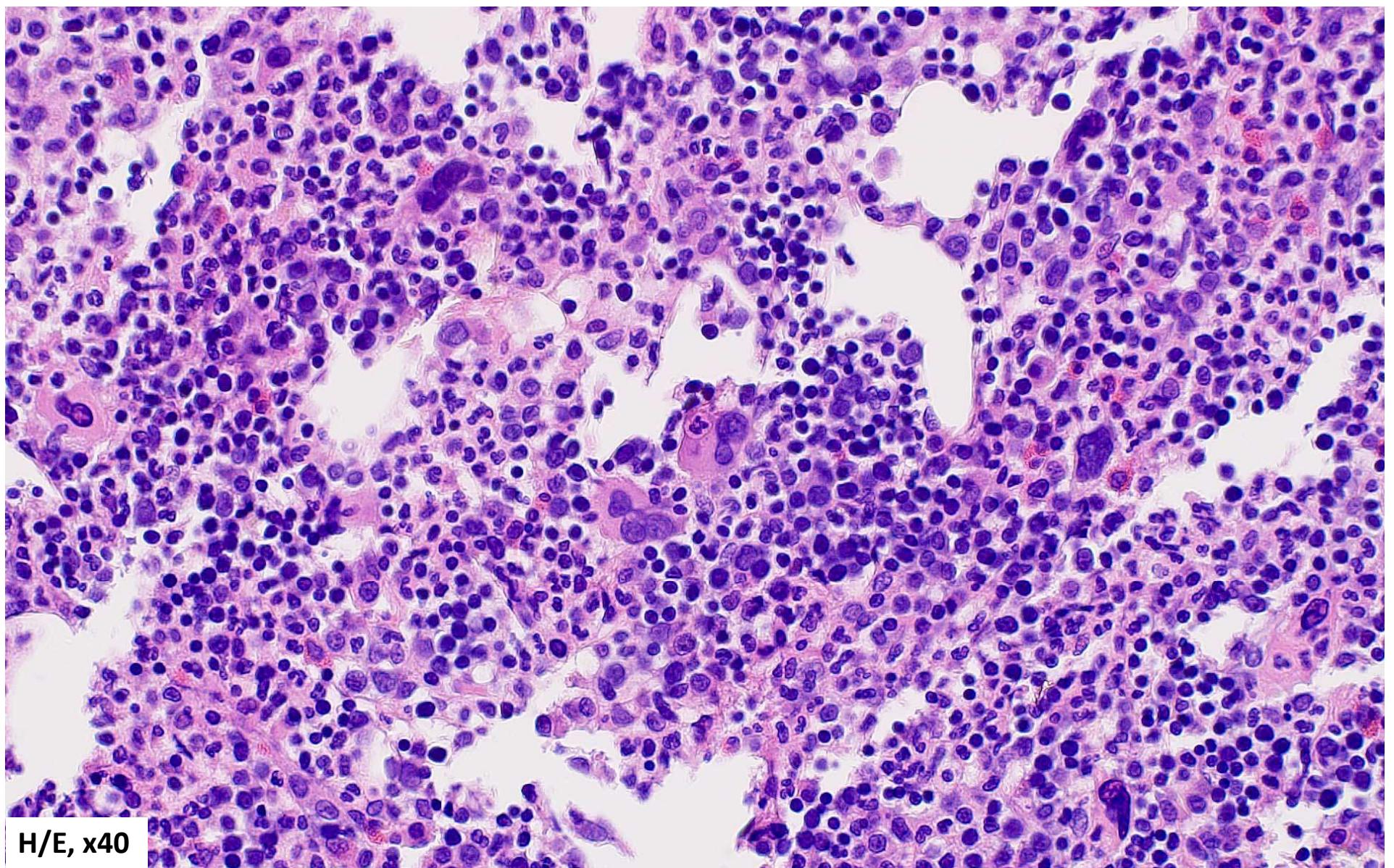
Nodular histiocytic/mesothelial hyperplasia, a benign entity posing diagnostic challenge

Hasan Basri Aydin, Anne Chen, Hwajeong Lee\*

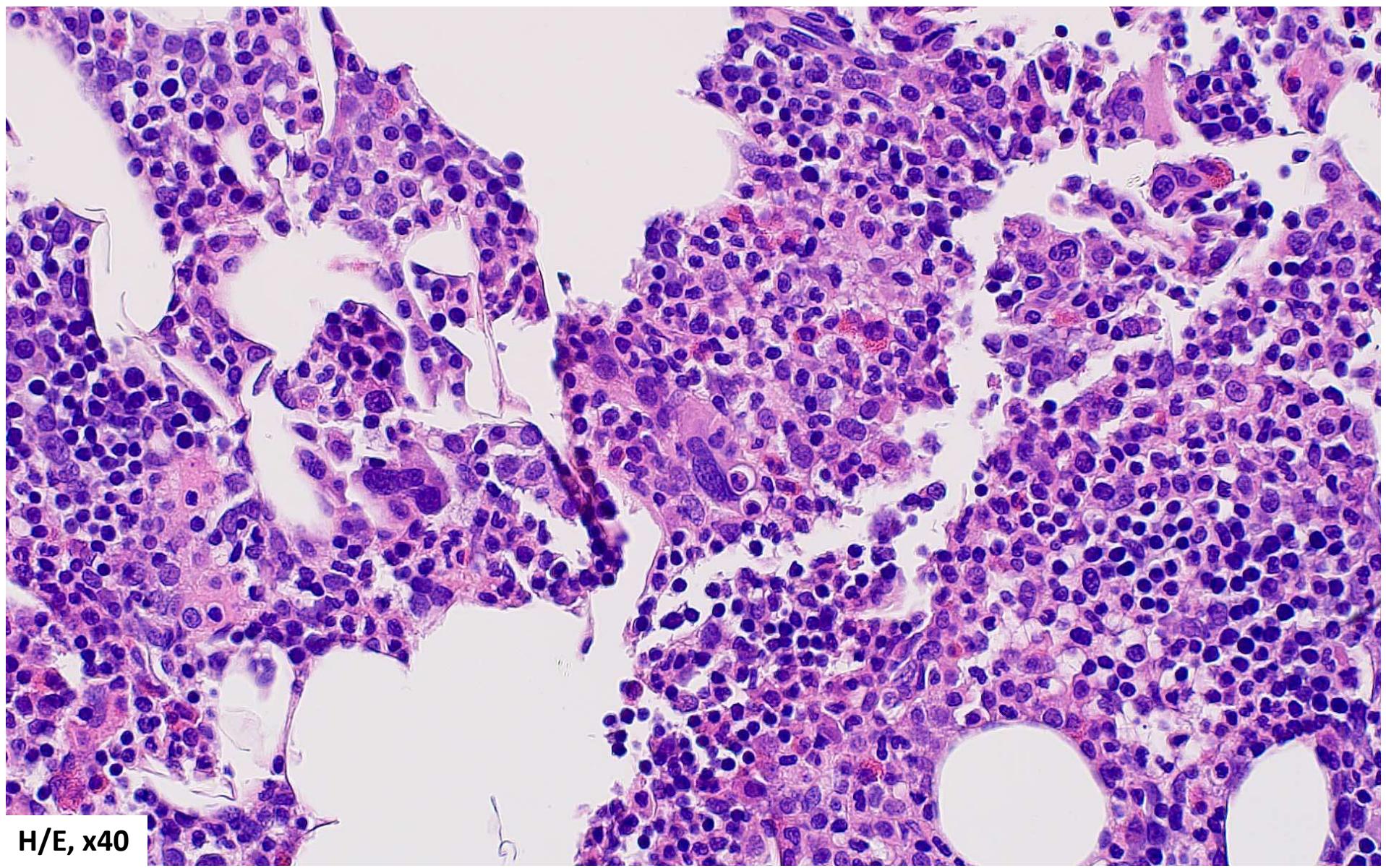
Department of Pathology and Laboratory Medicine, Albany Medical Center, Albany, NY 12208, USA

## Case 2

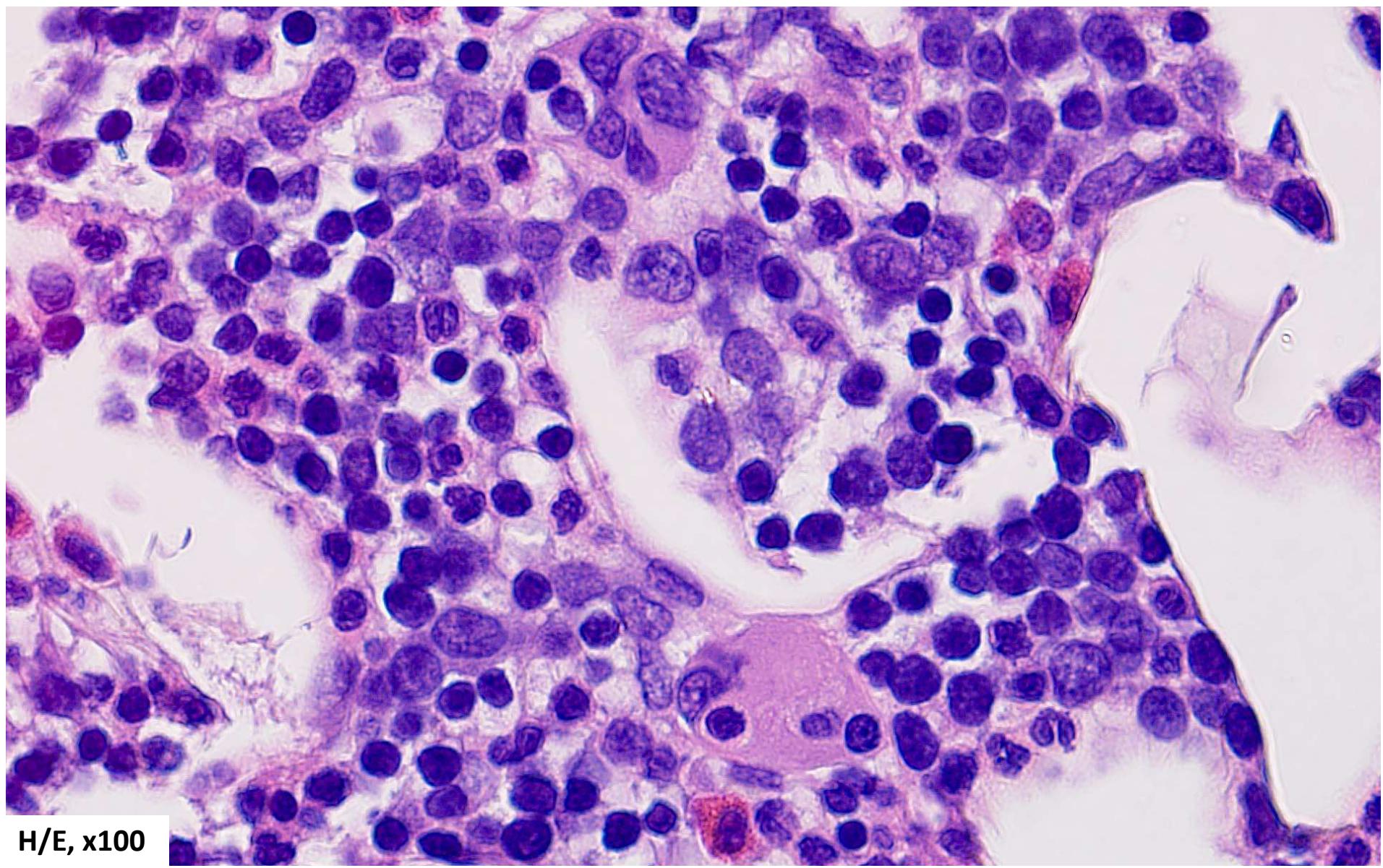
- 23-year-old man
- Leukopenia and thrombocytopenia for 4 years
- Complains of occasional epistaxis and easy bruising
- Hgb: 16.7 d/dL
- WBC:  $2.5 \times 10^9/L$
- Platelet:  $89 \times 10^9/L$
- Hepatomegaly (17.7 cm) and splenomegaly (15.2 cm)
- Vitamin B12: 2,841 pg/mL (reference: 232 -1,245 pg/mL)



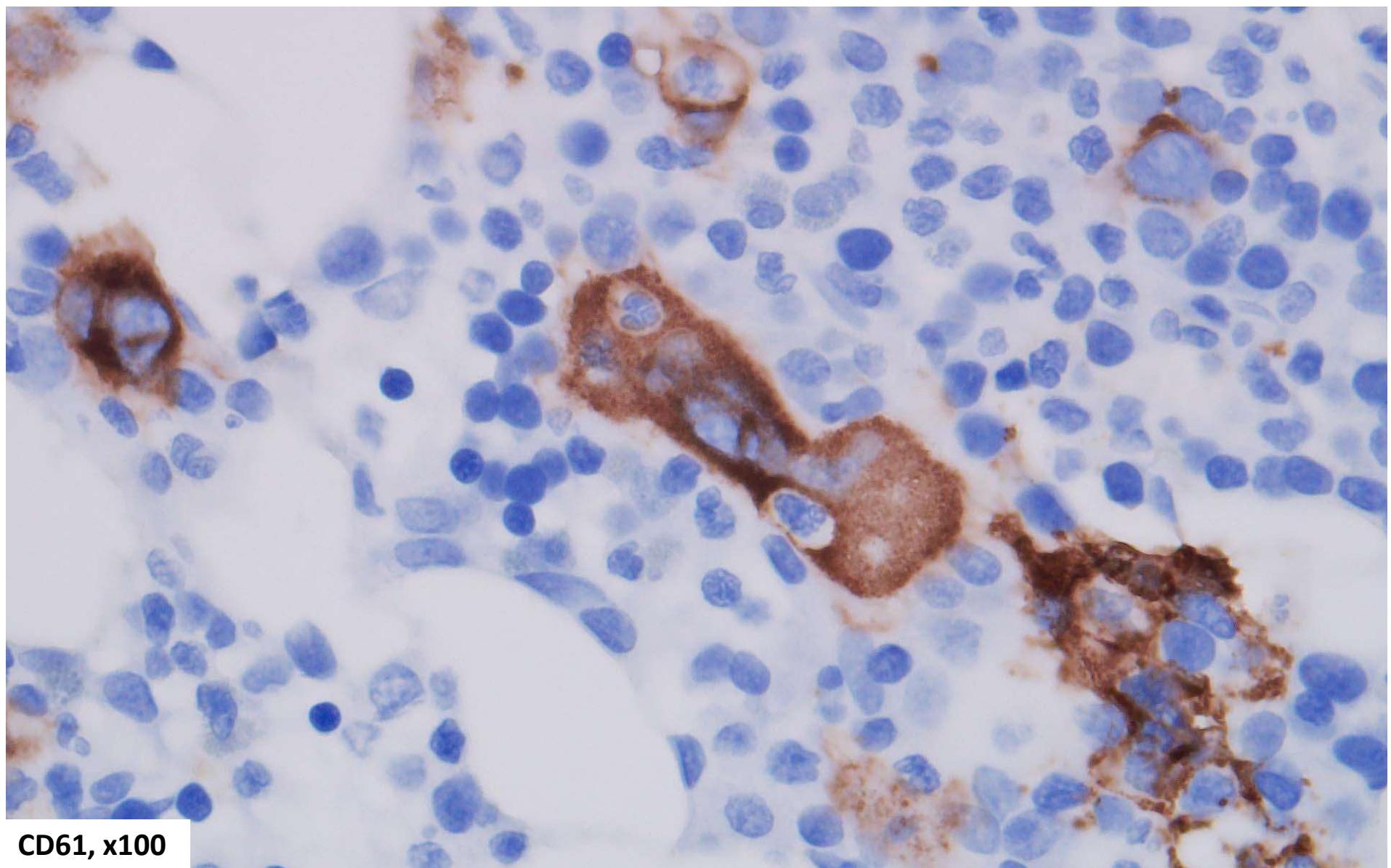
H/E, x40



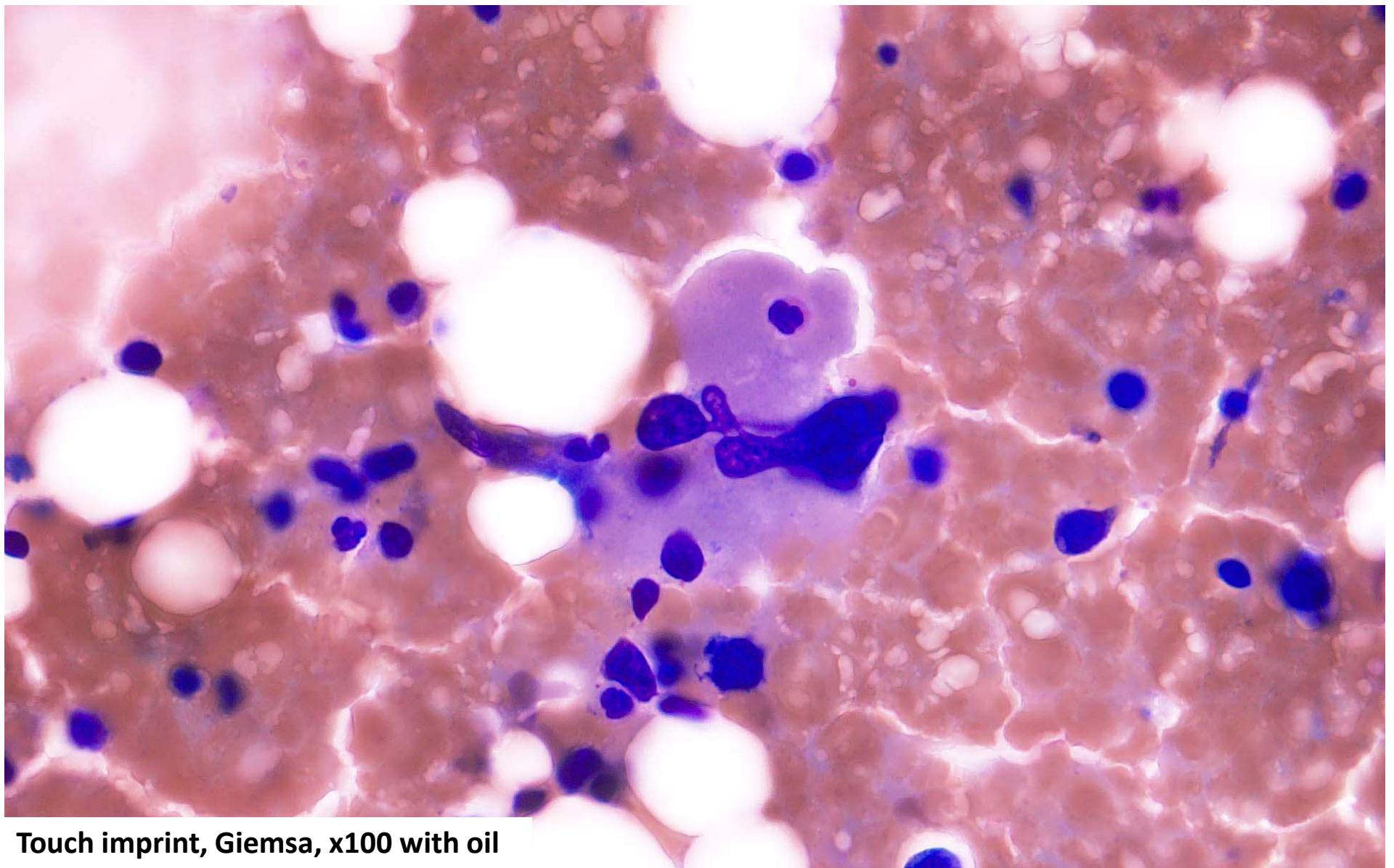
H/E, x40



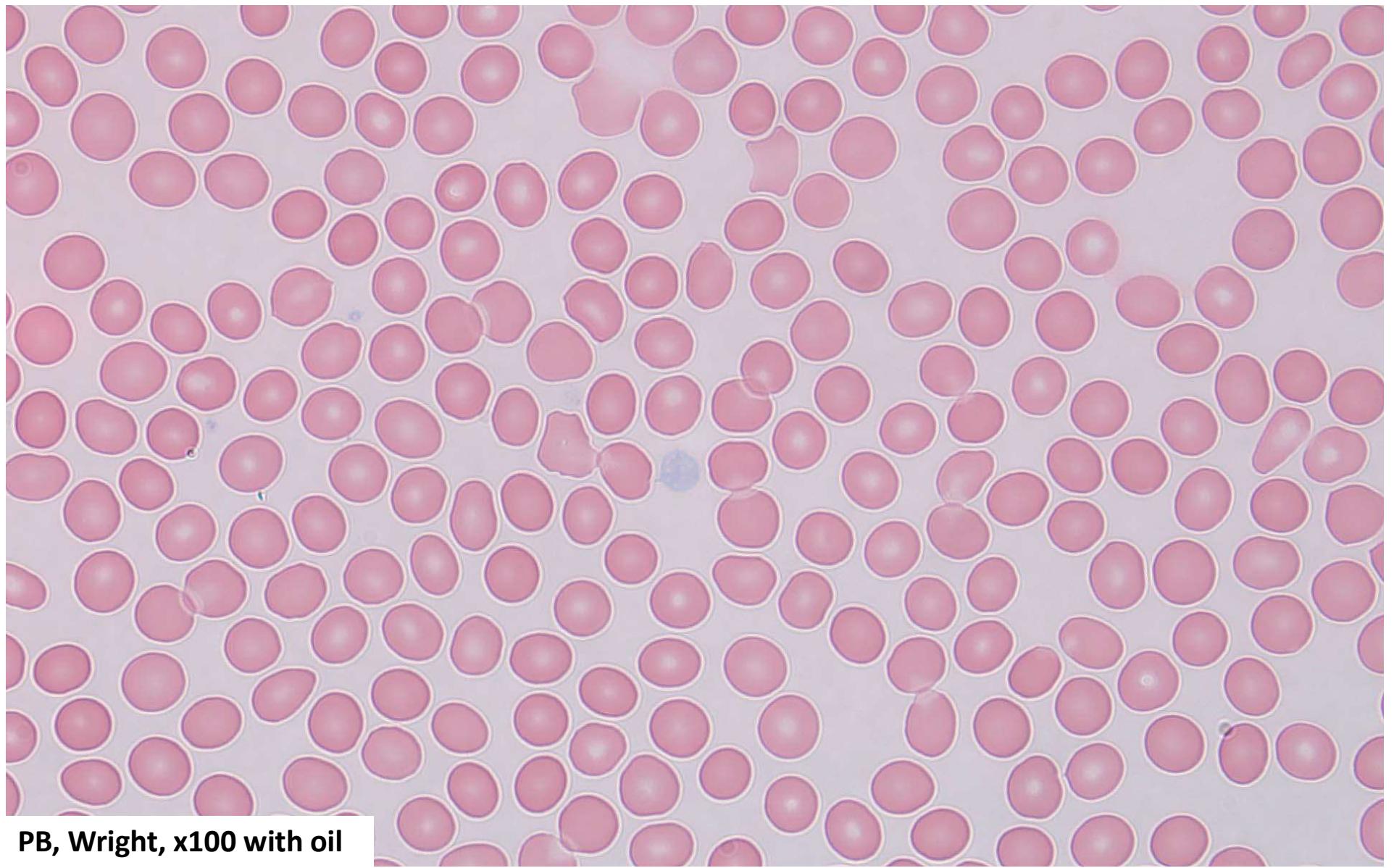
H/E, x100



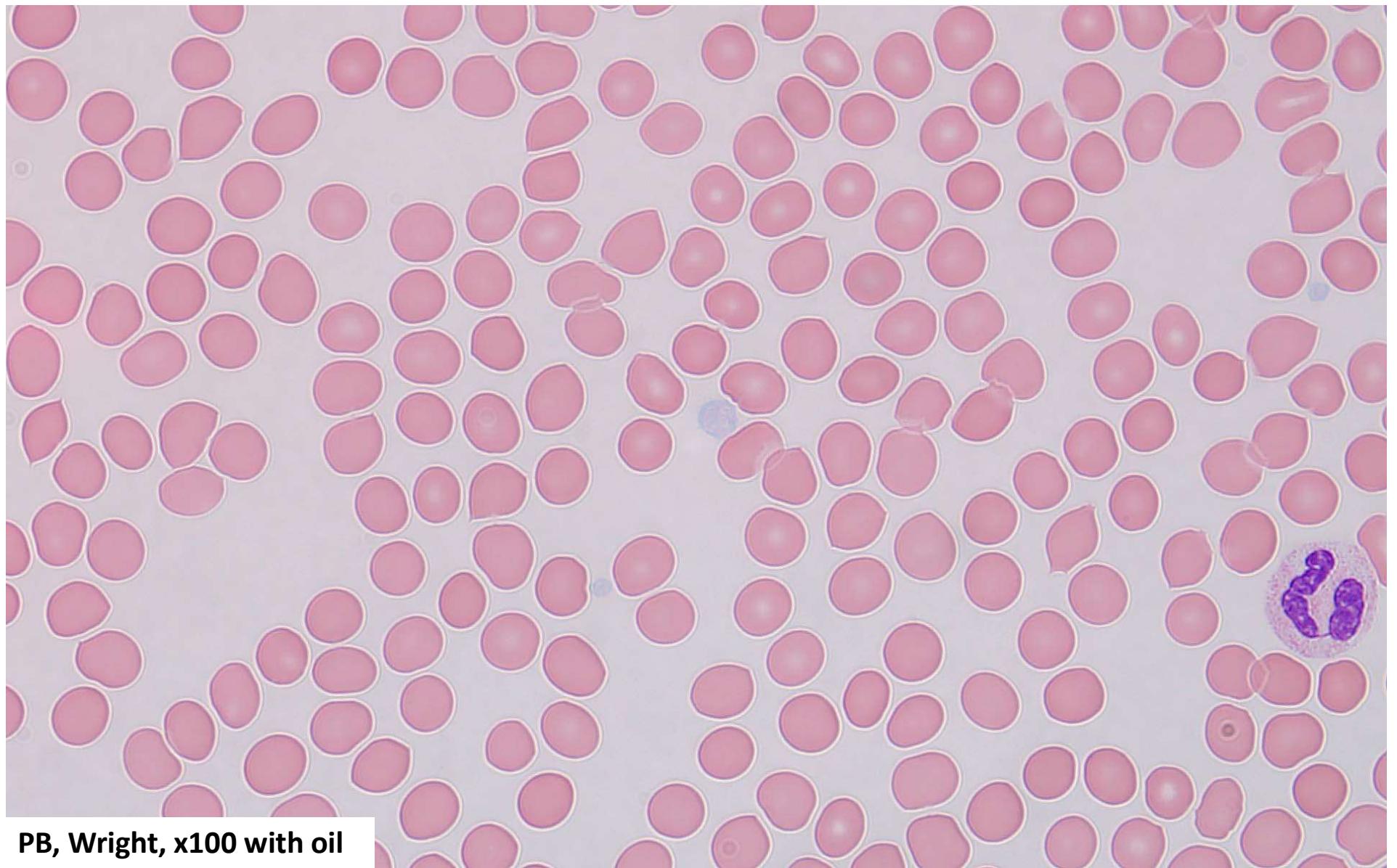
CD61, x100



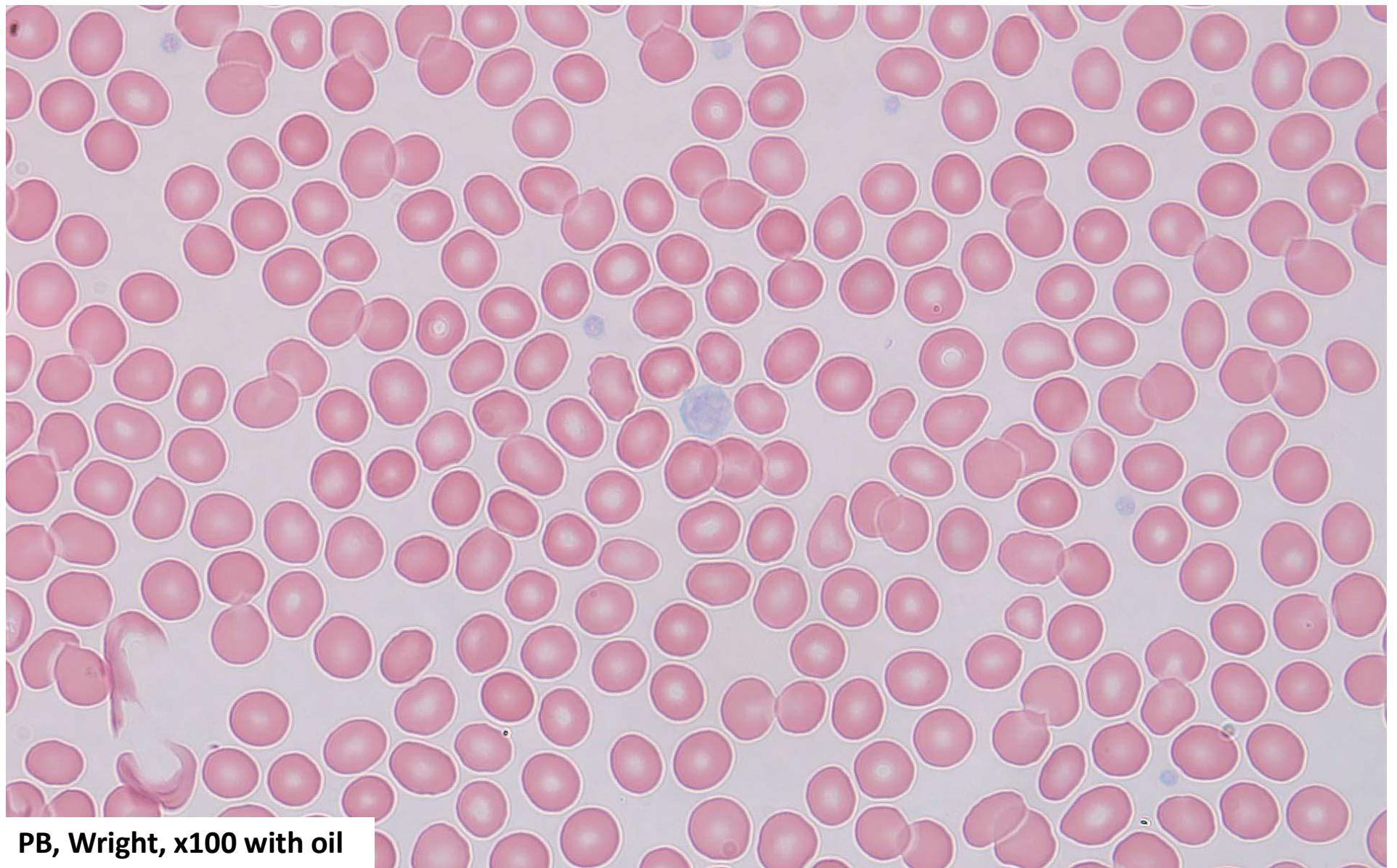
**Touch imprint, Giemsa, x100 with oil**



PB, Wright, x100 with oil



PB, Wright, x100 with oil



PB, Wright, x100 with oil

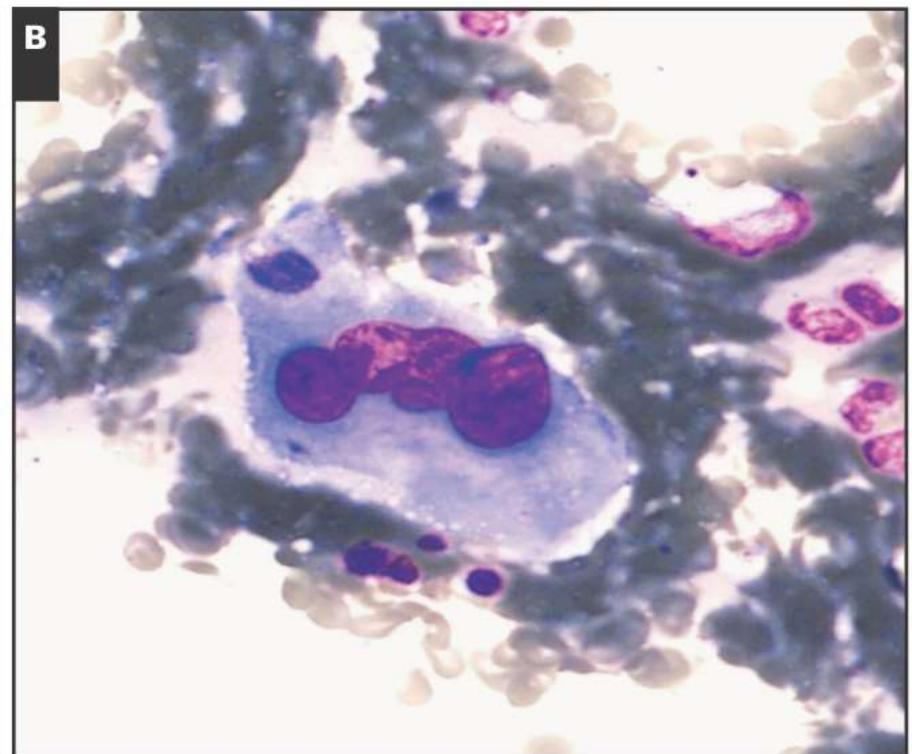
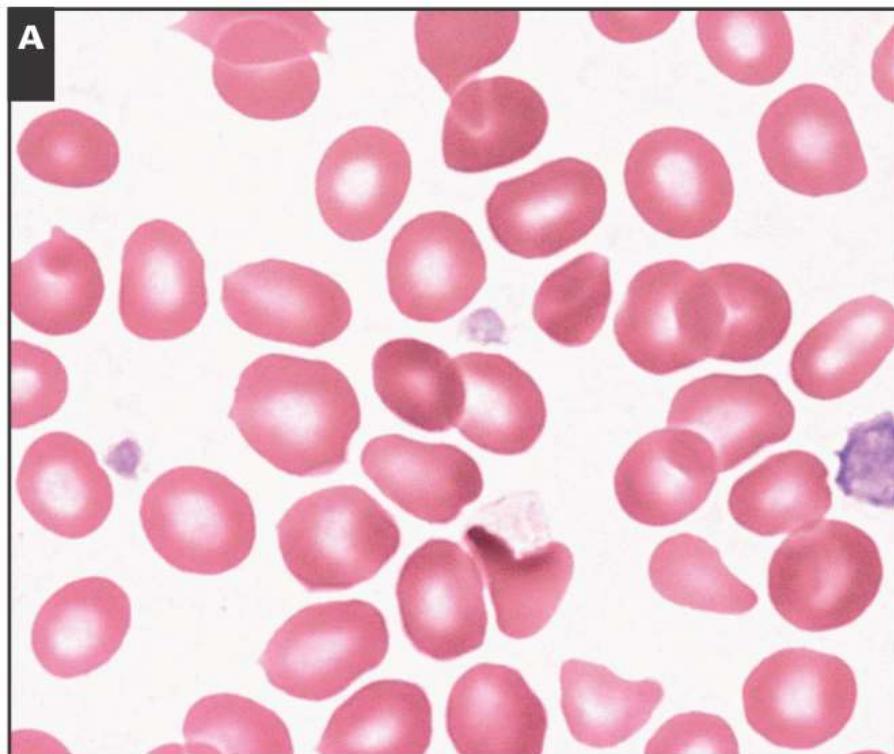
# Differential Diagnosis for macrothrombocytopenia

- Bernard-Soulier syndrome
- Gray platelet syndrome
- May-Hegglin anomaly

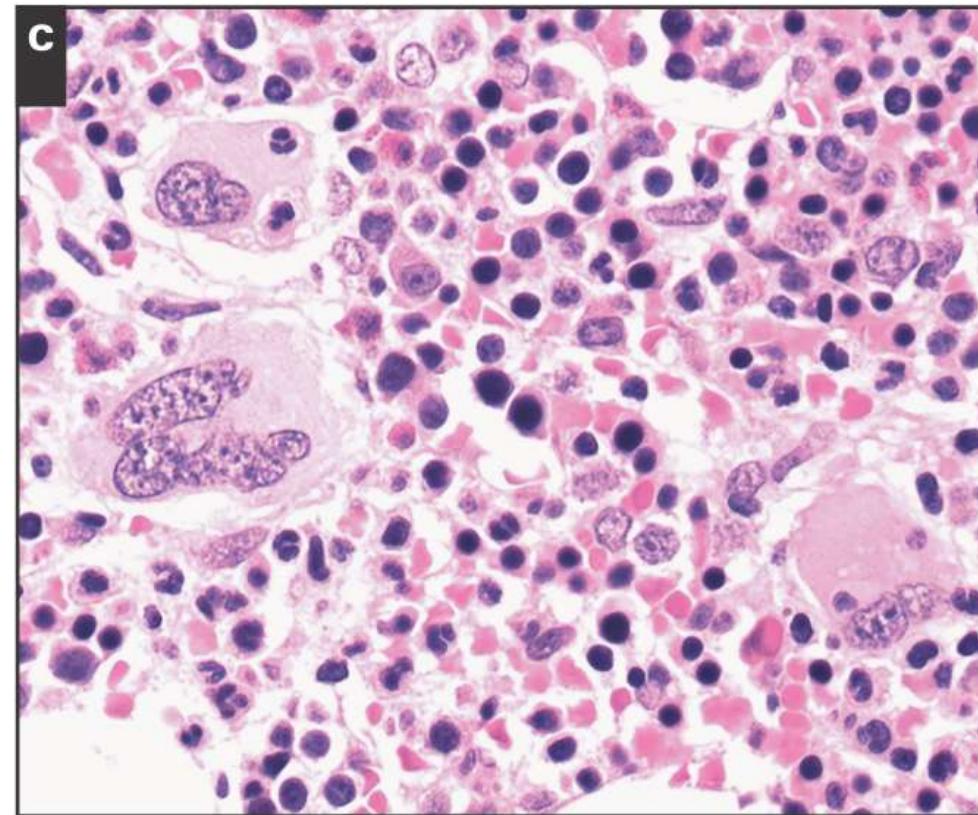
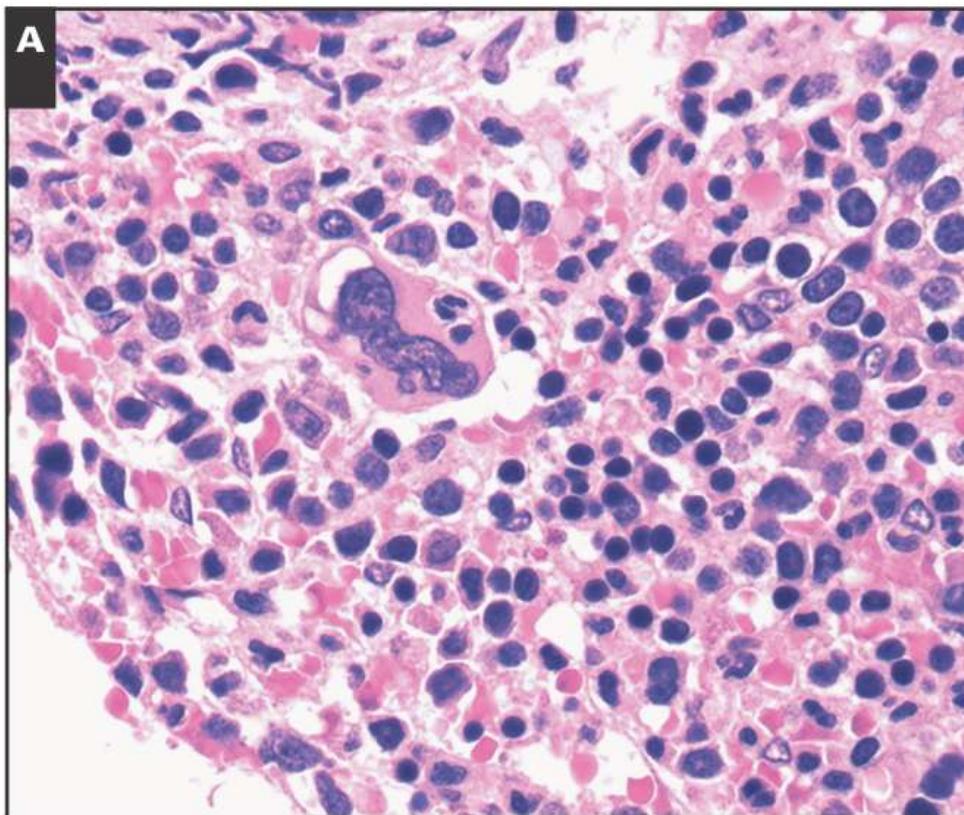
# Gray platelet syndrome (OMIM #139090)

- Autosomal recessive disorder
- Moderate macrothrombocytopenia
- Marked decrease or absence of platelet alpha-granules
- Mild to moderate bleeding
- Splenomegaly
- Bone marrow fibrosis
- Elevated vitamin B12
- *NBEAL2* mutations

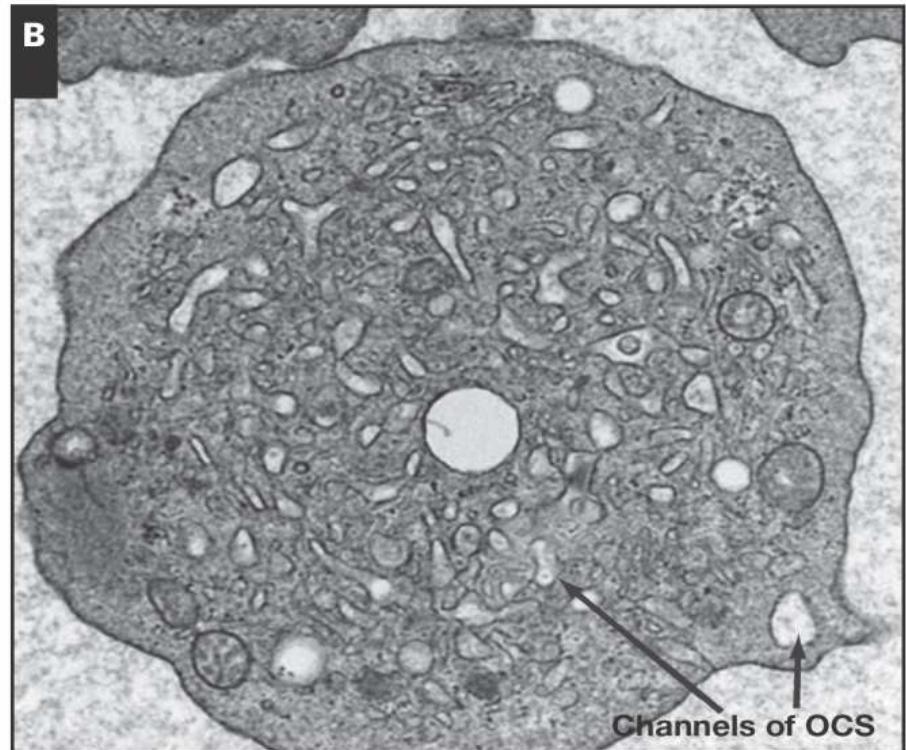
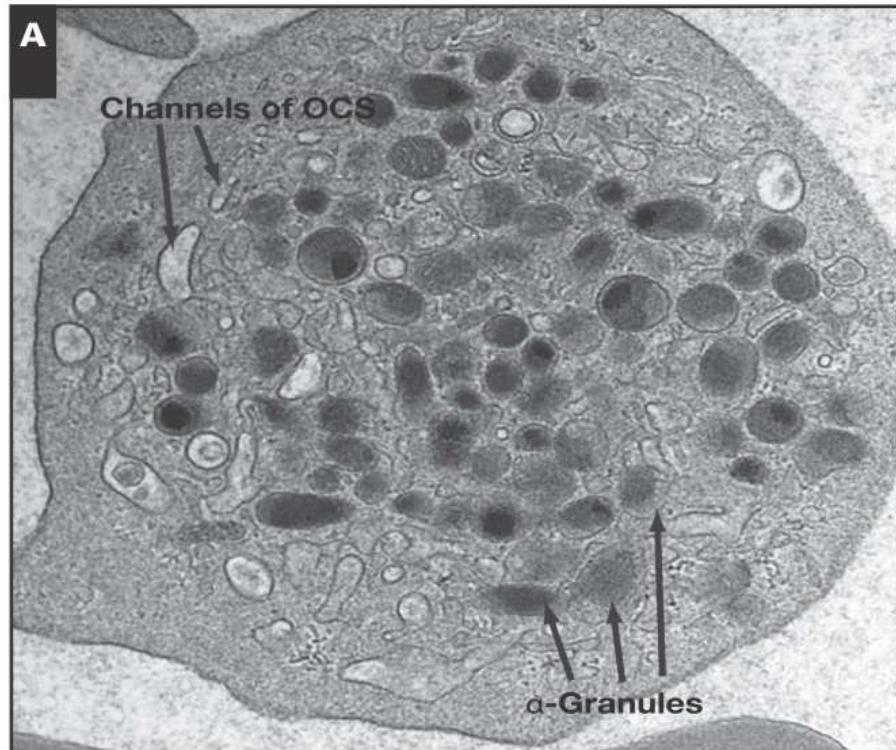
# Gray platelet syndrome



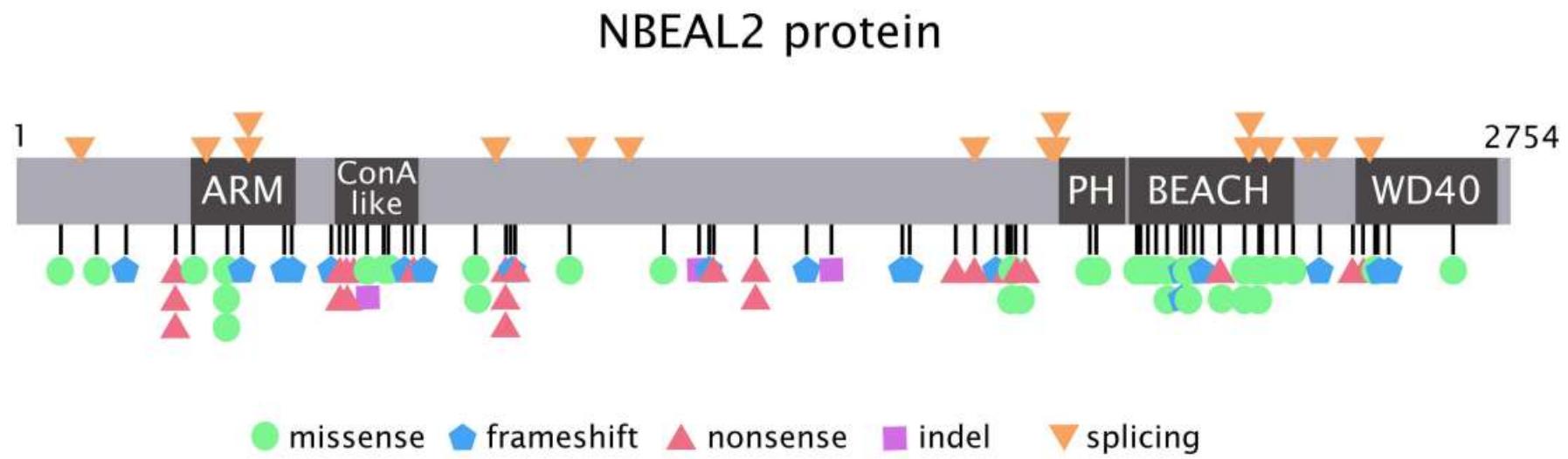
# Gray platelet syndrome



# Gray platelet syndrome



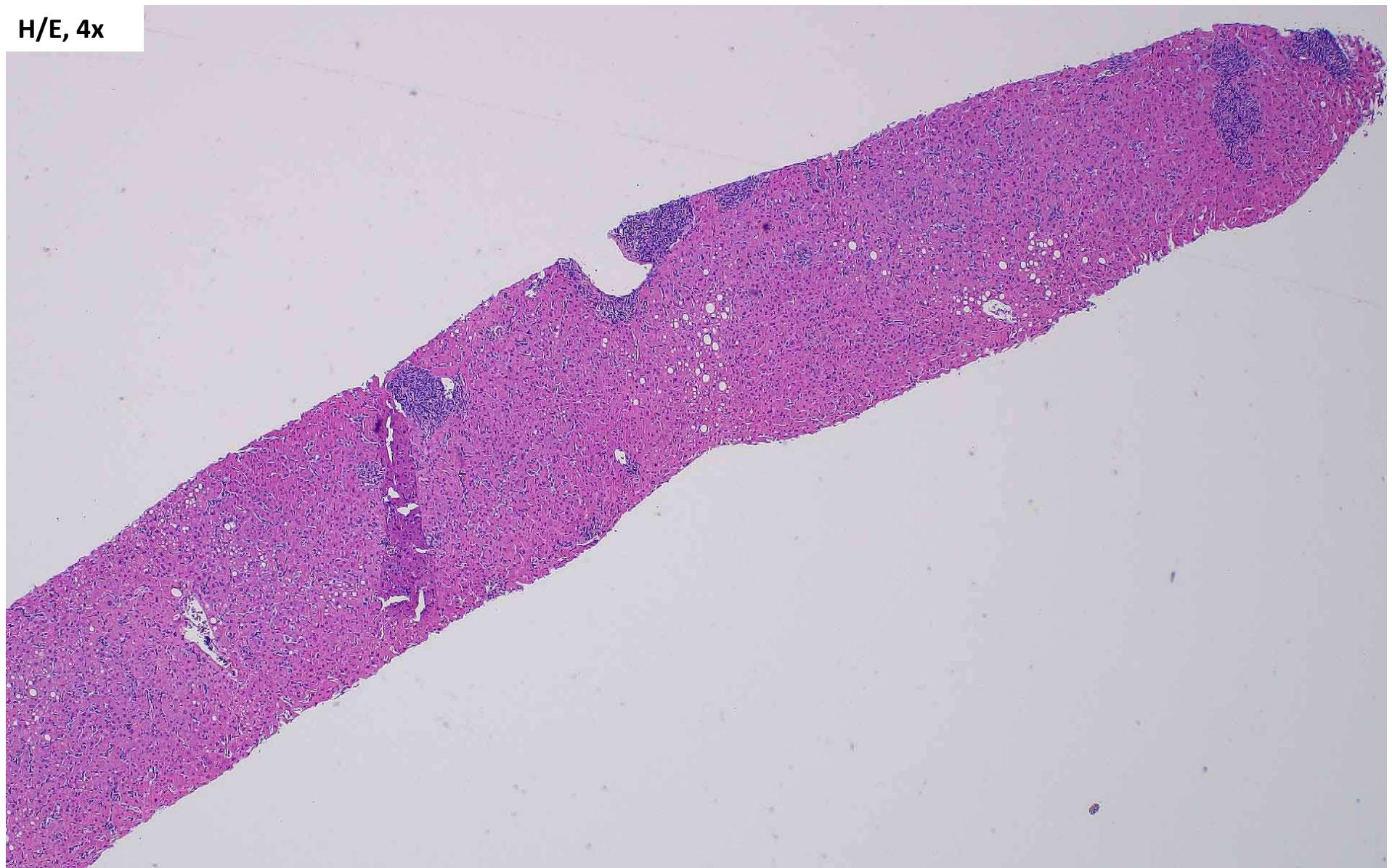
# Gray platelet syndrome



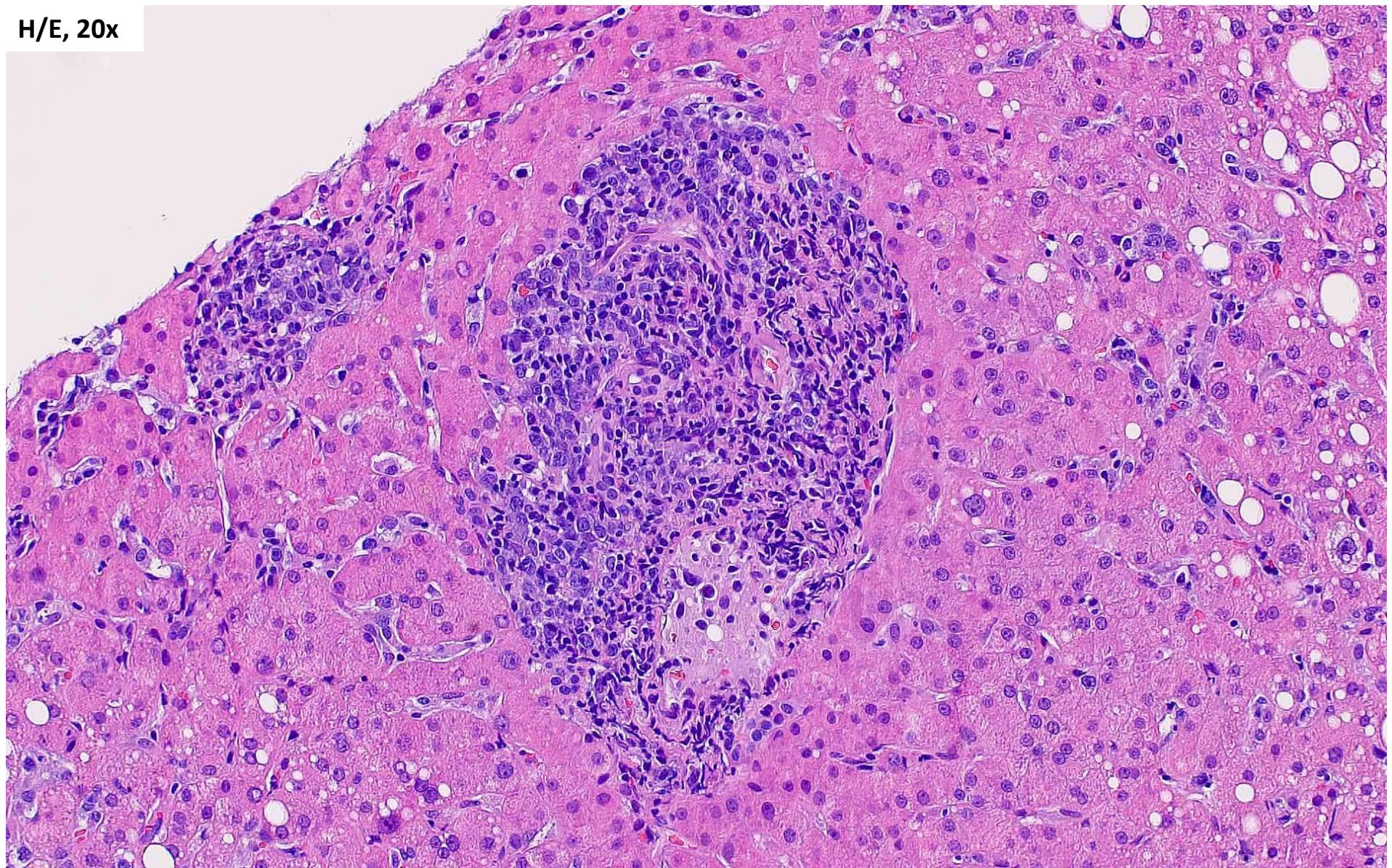
## Case 3

- 60 y/o man
- Drenching night sweat, fever, persistent N/V for 1 month
- CT scan
  - Hepatomegaly (23 cm) without mass or biliary ductal dilation
  - Splenomegaly (23%) with multiple infarcts
  - Adenopathy in the mediastinum, right hilum, retroperitoneum, porta hepatis
- PET-CT
  - Diffuse adenopathy
  - Splenomegaly
  - Small-to-moderate left pleural effusion
- Liver biopsy and bone marrow biopsy

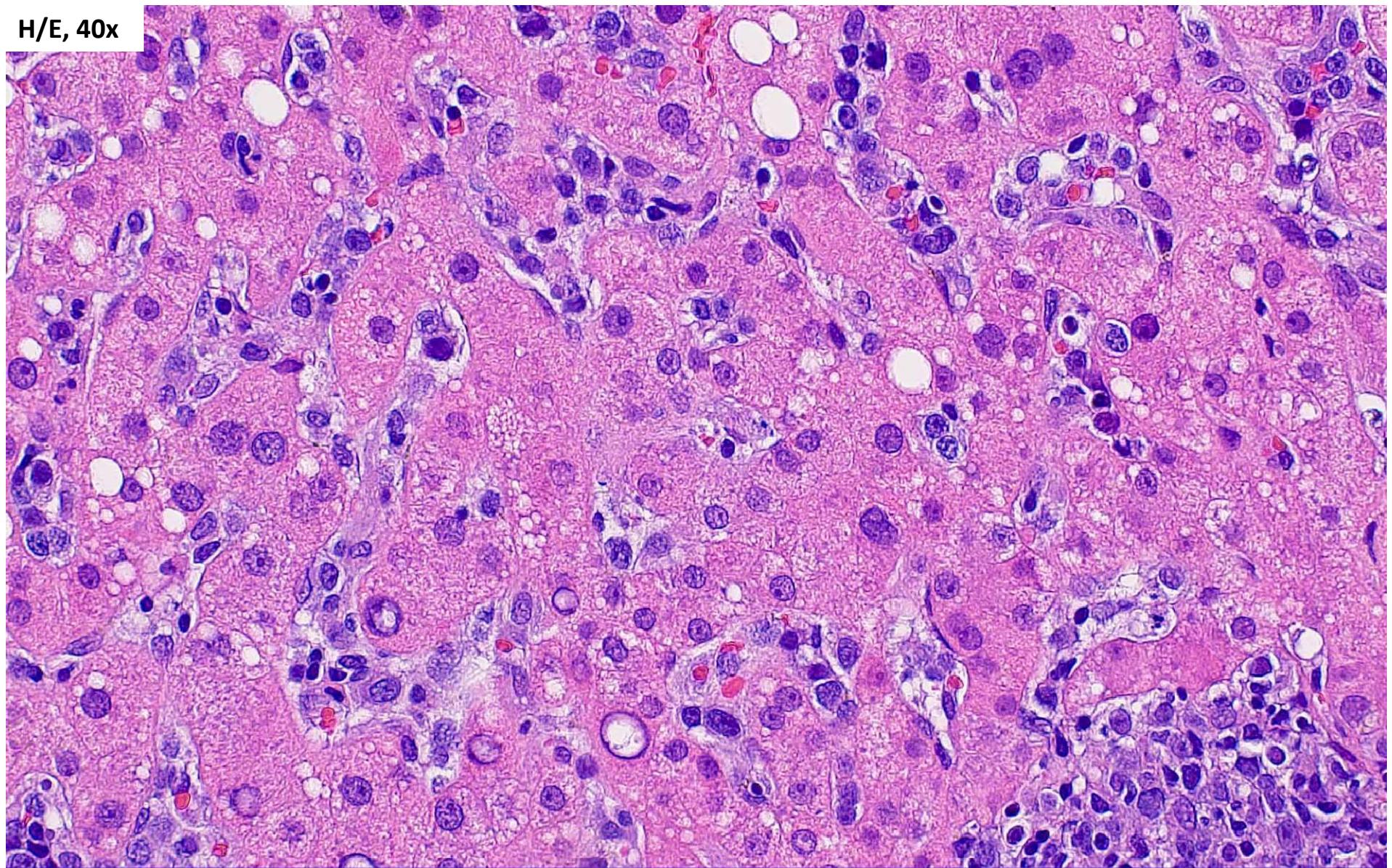
H/E, 4x



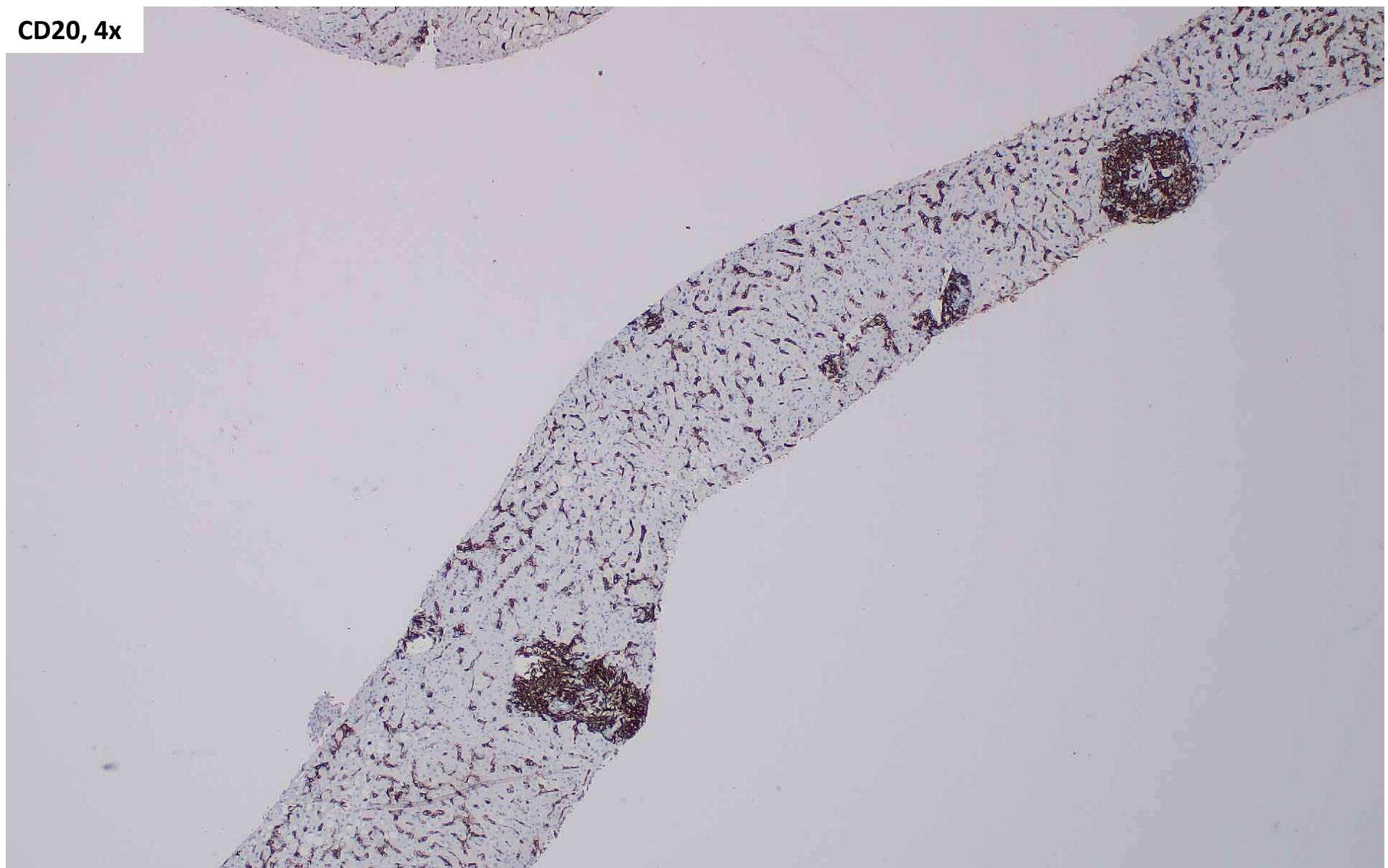
H/E, 20x



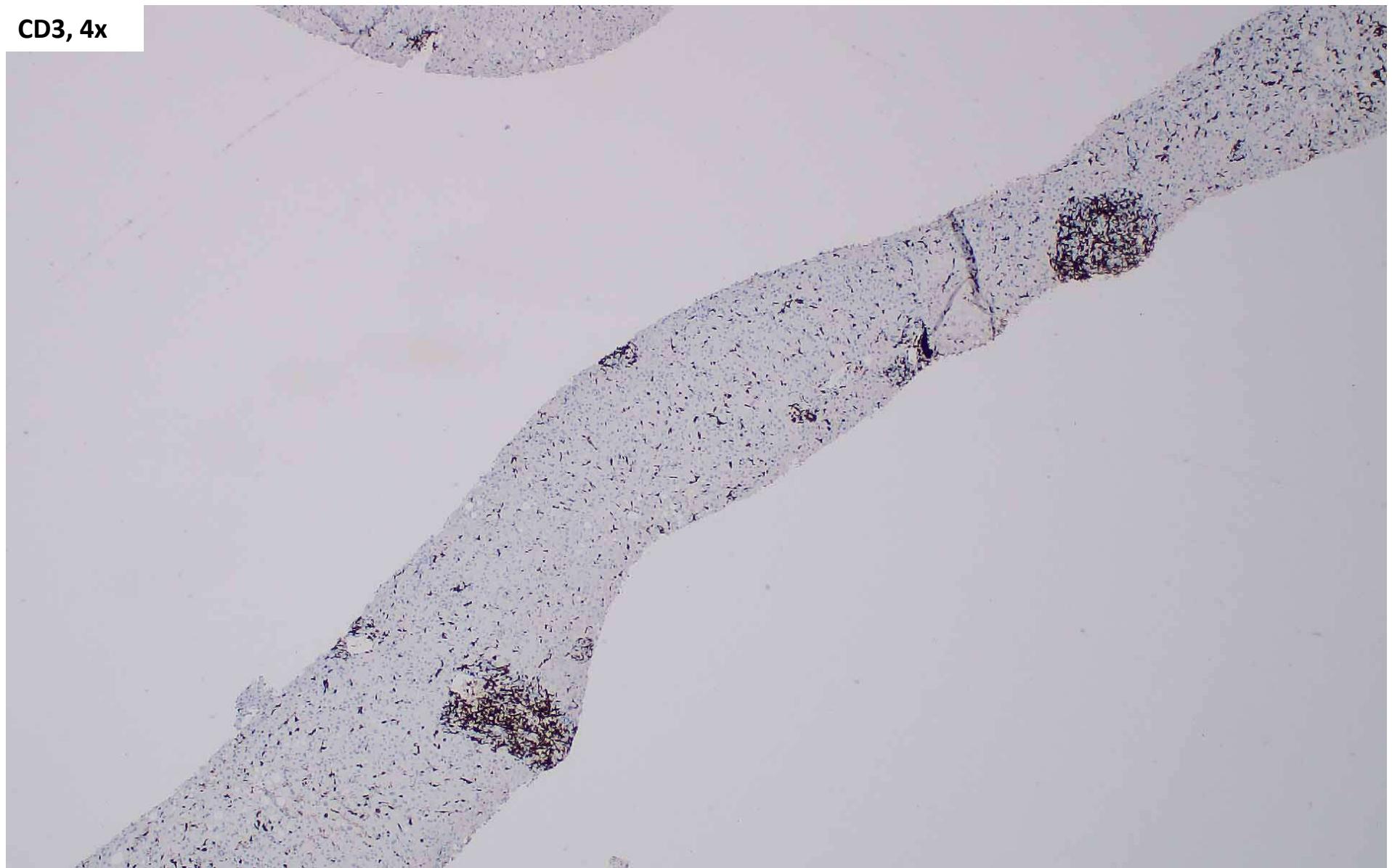
H/E, 40x



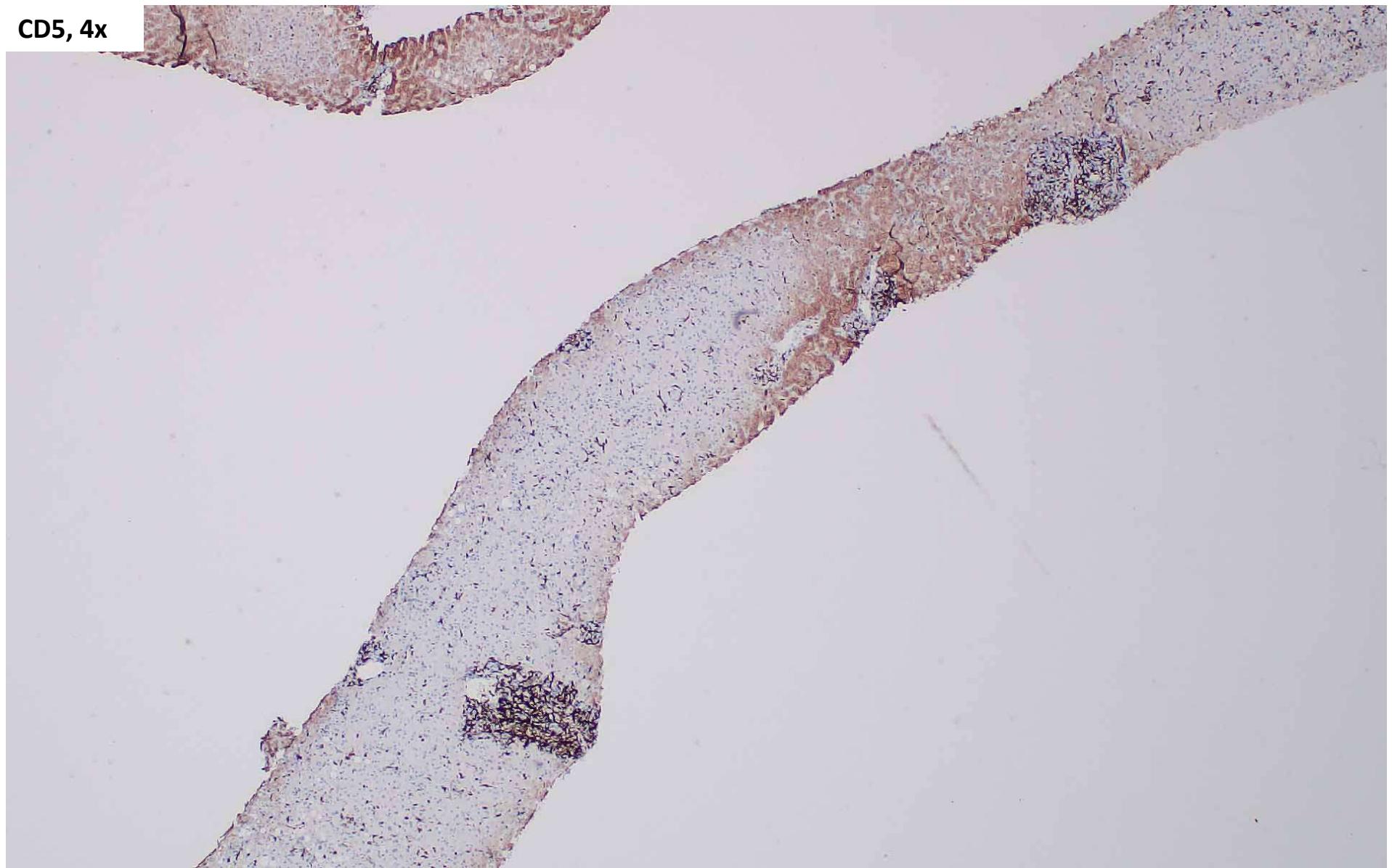
**CD20, 4x**



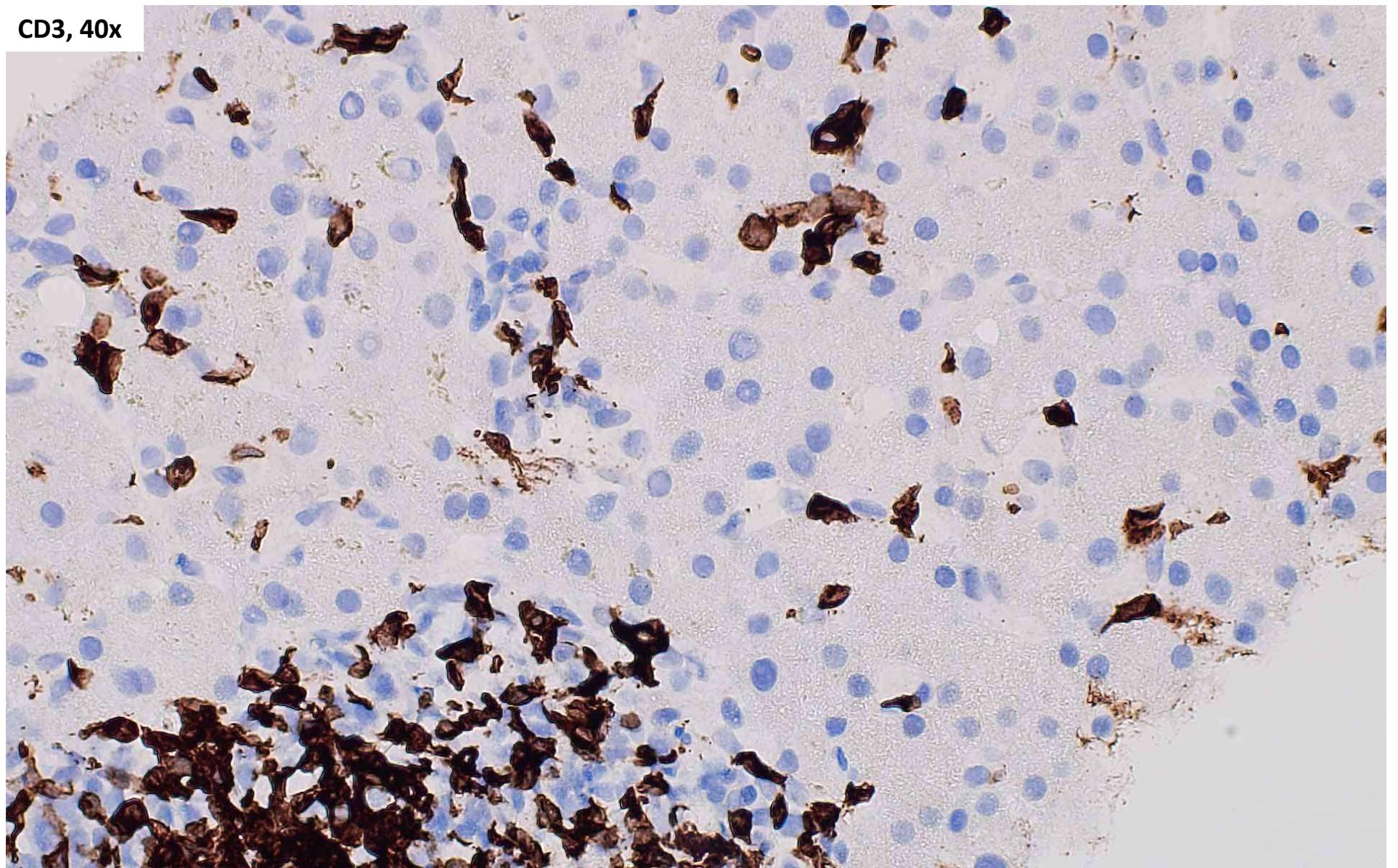
**CD3, 4x**



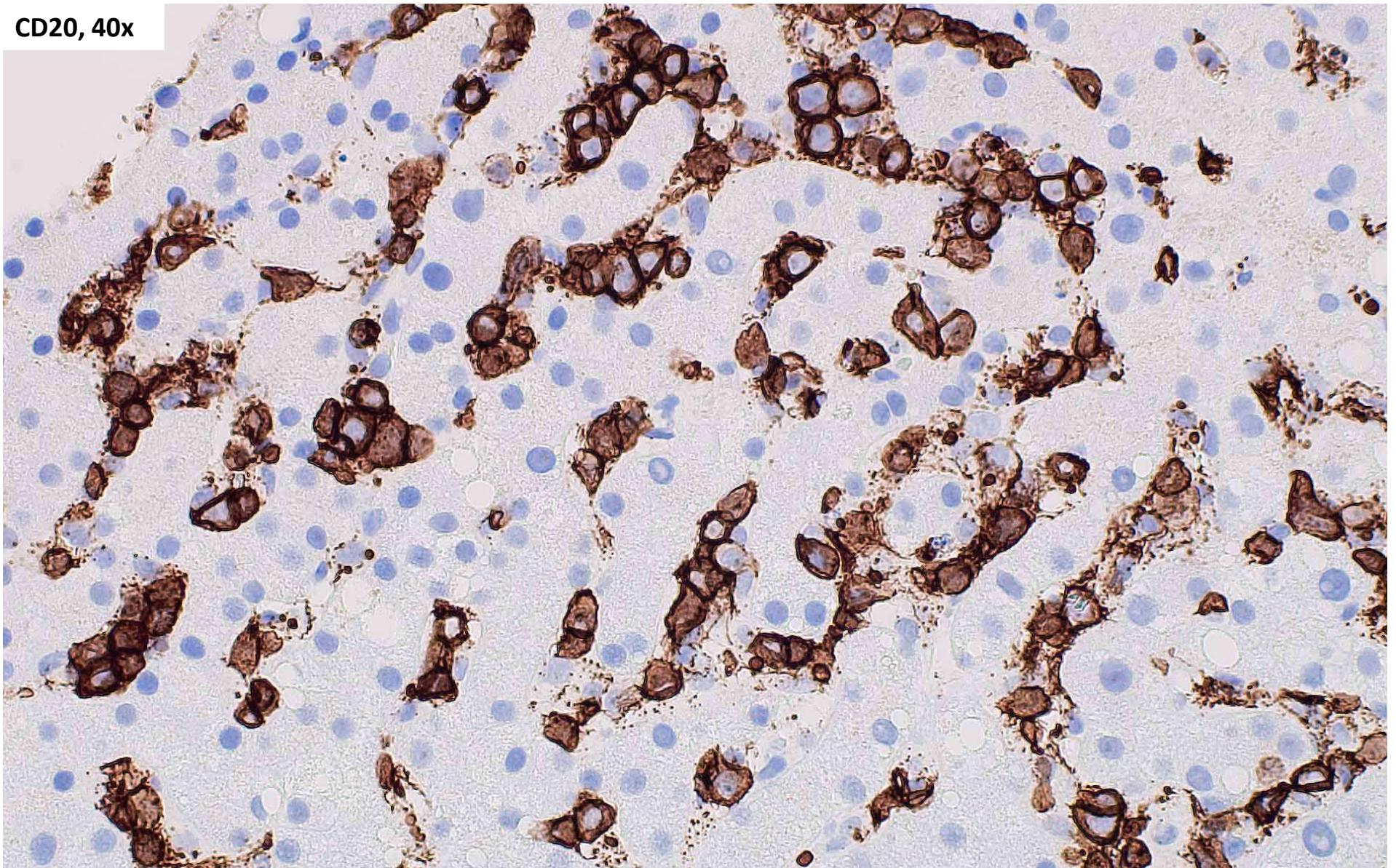
**CD5, 4x**



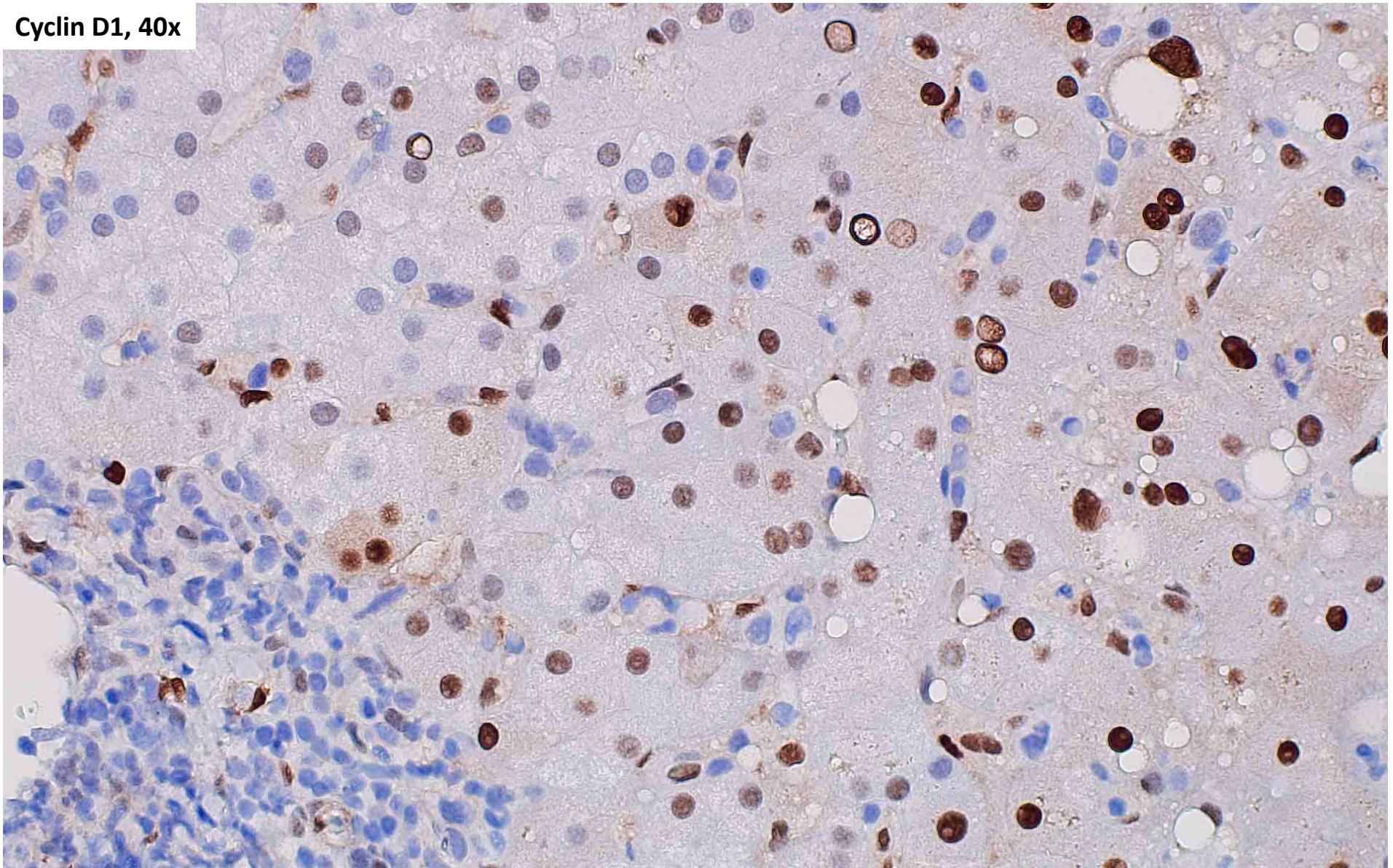
CD3, 40x



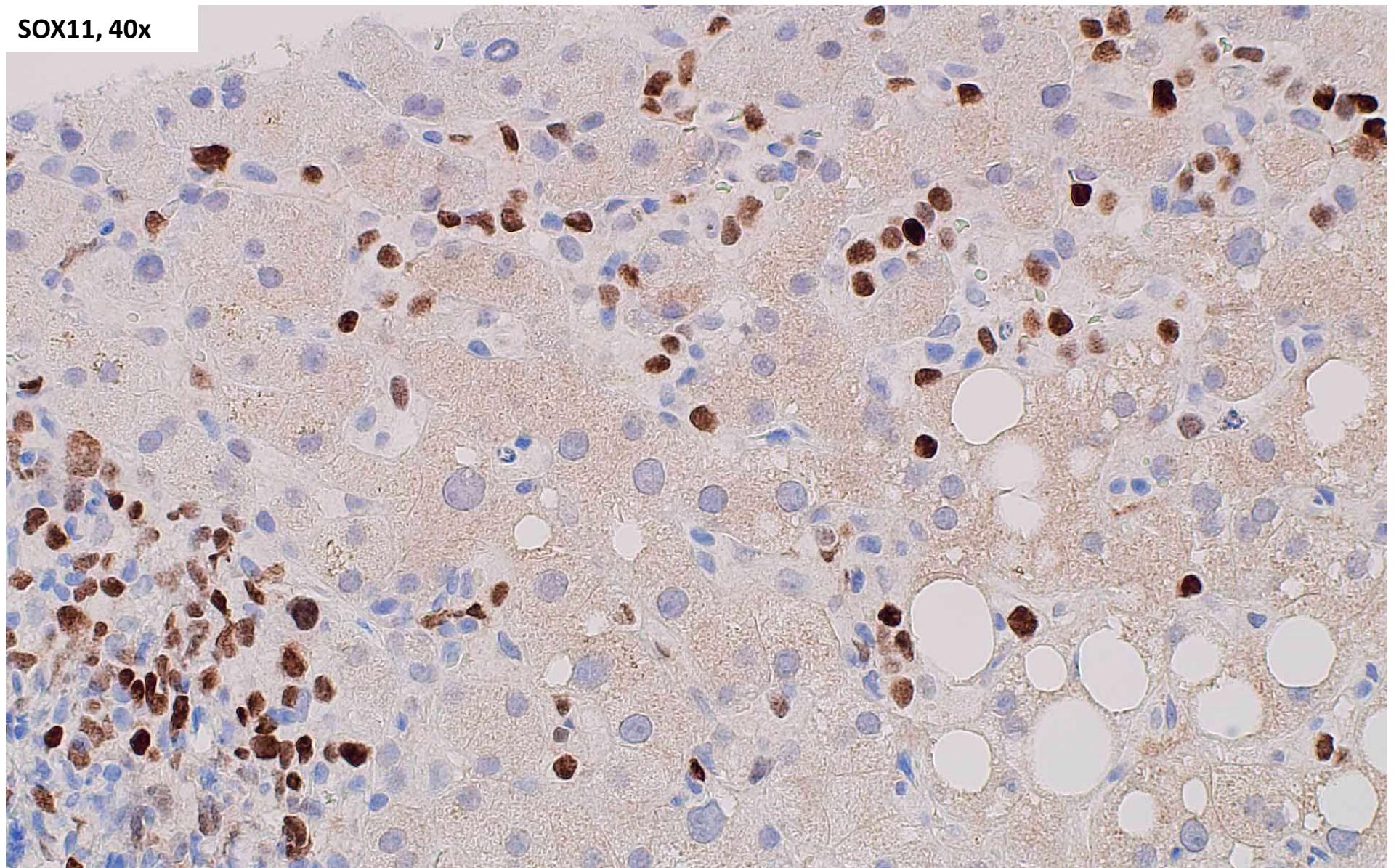
CD20, 40x



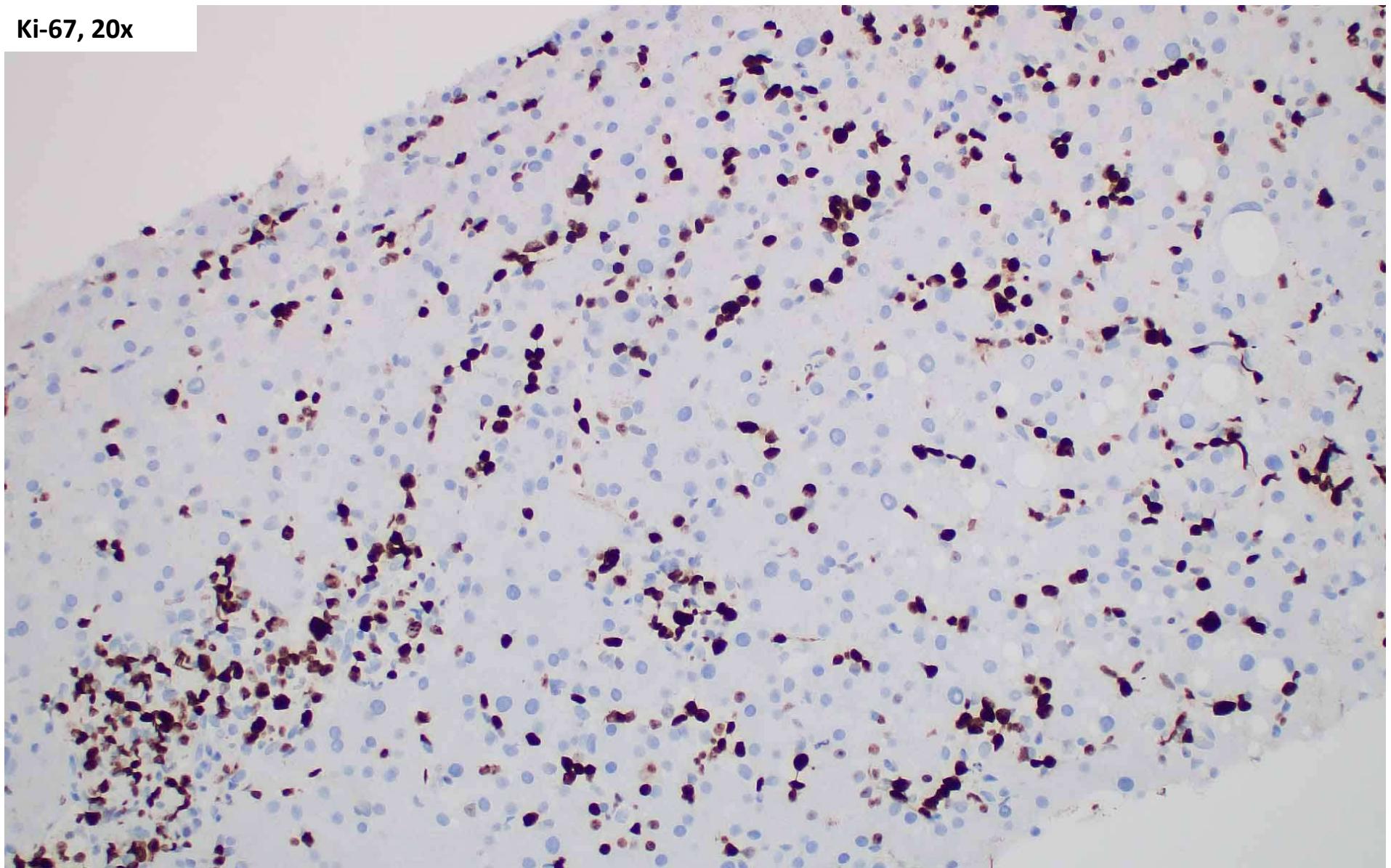
Cyclin D1, 40x



**SOX11, 40x**

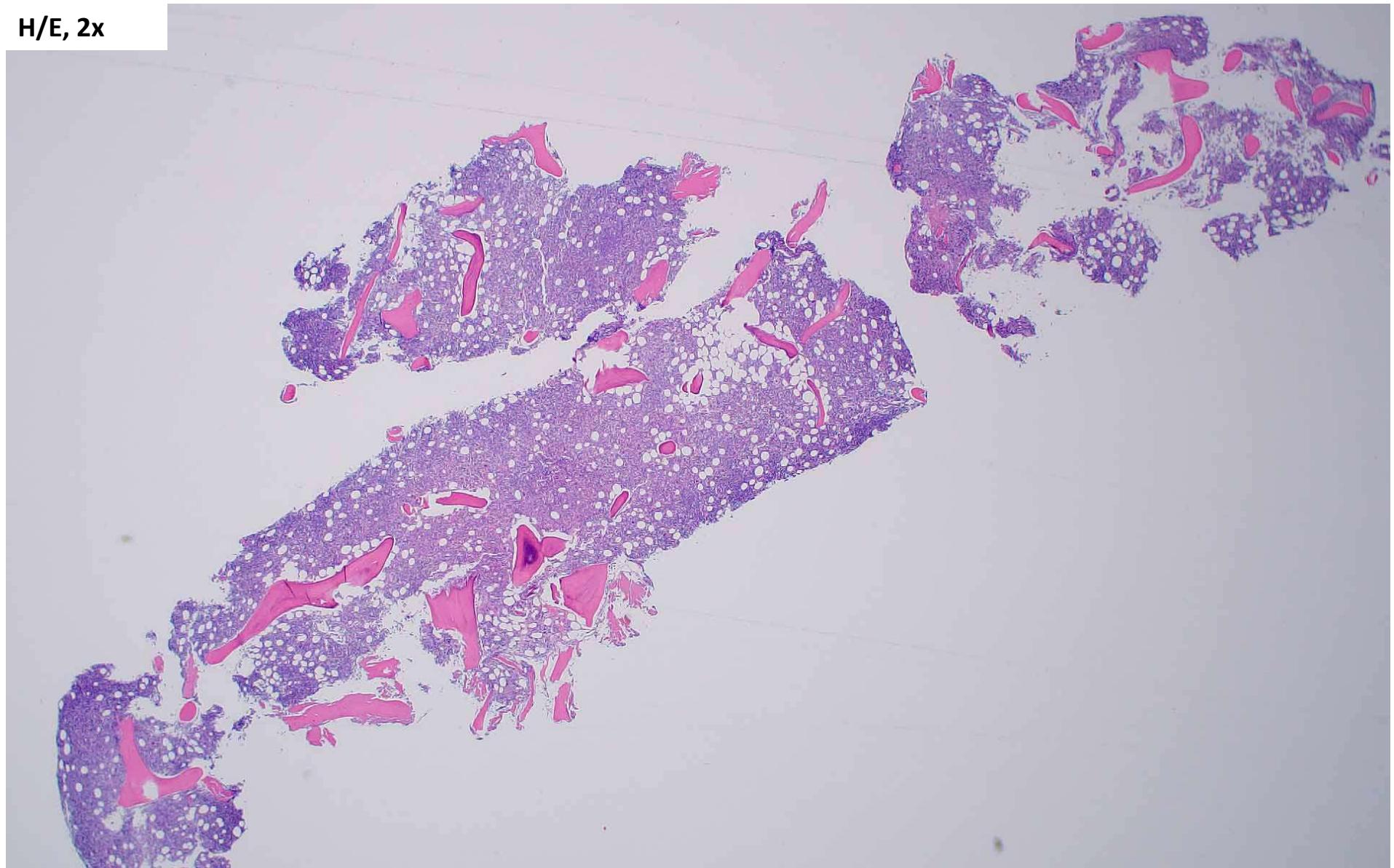


Ki-67, 20x

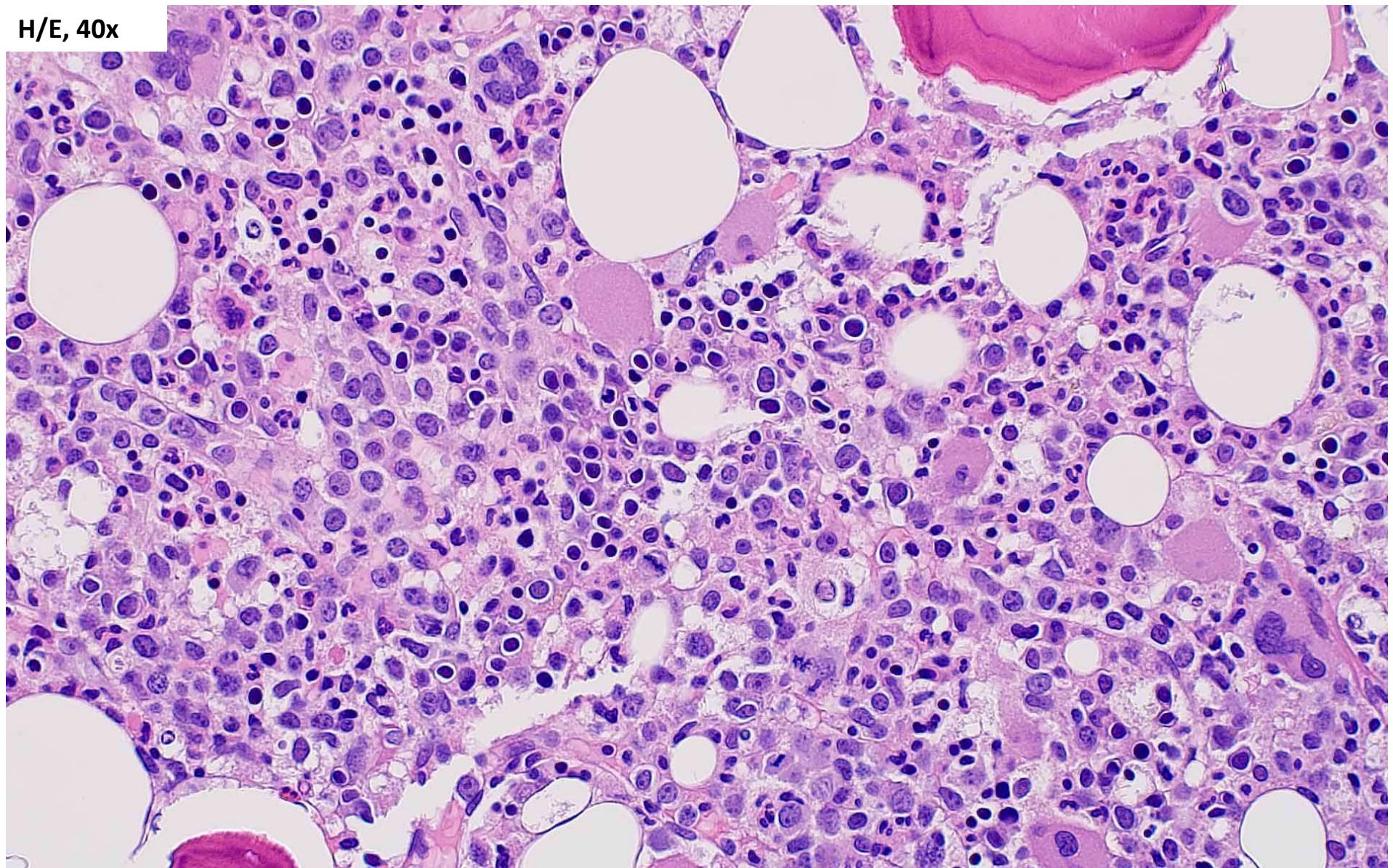


# Bone marrow biopsy

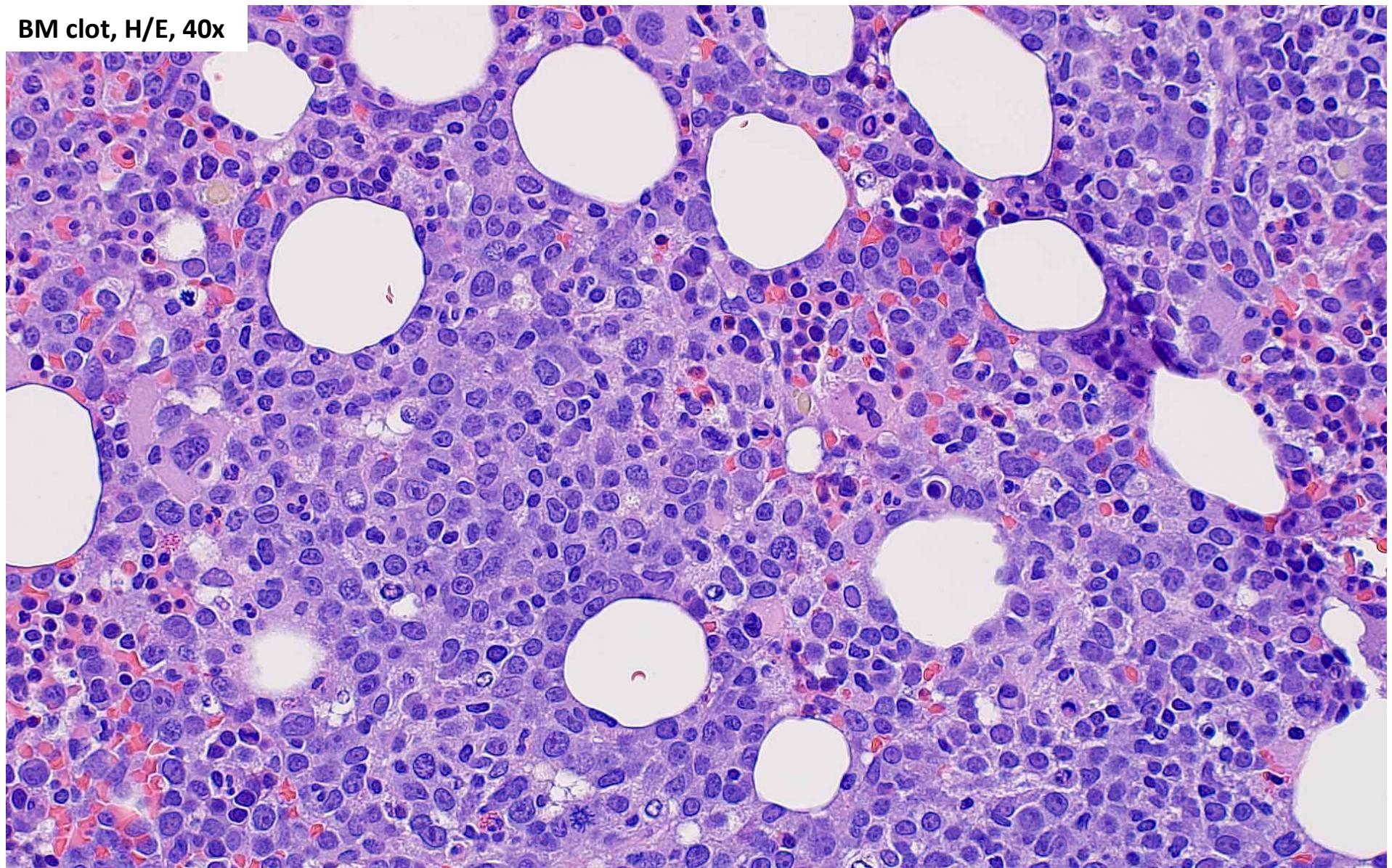
H/E, 2x



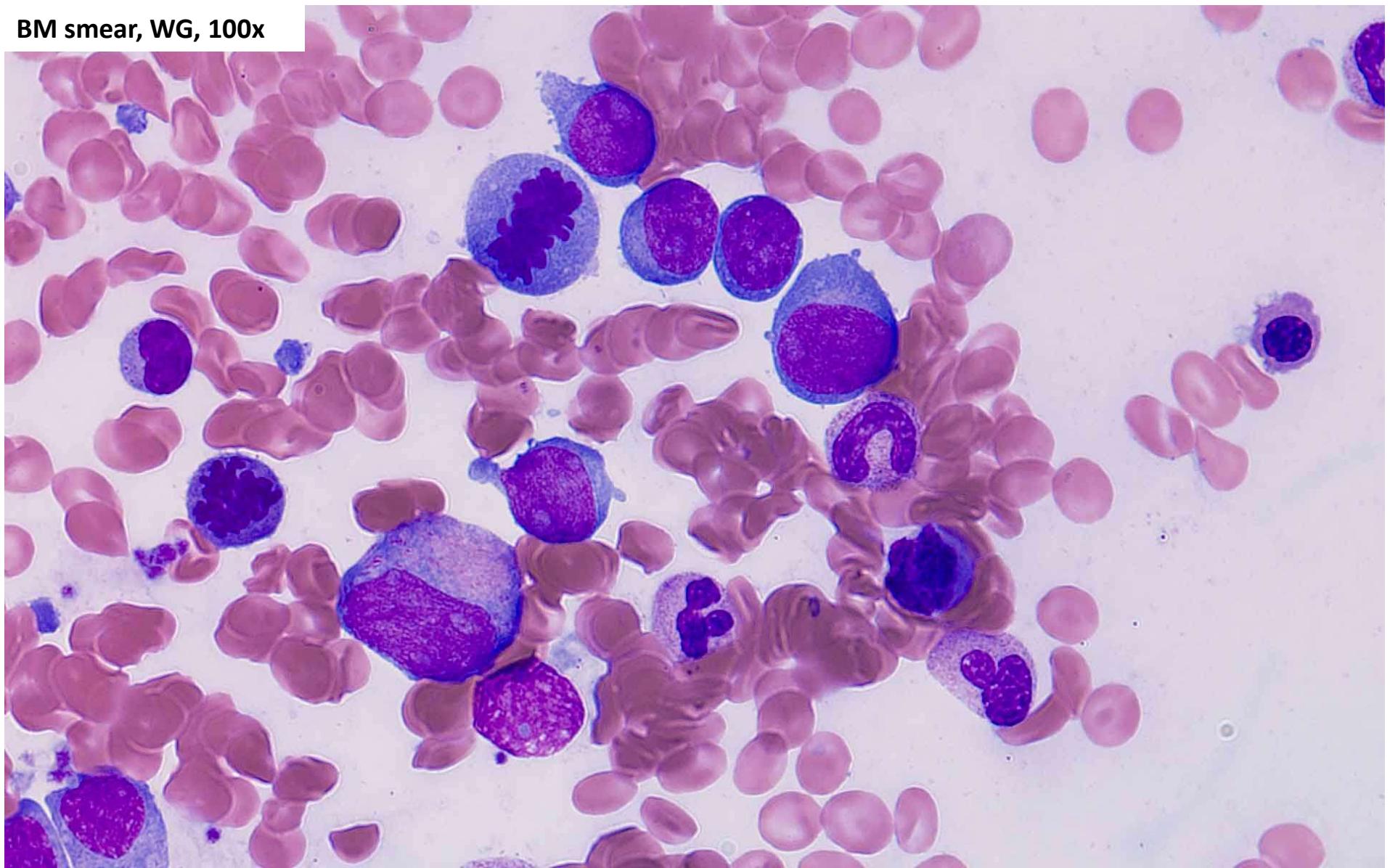
H/E, 40x



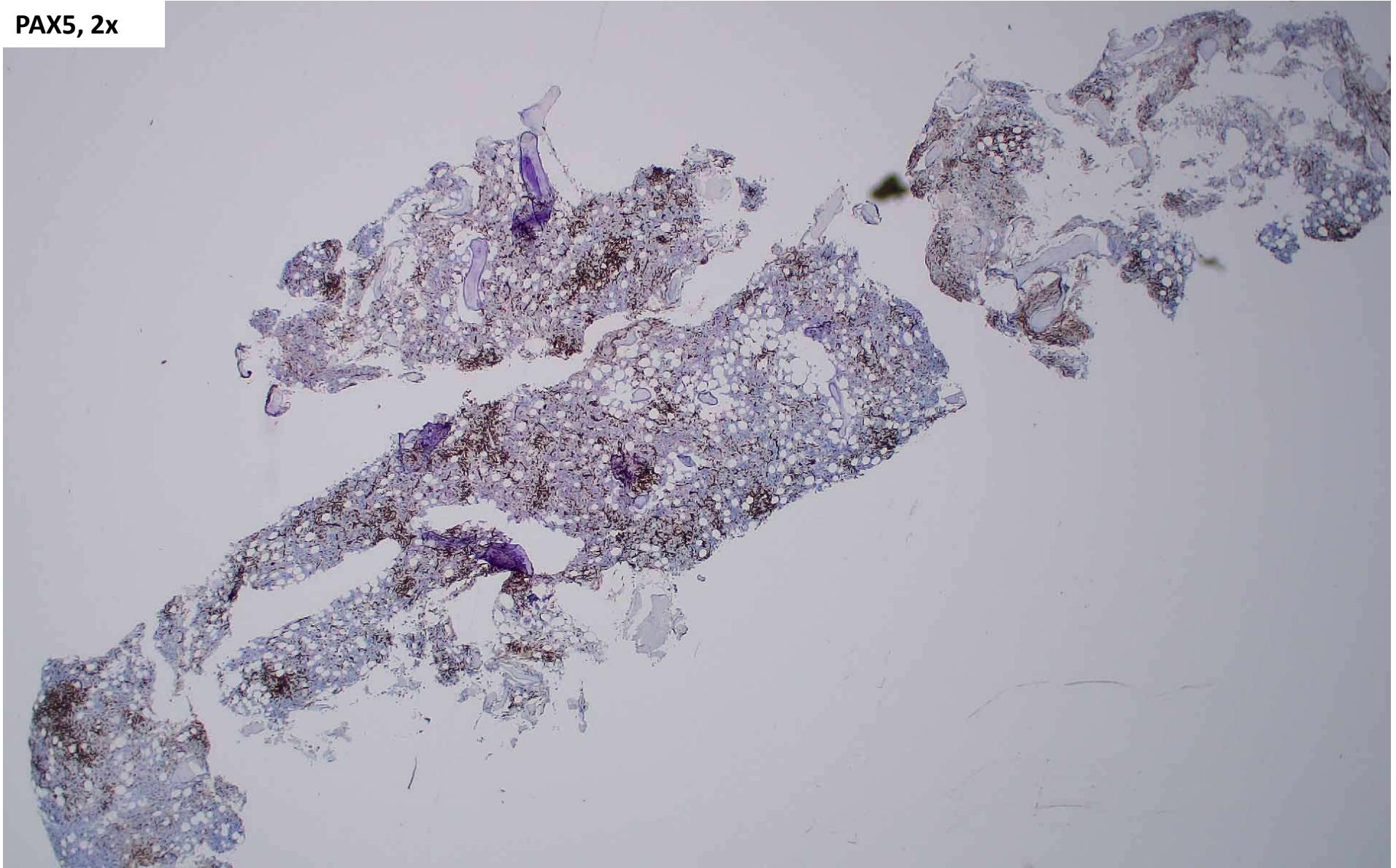
BM clot, H/E, 40x



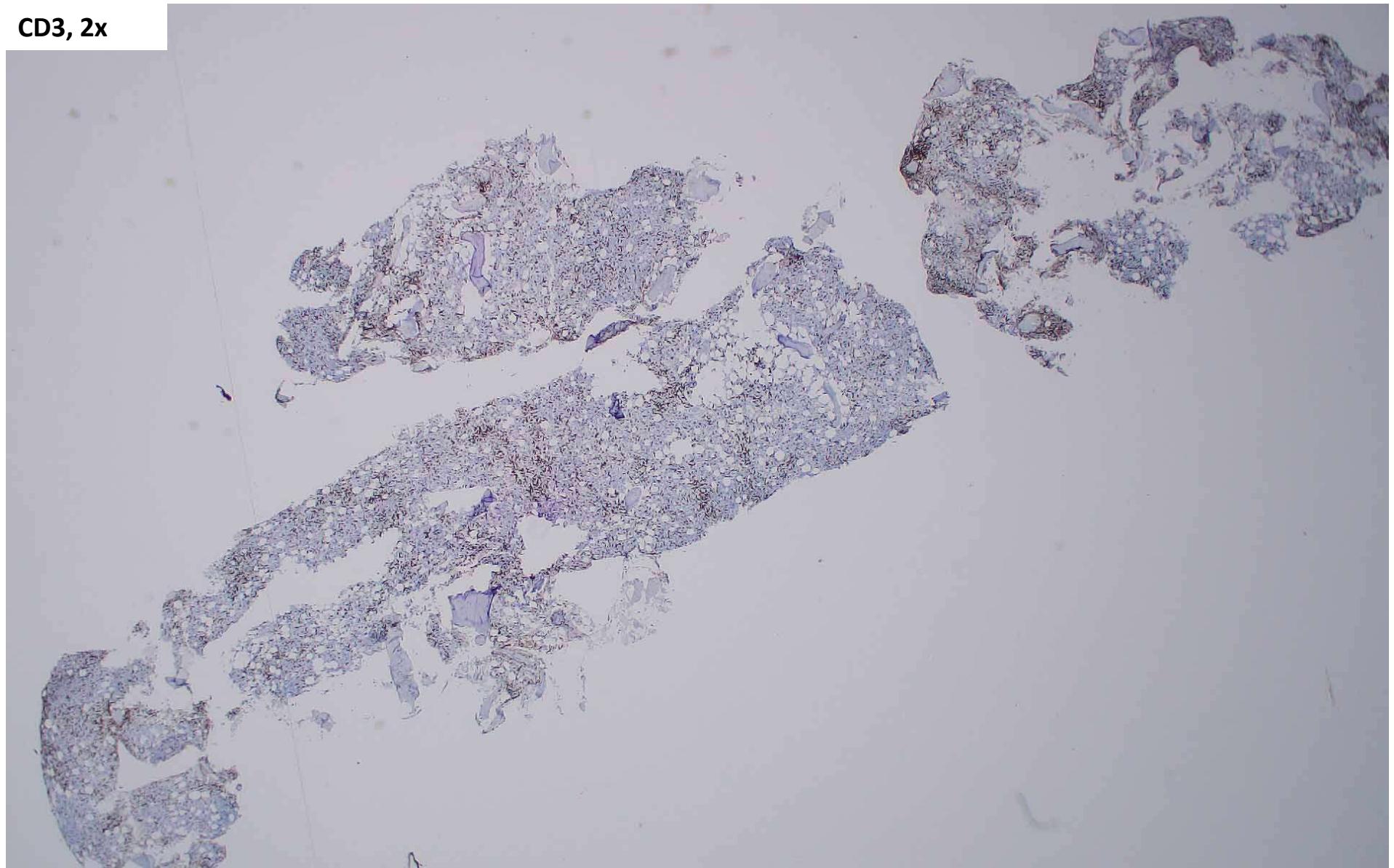
BM smear, WG, 100x



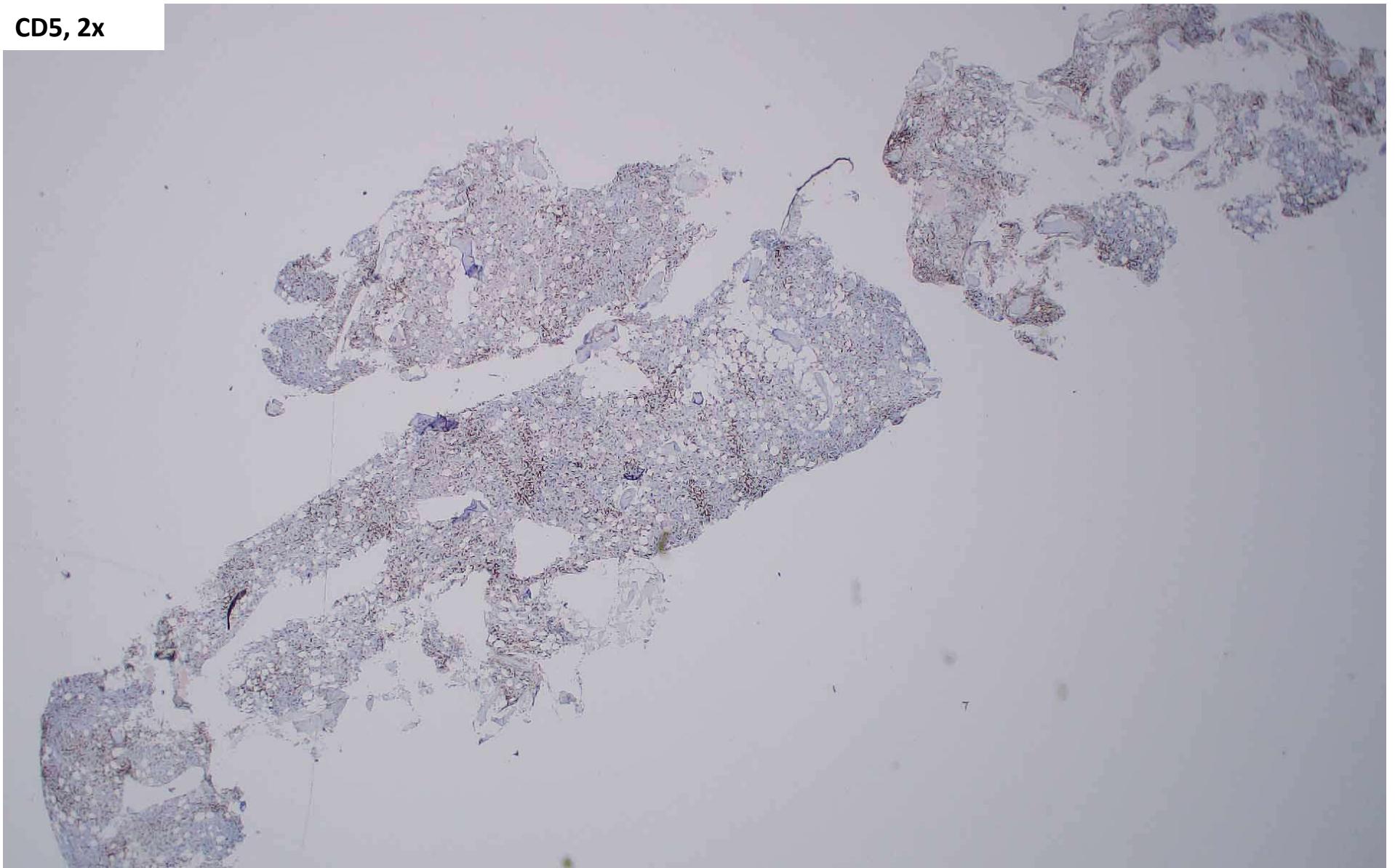
PAX5, 2x



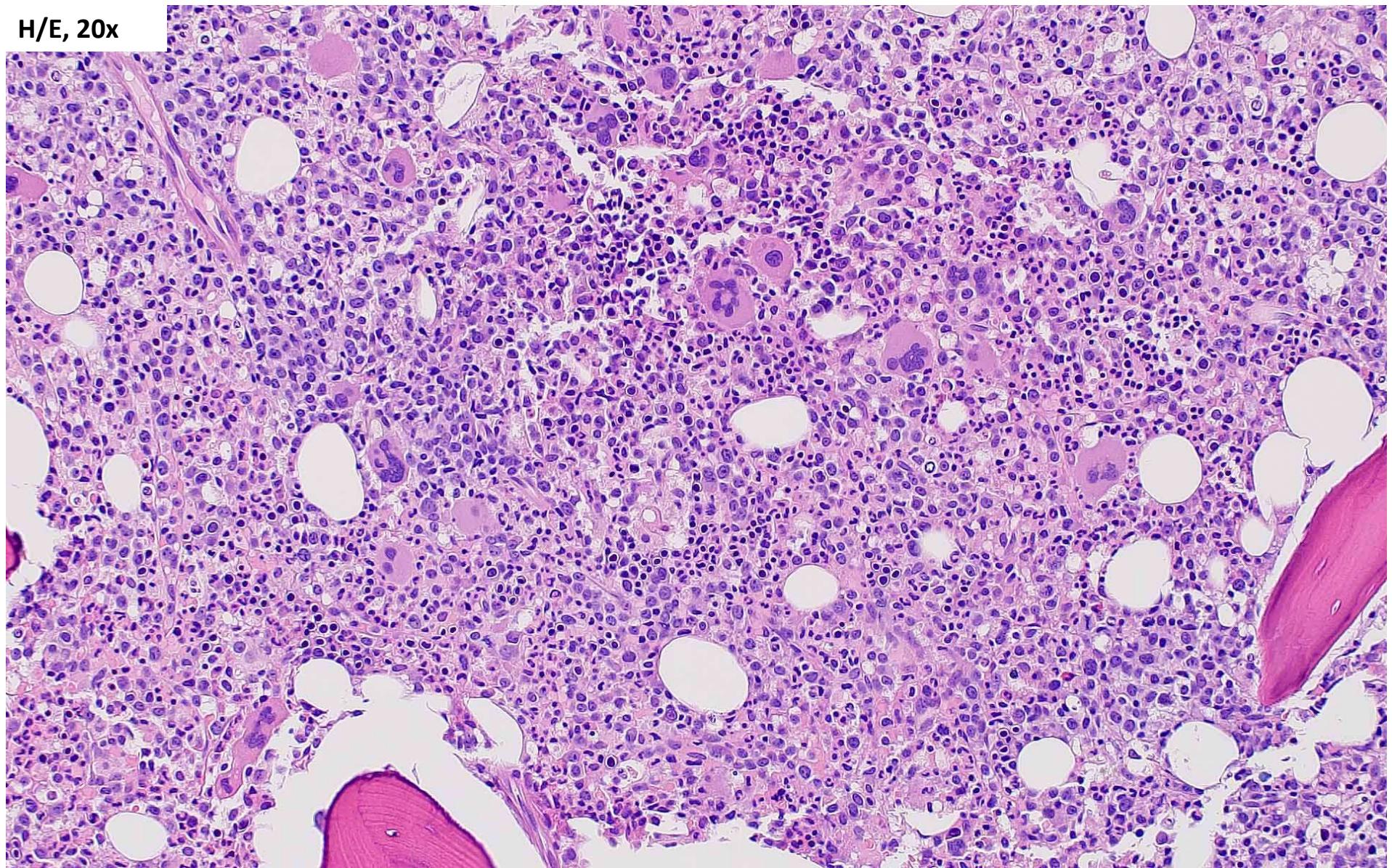
**CD3, 2x**



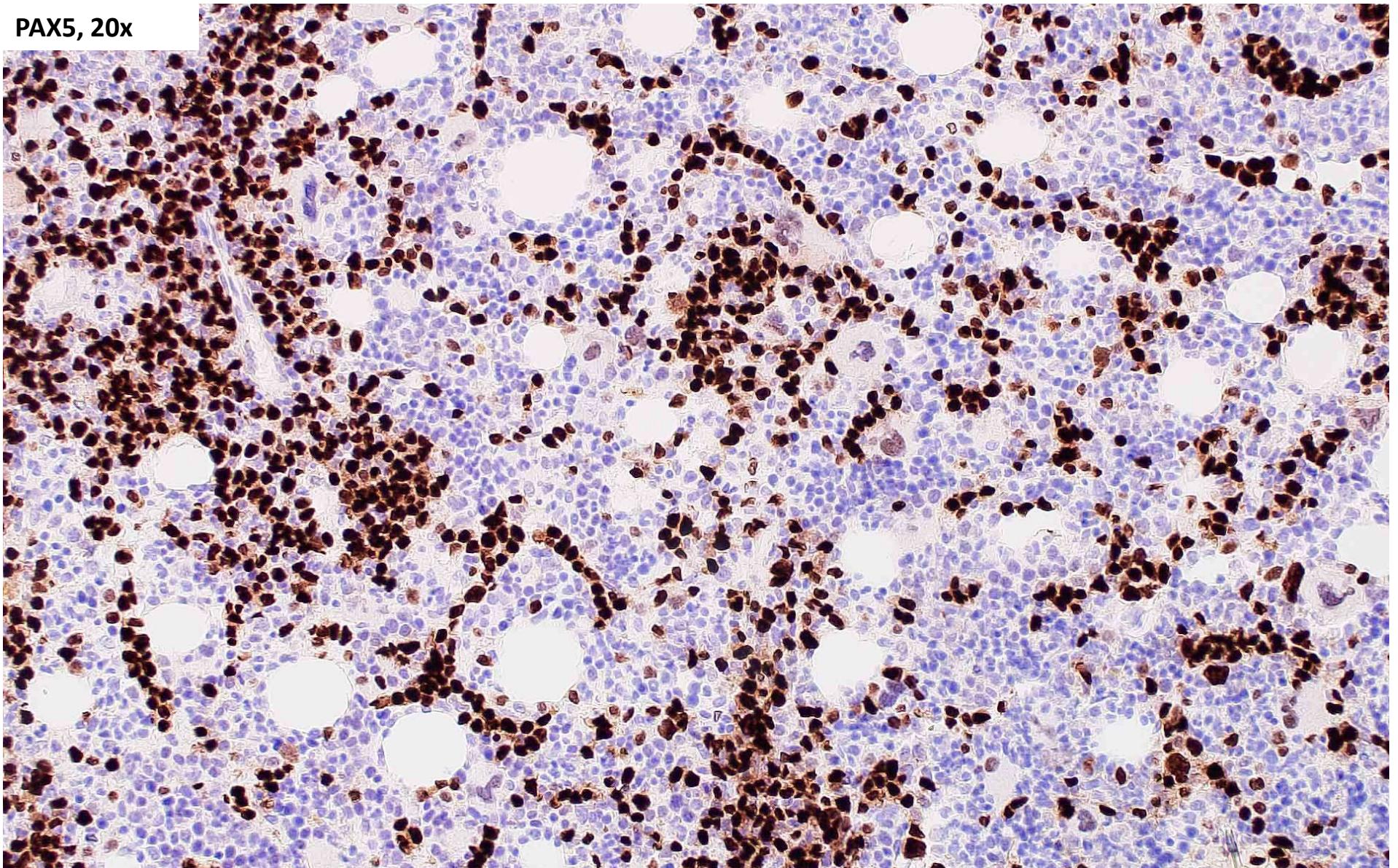
**CD5, 2x**



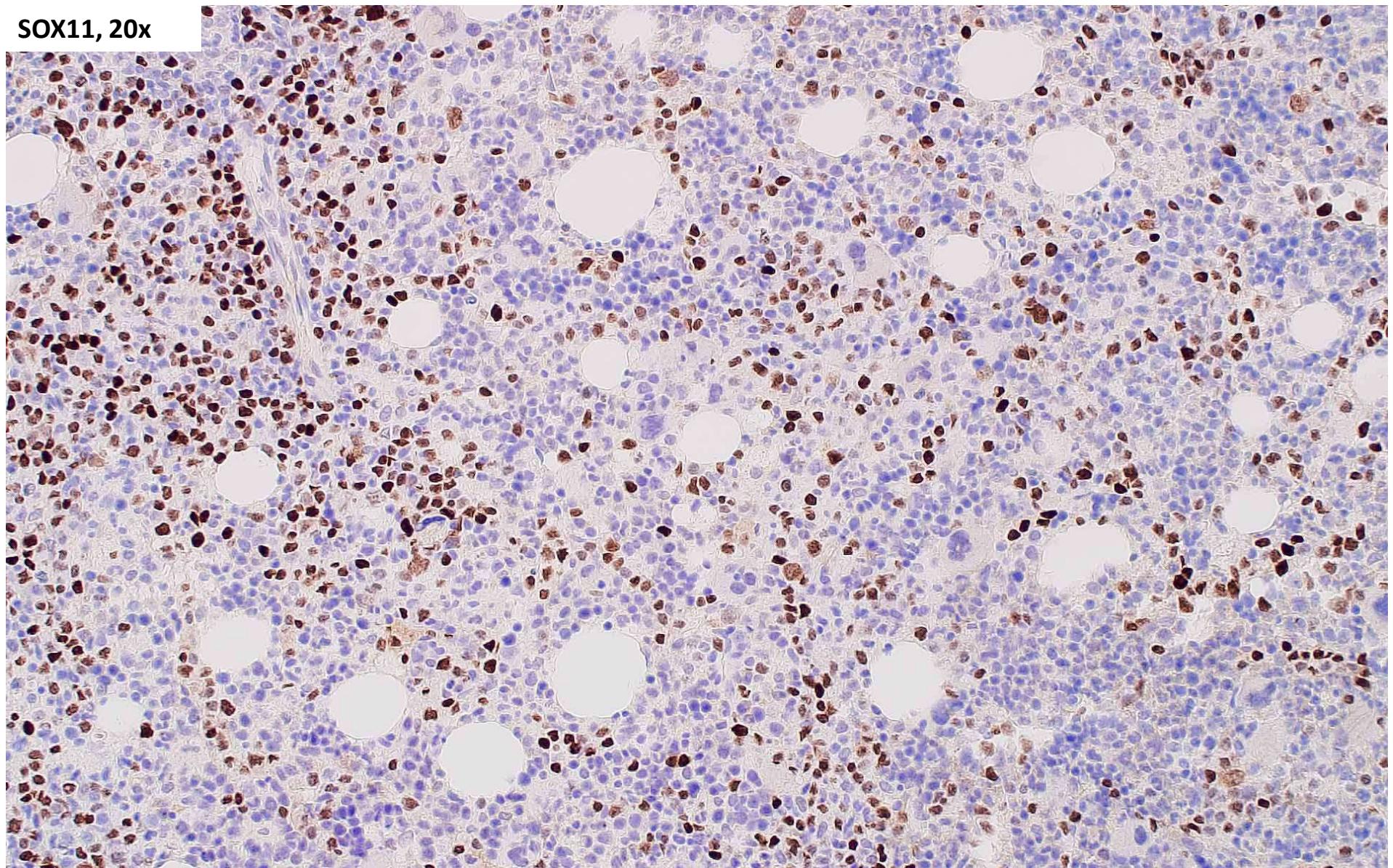
H/E, 20x



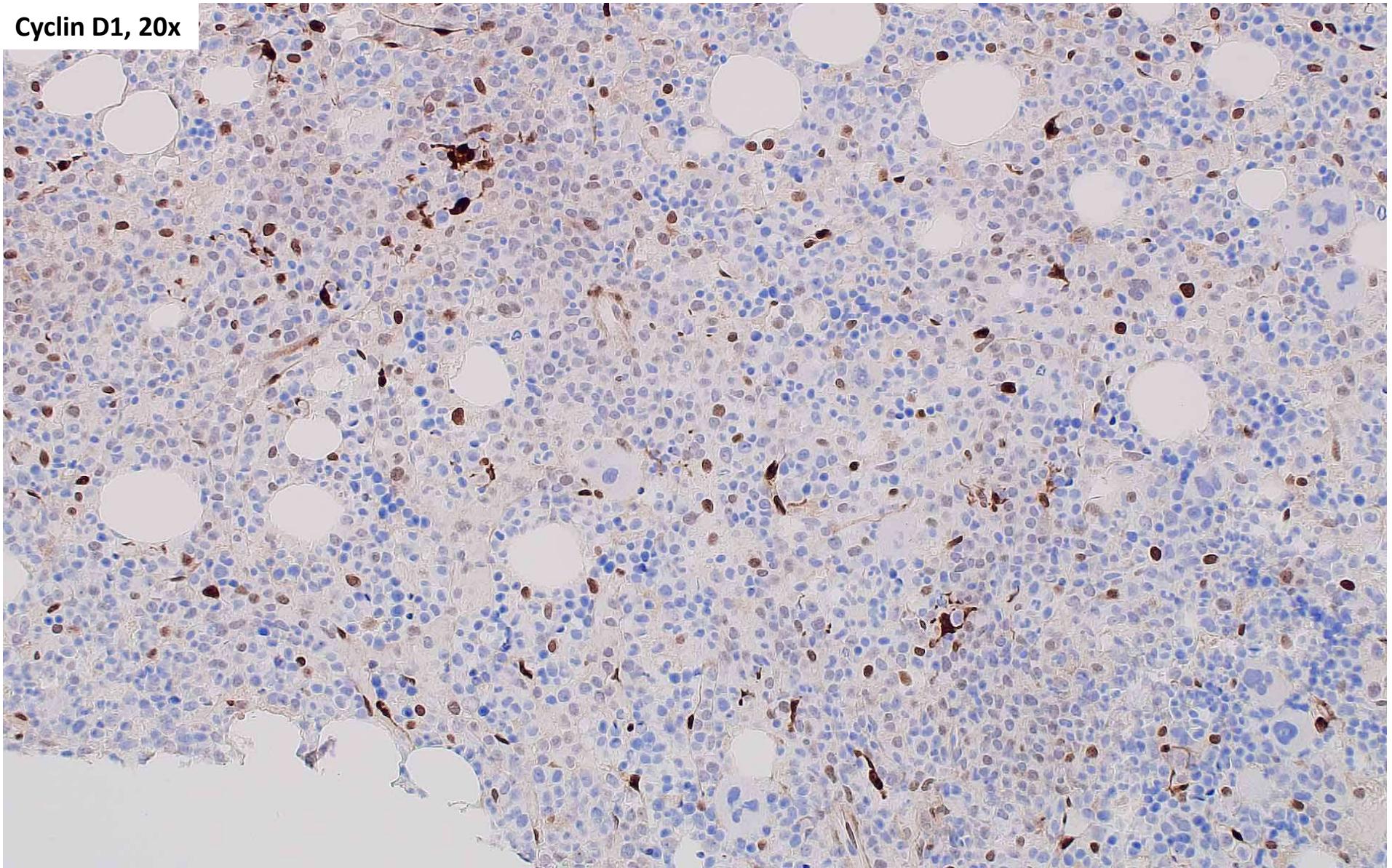
PAX5, 20x



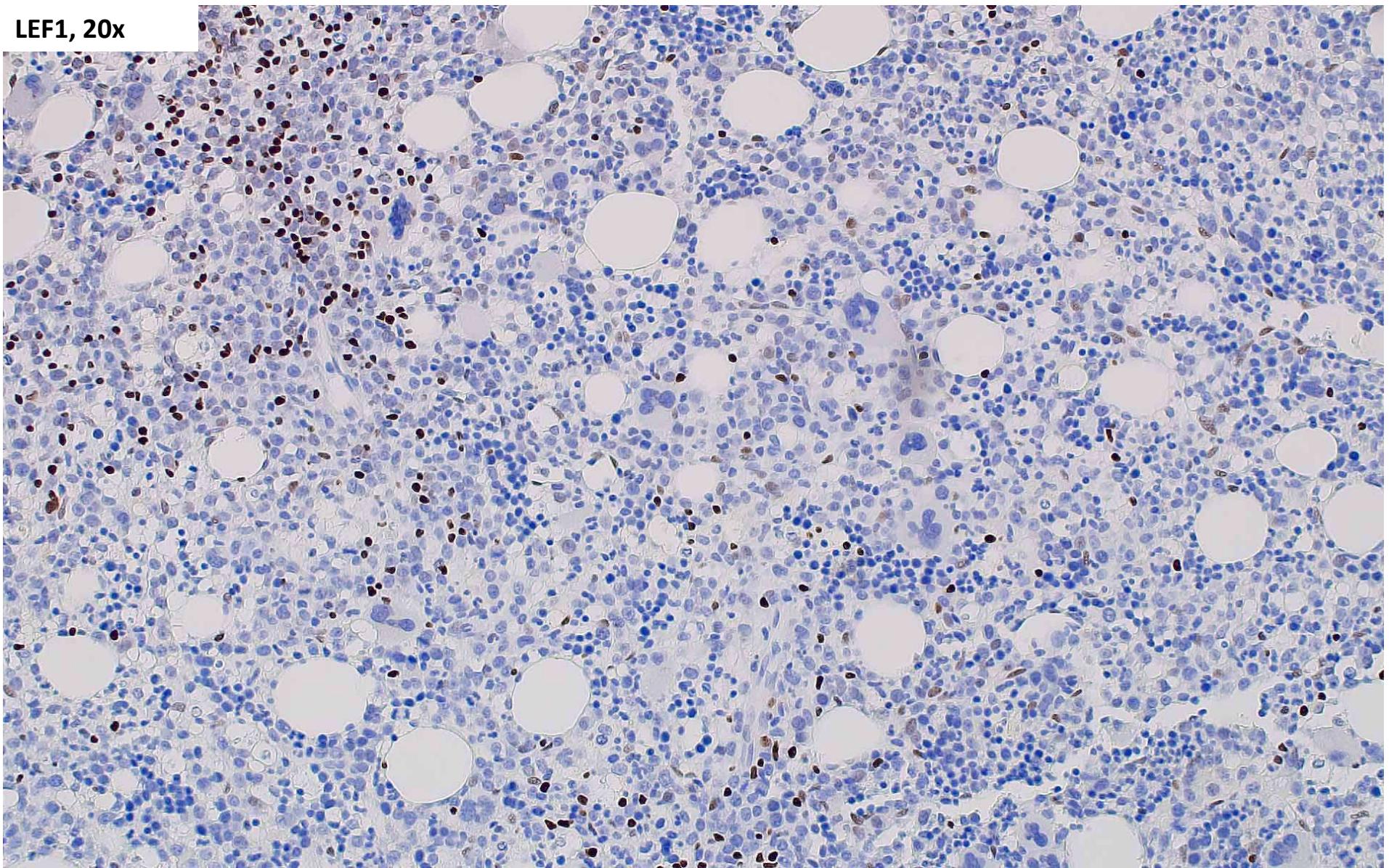
**SOX11, 20x**



Cyclin D1, 20x

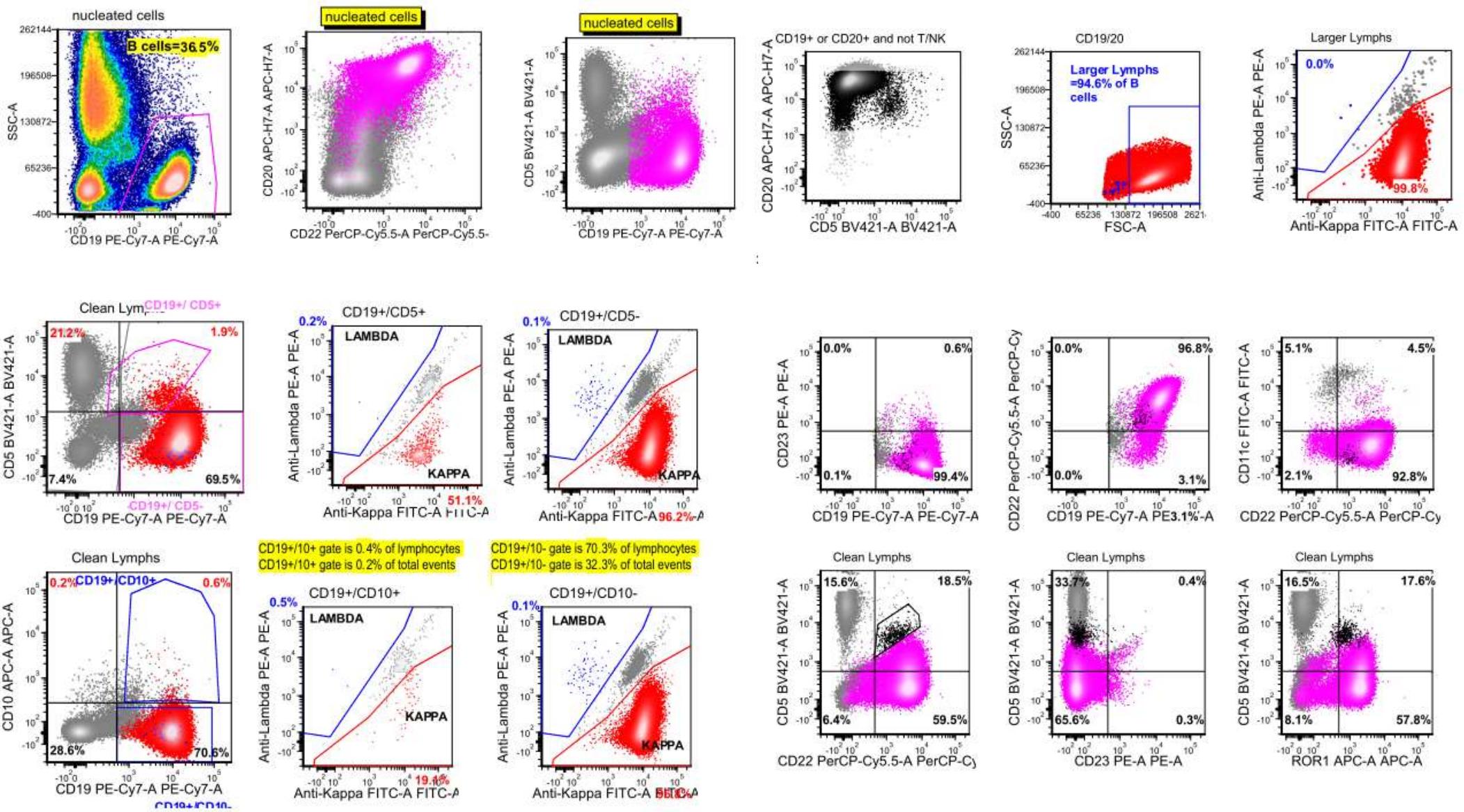


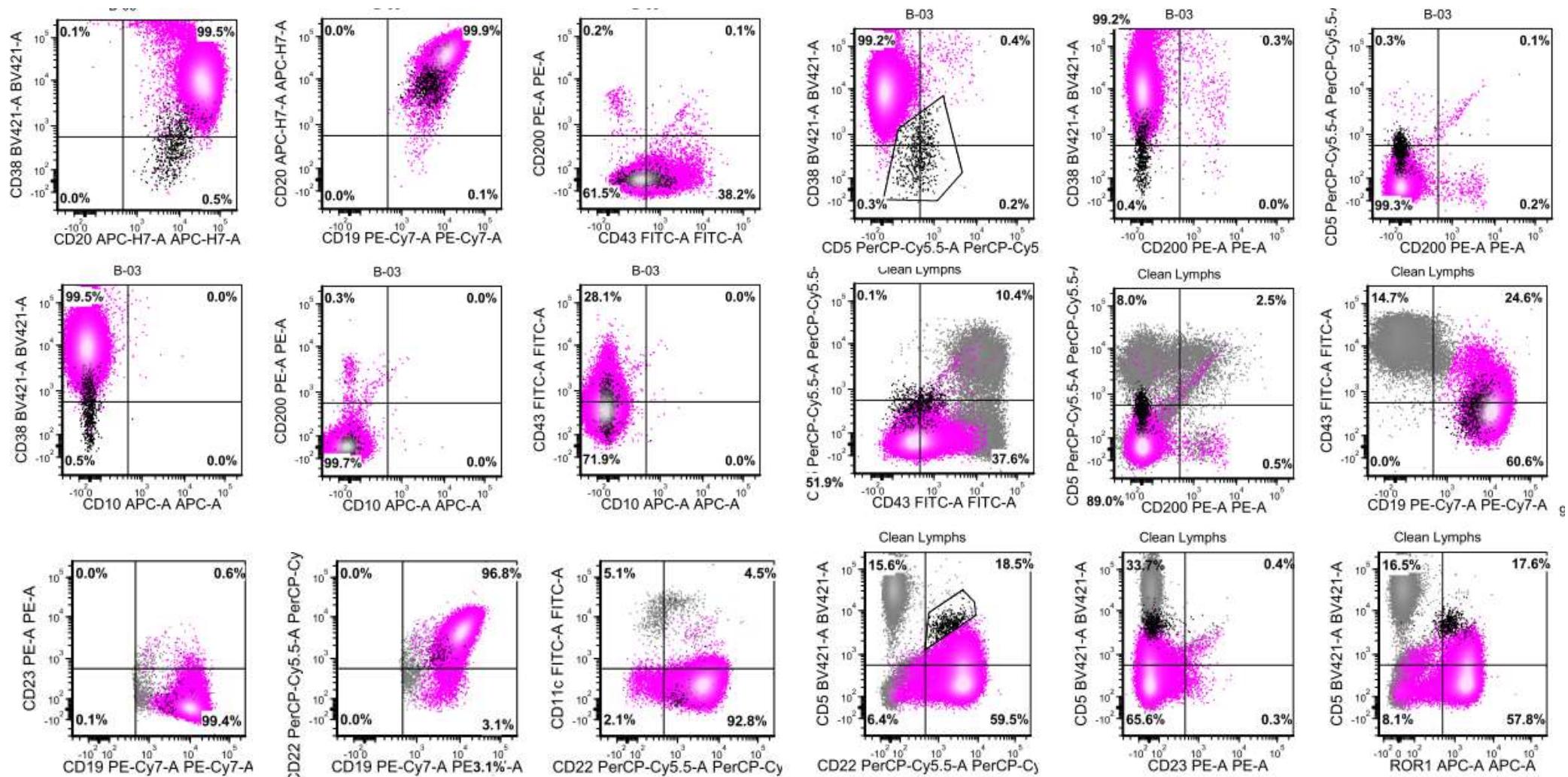
**LEF1, 20x**



## Ancillary studies (BM sample)

- Flow cytometry
- Chromosome analysis
- FISH for *IGH::CCND1* (dual color dual fusion probe)
- *IGH* somatic hypermutation
- NGS panel
- OGM





# Flow cytometric immunophenotype

- High forward scatter value
- Positive
  - CD19, CD20, CD22, CD38, CD43, ROR1, kappa
- Negative
  - CD5, CD10, CD11c, CD23, CD200, lambda
- A small subset of B-cells with CD5 expression (0.5% of total events)

# EndLymphoma panel

<i>ARID1A</i>	<i>CCND1</i>	<i>DIS3</i>	<i>GPR183</i>	<i>JAK1</i>	<i>MYD88</i>	<i>PLCG2</i>	<i>S1PR1</i>	<i>TBL1XR1</i>
<i>ASXL1</i>	<i>CCND3</i>	<i>DNMT3A</i>	<i>H1-2</i>	<i>JAK2</i>	<i>NF1</i>	<i>PLEKHG5</i>	<i>S1PR2</i>	<i>TCF3</i>
<i>ATM</i>	<i>CCR4</i>	<i>DUSP2</i>	<i>H1-4</i>	<i>JAK3</i>	<i>NFKB2</i>	<i>POLE</i>	<i>SAMHD1</i>	<i>TENT5C</i>
<i>B2M</i>	<i>CCR7</i>	<i>EGR1</i>	<i>H3C2</i>	<i>KIT</i>	<i>NFKBIA</i>	<i>POT1</i>	<i>SETD2</i>	<i>TET2</i>
<i>BAZ2A</i>	<i>CD274</i>	<i>EGR2</i>	<i>HRAS</i>	<b><i>KLF2</i></b>	<i>NFKBIE</i>	<i>PRDM1</i>	<i>SF3B1</i>	<i>TMEM30A</i>
<i>BCL10</i>	<i>CD28</i>	<i>ELF4</i>	<i>HUWE1</i>	<i>KLHL6</i>	<b><i>NOTCH1</i></b>	<i>PTEN</i>	<i>SGK1</i>	<i>TNFAIP3</i>
<i>BCL2</i>	<i>CD58</i>	<i>EP300</i>	<i>HVCN1</i>	<i>KMT2D</i>	<b><i>NOTCH2</i></b>	<i>PTPN1</i>	<i>SMARCA4</i>	<i>TNFRSF14</i>
<i>BCL6</i>	<i>CD79A</i>	<i>EWSR1</i>	<i>ID3</i>	<i>KRAS</i>	<i>NPM1</i>	<i>PTPN11</i>	<i>SMO</i>	<i>TP53</i>
<i>BCL7A</i>	<i>CD79B</i>	<i>EZH2</i>	<i>IDH1</i>	<i>LTB</i>	<i>NRAS</i>	<i>PTPRD</i>	<i>SOCS1</i>	<i>TRAF2</i>
<i>BCOR</i>	<b><i>CDKN2A</i></b>	<i>FAM50A</i>	<i>IDH2</i>	<i>LYN</i>	<b><i>NSD2</i></b>	<i>RASSF1</i>	<i>SOX11</i>	<i>TRAF3</i>
<i>BIRC3</i>	<i>CDKN2B</i>	<i>FAS</i>	<i>IFNGR1</i>	<i>MAP2K1</i>	<i>NXF1</i>	<i>RB1</i>	<i>SP140</i>	<i>TRAF6</i>
<i>BLNK</i>	<i>CHD2</i>	<i>FAT1</i>	<b><i>IGLL5</i></b>	<i>MAP3K14</i>	<i>P2RY8</i>	<i>RBMX</i>	<i>SPEN</i>	<i>U2AF1</i>
<i>BRAF</i>	<i>CHEK2</i>	<i>FBXW7</i>	<i>IKZF3</i>	<b><i>MAPK1</i></b>	<i>PAX5</i>	<i>RFTN1</i>	<i>SRSF2</i>	<i>UBR5</i>
<i>BRCC3</i>	<i>CIITA</i>	<i>FGFR3</i>	<i>IL2RG</i>	<i>MAX</i>	<i>PCBP1</i>	<i>RHOA</i>	<i>STAT3</i>	<i>VAV1</i>
<i>BTG1</i>	<i>CNOT3</i>	<i>FOXO1</i>	<i>IRAK1</i>	<i>MED12</i>	<i>PIK3CA</i>	<i>RIPK1</i>	<i>STAT5B</i>	<i>XPO1</i>
<i>BTG2</i>	<i>CREBBP</i>	<i>FYN</i>	<i>IRF4</i>	<i>MEF2B</i>	<i>PIK3R1</i>	<i>RPS15</i>	<i>STAT6</i>	<i>ZFAT</i>
<i>BTK</i>	<i>CXCR4</i>	<i>GNA13</i>	<i>IRF8</i>	<i>MFHAS1</i>	<i>PIM1</i>	<i>RRAGC</i>	<b><i>STK11</i></b>	<i>ZMYM3</i>
<i>CARD11</i>	<b><i>DDX3X</i></b>	<i>GNAS</i>	<i>ITPKB</i>	<i>MYC</i>	<i>PLCG1</i>	<i>RRAS</i>	<i>SYK</i>	<i>ZRSR2</i>

# EndLymphoma panel

## Variants of probable somatic origin (somatic mutations)

Gene	DNA change	Protein change	Location	VAF	Type
<i>CDKN2A</i>	c.238C>T	p.R80*	Exon 2	27%	SNV - Nonsense
<i>DDX3X</i>	c.908G>C	p.G303A	Exon 10	38%	SNV - Missense
<i>IGLL5</i>	c.166G>C	p.V56L	Exon 1	19%	SNV - Missense
<i>KLF2</i>	c.220_232del	p.Y74fs*63	Exon 2	17%	Deletion - Frameshift
<i>NOTCH2</i>	c.7081C>T	p.Q2361*	Exon 34	21%	SNV - Nonsense
<i>STK11</i>	c.511G>A	p.G171S	Exon 4	19%	SNV - Missense

## Variants of uncertain origin (germline versus somatic origin cannot be determined unequivocally)

Gene	DNA change	Protein change	Location	VAF	Type
<i>MAPK1</i>	c.17_22del	p.A6_A7del	Exon 1	45%	Deletion - Deletion

# Ancillary studies

- Flow cytometry
- Chromosome analysis
  - 46,XY[20]
- FISH for *IGH*::*CCND1* (dual color dual fusion probe)
  - Negative
- *IGH* somatic hypermutation
  - Unmutated (1.69% deviation from germline sequence, IGHV3-33)
- NGS panel
  - *CDKN2A*, *DDX3X*, *IGLL5*, *KLF2*, *NOTCH2*, *STK11* mutations
- OGM
  - t(2;12)(p11.2;p13.32) *IGK*::*CCND2*
  - t(8;14)(q24.21;q32.33) *IGH*::*MYC*

# Diagnosis

- Mantle cell lymphoma with *CCND2* rearrangement



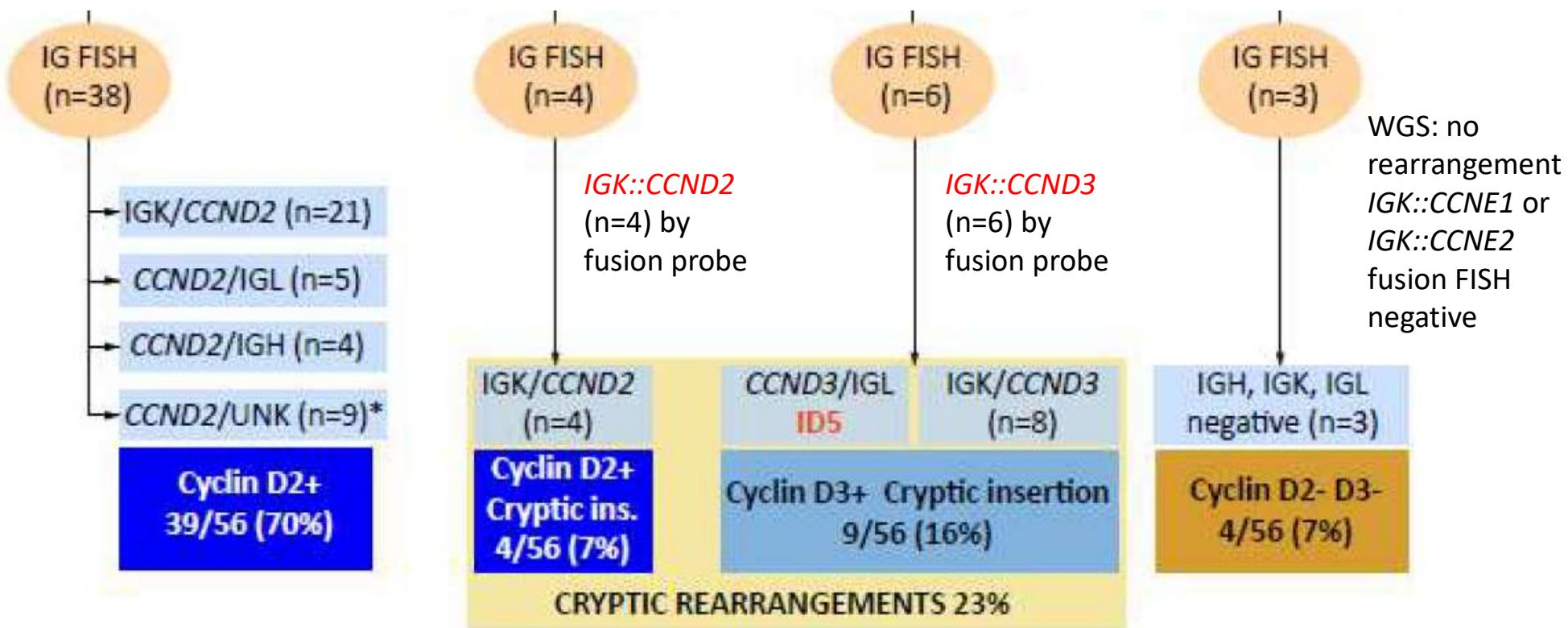
**LYMPHOID NEOPLASIA**

# **CCND2 and CCND3 hijack immunoglobulin light-chain enhancers in cyclin D1<sup>-</sup> mantle cell lymphoma**

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# *CCND2* and *CCND3* study in cyclin D1(-) MCL

- N=56 (28 patients included from the previous study)
- Inclusion criteria
  - Morphology typical for MCL
  - Immunophenotype (CD5+, CD23-)
  - Absence of cyclin D1 expression by IHC or t(11;14)(q13;q32)
  - Presence of SOX11 expression
- FISH for *CCND2*, *CCND3*, *CCNE1*, *CCNE2*, *IGH*, *IGK* and *IGL*
- NGS (WGS/WES) in 5 cases
- Gene expression and copy-number analysis



# Thank you

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